ELSEVIER

International Journal of Pharmaceutics 150 (1997) 1–200

international journal of pharmaceutics

Author Index

Volumes 1–149

A

A/Karim, E.F.I., M.H. Ahmed, R.B. Salama, Studies on the thermal and photochemical decomposition of mebendazole **142**, 251

A/Karim, E.I., K.E. Ibrahim, M.E. Adam, Studies on the photochemical decomposition of metronidazole **76**, 261

Aaiche, J.M., J.M. Cardot, S. Aiache, A comparative study of the release kinetics of trinitrine from transdermic therapeutic systems **55**, 147

Aaltonen, R., see Kahela, P. **14**, 103

Abd El Fattah, S., S.A.H. Khalil, Variations in dissolution rates of sugar-coated chlorpromazine tablets **18**, 225

Abdel-Hady, S.S., see Hosny, E.A. **142**, 163

Abdel-Hamid, M.E., see Tabba, H.D. **54**, 57

Abdel-Hamid, M.E., see Suleiman, M.S. **54**, 65

Abdel-Hay, M.H., Stability-indicating derivative spectrophotometric determination of frusemide **99**, 333

Abdel-Hay, M.H., see El-Yazbi, F.A. **27**, 139

Abdel-Hay, M.H., see Alkaysi, H.N. **87**, 73

Abdel-Rahman, A.A., see Ahmed, S.M. **96**, 5

Abdelnasser, M.A., see Desai, D.S. **103**, 69

Abdulhameed, M.E., see Suleiman, M.S. **50**, 71

Abe, K., T. Irie, H. Adachi, K. Uekama, Combined use of 2-hydroxypropyl-β-cyclodextrin and a lipophilic absorption enhancer in nasal delivery of the LHRH agonist, buserelin acetate, in rats **123**, 103

Abed, L., see Touitou, E. **27**, 89

Aberturas, M.R., see Guzman, M. **80**, 119

Aberturas, M.R., see Chacon, M. **141**, 81

Abi Khalil, F., M. Van Damme, M. Hanocq, P. Mauriac, J. Saint-Germain, D. Bouzard, J.F. Dauphin, Simultaneous determination of the ionisation and dimerization constants and the partition and ion pair extraction coefficients for 1,8-naphtyridine derivatives **140**, 1

Aboofazeli, R., C.B. Lawrence, S.R. Wicks, M.J. Lawrence, Investigations into the formation and characterization of phospholipid microemulsions. III. Pseudo-ternary phase diagrams of systems containing water–lecithin–isopropyl myristate and either an alkanoic acid, amine, alkanediol, polyethylene glycol alk **111**, 63

Aboofazeli, R., M.J. Lawrence, Investigations into the formation and characterization of phospholipid microemulsions. I. Pseudo-ternary phase diagrams of systems containing water–lecithin–alcohol–isopropyl myristate **93**, 161

Investigations into the formation and characterization of phospholipid microemulsions. II. Pseudo-ternary phase diagrams of systems containing water–lecithin–isopropyl myristate and alcohol: Influence of purity of lecithin **106**, 51

Aboofazeli, R., N. Patel, M. Thomas, M.J. Lawrence, Investigations into the formation and characterization of phospholipid microemulsions. IV. Pseudo-ternary phase diagrams of systems containing water–lecithin–alcohol and oil; the influence of oil **125**, 107

Abougela, I.K.A., see Grant, D.J.W. **17**, 77

Abra, R.M., see Schreier, H. **37**, 233

Abraham, I., see Krishnan, T.R. **80**, 277

Abrahamsson, B., M. Alpsten, U.E. Jonsson, P.J. Lundberg, A. Sandberg, M. Sundgren, A. Svenheden, J. Tolli, Gastrointestinal transit of a multiple-unit formulation (metoprolol CR/ZOK) and a non-disintegrating tablet with the emphasis on colon **140**, 229

Abrahamsson, B., see Wingstr, K. **60**, 151

Abrahamsson, B., see Sandberg, A. **68**, 167

Abramovici, B., see Lheritier, J. **123**, 273

Abu-Bakar, B.B., see Pryce-Jones, R.H. **86**, 231

Abu-Izza, K., see Lu, D.R. **129**, 243

Abu-Lafi, S., see Tabba, H.D. **54**, 57

Abu. Malooh, A., see Sheikh Salem, M. **141**, 257

Aburawi, S., S.H. Curry, R. Whelpton, Chemical denitration of nitroglycerin, and conversion of 1,2-dinitroglycerin to 1,3-dinitroglycerin **22**, 327

Acar, L., see Sarciaux, J.M. **120**, 127

Acar, N., see Onur, F. **78**, 89

Acarturk, F., O. Kislal, N. Celebi, The effect of some natural polymers on the solubility and dissolution characteristics of nifedipine **85**, 1

Acarturk, F., A. Sencan, Investigation of the effect of different adjuvants on felodipine release kinetics from sustained release monolithic films **131**, 183

Ackermann, C., see Cairncross, C. **27**, 361

Ackermann, C., see Du Plessis, J. **34**, 173

Ackermann, C., see Van der Merwe, E. **44**, 71

0378-5173/97/$17.00 © 1997 Elsevier Science B.V. All rights reserved.

PII S0378-5173(97)00262-7

Ackermann, C., see Siebert, J.G.H. **61**, 157; **65**, 29; **66**, 157
Ackermann, C., G.L. Flynn, Ether-water partitioning and permeability through nude mouse skin in vitro. I. Urea, thiourea, glycerol and glucose **36**, 61
Ackermann, C., G.L. Flynn, W.M. Smith, Ether-water partitioning and permeability through nude mouse skin in vitro. II. Hydrocortisone 21-*n*-alkyl esters, alkanols and hydrophilic compounds **36**, 67
Acquasaliente, M., see Dal Pozzo, A. **81**, 263
Acree, Jr. W.E., Thermochemical investigations of associated solutions. II. Calculation of iodine–benzene equilibrium constants from solute solubility in binary solvent mixtures **15**, 159
Solubility of biphenyl in binary solvent mixtures **18**, 47
Comments concerning 'Model for solubility estimation in mixed solvent systems' **127**, 27
Acree, Jr. W.E., W.E. Bacon, A.J. Leo, Octanol/water partition coefficients of 4-substituted benzylidene *t*-butylamine *N*-oxides **20**, 209
Acree, Jr. W.E., N.M. Pontikos, C.L. Judy, Solubility in binary solvent systems. VI. Prediction of naphthalene and biphenyl solubility based on the Wilson model **31**, 225
Acree, Jr. W.E., J.H. Rytting, J.T. Carstensen, Comment on solubility parameters from maxima in solubility/solvent plots **8**, 69
Acree, Jr. W.E., J.H. Rytting, Solubility in binary solvent systems. IV. Prediction of naphthalene solubility using the UNIFAC group contribution model **13**, 197
Acree, W.E., S.A. Tucker, Thermochemical investigations of associated solutions: 16. Comparison of the extended NIBS model and mobile order theory for solubility in systems containing solute–solvent complexation **101**, 199
Acree, Jr. W.E., see Judy, C.L. **27**, 39
Adachi, H., see Abe, K. **123**, 103
Adachi, I., see Liu, H.X. **118**, 55
Adachi, T., N. Koizumi, M. Nishio, Y. Ito, Degradation kinetics of KE-298 and chelating ability of its degradation products **124**, 129
Adair, C.G., see Jones, D.S. **131**, 83
Adam, M.E., see A Karim, E.I. **76**, 261
Adams, K.A., see Shenouda, L.S. **61**, 127
Adams, W.P., H.B. Kostenbauder, Phenoxybenzamine stability in aqueous ethanolic solutions. I. Application of potentiometric pH stat analysis to determine kinetics **25**, 293
Phenoxybenzamine stability in aqueous ethanolic solutions. II. Solvent effects upon kinetics **25**, 313
Addicks, W.J., G. Flynn, N. Weiner, R. Curl, A mathematical model to describe drug release from thin topical applications **56**, 243
Addicks, W.J., see Chiang, C.M. **49**, 109
Adelusi, S.A., Evaluation of the extraction of chloroquine from biological samples by different organic solvents **35**, 173
Ademola, J.I., see Montenegro, L. **140**, 51
Adesanya, K.I., see Eboka, C.J. **51**, 161, 263
Adeyemi, M.I., N. Pilpel, The tensile strengths of mixtures of oxytetracycline and lactose powders **15**, 293
Adeyemi, M.O., N. Pilpel, The effects of interacting variables on the tensile strength, disintegration and dissolution of oxytetracycline-lactose tablets **20**, 171
Adeyeye, C.M., J. Rowley, D. Madu, M. Javadi, S.S. Sabnis, Evaluation of crystallinity and drug release stability of directly compressed theophylline hydrophilic matrix tablets stored under varied moisture conditions **116**, 65
Adeyeye C.M., see Badawy, S.I.F. **128**, 45; **145**, 137
Adjei, A., R. Doyle, M. Pratt, R. Finley, E. Johnson, Bioavailability of leuprolide following intratracheal administration to beagle dogs **61**, 135
Adjei, A., J. Hui, R. Finley, T. Lin, J. Lancaster, F. Fort, Pulmonary bioavailability of leuprolide acetate following multiple dosing to beagle dogs: Some pharmacokinetic and preclinical issues **107**, 57
Adjei, A., S. Rao, J. Garren, G. Menon, M. Vadnere, Effect of ion-pairing on 1-octanol-water partitioning of peptide drugs. I: The nonapeptide leuprolide acetate **90**, 141
Adjei, A., see Annapragada, A. **127**, 219; **136**, 1
Adjei, A., see Gupta, P. **140**, 119; **147**, 207
Adjei, A., see Powers, M.E. **108**, 49
Adjei, A., see Qiu, Y. **115**, 95
Adjei, A., see Swanson, P.D. **142**, 33
Adler, J., see Melia, C.D. **100**, 263
Adler, J., see Aspden, T.J. **122**, 69
Adolfsson, A., C. Nystrom, Tablet strength, porosity, elasticity and solid state structure of tablets compressed at high loads **132**, 95
Adolfsson, A., see Olsson, H. **143**, 233
Advenier, C., see Efthymiopoulos, C. **95**, 127
Agache, P., see Makki, S. **133**, 245
Agarwal, S.P., see Attwood, D. **22**, 25
Agarwal, S.P., see Khanna, R. **138**, 67
Agbada, C.O., P. York, Dehydration of theophylline monohydrate powder—effects of particle size and sample weight **106**, 33
Agerbaek, H., see Pridal, L. **136**, 53
Aggarwal, N., see Mukherji, G. **71**, 187
Aggarwal, N., see Mukherji, G. **86**, 153
Agrawal, S.S., see Jain, G.K. **130**, 169
Agren, T., see Bjerga Bjaen, A.K. **92**, 183
Agren, T., see Henriksen, I. **119**, 231
Agren, T., see Tonneson, H.H. **90**, 221
Agyilirah, G.A., M. Green, R. DuCret, G.S. Banker, Evaluation of the gastric retention properties of a cross-linked polymer coated tablet versus those of a non-disintegrating tablet **75**, 241
Agyilirah, G.A., see Nixon, J.R. **6**, 277
Ah-Fat, N.M.W., D.Q.M. Craig, K.M.G. Taylor, An investigation into the effects of surfactants on phospholipid monolayers using a Langmuir-Blodgett film balance **107**, 239
Ahlbom, J., see Gottfries, J. **106**, 141
Ahlgren, R., see Wennergren, B. **53**, 35
Ahlgren, R., see Nicklasson, M. **69**, 255
Ahlneck, C., G. Alderborn, Moisture adsorption and tabletting. I. Effect on volume reduction properties and tablet strength for some crystalline materials **54**, 131

Ahlneck, C., G. Zografi, The molecular basis of moisture effects on the physical and chemical stability of drugs in the solid state **62**, 87
Ahlneck, C., see Alderborn, G. **73**, 249
Ahlneck, C., see Elamin, A.A. **108**, 213; **111**, 159; **119**, 25
Ahlneck, C., see Saleki-Gerhardt, A. **101**, 237
Ahlneck, C., see Sebhatu, T. **104**, 135; **146**, 101
Ahmed, B.M., see Salama, R.B. **17**, 325
Ahmed, B.M., see Ibrahim, K.E. **98**, 231
Ahmed, H., G. Buckton, D.A. Rawlins, The use of isothermal microcalorimetry in the study of small degrees of amorphous content of a hydrophobic powder **130**, 195
Ahmed, I., B. Chaudhuri, Evaluation of buffer systems in ophthalmic product development **44**, 97
Ahmed, I., T.F. Patton, Effect of pH and buffer on the precorneal disposition and ocular penetration of pilocarpine in rabbits **19**, 215
Selective intraocular delivery of liposome encapsulated inulin via the non-corneal absorption route **34**, 163
Ahmed, I., M. Sudoh, Disposition of timolol and inulin in the rabbit eye following corneal versus non-corenal absorption **38**, 9
Ahmed, I., see Francoeur, M. **16**, 203
Ahmed, I., see Jafari, M.R. **48**, 207
Ahmed, I., see Shanker, R.M. **119**, 149
Ahmed, M., J. Hadgraft, I.W. Kellaway, The effect of bile salts on the interfacial transport of phenothiazines **12**, 219
Phenothiazine transport across liquid–lipid, phospholipid and soft polymer membranes **13**, 227
Ahmed, M.H., see A Karim, E.F.I. **142**, 251
Ahmed, M.O., see Ahmed, S.M. **96**, 5
Ahmed, S.M., A.A. Abdel-Rahman, S.I. Saleh, M.O. Ahmed, Comparative dissolution characteristics of bropirimine-β-cyclodextrin inclusion complex and its solid dispersion with PEG 6000 **96**, 5
Ahmed, S.M., B. Casu, A. Cedro, M. Guerrini, E. Lanzarotti, D. Moltrasio, A. Naggi, G. Torri, Disruption of micellar aggregates of ganglioside GM-1 by complexation with α-cyclodextrin **109**, 99
Ahmed, S.M., A. Naggi, M. Guerrini, B. Focher, Inclusion complexes of bropirimine with β-cyclodextrin in solution and in solid state **77**, 247
Ahn, B.N., see Lee, H.J. **121**, 1; **131**, 91
Ahomaki, E., see Kiesvaara, J. **97**, 101
Ahuja, A., see Khanna, R. **138**, 67
Aiache, J.M., P. Edman, E. Beyssac, S. Aiache, R. Renoux, J.P. Kantelip, Kinetics of indomethacin release from suppositories. In vitro–in vivo correlation **39**, 235
Aiache, J.M., see Lootvoet G. **85**, 113
Aiache, J.M., S. Pradat, S. Aiache et al., Influence of monium on the gastrointestinal absorption of aspirin **16**, 241
Aiache, S., see Aiache, J.M. **16**, 241
Aiache, S., see Aiache, J.M. **39**, 235; **55**, 147
Aiba, T., see Hatanaka, T. **125**, 195
Aiba, T., see Panomsuk, S.P. **126**, 147
Aignasse M.F., P. Prognon, M. Stachowicz, R. Gheyouche, D. Pradeau, A new simple and rapid HPLC method for determination of DEHP in PVC packaging and releasing studies **113**, 241
Aioi, A., K. Kuriyama, T. Shimizu, M. Yoshioka, S. Uenoyama, Effects of vitamin E and squalene on skin irritation of a transdermal absorption enhancer, lauroylsarcosine **93**, 1
Aioi, A., T. Shimizu, K. Kuriyama, Effect of squalene on superoxide anion generation induced by a skin irritant, lauroylsarcosine **113**, 159
Aito, H., see Marvola, M. **53**, 145
Aizawa, K., see Watari, N. **21**, 85
Akaho, E., see Kagayama, A. **18**, 247
Akashi, M., see Sakuma, S. **149**, 93
Akazawa, M., T. Itoh, K. Masaki, B.T. Nghiem, N. Tsuzuki, R. Konishi, T. Higuchi, An automated method for continuously monitoring diffusion cells in skin penetration studies **50**, 53
Akbarieh, M., R. Tawashi, Morphic features of solid paratiles after micronization in the fluid energy mill **35**, 81
Akbarieh, M., see Thibert, R. **47**, 171
Akbarieh, M., see Kassem, M. **47**, 179
Akbarieh, M., see El-Ridy, M. **56**, 23
Akbuga J., Preparation and evaluation of controlled release furosemide microspheres by spherical crystallization **53**, 99
Furosemide-loaded ethyl cellulose microspheres prepared by spherical crystallization technique: Morphology and release characteristics **76**, 193
The effect of the physicochemical properties of a drug on its release from chitosonium malate matrix tablets **100**, 257
Use of chitosonium malate as a matrix in sustained-release tablets **89**, 19
Akbuga J., G. Durmaz, Preparation and evaluation of crosslinked chitosan microspheres containing furosemide **111**, 217
Akbuga, J., see Aydin, Z. **131**, 101; **137**, 133
Akbuga, J., see Sezer, A.D. **121**, 113
Akerele, J.O., see Okhamafe, A.O. **50**, 83; **68**, 11
Akhtar, J., see Hudson, A.J. **133**, 257; **136**, 23
Akhter S.A., S.L. Bennett, I.L. Waller, B.W. Barry, An automated diffusion apparatus for studying skin penetration **21**, 17
Akimoto J., K. Takayama, K. Isowa, T. Nagai, Evaluation of skin irritation of percutaneous absorption promoters by means of fractal dimension of rat skin structure **128**, 251
Erratum: Evaluation of skin irritation of percutaneous absorption promoters by means of fractal dimension of rat skin structure (Int. J. Pharm. 128 (1996) 251–260) **140**, 257
Akimoto K., S. Fujiwara, N. Awata, I. Sugimoto, The stability of 1-cyclopropylmethyl-5-(*o*-fluorophenyl)-7-chloro-1,3-dihydro-2H-1,4-benzodiazepin-2-one (KB-509) in aqueous solution **3**, 109
Akimoto M., A. Furuya, T. Maki, K. Yamada, T. Suwa, H. Ogata, Evaluation of sustained-release granules of chlorphenesin carbamate in dogs and humans **100**, 133
Akimoto M., A. Furuya, M. Nakamura, T. Maki, K. Yamada, T. Suwa, H. Ogata, Release and absorption characteristics of chlorphenesin carbamate sustained-release

formulations: In vitro–in vivo and in vivo dog–human correlations **117**, 31
Akimoto, M., see Yamada, K. **119**, 1
Akin, A., see Sener, E. **110**, 109
Akiyama Y., H.L. Luessen, A.G. de Boer, J.C. Verhoef, H.E. Junginger, Novel peroral dosage forms with protease inhibitory activis. I. Design of capsules with fast gel-forming and fast drug-releasing properties **136**, 155
Novel peroral dosage forms with protease inhibitol-gamma activis. II. Design of fast dissolving poly(acrylate) and controlled drug-releasing capsule formulations with trypsin inhibiting properties **138**, 13
Akiyama, K., see Yamashita, C. **70**, 225
Akojie F.O.B., J.O. Folayan, Antisickling potential of bat interferon **42**, 261
Al-Adham I.S.I., P. Gilbert, Effect of polyvinylpyrrolidone molecular weight upon the antimicrobial activity of povidone–iodine antiseptics **34**, 45
Al-Angary A.A., M.A. Al-Meshal, M.A. Bayomi, S.H. Khidr, Evaluation of liposomal formulations containing the antimalarial agent, arteether **128**, 163
Al-Angary A.A., M.A. Bayomi, S.H. Khidr, M.A. Al-Meshal, M. Al-Dardiri, Characterization, stability and in vivo targeting of liposomal formulations containing cyclosporin **114**, 221
Al-Angary A.A., S.H. Khidr, G.M. Mahrous, M.W. Gouda, In-vitro dissolution and in vivo absorption of marketed sustained release theophylline preparations **65**, R5
Al-Angary, A.A., see Al-Meshal, M.A. **64**, 155
Al-Angary, A.A., see Gouda, M.W. **37**, 227
Al-Angary, A.A., see El-Sayed, Y.M. **64**, 109; **109**, 107
Al-Angary, A.A., see Hosny, E.A. **112**, 87; **113**, 209
Al-Arab, M.M., see Tabba, H.D. **54**, 57
Al-Balla, S.R., see El-Sayed, Y.M. **109**, 107
Al-Dardiri, M., see Al-Angary, A.A. **114**, 221
Al-Dujaili H., A.T. Florence, E.G. Salole, In vitro assessment of the adhesiveness of film-coated tablets **34**, 67
The adhesiveness of proprietary tablets and capsules to porcine oesophageal tissue **34**, 75
Al-Gamal, S.S., see Al-Gohary, O.M. **55**, 47
Al-Gohary O.M., An in vitro study of the interaction between mebeverine hydrochloride and magnesium trisilicate powder **67**, 89
Al-Gohary O.M., S.S. Al-Gamal, A. Hammad, A.M. Molokhia, Effect of storage on tabletted microencapsulated aspirin granules **55**, 47
Al-Hassan, M.I., see Najjar, T.A. **98**, 51
Al-Hassan M.I., S.A. Bawazir, K.I. Al-Khamis, K.M. Matar, A. Tekle, The interaction potential of vigabatrin with phenytoin and carbamazepine **93**, 7
Al-Hassan M.I., S.A. Bawazir, K.M. Matar, A. Tekle, Effect of famotidine on theophylline pharmacokinetics in the rat **50**, 35
Al-Hmoud H., M. Efentakis, N.H. Choulis, A controlled release matrix using a mixture of hydrophilic and hydrophobic polymers **68**, R1
Al-Hmoud, H., see Buckton, G. **74**, 169
Al-Hmoud, H., see Efentakis, M. **70**, 153
Al-Humayyd M.S., Diltiazem and verapamil elevate plasma phenytoin concentrations in the rat **131**, 153
Effect of metoclopramide on lamotrigine absorption in rabbits **144**, 171
Al-Kassas R.S., C.A. Gilligan, A. Li Wan Po, Processing factors affecting particle size and in vitro drug release of sustained-release ibuprofen microspheres **94**, 59
Al-Kaysi, H.N., see Sheikh Salem, M. **141**, 257
Al-Khamis K., S.S. Davis, J. Hadgraft, S. Mills, The determination of thermodynamic activity by gas chromatography head space analysis and its use in studying release rates of drugs from topical preparations **10**, 25
Al-Khamis, K.I., see Al-Hassan, M.I. **93**, 7
Al-Khamis, K.I., see Kaka, J.S. **28**, 133
Al-Meshal M.A., Y.M. El-Sayed, A.A. Al-Angary, K.M. Lufti, M. Wafik Gouda, Interruption of the enterohepatic circulation of indomethacin by cholestyramine in rabbits **64**, 155
Al-Meshal, M.A., see Al-Angary, A.A. **114**, 221; **128**, 163
Al-Meshal, M.A., see Turkistani, A.A.A. **59**, R1
Al-Meshal, M.A., see El-Sayed, Y.M. **64**, 109; **109**, 107
Al-Meshal, M.A., see Hosny, E.A. **112**, 87
Al-Nimry S.S., S.M. Assaf, I.M. Jalal, N.M. Najib, Adsorption of ketotifen onto some pharmaceutical excipients **149**, 115
Al-Razzak L.A., V.J. Stella, Stability and solubility of 2-chloro-2′,3′-dideoxyadenosine **60**, 53
Al-Shamkhani A., R. Duncan, Synthesis, controlled release properties and antitumour activity of alginate-cis-aconityl-daunomycin conjugates **122**, 107
Al-Shamkhani, see Morgan, S.M. **122**, 121
Al-Shora A., Said S., Hammad A.L., Sustained release from inert matrices. II. Effect of polyethylene glycols on theophylline release **7**, 77
Al-Shora, H., see Said, S. **5**, 223; **6**, 11
Al-Shora H.I., M.A. Moustafa, E.M. Niazy, M. Gaber, M.W. Gouda, Interactions of procainamide, verapamil, guanethidine and hydralazine with adsorbent antacids and antidiarrhoeal mixtures **47**, 209
Al-Shora, H.I., see Moustafa, M.A. **34**, 207
Al-Shora, H.I., see Turkistani, A.A.A. **59**, R1
Al-Turk, W., see Shaheen, O. **38**, 123
Al-Turk W.A., I.A. Majeed, W.J. Murray, D.W. Newton, S. Othman, Some factors affecting the photodecomposition of nifedipine **41**, 227
Al-Turk, W.A., see Othman S. **41**, 197
Al Khouri Fallouh N., L. Roblot-Treupel, H. Fessi et al., Development of a new process for the manufacture of polyisobutylcyanoacrylate nanocapsules **28**, 125
Al Saidan, S.M.H., see Winfield, A.J. **8**, 211
Al Shora, H.I., see Wafik Gouda, H. **18**, 213
Alain M., L. Bardet, Energy of gelation of carbomer 940 **12**, 173
Alain M., J.N. McMullen, Molecular weight of Acacia **23**, 265
Alamercery, S., see Jacolot, A. **128**, 283

Alard, V., see Zeevi, A. **108**, 57
Alaya, M., see Bourget, P. **146**, 167
Albasini, A., see Costantino, L. **86**, 17
Albers, D.D., see Tu, Y.H. **61**, 119
Albers, E., see Muller, B.W. **79**, 273
Albers, W.M., see Duchateau, G.S.M.J.E. **34**, 131
Albertsson, A.C., see Singh, S.K. **142**, 199
Albery W.J., R.H. Guy, J. Hadgraft, Percutaneous absorption: Transport in the dermis **15**, 125
Albrecht, G., see Ter-Minassian-Saraga, L. **24**, 149
Albrecht, P., see Stella, V. **121**, 117
Alden M., M. Lyden, J. Tegenfeldt, Effect of counterions on the interactions in solid dispersions between polyethylene glycol, griseofulvin and alkali dodecyl sulphates **110**, 267
Alden M., J. Tegenfeldt, E. Sjokvis, Structure of solid dispersions in the system polyethylene glycol-griseofulvin with additions of sodium dodecyl sulphate **83**, 47
Alden M., J. Tegenfeldt, E. Sjokvist Saers, Structures formed by interactions in solid dispersions of the system polyethylene glycol-griseofulvin with charged and non charged surfactants added **94**, 31
Alden, M., see Sjokvist, E. **54**, 161; **69**, 53; **79**, 123
Alden, M., see Sjokvist Saers, E. **90**, 105
Alden, M., see Wulff, M. **142**, 189
Alderborn, G., C. Ahlneck, Moisture adsorption and tabletting. III. Effect on tablet strength-post compaction storage time profiles **73**, 249
Alderborn, G., P.O. Lang, A. Sagstrom, A. Kristensen, Compression characteristics of granulated materials. I. Fragmentation propensity and compactibility of some granulations of a high dosage drug **37**, 155
Alderborn, G., K. Pasanen, C. Nystrom, Studies on direct compression of tablets. XI. Characterization of particle fragmentation during compaction by permeametry measurements of tablets **23**, 79
Alderborn, G., see Ahlneck, C. **54**, 131
Alderborn, G., see Ek, R. **111**, 43
Alderborn, G., see Elamin, A.A. **108**, 213; **111**, 159
Alderborn, G., see Eriksson, M. **63**, 189, 99, 197; **109**, 59; **113**, 199; **148**, 149
Alderborn, G., see Jerwanska, E. **121**, 65; **149**, 131
Alderborn, G., see Johansson, B. **117**, 57; **132**, 207
Alderborn, G., see Newton, J.M. **93**, 249
Alderborn, G., see Sebhatu, T. **146**, 101
Alderborn, G., see Wikberg, M. **62**, 229, 63, 23; **69**, 239; **84**, 191
Alebic-Kolbah, T., see Zorc, B. **99**, 135
Alexander, J., see Baillie, A.J. **57**, 23
Alexander, J., see Carter, K.C. **53**, 129
Alexander, J., see Gogate, U.S. **40**, 235
Alexander, M., see Ghosh, T.K. **88**, 391
Alhaique, F., see Touitou, E. **78**, 85; **103**, 131
Alhaique, F., see Memoli, A. **117**, 159
Alhaique, F., see Touitou, E. **133**, 85
Ali, A.A., A. Farouk, Comparative studies on the bioavailability of ampicillin anhydrate and trihydrate **9**, 239
Ali, A.A., A.S. Gorashi, Absorption and dissolution of nitrofurantoin from different experimental formulations **19**, 297
Ali, H.M., Bioequivalence studies on two brands of bacampicillin **9**, 91
Comparative bioavailability of eight brands of ampicillin **7**, 301
The effect of Sudanese food and chloroquine on the bioavailability of ampicillin from bacampicillin tablets **9**, 185
Ali, H.M., A. Farouk, S.A.H. Khalil, M.A. El-Nakeeb, Correlation between microbiological and chemical assay of ampicillin in bioequivalence studies in man **9**, 233
Ali, H.M., A.M. Farouk, The effect of Sudanese diet on the bioavailability of ampicillin **6**, 301
Ali, H.M., M.M.A. Homeida, J. Ford, C.A. Truman, C.J.C. Roberts, A.A. Badwan, Paracetamol bioavailability from an elixir, a suspension and a new alcohol-free liquid dosage form in humans **42**, 155
Ali, S.I., see Moeller, H. **7**, 157
Ali, S.L., T. Strittmatter, Separation and determination of four ergot alkaloids, dihydroergotamine, dihydroergocornine, dihydroergocryptine and dihydroergocristine methanesulfonates by high performance liquid chromatography **4**, 111
Separation and quantitative determination of tetracycline, epitetracycline, epianhydrotetracycline and anhydrotetracycline by high-performance liquid chromatography **1**, 185
Separation and determination of four ergot alkaloids **6**, 183
Alireza Mortazavi, S., see Ghassabian, S. **130**, 49
Alkabes, M., see Touitou, E. **133**, 85
Alkan, M.H., see Groves, M.J. **3**, 81
Alkaysi, H.N., M.H. Abdel-Hay, M. Sheikh Salem, A.M. Gharaibeh, T.E. Na'was, Chemical and microbiological investigations of metal ion interaction with norfloxacin **87**, 73
Alkaysi, H.N., see Gharaibeh, A.E.M. **100**, 253
Allaben, L., see Wong, O. **76**, 171
Allain, H., see Cognez, P. **123**, 137
Allain, P., see Prudat-Christiaens, C. **141**, 109
Allan, G., see Brain, K.R. **71**, R9
Allan, G., see Hadgraft, J. **141**, 17
Allemann, E., R. Gurny, E. Doelker, Preparation of aqueous polymeric nanodispersions by a reversible salting-out process: Influence of process parameters on particle size **87**, 247
Allemann, E., see Quintanar-Guerrero, D. **143**, 133
Allen, J.C., see Morton, A.K. **9**, 321; **18**, 39
Allen, Jr. L.V., see Kale, N.J. **57**, 87
Allen, Jr. L.V., see Seth, S.C. **106**, 7
Allen, Jr. L.V., see Tu, Y.H. **51**, 157; **61**, 119
Allen, M.A., see Cho, M.J. **1**, 281
Allman, J., A.H.L. Ho, C. Ranson, J.K. Sugden, Effect of food dyes on radicals in aqueous media **115**, 241
Allsopp, M.A., G.J. Sewell, G.G. Rowl, C.M. Riley, R.L. Schowen, The degradation of carboplatin in aqueous solutions containing chloride or other selected nucleophiles **69**, 197

Allwood, M.C., The compatability of four trace elements in total parenteral nutrition infusions **16**, 57
The influence of buffering on the stability of erythromycin injection in small-volume infusions **80**, R7
The release of phthallate ester plasticizer from intravenous administration sets into fat emulsion **29**, 233
The stability of erythromycin injection in small-volume infusions **62**, R1
Allwood, M.C., P.W. Brown, Stability of ampicillin infusions in unbuffered and buffered saline **97**, 219
Allwood, M.C., H. Martin, The extraction of diethylhexylphthalate (DEHP) from polyvinyl chloride components of intravenous infusion containers and administration sets by paclitaxel injection **127**, 65
Allwood, M.C., J.H. Plane, The degradation of vitamin A exposed to ultraviolet radiation **19**, 207
The wavelength-dependent degradation of vitamin A exposed to ultraviolet radiation **31**, 1
Allwood, M.C., J.N. Staniforth, Sorption of parenteral nitrates during administration with a syringe pump and extension set **39**, 183
Allwood, M.C., see Holmes, C.J. **1**, 133; **2**, 325
Almeida, A.J., S. Runge, R.H. Muller, A.D. Woolfson, Peptide-loaded solid lipid nanoparticles (SLN): Influence of production parameters **149**, 255
Almeida, L.M., see Moreira, J.N. **147**, 153
Almog, S., see Touitou, E. **43**, 9
Almond, H., see Walkling, W.D. **4**, 39
Alonso, I.G., E.L. Marino, A. Dominguez-Gil, Pharmacokinetics of cefotiam after i.v. administration to healthy volunteers **19**, 345
Alonso, M.J., C. Losa, P. Calvo, J.L. Vila-Jato, Approaches to improve the association of amikacin sulphate to poly(alkylcyanoacrylate) nanoparticles **68**, 69
Alonso, M.J., P. Maincent, T. Garcia-Arias, J.L. Vila-Jato, A comparative biopharmaceutical study of fresh and ageing tolbutamide-polyethyleneglycols solid dispersions **42**, 27
Alonso, M.J., see Calvo, P. **103**, 283
Alonso, M.J., see Egea, M.A. **67**, 103
Alonso, M.J., see Henry-Michell, S. **35**, 121
Alonso, M.J., see Sanchez, A. **99**, 263
Alonso, M.J., see Torres, D. **59**, 9
Alpar, H.O., see Lewis, D.A. **22**, 137
Alpar, H.O., A.M. Bason, J.A. Hickman, F.M. Richards, W.N. Field, Estimation by FACS of the delivery of liposome encapsulated macromolecules into myeloid cells **62**, 133
Alpar, H.O., J.C. Bowen, M.R.W. Brown, Effectiveness of liposomes as adjuvants of orally and nasally administered tetanus toxoid **88**, 335
Alpar, O.H., J.B. Bamford, V. Walters, The in vitro incorporation and release of hydroxocobalamin by liposomes **7**, 349
Alpsten, M., see Abrahamsson, B. **140**, 229
Alsina, M.A., N. Sole, C. Mestres, M.A. Busquets, I. Haro, J.M. Garcia Anton, Physicochemical interaction of opioid peptides with phospholipids and membranes **70**, 111
Alsina, M.A., see Colome, C. **90**, 59
Alsina, M.A., see Egea, M.A. **67**, 103
Alsina, M.A., see Mestres, C. **76**, 145
Alsina, M.A., see Reig, F. **44**, 257, 269
Altomare, C., A. Carotti, S. Cellamare, M. Ferappi, Lipophilicity measurements of benzenesulfonamide inhibitors of carbonic anhydrase by reversed-phase HPLC **56**, 273
Altomare, C., A. Carotti, S. Cellamare, M. Ferappi, R. Cagiano, G. Renna, QSAR analysis of chemical and serum-catalyzed hydrolysis of phenyl ester prodrugs of nipecotic acid **48**, 91
Altomare, C., see Trapani, G. **116**, 95
Altorfer, H., see Werner, I.A. **63**, 155
Alvarez-Fuentes J., I. Carabello, A. Boza, J.M. Llera, M.A. Holgado, M. Fernandez-Arevalo, Study of a complexation process between naltrexone, Eudragit^{R} L as an oral controlled release system **148**, 219
Alvarez-Fuentes J., M. Fernandez-Arevalo, M.A. Holgado, I. Caraballo, A.M. Rabasco, J.A. Mico, O. Rojas, A. Ortega-Alvaro, Preclinical study of a controlled release oral morphine system in rats **139**, 237
Alvarez-Fuentes, J., see Holgado, M.A. **114**, 13; **118**, 151; **142**, 143
Alvarez, A.I., see Del Estal, J.L. **91**, 105; **102**, 257
Alvarez de Eulate, J., see Veiga, M.D. **110**, 223
Alway, B., R. Sangchantra, P.J. Stewart, Modelling the dissolution of diazepam in lactose interactive mixtures **130**, 213
Aly, Z.H., see Zaghloul, I.Y. **131**, 131
Am Ende, M.T., C.L. Bell, N.A. Peppas, G. Massimo, P. Colombo, Measurement of the swelling force in ionic polymeric networks: II. Swelling force and disintegration of controlled release dosage formulations using pH-sensitive components **120**, 33
Amagai, Y., see Watanabe, Y. **64**, 147
Amagase, H., see Murakami, T. **49**, 173; **54**, 259
Ambroggi, G., see Leroy, P. **82**, 157
Amdidouche, D., H. Darrouzet, D. Duchene, M.C. Poelman, Inclusion of retinoic acid in β-cyclodextrin **54**, 175
Amdidouche, D., P. Montassier, M.C. Poelman, D. Duchene, Evaluation by laser Doppler velocimetry of the attenuation of tretinoin induced skin irritation by β-cyclodextrin complexation **111**, 111
Amdidouche, D., see Leomy, C. **115**, 235
Amicosante, G., see Di Giulio, A. **74**, 183; **144**, 27
Amidon, G., see Johnston, T.P. **59**, 95
Amidon, G., see Vadnere, M. **22**, 207
Amidon, G.E., K.R. Middleton, Accelerated physical stability testing and long-term predictions of changes in the crushing strength of tablets stored in blister packages **45**, 79
Amidon, G.L., see Johnston, T.P. **52**, 139
Amidon, G.L., see Lin, S.Y. **95**, 57
Amidon, G.L., see Oh, D.M. **85**, 181
Amidon, G.L., see Sinko, C.M. **55**, 247
Amidon, G.L., see Skelly, J.P. **63**, 83
Amidon, G.L., see Yu, L.X. **140**, 111

Ammar, H.O., M.S. El-Ridy, M. Ghorab, M.M. Ghorab, Evaluation of the antischistosomal effect of praziquantel in a liposomal delivery system in mice **103**, 237
Ammoury, N., see Fessi, H. **55**, R1
Amperiadou, A., M. Georgarakis, Controlled release salbutamol sulphate microcapsules prepared by emulsion solvent-evaporation technique and study on the release affected parameters **115**, 1
Amselem, S., D. Friedman, A. Yogev, W.R. Anderson, D.O. Helton, T. Loftsson, N. Bodor, E. Pop, M.E. Brewster, Formulation development for a zidovudine chemical delivery system 2. Towards oral and non-parenteral dosage forms **125**, 31
Amselem, S., see Brewster, M.E. **125**, 17
Amstrong, D.J., see Perez-Marcos, B. **111**, 251
Anderberg, E.K., M. Bisrat, C. Nystrom, Physicochemical aspects of drug release. VII. The effect of surfactant concentration and drug particle size on solubility and dissolution rate of felodipine, a sparingly soluble drug **47**, 67
Anderberg, E.K., C. Nystrom, Physicochemical aspects of drug release. X. Investigation of the applicability of the cube root law for characterization of the dissolution rate of fine particulate materials **62**, 143
Anderberg, E.K., see Bisrat, M. **80**, 191
Anders, R., H.P. Merkle, Evaluation of laminated muco-adhesive patches for buccal drug delivery **49**, 231
Andersen, C., see Christrup, L.L. **66**, 169
Andersen, F.M., H. Bundgaard, A. Dominguez-Gil, Inclusion complexation of metronidazole benzoate with β-cyclodextrin and its depression of anhydrate–hydrate transition in aqueous suspensions **19**, 189
Andersen, F.M., H. Bundgaard, H.B. Mengel, Formation, bioavailability and organoleptic properties of an inclusion complex of femoxetine with β-cyclodextrin **21**, 51
Andersin, R., S. Tammilehto, Photochemical decomposition of midazolam. II. Kinetics in ethanol **56**, 175
Photochemical decomposition of midazolam. IV. Study of pH-dependent stability by high-performance liquid chromatography **123**, 229
Anderson, B.D., W.W. Chu, R.E. Galinsky, Reduction of first-pass metabolism of propranolol after oral administration of ester prodrugs **43**, 261
Anderson, B.D., J.H. Rytting, T. Higuchi, Vapor pressure studies of self-association of alcohols in isooctane. I. The effect of chain length **1**, 15
Anderson, B.D., M.B. Wygant, T.X. Xiang, W.A. Waugh, V.J. Stella, Preformulation solubility and kinetic studies of 2′,3′-dideoxypyrine nucleosides: Potential anti-AIDS agents **45**, 27
Anderson, B.D., see Darrington, R.T. **59**, 35
Anderson, B.D., see Oh, I. **73**, 23
Anderson, B.D., see Xiang, T.X. **59**, 45
Anderson, C., see Wong, O. **76**, 171
Anderson, F.M., B. Bundgaard, The influence of cyclodextrin complexation on the stability of betamethasone-17-valerate **20**, 155
Anderson, G.J., see Toth, I. **77**, 13
Anderson, L., see McCafferty, D.F. **38**, 59
Anderson, W., see Pop, E. **84**, 39
Anderson, W.K., see Stella, V.J. **71**, 157
Anderson, W.R., see Amselem, S. **125**, 31
Anderson, W.R., see Brewster, M.E. **125**, 17
Anderson, W.R., see Pop, E. **140**, 33
Andersson, J., see Moren, F. **6**, 295
Andersson, M., see Sturesson, C. **89**, 235
Ando, H., T. Ohwaki, M. Ishii et al., Crystallization of theophylline in tablets **34**, 153
Ando, H., see Aoki, S. **95**, 67
Ando, H.Y., see Simon, B.H. **121**, 233
Ando, T., see Hodoshima, N. **146**, 81
Andonovski, B., see Dimitrovska, A. **134**, 213
Andrady, A.L., see Pitt, C.G. **45**, 1
Andremont, A., see Henry-Michell, S. **35**, 121
Andres, C., P. Reginault, M.H. Rochat, B. Chaillot, Y. Pourcelot, Particle-size distribution of a powder: Comparison of three analytical techniques **144**, 141
Andreux, J.P., see Leroy-Lechat, F. **101**, 97
Andrews, A.H., see Hempenstall, J.M. **23**, 131
Andrews, A.H., see Irwin, W.J. **20**, 25
Andrianoff, N., see Sebert, P. **99**, 37
Andry, M.C., F. Edwards-Levy, M.C. Levy, Free amino group content of serum albumin microcapsules. III. A study at low pH values **128**, 197
Andry, M.C., see Edwards-Levy, F. **96**, 85; **103**, 253
Andry, M.C., see Levy, M.C. **62**, 27; **69**, R1
Angberg, M., see Sebhatu, T. **104**, 135
Angberg, M., C. Nystrom, S. Castensson, Evaluation of heat-conduction microcalorimetry in pharmaceutical stability studies VII. Oxidation of ascorbic acid in aqueous solution **90**, 19
Evaluation of heat-conduction microcalorimetry in pharmaceutical stability studies. II. Methods to evaluate the microcalorimetric response **61**, 67
Evaluation of heat-conduction microcalorimetry in pharmaceutical stability studies. III. Crystallographic changes due to water vapour uptake in anhydrous lactose powder **73**, 209
Evaluation of heat-conduction microcalorimetry in pharmaceutical stability studies. IV. The influence of microcrystalline cellulose on the hydration rate of anhydrous lactose **77**, 269
Evaluation of heat-conduction microcalorimetry in pharmaceutical stability studies. V. A new approach for continuous measurements in abundant water vapour **81**, 153
Evaluation of heat-conduction microcalorimetry in pharmaceutical stability studies. VI. Continuous monitoring of the interaction of water vapour with powders and powder mixtures at various relative humidis **83**, 11
Anglade, P., see Postaire, E. **55**, 99
Anguiano-Igea, S., F.J. Otero-Espinar, J.L. Vila-Jato, J. Blanco-Mendez, Improvement of clofibrate dissolution by complexation with cyclodextrin **135**, 161
Anguiano-Igea, S., see Blanco-Fuente, H. **138**, 103, 142, 169
Anguiano-Igea, S., see Ganza-Gonzalez, A. **106**, 179

Anguiano-Igea, S., see Garcia-Gonzalez, N. **100**, 25, 65; **104**, 107
Anguiano-Igea, S., see Otero-Espinar, F.J. **75**, 37; **79**, 149; **70**, 35
Anie, S.J., J.T. Fell, R.D. Waigh, B. Wood, Gastrointestinal transit studies in the rat using ^{19}F NMR **76**, 183
Anie, S.J., J.T. Fell, R.D. Waigh, Visualisation of a fluorinated pharmaceutical formulation in the anaesthetised rat using ^{19}F- and ^{1}H NMR imaging **69**, 181
Anigbogu, A.N.C., A.C. Williams, B.W. Barry, H.G.M. Edwards, Fourier transform Raman spectroscopy of interactions between the penetration enhancer dimethyl sulfoxide and human stratum corneum **125**, 265
Anik, S.T., J.Y. Hwang, Adsorption of D-Nal(2)^{6}LHRH, a decapeptide, onto glass and other surfaces **16**, 181
Annapragada, A., A. Adjei, An analysis of the Fraunhofer Diffraction method for particle size distribution analysis and its application to aerosolized sprays **127**, 219
Numerical simulation of milling processes as an aid to process design **136**, 1
Annapragada, A., see Swanson, P.D. **142**, 33
Antal, E.J., C.F. Dick, C.E. Wright III, I.R. Welshman, E.M. Block, Comparative bioavailability of two medroxyprogesterone acetate suspensions **54**, 33
Antal, I., see Dredan, J. **145**, 61
Antikainen, O., see Juslin, L. **123**, 247; 257
Antimisiaris, S., see Macheras, P. **54**, 123
Antipas, A.S., D. Vander Velde, V.J. Stella, Factors affecting the deamidation of vancomycin in aqueous solutions **109**, 261
Anto, R.J., G. Kuttan, K.V. Dinesh Babu, K.N. Rajasekharan, R. Kuttan, Anti-tumour and free radical scavenging activity of synthetic curcuminoids **131**, 1
Antolic, S., see Zorc, B. **99**, 135
Antonin, K.H., R. Rak, P.R. Bieck, R. Preiss, U. Schenker, J. Hastewell, R. Fox, M. Mackay, The absorption of human calcitonin from the transverse colon of man **130**, 33
Antonin, K.H., see Hastewell, J. **126**, 245
Anwar, H., see Owusu-Ababio, G. **122**, 9
Aoki, H., K. Fuji, K. Miyajima, Effects of blood on the uptake of charged liposomes by perfused rat liver: Cationic glucosamine-modified liposomes interact with erythrocyte and escape phagocytosis by macrophages **149**, 15
Aoki, H., C. Sun, K. Fuji, K. Miyajima, Disposition kinetics of liposomes modified with synthetic aminoglycolipids in rats **115**, 183
Aoki, S., H. Ando, K. Tatsuishi, K. Uesugi, H. Ozawa, Determination of the mechanical impact force in the in vitro dissolution test and evaluation of the correlation between in vivo and in vitro release **95**, 67
Aoki, S., T. Ohwaki, K. Uesugi, K. Tatsuishi, H. Ozawa, M. Kayano, Preparation of a novel type of controlled-release carrier and evaluation of drug release from the matrix tablet and its physical properties **85**, 29
Aoki, S., K. Uesugi, K. Tatsuishi, H. Ozawa, M. Kayano, Evaluation of the correlation between in vivo and in vitro release of phenylpropanolamine HCl from controlled-release tablets **85**, 65
Aoyagi, N., see Ogata, H. **23**, 277, 289; **29**, 113, 121
Apichartpichean, R., see Hasegawa, T. **56**, 235; **58**, 129
Apostolelli, E., see Macheras, P. **36**, 73
Appel, M., see Tabosa de Egito, E.S. **145**, 17
Apprich, I., see Bernkop-Schnurch, A. **146**, 247
Aprahamian, M., see Damge, C. **36**, 121
Arafat, T., see Sallam, E. **67**, 247
Arafat, T.A., see Sheikh Salem, M. **141**, 257
Arakawa, T., see Niven, R.W. **109**, 17; **127**, 191
Araki, M., S. Yamashita, T. Nadai, M. Masada, Y. Kurosaki, T. Nakayama, T. Kimura, Interaction of percutaneous absorption enhancer with stratum corneum of hamster cheek pouch; an electrophysiological study **81**, 39
Araki, Y., see Fukuda, T. **36**, 1
Araki, Y., see Gomita, Y. **54**, 89
Aramaki, Y., see Hara, T. **42**, 69; **67**, 123
Aramaki, Y., see Kikuchi, S. **42**, 193
Aramaki, Y., see Niibuchi, J.J. **30**, 181
Aramaki, Y., see Tsuchiya, S. **14**, 279
Arase, K., see Mukai, E. **25**, 95
Arbeille, B., see Matchet, L. **133**, 39
Arbin, A., S. Jacobsson, K. Hanninen et al., Studies on contamination of intravenous solutions from PVC-bags with dynamic headspace GC-MS, LC-diode array techniques **28**, 211
Archer, W.I., see Kohli, R. **26**, 275; **36**, 91
Archimbault, P., see Leroy, P. **82**, 157
Archondikis, A., G. Papaioannou, Comparative study of two dissolution methods for indomethacin suppositories from fatty and water-soluble bases **55**, 217
Arellano, A., S. Santoyo, C. Martin, P. Ygartua, Enhancing effect of terpenes on the in vitro percutaneous absorption of diclofenac sodium **130**, 141
Arellano, A., see Santoyo, S. **117**, 219
Arenare, L., see Palagiano, F. **149**, 171
Arends-Scholte, A.W., see Te Wierik, G.H.P. **134**, 27
Arias-Blanco, M.J., P. Munoz, J.R. Moyano, Gines J.M., Novak C., Preliminary study of different omeprazole-γ-CD co-grinded systems **143**, 113
Arias-Blanco, M.J., see Gines, J.M. **143**, 247
Arias-Blanco, M.J., see Moyona, J.R. **148**, 211
Arias, M.J., J.M. Gines, J.R. Moyano, J.I. Perez-Martinez, A.M. Rabasco, Influence of the preparation method of solid dispersions on their dissolution rate: Study of triamterene-D-mannitol system **123**, 25
Arias, M.J., see Gines, J.M. **126**, 287
Arias, M.J., see Moyano, J.R. **114**, 95
Arikawa, K., see Sakuma, S. **149**, 93
Arima, H., T. Irie, K. Uekama, Differences in the enhancing effects of water soluble β-cyclodextrins on the release of ethyl 4-biphenylyl acetate, an anti-inflammatory agent from an oleaginous suppository base **57**, 107
Arima, H., see Irie, T. **84**, 129
Arimori, K., see Matsuo, M. **138**, 225
Arita, T., see Kohri, N. **49**, 213
Arita, T., see Nakano, M. **3**, 195
Aritomi, H., see Irie, T. **84**, 129

Armstrong, D.J., see Ford, J.L. **71**, 95, 213
Armstrong, D.J., see Mitchell, K. **66**, 233; **100**, 155, 165, 175
Armstrong, Elliott D.J.P.N.C., see Mitchell, K. **100**, 143
Armstrong, J.K., see Buckton, G. **83**, 115; **110**, 179
Armstrong, N.A., Time-dependent factors involving in powder compression and tablet manufacture **49**, 1
Armstrong, N.A., T. Gebre-Mariam, K.C. James, An apparatus for investigating drug migration into gelatin capsule shells **34**, 125
An in vitro model to simulate drug release from oily media **41**, 115
Migration of ephedrine and salicylic acid from lipid mixtures containing isopropyl myristate **41**, 205
Armstrong, N.A., K.C. James, Drug release from lipid-based dosage forms. I. Drug release from lipid-based dosage forms **6**, 185
II. quinolinemethanol◇ **6**, 195
Armstrong, N.A., A.N. Patel, T.M. Jones, Relationship between porosity and water content of dicalcium phosphate tablets **48**, 173
Armstrong, N.A., see Moreton, R.C. **122**, 79
Arnardottir, H.B., S.J. Sveinsson, T. Kristmundsdottir, The release of clindamycin phosphate from a suspension of different types of liposomes and selected topical dosages forms **134**, 71
The use of a high intensity ultrasonic processor equipped with a flow cell in the production of reverse-phase liposomes **117**, 237
Arnaud, P., C. Elkoubi, C. Renaux, A. Le Hir, Computerized linearization test for tablet dissolution kinetics **80**, 129
Arnaud, P., I. Guillard, D. Brossard, J.C. Chaumeil, Gastrointestinal diffusion tablet: Influence of polyoxyethyleneglycol 400 **129**, 279
Arnaud, P., see Benkhelifa, S. **145**, 115
Arnaud, P., see Fontan, J.E. **73**, 17; **82**, 67
Arnaud, P., see Jacolot, A. **128**, 283
Arnaud, P., see Penaud, J.F. **142**, 1
Arnaud, P., see Prudat-Christiaens, C. **141**, 109
Arnaud, P., see Valiere, C. **138**, 125
Arnaud, P., see Zuber, M. **47**, 31
Arndt, J.W., see Blok, D. **73**, 171
Arndts, D.E., see Harron, D.W.G. **12**, 277
Arnold, E., see Bundgaard, H. **18**, 79
Aronhime, M., see Benita, S. **33**, 71
Aroztegui, M., see Halbaut, L. **147**, 31
Arppe, J., see Vidgren, M. **115**, 209
Arrowsmith M., J. Hadgraft, I.W. Kellaway, The in vitro release of steroids from liposomes **14**, 191
The in vivo release of cortisone esters from liposomes and the intramuscular clearance of liposomes **20**, 347
The interaction of cortisone esters with liposomes as studied by differential scanning calorimetry **16**, 305
The stability of a prodrug, liposome and a prodrug-liposome complex in simulated biological conditions **17**, 91
Arsac, C., see Walchshofer, N. **100**, 271
Artico, R., see Martini, A. **127**, 239
Artursson, P., A. Berg, P. Edman, Biochemical and cellular effects of degraded starch microspheres on macrophages **52**, 183
Artursson, P., I. Sjoholm, Effect of opsonins on the macrophage uptake of polyacrylstarch microparticles **32**, 165
Artursson, P., see Buur, A. **129**, 223
Artursson, P., see Edman, P. **34**, 225
Artursson, P., see Fahlvik, A.K. **65**, 249
Artursson, P., see Flinn, N. **138**, 167; **143**, 125
Artursson, P., see Jorgensen, L. **95**, 209
Artursson, P., see Karlsson, J. **71**, 55; **99**, 209
Artursson, P., see Lennernas, H. **127**, 103
Artursson, P., see Lundin, S. **64**, 181; **76**, R1
Artursson, P., see Stjarnkvist, P. **40**, 215
Artursson, P., see Toth, I. **102**, 223; **105**, 241
Arwidsson, H., B. Johansson, Application of intrinsic viscosity and interaction constant as a formulation tool for film coating. III. Mechanical studies on free ethyl cellulose films, cast from organic solvents **76**, 91
Arwidsson, H., M. Nicklasson, Application of intrinsic viscosity and interaction constant as a formulation tool for film coating. II. Studies on different grades of ethyl cellulose in organic solvent systems **58**, 73
Arwidsson, H.G., see Stubberud, L. **114**, 55; **134**, 79
Asano, M., H. Fukuzaki, M. Yoshida, M. Kumakura, T. Mashimo, H. Yuasa, K. Imai, H. Yamanaka, U. Kawaharada, K. Suzuki, In vivo controlled release of a luteinizing hormone-releasing hormone agonist from poly(DL-lactic acid) formulations of varying degradation patttern **67**, 67
Asano, M., see Imasaka, K. **68**, 87; **81**, 31
Asano, M., see Yoshida, M. **115**, 61
Ashby, L.J., A.E. Beezer, G. Buckton, In vitro dissolution testing of oral controlled release preparations in the presence of artificil foodstuffs. I. Exploration of alternative methodology: Microcalorimetry **51**, 245
Ashby, L.J., see de Morais, S.M. **66**, 107
Ashford, M., J.T. Fell, D. Attwood, H. Sharma, P.J. Woodhead, An in vivo investigation into the suitability of pH dependent polymers for colonic targeting **95**, 193
Ashford, M., J.T. Fell, D. Attwood, P.J. Woodhead, An in vitro investigation into the suitability of pH-dependent polymers for colonic targeting **91**, 241
Ashton, P., K.A. Walters, K.R. Brain, J. Hadgraft, Surfactant effects in percutaneous absorption. II. Effects on protein and lipid structure of the stratum corneum **87**, 265
Surfactant effects of percutaneous absorption. I. Effects on the transdermal flux of methyl nicotinate **87**, 261
Ashton, P., see Steffansen, B. **132**, 243
Aso, Y., S. Yoshioka, T. Terao, Effects of storage on the physicochemical properties and release characteristics of progesterone-loaded poly(L-lactide) microspheres **93**, 153
Aspden, T.J., J. Adler, S.S. Davis, O. Skaugrud, L. Illum, Chitosan as a nasal delivery system: Evaluation of the effect of chitosan on mucociliary clearance rate in the frog palate model **122**, 69
Assaf, S.M., see Al-Nimry, S.S. **149**, 115

Assari, M., see Dehpour, A.R. **119**, 133
Astre, C., see Martel, P. **149**, 37
Asuero, A.G., M.J. Navas, M.A. Herrador, A.F. Recamales, Spectrophotometric evaluation of acidity constants of isonicotinic acid **34**, 81
Asuero, A.G., A bilogarithmic method for the evaluation of acidity constants of amphoteric substances from solubility measurements **52**, 129
A hyperbolic cosine method for the spectrophotometric evaluation of acidity constants of two-step overlapping equilibria **88**, 15
Buffer index in the titration of a monoprotic acid with a strong base **87**, 11
Evaluation of acidity constants of dibasic acids and diacid bases from liquid–liquid distribution measurements **89**, 103
Evaluation of partition coefficient and acidity constants of amphoteric substances by graphic differentiation of the distribution ratio against pH curve **45**, 157
On the buffer index of a single weak monoprotic acid system **134**, 1
Asuero, A.G., see Gonzalez, A.G. **72**, 193; **84**, R1; **93**, 183, BOLD 120, 215, **108**, 149
Asuncion Alsina, M., see Fonseca, M.J. **133**, 265
Atanasiu, R., A. Radu, The interaction of cationic nanoparticles with mouse capillary endothelial cells in situ and in vivo **90**, 119
Atassi, G., see Dubois, J.G. **35**, 219
Atcheva, M., M. Simeonova, M. Boeva, C. Konstantinov, Effect of polybutylcyano acrylate nanoparticles on primary immune response of mice to sheep erythrocytes **103**, 303
Athanasiou, A., see Avgoustakis, K. **79**, 67
Athanassiou, G.C., D.M. Rekkas, N.H. Choulis, Bioavailability of sulphamethoxazole-trimethoprim spheroidal granules **85**, 209
Correlation of in vitro dissolution data with in vivo plasma concentrations, for three, orally administered, formulations of sulphamethoxazole-trimethoprim, by statistical moments analysis **90**, 51
Development of sulphamethoxazole-trimethoprim spheroidal granules: Factors affecting drug release in vitro **72**, 141
Athersuch, A., see Washington, C. **64**, 217
Atkin, B.P., see Watts, P.J. **76**, 55
Atkins, P.J., T.O. Herbert, N.B. Jones, Kinetic studies on the decomposition of erythromycin A in aqueous acidic and neutral buffers **30**, 199
Atsumi, R., see Takakura, Y. **37**, 145
Attia, M.A., M.A. Kassem, S.M. Safwat, In vivo performance of [^{3}H]dexamethasone ophthalmic film delivery systems in the rabbit eye **47**, 21
Attia, M.A., see Habib, F.S. **26**, 243
Attia, M.A., see Kassem, M.A. **29**, 243; **32**, 47
Attwood, D., S.P. Agarwal, Self-association of disodium cromoglycate in dilute aqueous solution **22**, 25
Attwood, D., J.H. Collett, C.A. O'Connor, Influence of gamma irradiation on the rheological properties of gels of the poloxamine, Synperonic T908 **70**, 147
The effect of gamma irradiation on the surface, rheological and micellar behaviour of the block copolymer, Tetronic 908 **65**, 201
Attwood, D., J.H. Collett, C.J. Tait, The micellar properties of the poly(oxyethylene)–poly(oxypropylene) copolymer Pluronic F127 in water and electrolyte solution **26**, 25
Attwood, D., L. Johansen, J.A. Tolley, J. Rassing, A new ultrasonic method for the measurement of the diffusion coefficient of drugs within hydrogel matrices **9**, 285
Attwood, D., G. Ktistis, A light scattering study on oil-in-water microemulsions **52**, 165
Attwood, D., C. Mallon, G. Ktistis, C.J. Taylor, A study on factors influencing the droplet size in nonionic oil-in-water microemulsions **88**, 417
Attwood, D., C. Mallon, C.J. Taylor, Phase studies on oil-in-water phospholipid microemulsions **84**, R5
Attwood, D., H.K. Patel, Composition of mixed micellar systems of cetrimide and chlorhexidine digluconate **49**, 129
Attwood, D., see Ashford, M. **91**, 241; **95**, 193
Attwood, D., see Barnwell, S.G. **88**, 423; **89**, 245; **128**, 145
Attwood, D., see Burns, S.J. **110**, 291; **121**, 37; **134**, 223; **141**, 9
Attwood, D., see Burrows, R. **111**, 283
Attwood, D., see Cole, S.K. **80**, 63; **88**, 211
Attwood, D., see Higginbottom, S. **109**, 173
Attwood, D., see Martini, L. **113**, 223
Attwood, D., see Miyazaki, S. **118**, 257
Attwood, D., see Rassing, J. **13**, 47
Attwood, D., see Ruth, H.S. **116**, 253
Attwood, D., see Wakerly, Z. **129**, 73
Attwood, D., see Wood, J.M. **7**, 189
Audran, M., J.L. Chanal, G. Berge et al., Correlations between structure and pharmacokinetics of a series of cresotamide derivatives **20**, 129
Audran, M., J.L. Chanal, G. Berge, R. Marignan, Correlation between structure and serum albumin binding in a series of cresotinamide derivatives **15**, 273
Audus, K.L., M.R. Tavakoli-Saberi, Aminopeptidases of newborn bovine nasal turbinate epithelial cell cultures **76**, 247
Audus, K.L., see Borchard, G. **110**, 29
Audus, K.L., see Reardon, P.M. **94**, 161
Audus, K.L., see Rim, S. **32**, 79
Audus, K.L., see Sheng, H. **131**, 159
Audus, K.L., see Tavakoli-Saberi, M.R. **56**, 135
Audus, K.L., see Zheng, H. **101**, 121; **129**, 103
Augsburger, L.L., see Baichwal, A.R. **26**, 191
Augsburger, L.L., see Botzolakis, J.E. **12**, 341
Augsburger, L.L., see Cropp, J.W. **71**, 127
Augsburger, L.L., see Mehta, A.M. **7**, 327
Augsburger, L.L., see Scott, D.C. **84**, 49
Augsburger, L.L., see Torrado, J.J. **106**, 149
Aulton, M.E., see Ferrari, F. **121**, 11
Aulton, M.E., see Okutgen, E. **119**, 193
Aulton, M.E., see Wong, D.Y.T. **72**, 233
Aungst, B.J., Site-dependence and structure-effect relationships for alkylglycosides as transmucosal absorption promoters for insulin **105**, 219

Aungst, B.J., F. Hirayama, E. Shefter, E.G. Shami, Prodrugs for improved oral nalbuphine bioavailability: Inter-species differences in the disposition of nalbuphine and its acetylsalicylate and anthranilate esters **38**, 199
Aungst, B.J., S. Phang, Metabolism of a neurotensin (8–13) analog by intestinal and nasal enzymes, and approaches to stabilize this peptide at these absorption sites **117**, 95
Aungst, B.J., N.J. Rogers, E. Shefter, Enhancement of naloxone penetration through human skin in vitro using fatty acids, fatty alcohols, surfactants, sulfoxides and amides **33**, 225
Aungst, B.J., N.J. Rogers, Comparison of the effects of various transmucosal absorption promoters on buccal insulin delivery **53**, 227
Aungst, B.J., see Hussain, M.A. **36**, 127; **105**, 7
Auramo, P., see Laine, E. **43**, 241
Auriola, S., see Jarvinen, T. **75**, 249; **79**, 233
Ausborn, M., see Engel, A. **107**, 99
Ausborn, M., see Schreier, H. **87**, 183
Austin, P., see Collier, P.J. **66**, 201; **74**, 195
Avgerinos, A., S. Malamataris, Bioavailability of controlled release indomethacin microspheres and pellets **63**, 77
Avgerinos, A., A. Noormohammadi, A.J. Hutt, Disposition of ibuprofen enantiomers following the oral administration of a novel controlled release formulation to healthy volunteers **68**, 97
Avgerinos, A., see Malamataris, S. **62**, 105
Avgoustakis, K., A. Athanasiou, P.P. Georgakopoulos, Effect of helix characteristics on the dissolution rate of hard gelatin capsules **79**, 67
Avgoustakis, K., J.R. Nixon, Biodegradable controlled release tablets. I. Preparative variables affecting the properties of poly(lactide-co-glycolide) copolymers as matrix forming material **70**, 77
Biodegradable controlled release tablets. II. Preparation and properties of poly(lactide-co-glycolide) powders **99**, 239
Biodegradable controlled release tablets. III. Effect of polymer characteristics on drug release from heterogeneous poly(lactide-co-glycolide) matrices **99**, 247
Avramoff, A., see Hoffman, A. **111**, 197
Awad, R., see Sallam, E. **67**, 247
Awata, N., see Akimoto, K. **3**, 109
Awata, N., see Nakada, Y. **89**, 169
Awataguchi, M., see Naito, S.I. **24**, 127
Awazu, S., see Kajii, H. **33**, 253
Awidi, A., see Othman, S. **41**, 197
Awidi, A., see Shaheen, O. **38**, 123
Aydin, Z., J. Akbuga, Chitosan beads for the delivery of salmon calcitonin: Preparation and release characteristics **131**, 101
Preparation and evaluation of pectin beads **137**, 133
Ayres, J.W., see Borin, M.T. **54**, 199
Ayres, J.W., see Hossain, M. **133**, 223
Ayres, J.W., see Lee, B.J. **124**, 119
Ayres, J.W., see Tavipatana, W. **30**, 143; **38**, 139
Azemar, N., see Garcia-Celma, M.J. **105**, 77
Azmin, M.N., A. Setanoians, R.G.G. Blackie et al., Formulation of 1,3,5-triglycidyl-*S*-triazinetrione (α-TGT) for intravenous injection **10**, 109
Aznarte, P., see Bedmar, M.C. **138**, 95

B

Babalola, C.P., see Onyeji, C.O. **100**, 249
Babhair, S., A. Hussain, *O*-acylsalicylamides as possible prodrugs for salicylamide. I. Kinetics and mechanisms of their degradation and reaction with enzymes and sodium bisulfite **13**, 273
Babhair, S.A., see Gouda, M.W. **37**, 227; **22**, 257
Babhair, S.A., see Moustafa, M.A. **36**, 185
Babler, M., see Kararli, T.T. **102**, 177
Babu, S.R., see Luner, P.E. **128**, 29
Babu, S.R., see Radebaugh, G.W. **57**, 95
Bacelle, L., see Richardson, J.L. **115**, 9
Bachmann, D., M. Brandl, G. Gregoriadis, Preparation of liposomes using a Mini-Lab 8.30 H high-pressure homogenizer **91**, 69
Backensfeld, T., B.W. Muller, K. Kolter, Interaction of NSA with cyclodextrins and hydroxypropyl cyclodextrin derivatives **74**, 85
Bacon, W.E., see Acree, Jr. W.E. **20**, 209
Badawi, A.A., see Fouli, A.M. **14**, 95
Badawi, A.A., A.M. Fouli, A.A. El-Sayed, Drug release from matrices made of polymers with reacting sites **6**, 55
Badawy, S.I.F., M.M. Ghorab, C.M. Adeyeye, Characterization and bioavailability of danazol-hydroxypropyl β-cyclodextrin coprecipitates **128**, 45
Bioavailability of danazol-hydroxypropyl-β-cylodextrin complex by different routes of administration **145**, 137
Badian, M., Normand Y. Le, W. Rupp, R. Zapf, There is no interaction between dipyrone (metamizol) and the anticoagulants, phenprocoumon and ethylbiscoumacetate, in normal caucasian subjects **18**, 9
Badwan, A.A., Khordagui El, A.M. Saleh, S.A. Khalil, The solubility of benzodiazepines in sodium salicylate solution and a proposed mechanism for hydrotropic solubilization **13**, 67
Badwan, A.A., O.A. Jawan, L. Owais, Metoclopramide determination by amperometric and potentiometric titrations—application to some pharmaceutical preparations **28**, 41
Badwan, A.A., see Saleh, A.M. **17**, 115
Badwan, A.A., see Sheikh, Salem M. **141**, 257
Bae, E.J., see Yoon, E.J. **97**, 213
Bae, Y.H., see Cremers, H.F.M. **110**, 117; **120**, 51
Bae, Y.H., see Kwon, G.S. **79**, 191
Baert, L., G.R.B. Down, A comparison of two methods of instrumenting a small-scale basket extruder **107**, 219
Baert, L., J.P. Remon, Influence of amount of granulation liquid on the drug release rate from pellets made by extrusion spheronisation **95**, 135
Water vapour permeation of aqueous based ethylacrylate methylmethacrylate copolymer films **99**, 181
Baert, L., J.P. Remon, J.A.C. Elbers, Bommel E.M Van, Comparison between a gravity feed extruder and a twin screw extruder **99**, 7

Baert, L., J.P. Remon, P. Knight, J.M. Newton, A comparison between the extrusion forces and sphere quality of a gravity feed extruder and a ram extruder **86**, 187
Baert, L., H. Vermeersch, J.P. Remon, J. Smeyers-Verbeke, D.L. Massart, Study of parameters important in the spheronisation process **96**, 225
Baert, L., see Van, der Meeren **106**, 89
Baert, L., see Vervaet, C. **107**, 29; **108**, 207; **116**, 131
Bagger Hansen, A., see Bundgaard, H. **3**, 341
Baggerman, C., J.A. Loos, H.E. Junginger, Endotoxin removal from large-volume parenterals by various adsorbents **27**, 17
Baggot, J.D., see Pope, D.G. **14**, 123
Baghal, T., see Sallam, E. **67**, 247
Bahrendt, W.A., see Beckett, A.H. **52**, 37
Bai, J.P.F., The regional differences in the mucosal-cell lysosomal proteases within the rat small intestine **107**, 133
Effects of bile salts on brush-border and cytosolic proteolytic activis of intestinal enterocytese **111**, 147
Stability of neurotensin and acetylneurotensin 8–13 in brush-border membrane, cytosol, and homogenate of rat small intestine **112**, 133
aichwal, A.R., L.L. Augsburger, Development and validation of a modified annular shear cell (MASC) to study frictional properties of lubricants **26**, 191
Baichwal, A.R., see Staniforth, J.N. **40**, 267
Baichwal, A.R., see Tobyn, M.J. **128**, 113
Baie, S., see Mahrag, Tur K. **148**, 63
Bailey, A.J.G., A. Cole, J. Goodfield et al., The complexation of transition series metal ions by nalidixic acid and related methoxyquinolones: Its influence on partition coefficients with reference to antibacterial activity **22**, 283
Bailey, C.J., see Nicklin, P. **128**, 155
Baillie, A.J., T.F. Dolan, J. Alexander, K.C. Carter, Visceral leishmaniasis in the BALB/c mouse: Sodium stibogluconate treatment during acute and chronic stages of infection **57**, 23
Baillie, A.J., see Carter, K.C. **53**, 129
Baillie, A.J., see Collins, M. **83**, 251
Baillie, A.J., see Mendelow, A.Y. **49**, 29
Baillie, A.J., see Onaga, I.C. **7**, 89; **12**, 323
Bains, D., S.L. Boutell, J.M. Newton, The influence of moisture content on the preparation of spherical granules of barium sulphate and microcrystalline cellulose **69**, 233
Bak, A., see Friis, G.J. **136**, 61
Bakalova, M.V., see Porter, C.J.H. **141**, 227
Baker, E., see Hoelgaard, A. **43**, 233
Baker, N.D., R.J. Griffin, W.J. Irwin, The percutaneous absorption of *m*-azidopyrimethamine: A soft antifolate for topical use **65**, 115
Baker, N.D., R.J. Griffin, W.J. Irwin, J.A. Slack, The reduction of aryl azides by dithiothreitol: A model for bioreduction of aromatic azido-substituted drugs **52**, 231
Bakhouya, A., M. Saidna, J.M. Vergnaud, Calculation of the antibiotic level in the plasma with an oral erosion-controlled dosage form **143**, 143
Calculation of the blister fluid-time history with ciprofloxacin administered orally or by infusion **146**, 225
Bakker, S., see Moolenaar, F. **5**, 127
Balant, L.P., see Jonkman, J.H.G. **25**, 113
Balboni, G., see Damge, C. **36**, 121
Baldvinsdottir, J., see Brewster, M.E. **75**, R5
Baldvinsdottir, J., see Loftsson, T. **79**, 107; **98**, 225
Baldwin, M.A., see Toth, I. **73**, 259
Baldwin, R.W., see Pimm, M.V. **79**, 77
Ballantine, J., A.D. Woolfson, Determination of catecholamines in pharmaceutical preparations by differential pulse polarography at the glassy carbon electrode **3**, 239
Bamba, M., F. Puisieux, J.P. Marty, J.T. Carstensen, Release mechanisms in gelforming sustained release preparations **2**, 307
Bambagiotti-Alberti, M., see Vincieri, F.F. **48**, 119
Bamford, J.B., see Alpar, O.H. **7**, 349
Bando, H., M. Sahashi, S. Mohri, F. Yamashita, Y. Takakura, M. Hashida, In vivo skin penetration enhancement of acyclovir by theoretical design of prodrug-enhancer combination **145**, 103
Bando, H., M. Sahashi, T. Takagi, F. Yamashita, Y. Takakura, M. Hashida, Analysis of in vitro skin penetration of acyclovir prodrugs based on a diffusion model with a metabolic process **135**, 91
Banerjee, P.S., W.A. Ritschel, Transdermal permeation of vasopressin. I. Influence of pH, concentration, shaving and surfactant on in vitro permeation **49**, 189
Transdermal permeation of vasopressin. II. Influence of Azone on in vitro and in vivo permeation **49**, 199
Banga, A.K., M. Katakam, R. Mitra, Transdermal iontophoretic delivery and degradation of vasopressin across human cadaver skin **116**, 211
Banga, A.K., Y.W. Chien, Systemic delivery of therapeutic peptides and proteins **48**, 15
Banker, G.S., see Agyilirah, G.A. **75**, 241
Banker, G.S., see Davis, M. **28**, 157
Banker, G.S., see Kumar, V. **79**, 61
Banker, G.S., see Wei, S. **142**, 175
Bankowsky, K., see Toth, I. **125**, 117
Banning, D., see Craig, D.Q.M. **114**, 103
Bao, Y.T., see Pitt, C.G. **45**, 1
Barary, M.H., see El-Yazbi, F.A. **27**, 139
Barary, M.H., see Wahbi, A.M. **92**, 15
Barbato, F., see Palagiano, F. **149**, 171
Barbe, C., see Halbaut, L. **130**, 203; **147**, 31
Barber, R.F., P.N. Shek, Tear-induced release of liposome-entrapped agents **60**, 219
Barber, R.F., see Jurima-Romet, M. **63**, 227; **88**, 201
Barbetti, P., see Fardella, G. **121**, 123
Barbosa, J., R. Berges, I. Toro, V. Sanz-Nebot, Dissociation constants and preferential solvation of fluoroquinolones in hydroorganic mixtures used in LC **149**, 213
Bard, J., see Wu-Pong, S. **143**, 259
Bardet, L., see Alain, M. **12**, 173
Bardet, L., see Bourret, E. **12**, 19
Bardin, C., see Cohen, T. **137**, 249

Barker, B.S.A., see Craig, D.Q.M. **114**, 103
Barker, N., J. Hadgraft, Facilitated percutaneous absorption: A model system **8**, 193
Barker, S.A., K.M.G. Taylor, M.D. Short, The deposition and clearance of liposome entrapped ^{99m}Tc-DTPA in the human respiratory tract **102**, 159
Barnes, A.R., Determination of caffeine and potassium sorbate in a neonatal oral solution by HPLC **80**, 267
Barnes, A.R., D.J. Makohon, Correlation of degradation in metronidazole infusion with F_0 applied during steam sterilization at 122°C **92**, 233
Barnes, A.R., S. Nash, HPLC determination of bendrofluazide in capsules **94**, 231
Comparison of aerodynamic particle size from 250 μg per dose beclomethasone dipropionate metered-dose inhalers **139**, 255
Barnes, A.R., J.K. Sugden, Detection of photolytically produced hydroxyl radicals by reaction with 2,2,2-trichloroethanol **44**, 31
Barnes, A.R., see Hill, D.G. **147**, 127
Barnes, D., see Kochhar, S.K. **112**, 225; **115**, 35
Barnett, M.I., see Bisrat, M. **80**, 191
Barnett, M.I., see Britten, J.R. **71**, R5
Barnhill, J., see Boxenbaum, H. **21**, 135
Barnwell, S.G., S.J. Burns, S. Higginbottom, I. Whelan, D. Corness, G. Hay, E. Rosenberg, D. Attwood, Demonstration of the importance of biphasic oleic acid delivery for enhancing the bioavailability of propranolol in healthy volunteers **128**, 145
Barnwell, S.G., T. Laudanski, M. Dwyer, M.J. Story, P. Guard, S. Cole, D. Attwood, Reduced bioavailability of atenolol in man: The role of bile acids **89**, 245
Barnwell, S.G., T. Laudanski, M.J. Story, C.B. Mallinson, R.J. Harris, S.K. Cole, M. Keating, D. Attwood, Improved oral bioavailability of propranolol in healthy human volunteers using a liver bypass drug delivery system containing oleic acid **88**, 423
Barnwell, S.G., see Burns, S.J. **110**, 291; **121**, 37; **134**, 223; **141**, 9
Barnwell, S.G., see Cole, S.K. **80**, 63
Barnwell, S.G., see Higginbottom, S. **109**, 173
Barnwell, S.G., see White, D.G. **69**, 169
Barone, J.S., see Faulkner, R.D. **43**, 53
Barr, W.H., see Peck, C.C. **82**, 9
Barr, W.H., see Skelly, J.P. **63**, 83
Barracchini, A., see Di, Giulio A. **144**, 27
Barratt, G., see Guterres, S.S. **113**, 57
Barratt, G., see Tabosa, de Egito E **145**, 17
Barratt, G., see Yu, W. **89**, 139
Barrow, L., see Watts, P.J. **87**, 215
Barry, B.W., M.C. Meyer, The rheological properties of Carbopol gels. I. Continuous shear and creep properties of Carbopol gels **2**, 1
The rheological properties of Carbopol gels. II. Oscillatory properties of Carbopol gels **2**, 27
Barry, B.W., see Akhter, S.A. **21**, 17
Barry, B.W., see Anigbogu, A.N.C. **125**, 265
Barry, B.W., see Bond, J.R. **41**, 91
Barry, B.W., see Cornwell, P.A. **94**, 189; **117**, 101; **127**, 9
Barry, B.W., see Goodman, M. **57**, 29
Barry, B.W., see Kadir, R. **60**, 139; **70**, 87
Barry, B.W., see Megrab, N.A. **116**, 101
Barry, B.W., see Michniak-Mikolajczak, B.B. **31**, 247
Barry, B.W., see Moghimi, H.R. **131**, 103, 117; **145**, 37, 49; **146**, 41
Barry, B.W., see Roe, J.M. **14**, 159
Barry, B.W., see Southwell, D. **18**, 299; **22**, 291
Barry, B.W., see Williams, A.C. **56**, 43; **74**, 157; **81**, R11; **86**, 69
Barry, B.W., see Woodford, R. **26**, 145
Barry, B.W., see Yamane, M.A. **116**, 237
Bartel, R.L., see Shah, P.K. **68**, 285
Barthelemy, C., see Santi, P. **68**, 43
Barthelemy, J., see Chopineau, J. **73**, 191
Barthes, D., see Rochard, E. **101**, 257
Barzegar-Jalali, A., see Barzegar-Jalali, M. **120**, 111
Barzegar-Jalali, M., Determination of absolute drug bioavailability without intravenous administration by renal clearance perturbation using urinary excretion data **5**, 253
Calculation of infinity value of cumulative drug amount excreted via urine in bi-exponential processes **8**, 63
Prediction of infinity value of cumulative drug amount excreted via urine in tri-exponential processes **8**, 235
Estimation of parameters of one compartment open model with constant intravenous infusion using early blood level data **9**, 349
Some methods for estimating absorption rate constant of linear one-compartment open models **13**, 103
On the calculation of absorption rate constant of linear one-compartment open models using peak blood level or peak urinary excretion rate and post-absorptive data **13**, 99
A theoretical justification for the superposition principle **13**, 239
Theoretical justification for the midpoint back-extrapolation method of estimating apparent volume of distribution of drugs after a short intravenous infusion **18**, 353
Some applications of the parallel line method **19**, 101
A model for linearizing drug dissolution data **63**, R9
Determination of absolute drug bioavailability without intravenous administration **65**, R1
Barzegar-Jalali, M., A. Barzegar-Jalali, Bioavailability determination for drugs exhibiting variable renal excretion rate constant between treatments **120**, 111
Barzegar-Jalali, M., J. Hanaee, Model for solubility estimation in mixed solvent systems **109**, 291
Barzegar-Jalali, M., A. Jouyban-Gharamaleki, Models for calculating solubility in binary solvent systems **140**, 237
Barzegar-Jalali, M., A. Jouyban-Gharamaleki, J. Hanaee, M.H. Chookhachizadeh-Moghaddam, Comparison of double log–log, mixture response-surface and combined NIBS/Redlich–Kister solubility models **144**, 127
Barzegar-Jalali, M., J.H. Richards, The effect of suspending agents on the release of aspirin from aqueous suspensions in vitro **2**, 195

The effects of various suspending agents on the bioavailability of aspirin and salicylic acid in the rabbit **3**, 133
Simulation of the effects of gastrointestinal transit rate on drug availability from aspirin suspensions using a modified dialysis method **4**, 127
Kinetics of drug dissolution and dialysis from suspension dosage forms **4**, 171
Barzegar-Jalali, M., M. Toomanian, Theoretical consideration for the cases where absorption rate constant approaches elimination rate constant in the linear one-compartment open models **12**, 351
Basci, N., see Onur, M.A. **88**, 313
Bashal, A., see Payne, R.S. **145**, 165
Bashir, A.K., see Tanira, M.O.M. **102**, 267
Basly, J.P., M. Bernard, Radiosterilization dosimetry by ESR spectroscopy: Ritodrine hydrochloride and comparison with other sympathomimetics **149**, 85
Basly, J.P., J.L. Duroux, M. Bernard, Gamma radiation induced effects on metronidazole **139**, 219
Gamma irradiation sterilisation of orciprenaline and fenoterol **142**, 125
Radiosterilization dosimetry by ESR spectroscopy: Application to terbutaline **142**, 247
Basly, J.P., see Fauconnet, A.L. **144**, 123
Bason, A.M., see Alpar, H.O. **62**, 133
Bassam, F., P. York, R.C. Rowe, R.J. Roberts, Young's modulus of powders used as pharmaceutical excipients **64**, 55
Bassam, F., see York, P. **66**, 143
Bassani, V., J. Casadebaig, M. Jacob, C. Menut, I. Lechat, G. Lamaty, Preparation of a low-alcohol extract of *Rosmarinus officinalis* using a reverse osmosis membrane **63**, 57
Bassani, V., see Bassani, V. **63**, 57
Bassani, V., see Doko, M.B. **76**, 199
Basseleur, S.W.J., see Duchateau, G.S.M.J.E. **39**, 87
Bassignot, P., see Makki, S. **133**, 245
Bassini, V.L., see Petrovick, P.D. **76**, 49
Bastami, S.M., M.J. Groves, Some factors influencing the in vitro release of phenytoin from formulations **1**, 151
Bastide, M., see Mazuel, G. **16**, 97
Bastide, P., see Chopineau, J. **73**, 191
Basu, K., D.O. Kildsig, A.K. Mitra, Synthesis and kinetic stability studies of progesterone derivatives **47**, 195
Basu, K., A.K. Mitra, Effects of 3-hydrazone modification on the metabolism and protein binding of progesterone **65**, 109
Baswell, B., see Pollack, G.M. **21**, 3
Baszkin, A., see Klang, S.H. **132**, 33
Baszkin, A., see Weingarten, C. **75**, 171
Bataille, B., see Delalonde, M. **146**, 159; **130**, 147
Bataille, B., see Sonanglio, D. **115**, 53
Bateman, N.E., see D'Onofrio, G.P. **2**, 91
Bateman, S.D., M.H. Rubinstein, R.C. Rowe, R.J. Roberts, P. Drew, A.Y.K. Ho, A comparative investigation of compression simulators **49**, 209
Bateman, S.D., see Bateman, S.D. **49**, 209
Bauer-Brandl, A., Polymorphic transitions of cimetidine during manufacture of solid dosage forms **140**, 195
Erratum: Polymorphic transition of cimetidine during manufacture of solid dosage forms (Int. J. Pharm. 140 (1996) 195) **145**, 253
Bausch, A., H. Leuenberger, Wet spherical agglomeration of proteins as a new method to prepare parenteral fast soluble dosage forms **101**, 63
Baveja, S.K., S.S. Davis, K.P. Devi, Zero-order release hydrophilic matrix tablets of β-adrenergic blockers **39**, 39
Baveja, S.K., K.V. Ranga Rao, Sustained release tablet formulation of centperazine **31**, 169
Baveja, S.K., K.V. Ranga Rao, Devi K. Padmalatha, Sustained release tablet formulation of diethylcarbamazine. III **27**, 157
Baveja, S.K., K.V. Ranga Rao, R. Kumar, Devi K. Padmalatha, Sustained release tablet formulation of diethylcarbamazine. II **24**, 355
Baveja, S.K., K.V. Ranga Rao, A. Singh, Sustained release tablet formulation of diethylcarbamazine **19**, 229
Baveja, S.K., K.V.R. Rao, A. Singh, V.K. Gombar, Release characteristics of some bronchodilators from compressed hydrophilic polymeric matrices and their correlation with molecular geometry **41**, 55
Baveja, S.K., K.V.R. Rao, K.P. Devi, Relationship between gum content and half-life of soluble β-blockers from hydrophilic matrix tablets **47**, 133
Bawarshi-Nassar, R., see Truelove, J. **19**, 17
Bawazir, S.A., see Al-Hassan, M.I. **50**, 35; **93**, 7
Baykara, T., see Yuksel, N. **140**, 145
Baylac, G., see Delalonde, M. **130**, 147; **146**, 159
Bayley, D.L., see Steward, A. **104**, 145
Baylor, R.B., see Richards, R.M.E. **92**, 237
Bayomi, M.A., see Al-Angary, A.A. **114**, 221; **128**, 163
Bayon, L.G., see Cremers, H.F.M. **120**, 51
Bayoumi, S.M., J.J. Vallner, J.T. DiPiro, Quantitation of cefazolin sodium in plasma and tissues by high-performance liquid chromatography **30**, 57
Bazin, J.E., see Levadoux, E. **127**, 255
Beall, H.D., J.J. Getz, K.B. Sloan, The estimation of relative water solubility for prodrugs that are unstable in water **93**, 37
Beall, H.D., K.B. Sloan, Transdermal delivery of 5-fluorouracil (5-FU) by 1-alkylcarbonyl-5-FU prodrugs **129**, 203
Beall, H.D., see Sloan, K.B. **73**, 97; **93**, 27
Beall, H.R., Prankerd, K. Sloan, Transdermal delivery of 5-fluorouracil (5-FU) through hairless mouse skin by 1-alkyloxycarbonyl-5-FU prodrugs: Physicochemical characterization of prodrugs and correlations with transdermal delivery **111**, 223
Beastall, J.C., J. Hadgraft, C. Washington, Mechanism of action of Azone as a percutaneous penetration enhancer: Lipid bilayer fluidity and transition temperature effects **43**, 207
Beccaro, M., see Rochira, M. **144**, 19
Bechard, S., J.N. McMullen, Pectin-gelatin microglobules: Effect of a cross-linking agent (formaldehyde) on in vitro dissolution rate **31**, 91

Diffusional release of a solute from a porous polymeric matrix **32**, 71
Bechard, S.R., L. Levy, S.D. Clas, Thermal, mechanical and functional properties of cellulose acetate phthalate **114**, 205
Bechard, S.R., O. Quraishi, E. Kwong, Film coating: Effect of titanium dioxide concentration and film thickness on the photostability of nifedipine **87**, 133
Bechgaard, E., L. Jorgensen, R. Larsen, S. Gizurarson, J. Carstensen, A. Hvass, Insulin and didecanoyl-L-α-phosphatidylcholine: In vitro study of the transport through rabbit nasal mucosal tissue **89**, 147
Bechgaard, E., S. Gizurarson, R.K. Hjortkjaer, A.R. Sorensen, Intranasal administration of insulin to rabbits using glycofurol as an absorption promoter **128**, 287
Bechgaard, E., S. Gizurarson, L. Jorgensen, R. Larsen, The viability of isolated rabbit nasal mucosa in the Ussing chamber, and the permeability of insulin across the membrane **87**, 125
Bechgaard, E., K.J. Riis, L. Jorgensen, The development of an Ussing chamber technique for isolated human vaginal mucosa, and the viability of the in vitro system **106**, 237
Bechgaard, E., see Bindseil, E. **119**, 37
Bechgaard, E., see Gizurarson, S. **65**, 243
Bechgaard, E., see Jorgensen, L. **95**, 209; **99**, 165; **107**, 231; **113**, 83
Becirevic-Lacan, M., see Filipovic-Grcic, J. **135**, 183
Beck, M.I., I. Tomka, E. Waysek, Physico-chemical characterization of zein as a film coating polymer. A direct comparison with ethyl cellulose **141**, 137
Becker, A., see Gu, L. **41**, 95
Becker, A.R., see Powell, M.F. **35**, 61; **44**, 225
Beckert, T.E., K. Lehmann, P.C. Schmidt, Compression of enteric-coated pellets to disintegrating tablets **143**, 13
Beckett, A.H., W.A. Bahrendt, B.W. Hadzija, Some aspects on the bioavailability of a controlled release clonidine formulation in man **52**, 37
Beckett, A.H., see Hadzija, B.W. **67**, 185
Beckett, A.H., see Kyroudis, A. **43**, 145
Bedmar, M.C., A. Cerezo, P. Aznarte, R. Briz, L. Garcia, P.J. Hernandez, Hydrophobicity parametrization of progesterone and corticosteroid derivatives by high performance liquid chromatographic technique. I. Linear case **138**, 95
Bedmar, M.C., see Bedmar, M.C. **138**, 95
Beesley, J., see Lashmar, U.T. **91**, 59
Beezer, A.E., G.G. Fox, C.A. Gooch, W.H. Hunter, R.J. Miles, B.V. Smith, Microcalorimetric studies of the interaction of *m*-hydroxybenzoates with *E. coli* and with *S. aureus*. Demonstration of a Collander relationship for biological response **45**, 153
Beezer, A.E., J. Karlsen, W.H. Hunter, M.C.P. Lima, B.V. Smith, Quantitative structure-activity relationships: A group additivity scheme for biological response of *E. coli* to the action of *o*-, *m*- and *p*-alkoxyphenol **38**, 251
Beezer, A.E., P.L.O. Volpe, C.A. Gooch et al., Quantitative structure-activity relationships: Microcalorimetric determination of a group additivity scheme for biological response **29**, 237
Beezer, A.E., see Ashby, L.J. **51**, 245
Beezer, A.E., see Blair, T.C. **63**, 251
Beezer, A.E., see Buckton, G. **41**, 139; **47**, 121; **56**, 151; **72**, 181; **73**, 1; **82**, R7; **110**, 179
Beezer, A.E., see De, Morais S.M **66**, 107
Beezer, A.E., see Forster, S. **72**, 29
Beezer, A.E., see Montanari, M.L.C. **85**, 199
Beezer, A.E., see Willson, R.J. **132**, 45
Begtrup, M., see Fredholt, K. **123**, 209
Begtrup, M., see Thomsen, K.F. **112**, 143
Begue, J.M., see Rolland, A. **53**, 67
Behari, J.R., G. Gregoriadis, A comparative study of free and liposome-entrapped diethylenetriaminepentaacetic acid used in the treatment of mice loaded with cadmium **79**, 213
Behl, C.R., see Park, J.Y. **2**, 215
Behl, C.R., see Seta, Y. **81**, 89
Behl, C.R., see Shah, V.P. **82**, 21
Beier, W., see Jonkman, J.H.G. **43**, 139
Beihn, R.M., see Jay, M. **14**, 343
Beihn, R.M., see Parr, A. **32**, 251
Beijen, J.H., see Van der Houwen, O.A.G.J. **99**, 73
Beijersbergen Van Henegouwen, G.M.J, H. Van de Zijde, Van de Griend, H. de Vries, Photochemical decomposition of diphenhydramine in water **35**, 259
Beijersbergen Van Henegouwen, G.M.J, see Busker, R.W. **36**, 113
Beijersbergen Van Henegouwen, G.M.J, see Cornelissen, P.J.G. **3**, 205
Beijersbergen Van Henegouwen, G.M.J., see de Vries, H. **20**, 265
Beijersbergen Van Henegouwen, G.M.J, see Sedee, A.G.J. **15**, 149
Beijnen, J.H., R.H. Fokkens, H. Rosing, W.J.M. Underberg, Degradation of mitomycin C in acid phosphate and acetate buffer solutions **32**, 111
Beijnen, J.H., J.J.M. Holthuis, H.G. Kerkdijk, O.A.G.J. Van der Houwen, A.C.A. Paalman, A. Bult, W.J.M. Underberg, Degradation kinetics of etoposide in aqueous solution **41**, 169
Beijnen, J.H., R.P. Potman, R.D Van Ooijen et al., Structure elucidation and characterization of daunorubicin degradation products **34**, 247
Beijnen, J.H., W.J.M. Underberg, Degradation of mitomycin C in acidic solution **24**, 219
Beijnen, J.H., O.A.G.J. Van der Houwen, W.J.M. Underberg, Aspects of the degradation kinetics of doxorubicin in aqueous solution **32**, 123
Beijnen, J.H., O.A.G.J. Van der Houwen, M.C.H. Voskuilen, W.J.M. Underberg, Aspects of the degradation kinetics of daunorubicin in aqueous solution **31**, 75
Beijnen, J.H., see Bekers, O. **52**, 239; **56**, 103; **68**, 271; **72**, 123
Beijnen, J.H., see de Vries, J.D. **100**, 181
Beijnen, J.H., see Suenaga, A. **59**, 121; **82**, 29
Beijnen, J.H., see Underberg, W.J.M. **50**, 175
Beijnen, J.H., see Van der Houwen, O.A.G.J. **67**, 155; **89**, R5; **105**, 249; **109**, 191; **112**, 97
Beijnen, J.H., see Vendrig, D.E.M.M. **43**, 131; **50**, 189

Bekers, O., J.H. Beijnen, Bramel E.H Groot, M. Otagiri, A. Bult, W.J.M. Underberg, Stabilization of mitomycins on complexation with cyclodextrins in aqueous acidic media **52**, 239
Bekers, O., J.H. Beijnen, Y.A.G. Kempers, A. Bult, W.J.M. Underberg, Effects of cyclodextrins on *N*-trifluoroacetyldoxorubicin-14-valerate (AD-32) stability and solubility in aqueous media **68**, 271
Bekers, O., J.H. Beijnen, G. Storm, A. Bult, W.J.M. Underberg, Chemical stability of *N*-trifluoroacetyldoxorubicin-14-valerate (AD-32) in aqueous media and after liposome encapsulation **56**, 103
Bekers, O., J.H. Beijnen, B.J. Vis, A. Suenaga, M. Otagiri, A. Bult, W.J.M. Underberg, Effect of cyclodextrin complexation on the chemical stability of doxorubin and daunorubicin in aqueous solutions **72**, 123
Bekers, O., see Suenaga, A. **59**, 121; **82**, 29
Bekers, O., see Underberg, W.J.M. **50**, 175
Bekers, O., see Van der Houwen, O.A.G.J. **67**, 155; **89**, R5; **99**, 73; **105**, 249; **112**, 97
Beksac, S., see Onur, M.A. **88**, 313
Belaid, K.A., see Irwin, W.J. **48**, 159
Belbella, A., C. Vauthier, H. Fessi, J.P. Devissauguet, F. Puisieux, In vitro degradation of nanospheres from poly(D,L-lactides) of different molecular weights and polydispersis **129**, 95
Bell, A.E., see Martin, G.P. **44**, 57
Bell, C.L., see am, Ende M.T. **120**, 33
Bell, C.L., N.A. Peppas, An apparatus to measure polymer swelling under load **134**, 167
Bell, G.D., see White, D.A. **40**, 151
Bell, J., see Grant, D.J.W. **14**, 251
Bellemare, M., see Juhasz, J. **77**, 309
Belloto, Jr. R.J., A.M. Dean, M.A. Moustafa et al., Statistical techniques applied to solubility predictions and pharmaceutical formulations: an approach to problem solving using mixture response surface methodology **23**, 195
Bellotti, A., see Catellani, P.L. **51**, 63
Belt, R.J., see Patton, T.F. **10**, 77
Beltrami, V., see Gander, B. **58**, 63
Ben-Yossef, R., see Touitou, E. **70**, 159
Benaji, B., T. Dine, F. Goudaliez, M. Luyckx, C. Brunet, M.L. Mallevais, B. Gressier-Cazin, M. Cazin J.C., Compatibility study of methotrexate with PVC bags after repackaging into two types of infusion admixtures **105**, 83
Benameur, H., G. De Gand, R. Brasseur, J.P Van Vooren, F.J. Legros, Liposome-incorporated dexamethasone palmitate: Chemical and physical properties **89**, 157
Bendas, B., U. Schmalfuss, R. Neubert, Influence of propylene glycol as cosolvent on mechanisms of drug transport from hydrogels **116**, 19
Bendas, C., see Neubert, R. **75**, 89
Bendas, G., see Engel, A. **107**, 99
Benedetti, A., see Stella, V.J. **71**, 157
Benedetti, L., see Ghezzo, E. **87**, 21
Benedetti, L., see Hume, L.R. **111**, 295
Benedetti, L., see Kyyronen, K. **80**, 161
Benedetti, L., see Richardson, J.L. **115**, 9
Benedetti, L., see Sanzgiri, Y.D. **107**, 91
Benet, L.Z., see Gumbleton, M. **71**, 175
Benet, L.Z., see Noonan, P.K. **12**, 331
Benet, L.Z., see Peck, C.C. **82**, 9
Benet, L.Z., see Skelly, J.P. **63**, 83
Benghalem, A., see Sebert, P. **110**, 285; **122**, 129
Benhamou, D., see Bonhomme, L. **84**, 33
Benita, S., M. Donbrow, Dissolution rate control of the release kinetics of water-soluble compounds from ethyl cellulose film-type microcapsules **12**, 251
Benita, S., P.H. Dor, M. Aronhime, G. Marom, Permeability and mechanical properties of a new polymer: Cellulose hydrogen phthalate **33**, 71
Benita, S., D. Friedman, M. Weinstock, Physostigmine emulsion: A new injectable controlled release delivery system **30**, 47
Benita, S., see Fessi, H. **55**, R1
Benita, S., see Gross, S.T. **29**, 213
Benita, S., see Klang, S.H. **132**, 33
Benita, S., see Levy, M.Y. **54**, 103; **66**, 29
Benita, S., see Magenheim, B. **94**, 115
Benita, S., see Pathak, Y.V. **65**, 169
Benita, S., see Santos Magalhaes, N.S. **76**, 225
Benita, S., see Weingarten, C. **75**, 171
Benita, S., see Zeevi, A. **108**, 57
Benjamin, E.J., D.L. Conley, On-line HPLC method for clean-up and analysis of hydrocortisone and sulconazole nitrate in a cream **13**, 205
Benjamin, E.J., B.A. Firestone, J.A. Schneider, L.H. Lin, Stabilization of the dipropionate ester of DHPG, 9-[(1,3-dihydroxy-2-propoxy)-methyl]guanine against enzymatic hydrolysis using complexation **35**, 73
Benjamin, E.J., M. Lee, J. Tom et al., Stabilization of sulconazole nitrate in a topical powder formulation **14**, 209
Benjamin, E.J., see Shek, E. **11**, 257
Benkerrour, L., D. Duchene, F. Puisieux, J. Maccario, Granule and tablet formulae study by principal component analysis **19**, 27
Benkhelifa, S., see Benkhelifa, S. **145**, 115
Benkhelifa, S., M.O. Decroix, P. Arnaud, J.F. Huneau, D. Tome, J.C. Chaumeil, Transport of cephalosporins across artificial membranes and rabbit ileum **145**, 115
Bennett, C.E., J.G. Hardy, C.G. Wilson, The influence of posture on the gastric emptying of antacids **21**, 341
Bennett, S.L., see Akhter, S.A. **21**, 17
Benoist, C., see Regnault, C. **132**, 263
Benoit, E., P. Maincent, J. Bessiere, Applicability of dielectric measurements to the adsorption of drugs onto nanoparticles **84**, 283
Benoit, E.F., O.M. Prot, P.D. Maincent, J. Bessiere, Rapid quantitative determination of the sulphate groups of dextran sulphate **118**, 121
Benoit, J.P., F. Courteille, C. Thies, A physicochemical study of the morphology of progesterone-loaded poly (D,L-lactide) microspheres **29**, 95
Benoit, J.P., see Dubernet, C. **35**, 145; **64**, 99

Benoit, J.P., see Le, Ray A.M. **106**, 201
Benoit, J.P., see Mestiri, M. **89**, 229
Benoit, J.P., see Pech, B. **128**, 179
Benoit, J.P., see Pense, A.M. **81**, 111
Benoit, J.P., see Ruiz, J.M. **49**, 69
Benoit, J.P., see Zinutti, C. **133**, 97
Benson, H.A.E., J.C. McElnay, R. Harland, Phonophoresis of lignocaine and prilocaine from Emla cream **44**, 65
Bentele, V., U.E. Berg, J. Kreuter, Molecular weights of poly(methyl methacrylate) nanoparticles **13**, 109
Bentele, V., see El Egakey, M.A **13**, 349
Bentley, M.V.L.B., E.R.M. Kedor, R.F. Vianna, J.H. Collett, The influence of lecithin and urea on the in vitro permeation of hydrocortisone acetate through skin from hairless mouse **146**, 255
Berg, A., see Artursson, P. **52**, 183
Berg, U.E., see Bentele, V. **13**, 109
Berge, G., see Audran, M. **15**, 273; **20**, 129
Bergenstahl, B., see Sjostrom, B. **84**, 107; **88**, 53; **94**, 89
Berger, S., see Waser, P.G. **39**, 213
Berger, Y., see Boulenc, X. **123**, 13, 71
Berger, Y., see Caro, I. **116**, 147
Bergers, J.J., see Steerenberg, P.A. **40**, 51
Berges, L., see Chacon, M. **141**, 81
Berges, R., see Barbosa, J. **149**, 213
Bergougnoux, L., see Irache, J.M. **126**, 103
Bergren, M.S., An automated controlled atmosphere microbalance for the measurement of moisture sorption **103**, 103
Bergsma, J., see Te, Wierik G.H **134**, 27
Bergstrom, K., see Vidgren, M. **96**, 183
Bergum, J.S., see Desai, D.S. **110**, 249, 257
Bergum, J.S., see Patel, N.K. **110**, 203
Beris, H., see Johnston, T.P. **100**, 279
Berlo, J.A., see Ntawukuliyayo, J.D. **139**, 79
Bermejo, M.V., A.T. Perez-Varona, M.J. Segura-Bono, A. Martin-Villodre, J.M. Pla-Delfina, T.M. Garrigues, Compared effects of synthetic and natural bile acid surfactants on xenobiotic absorption. I. Studies with polysorbate and taurocholate in rat colon **69**, 221
Bermejo, M.V., see Bermejo, M.V. **69**, 221
Bermejo, M.V., see Garrigues, T.M. **64**, 127; **101**, 209
Bermejo, M.V., see Garrigues Pelufo, T.M **79**, 135
Bermejo, M.V., see Segura-Bono, M.J. **107**, 159
Bernabei, M.T., see Forni, F. **60**, 83
Bernad, M.J., see Veiga, M.D. **89**, 119; **93**, 215
Bernard, E., J.L. Dubois, J. Wepierre, Percutaneous absorption of a new antiandrogen included in liposomes or in solution **126**, 235
Bernard, M., see Basly, J.P. **139**, 219; **142**, 125, 247; **149**, 85
Bernard, M., see Fauconnet, A.L. **144**, 123
Bernatchez, S.F., C. Tabatabay, R. Gurny, Sodium hyaluronate as a vehicle for an improved tolerance of 5-fluorouracil administered subconjunctivally to rabbits **106**, 161
Berner, B., see Nangia, A. **99**, 67
Berner, B., see Sznitowska, M. **99**, 43; **123**, 41
Berner, B., see Wilding, I.R. **132**, 81
Berner, B., see Yuk, S.H. **77**, 221, 231
Berney, B.M., P.B. Deasy, Evaluation of carbopol 934 as a suspending agent for sulphadimidine suspensions **3**, 73
Bernkop-Schnurch, A., I. Apprich, Synthesis and evaluation of a modified mucoadhesive polymer protecting from α-chymotrypsinic degradation **146**, 247
Bernkop-Schnurch, A., K. Dundalek, Novel bioadhesive drug delivery system protecting (poly)peptides from gastric enzymatic degradation **138**, 75
Bernkop-Schnurch, A., see Valenta, C. **148**, 131
Beronius, P., see Sjoberg, H. **141**, 63
Bertaccini, A., see Gatti, R. **115**, 113
Bertelsen, F., F.N. Christensen, P. Holm, K. Jorgensen, Comparison of organic solvent-based ethylcellulose coatings on KCl crystals applied by top and bottom spraying in fluidized-bed equipment **111**, 117
Berthold, A., see Langer, K. **137**, 67
Bertoni, M., see Bonferoni, M.C. **117**, 41
Bertoni, M., see Ferrari, F. **109**, 115; **112**, 29; **117**, 49; **121**, 11; **136**, 71
Besnard, J.C., see Machet, L. **72**, R1
Bessiere, J., see Benoit, E. **84**, 283
Bessiere, J., see Benoit, E.F. **118**, 121
Betageri, G.V., L.S. Burrell, Stability of antibody-bearing liposomes containing dideoxyinosine triphosphate **98**, 149
Betageri, G.V., K.R. Makarla, Enhancement of dissolution of glyburide by solid dispersion and lyophilization techniques **126**, 155
Betageri, G.V., D.L. Parsons, Drug encapsulation and release from multilamellar and unilamellar liposomes **81**, 235
Betageri, G.V., J.A. Rogers, Thermodynamics of partitioning of -blockers in the *n*-octanol-buffer and liposome systems **36**, 165
Beten, D.B., Gansbeke B Van, A. Schoutens, A.J. Moes, Evaluation of the gastric behaviour of coevaporate particles under fasting and non-fasting conditions **123**, 145
Beten, D.B., M. Gelbcke, B. Diallo, A.J. Moes, Interaction between dipyridamole and Eudragit S **88**, 31
Beten, D.B., A.J. Moes, Controlled-release coevaporates of dipyridamole prepared with acrylic polymers **103**, 243
Bethke, H., HPLC-separation of codergocrine-mesylate (dihydroergotoxine-methanesulfonate) **6**, 87
Betteridge, R.F., A.G. Bosanquet, Automated stability determination of anticancer drugs incubated in cell culture medium **71**, 73
Betteridge, R.F., A.L. Culverwell, A.G. Bosanquet, Stability of tauromustine (TCNU) in aqueous solutions during preparation and storage Urea analogues in propylene glycol as penetration enhancers in human skin **56**, 37
Bettinetti, G.P., see Giordano, F. **62**, 153
Bettini, R., see Hariharan, D. **112**, 47
Beugre, T., see Bonhomme, L. **65**, R9; **84**, 33
Beutner, D., see Hadgraft, J. **73**, 125; **89**, R1
Bevan, B., see Clemessy, M. **101**, 219
Beyssac, E., see Aiache, J.M. **39**, 235
Beyssac, E., see Lootvoet, G. **85**, 113

Bhalla, H.L., M.R. Menon, N.G.S. Gopal et al., Radiation sterilization of polyethylene glycols **17**, 351
Bhardwaj, R., see Surendran, N. **135**, 81
Bhaskar, R., R.S.R. Murthy, B.D. Miglani, K. Viswanathan, Novel method to evaluate diffusion controlled release of drug from resinate **28**, 59
Bhat, P.G., D.R. Flanagan, M.D. Donovan, The limiting role of mucus in drug absorption: Drug permeation through mucus solution **126**, 179
Drug binding to gastric mucus glycoproteins **134**, 15
Bhatnagar, S., see Vyas, S.P. **69**, 5
Bhatt, P.P., M.S. Hanna, P. Szeptycki, T. Higuchi, Finite dose transport of drugs in liquid formulations through stratum corneum: Analytical solution to a diffusion model **50**, 197
Bhatt, P.P., J.H. Rytting, E.M. Topp, Influence of Azone^{R} and lauryl alcohol on the transport of acetaminophen and ibuprofen through shed snake skin **72**, 219
Bhattachar, S.N., J.H. Rytting, T. Itoh, T. Nishihata, The effects of complexation with hydrogenated phospholipid on the transport of salicylic acid, diclofenac and indomethacin across snake stratum corneum **79**, 263
Bhise, S.B., C.V.S. Subrahmanyam, A.K. Malhotra, R.C. Srivastava, Liquid membrane phenomenon in anpileptic drugs **24**, 297
Bhise, S.B., C.V.S. Subrahmanyam, R.C. Srivastava, Liquid membrane phenomenon in antihistamines **17**, 263
Liquid membrane phenomena in cimetidine, ranitidine and disodium cromoglycate **21**, 27
Bhise, S.B., C.V.S. Subrahmanyam, R.K. Sharma, R.C. Srivastava, Liquid membrane phenomena in antiarrhythmic action **28**, 145
Bialer, M., Estimation of pharmacokinetic parameters in extravascular multiple dose administration in the one-compartment open body model with equal absorption and elimination first-order rate constants **13**, 345
Bialer, M., M. Friedman, J. Dubrovsky, Effect of sustained release on the pharmacokinetics of valproic acid in the dog **20**, 53
Bialer, M., Z. Hussein, Y. Herishanu, Y. Melnik, An alternative method for calculating absorption and elimination rate constants in first-order processes: Application to valproic acid **16**, 285
Bialer, M., A. Rubinstein, J. Dubrovsky et al., A comparative pharmacokinetic study of valpromide and valproic acid after intravenous administration in humans **23**, 25
Bialer, M., see Hussein, Z. **37**, 97
Bialik, W., K.A. Walters, K.R. Brain, J. Hadgraft, Some factors affecting the in vitro penetration of ibuprofen through human skin **92**, 219
Bianchi, L.M., see Saettone, M.F. **126**, 79
Bibby, D.C., W.N. Charman, S.A. Charman, M.N. Iskander, C.J.H. Porter, Synthesis and evaluation of 5′ alkyl ester prodrugs of zidovudine for directed lymphatic delivery **144**, 61
Bidah, D., E.M. Ouriemchi, J.M. Vergnaud, Diffusional process of drug delivery from a dosage form with a Gelucire matrix **80**, 145
Bidah, D., J.M. Vergnaud, Kinetics of in vitro release of sodium salicylate dispersed in Gelucire **58**, 215
New oral dosage form with two polymers: Gelucire and Sumikagel **72**, 35
Dosage forms with a polymer matrix and a swelling polymer **77**, 81
Biddiscombe, M., see Zainudin, B.M.Z. **51**, 67
Biddiscombe, M.F., R. Melchor, V.H.F. Mak, R.J. Marriott, A.J. Taylor, M.D. Short, S.G. Spiro, The lung deposition of salbutamol, directly labelled with technetium-99m, delivered by pressurised metered dose and dry powder inhalers **91**, 111
Biddiscombe, M.F., see Biddiscombe, M.F. **91**, 111
Bidgood, A.M., see Musson, D.G. **99**, 85
Bieck, P.R., see Antonin, K.H. **130**, 33
Bilbao-Revoredo, O., see Sutinen, R. **57**, 155
Bilbruck, J., G.W. Hanlon, G.P. Martin, The effects of poly-HEMA coating on the adhesion on bacteria to polymer monofilaments **99**, 293
Bildet, J., see Fawaz, F. **132**, 271
Bildet, J., see Guyot, M. **123**, 53
Bilia, A., V. Carelli, Colo G. Di, E. Nannipieri, In vitro evaluation of a pH-sensitive hydrogel for control of GI drug delivery from silicone-based matrices **130**, 83
Billany, M.R., S. Denman, S. Jameel, J.K. Sugden, Topical antirheumatic agents as hydroxyl radical scavengers **124**, 279
Billany, M.R., K. Khatib, M. Gordon, J.K. Sugden, Alcohols and ethanolamines as hydroxyl radical scavengers **137**, 143
Billany, M.R., J. McMullan, C. Ranson, J.K. Sugden, Hydroxyl and hydroperoxy radical scavenging by copper(II) chelates related to non-steroidal anti-inflammatory drugs **121**, 107
Bindoni, M., see Panico, A.M. **98**, 19
Bindra, D.S., see Gorecka, B.A. **125**, 55
Bindra, D.S., see Safadi, M. **90**, 239
Bindschaedler, C., see Ibrahim, H. **77**, 211; **87**, 239
Bindseil, E., E. Bechgaard, L. Jorgensen, R. Larsen, Morphological examination of rabbit nasal mucosa after exposure to acetylsalicylic acid, glycofurol 75 and ephedrine **119**, 37
Binns, J.S., see Davies, M.C. **66**, 223
Birmancevic, M., see Sekulovic, D. **32**, 269
Bisrat, M., E.K. Anderberg, M.I. Barnett, C. Nystrom, Physicochemical aspects of drug release. XV. Investigation of diffusional transport in dissolution of suspended, sparingly soluble drugs **80**, 191
Bisrat, M., C. Nystrom, Physicochemical aspects of drug release. VIII. The relation between particle size and surface specific dissolution rate in agitated suspensions **47**, 223
Bisrat, M., C. Nystrom, P. Edman, Influence of dissolution rate of sparingly soluble drugs on corneal permeability in vitro **63**, 49
Bisrat, M., see Anderberg, E.K. **47**, 67
Bittner, S., see Kararli, T.T. **102**, 177
Bitz, C., E. Doelker, Influence of the preparation method on residual solvents in biodegradable microspheres **131**, 171
Biviano, F., see Ghezzo, E. **87**, 21

Bjankova, R., see Okhamafe, A.O. **39**, 1
Bjerga Bjaen, A.K, K. Nord, S. Furuseth, T. Agren, H.H. Tonnesen, J. Karlsen, Polymorphism of chloroquine diphosphate **92**, 183
Bjerre, C., E. Bjork, O. Camber, Bioavailability of the sedative propiomazine after nasal administration in rats **144**, 217
Bjerre, C., see Gren, T. **141**, 53
Bjork, E., S. Bjurstrom, P. Edman, Morphologic examination of rabbit nasal mucosa after nasal administration of degradable starch microspheres **75**, 73
Bjork, E., P. Edman, Degradable starch microspheres as a nasal delivery system for insulin **47**, 233
Characterization of degradable starch microspheres as a nasal delivery system for drugs **62**, 187
Bjork, E., see Bjerre, C. **144**, 217
Bjork, E., see Edman, P. **34**, 225
Bjornsdottir, S., see Loftsson, T. **57**, 63
Bjurstrom, S., see Bjork, E. **75**, 73
Bjurstrom, S., see Pereswetoff-Morath, L. **128**, 9
Blaauwgeers, H.J.A., see Sedee, A.G.J. **15**, 149
Black, M., see Vidgren, M. **115**, 209
Blackburn, H.D., see Roberts, M.S. **2**, 295
Blackett, P.M., G. Buckton, A microcalorimetric study of surfactant aggregation and surfactant–drug interaction in a model inhalation aerosol system **125**, 133
Blackhall, N.W., see Jenkins, P.G. **102**, 261
Blackie, R.G.G., see Azmin, M.N. **10**, 109
Blair, T.C., G. Buckton, A.E. Beezer, S.F. Bloomfield, The interaction of various types of microcrystalline cellulose and starch with water **63**, 251
Blair, T.C., G. Buckton, S.F. Bloomfield, On the mechanism of kill of microbial contaminants during tablet compression **72**, 111
Blake, J.M., see Quigley, J.M. **54**, 155
Blanchard, J., D.F. Perry, P.D. Walson, Factors influencing the transfer of xanthines across the everted rat jejunum **18**, 259
Blanchard, J., see Rubino, J.T. **26**, 165
Blanchard, J., see Surendran, N. **135**, 81
Blanchard, J., see Ugwu, S.O. **102**, 193
Blanchon, S., G. Couarraze, F. Rieg-Falson, G. Cohen, F. Puisieux, Permeability of progesterone and a synthetic progestin through methacrylic films **72**, 1
Blanco-Medez, J., see Otero-Espinar, F. **70**, 35
Blanco-Prieto, M.J, see Blanco, Prieto M.J **111**, 137
Blanco-Prieto, M.J, F. Delie, E. Fattal, A. Tartar, F. Puisieux, A. Gulik, P. Couvreur, Characterization of V3 BRU peptide-loaded small PLGA microspheres prepared by a (w_1/o)w_2 emulsion solvent evaporation method **111**, 137
Blanco-Fuente, H., S. Anguiano-Igea, F.J. Otero-Espinar, J. Blanco-Mendez, In-vitro bioadhesion of carbopol hydrogels **142**, 169
Blanco-Fuente, H., B. Vila-Dorrio, S. Anguiano-Igea, F.J. Otero-Espinar, J. Blanco-Mendez, Tanned leather: A good model for determining hydrogels bioadhesion **138**, 103
Blanco-Fuente, H., see Garcia-Gonzalez, N. **100**, 25, 65; **104**, 107
Blanco-Mendez, J., see Anguiano-Igea, S. **135**, 161
Blanco-Mendez, J., see Blanco-Fuente, H. **138**, 103; **142**, 169
Blanco-Mendez, J., see Ganza-Gonzalez, A. **106**, 179
Blanco-Mendez, J., see Garcia-Gonzalez, N. **100**, 25, 65; **104**, 107
Blanco-Mendez, J., see Otero-Espinar, F.J. **75**, 37; **79**, 149
Blanford, M.F., see Wu, C.C. **8**, 323
Blanque, D., H. Sternagel, F. Podczeck, J.M. Newton, Some factors influencing the formation and in vitro drug release from matrix pellets prepared by extrusion/spheronization **119**, 203
Bleich, J., P. Kleinebudde, B.W. Muller, Influence of gas density and pressure on microparticles produced with the ASES process **106**, 77
Bleich, J., B.W. Muller, W. Wassmus, Aerosol solvent extraction system—a new microparticle production technique **97**, 111
Block, E.M., see Antal, E.J. **54**, 33
Blok, D., J.W. Arndt, Haan F.H.N De, P. Vermeij, H.E. Junginger, E.K.J. Pauwels, Scintigraphic investigation of the gastric emptying of 3 mm pellets in human volunteers **73**, 171
Blok, D., see Blok, D. **73**, 171
Blom-Roosemalen, M.C.M., see Noach, A.B.J. **90**, 229
Bloom, E., see Nangia, A. **99**, 67
Bloomfield, S.F., see Blair, T.C. **63**, 251; **72**, 111
Bloomfield, S.F., see Darwish, R.M. **119**, 183; **147**, 51
Bloor, J.R., see Wilding, I.R. **83**, 155
Blume, A., M. Jansen, M. Ghyczy, J. Gareiss, Interaction of phospholipid liposomes with lipid model mixtures for stratum corneum lipids **99**, 219
Bluhm, D.P., R.S. Summers, M.M.J. Lowes, H.H. Durrheim, Lipid emulsion content and vitamin A stability in TPN admixtures **68**, 277
Influence of container on vitamin A stability in TPN admixtures **68**, 281
Blunk, T., see Muller, R.H. **84**, 1
Bocci, V., F. Corradeschi, A. Naldini, E. Lencioni, Enteric absorption of human interferons α and β in the rat **34**, 111
Bocci, V., M. Muscettola, A. Naldini, The lymphatic route. IV. Pharmacokinetics of human recombinant interferon α_2 and natural interferon-β administered intradermally in rabbits **32**, 103
Bocci, V., A. Naldini, F. Corradeschi, E. Lencioni, Colorectal administration of human interferon-α **24**, 109
Bodde, H.E., M.A.M. Kruithof, J. Brussee, H.K. Koerten, Visualization of normal and enhanced $HgCl_2$ transport through human skin in vitro **53**, 13
Bodde, H.E., J.G.H. Joosten, A mathematical model for drug release from a two-phase system to a perfect sink **26**, 57
Bodde, H.E., see Bouwstra, J.A. **52**, 47
Bodde, H.E., see de Haan, F.H.N **56**, 75
Bodde, H.E., see de Vries, M. **76**, 25
Bodde, H.E., see Le Brun, P.P.H **49**, 141
Bodde, H.E., see Tiemessen, H.L.G.M. **53**, 119; **56**, 87
Bodmeier, R., J.W. McGinity, Solvent selection in the preparation of poly(DL-lactide) microspheres prepared by the solvent evaporation method **43**, 179

Bodmeier, R., K.H. Oh, H. Chen, The effect of the addition of low molecular weight poly(DL-lactide) on drug release from biodegradable poly(DL-lactide) drug delivery systems **51**, 1
Bodmeier, R., O. Paeratakul, Propranolol HCl release from acrylic films prepared from aqueous latexes **59**, 197
Process and formulation variables affecting the drug release from chlorpheniramine maleate-loaded beads coated with commercial and self-prepared aqueous ethyl cellulose pseudolatexes **70**, 59
Dry and wet strengths of polymeric films prepared from an aqueous colloidal polymer dispersion, Eudragit RS30D **96**, 129
The distribution of plasticizers between aqueous and polymer phases in aqueous colloidal polymer dispersions **103**, 47
Bodmeier, R., see Chang, C.M. **130**, 187; **147**, 135
Bodmeier, R., see Herrmann, J. **126**, 129
Bodmeier, R., see Remunan-Lopez, C. **135**, 63
Bodor, N., J.J. Kaminski, R.G. Roller, Improved delivery through biological membranes. VI. Potent sympathomimetic adrenalone derivatives **1**, 189
Bodor, N., J. McCornack, M.E. Brewster, Improved delivery through biological membranes. XXII. Synthesis and distribution of brain-selective estrogen delivery systems **35**, 47
Bodor, N., K.B. Sloan, Improved delivery through biological membranes. XII. The effect of the incorporation of biphasic solubilizing groups into prodrugs of steroids **15**, 235
Bodor, N., K.B. Sloan, R.J. Little et al., Soft drugs 4. 3-Spirothiazolidines of hydrocortisone and its derivatives **10**, 307
Bodor, N., V. Venkatraghavan, D. Winwood, K. Estes, M.E. Brewster, Improved delivery through biological membranes. XLI. Brain-enhanced delivery of chlorambucil **53**, 195
Bodor, N., J. Zupan, S. Selk, Improved delivery through biological membranes. VII. Dermal delivery of cromoglycic acid (cromolyn) via its prodrugs **7**, 63
Bodor, N., see Amselem, S. **125**, 31
Bodor, N., see Brewster, M.E. **59**, 231; **68**, 215; **75**, R5; **125**, 17
Bodor, N., see El-Koussi, A.A. **53**, 189
Bodor, N., see Estes, K.S. **65**, 101
Bodor, N., see Kaminski, J.J. **3**, 151
Bodor, N., see Kourounakis, A. **141**, 239
Bodor, N., see Loftsson, T. **57**, 63
Bodor, N., see Pop, E. **84**, 39
Bodor, N., see Sloan, K.B. **12**, 299
Bodor, N.S., see Dietzel, K. **59**, 225
Boersma, T., see Te, Wierik G.H. **134**, 27
Boeva, M., see Atcheva, M. **103**, 303
Bogardus, J.B., N.R. Palepu, Ionization and solubility of an amphoteric -lactam antibiotic **4**, 159
Bogardus, J.B., see Jay, M. **19**, 97
Bogdanov, B., see Panzova, B. **65**, 35; **70**, 187
Bogdanova, S., M. Rollet, E. Minkov, L. Rainova, Influence of polyvinylpyrrolidone on the in vitro diffusion characteristics of tetraminol **38**, 99
Bogdanova, S., see Dimitrova, E. **93**, 21
Bogner, R.H., see Wang, J.Z.Y. **113**, 113; **119**, 81
Bohm, A., see Keipert, S. **142**, 153
Bohner, V., see Luessen, H.L. **141**, 39
Boldyrev, V.V., see Shakhtshneider, T.P. **130**, 25
Bolhuis, B.G., see Moolenaar, F. **9**, 353
Bolhuis, G.K., see Proost, J.H. **13**, 287
Bolhuis, G.K., see Riepma, K.A. **66**, 47; **76**, 9; **85**, 121
Bolhuis, G.K., see Van der Voort Maarschalk, K. **140**, 185
Bolhuis, G.K., see Van Kamp, H.V. **28**, 229
Bolhuis, G.K., see Vromans, H. **35**, 29; **39**, 207
Bolhuis, G.K., see Waser, P.G. **39**, 213
Bolhuis, G.K., see Zuurman, K. **102**, 1; **119**, 65
Bolling, S.F., see Johnston, T.P. **52**, 139
Bolos, J., see Girona, V. **41**, 241
Bolton, B.A., see Resetarits, D.E. **2**, 113
Bolton, R., see de Morais, S.M **66**, 107
Bolton, S., see Khan, M.A. **102**, 185
Bona, M., see Burgot, G. **129**, 295
Bond, J.R., B.W. Barry, Damaging effect of acetone on the permeability barrier of hairless mouse skin compared with that of human skin **41**, 91
Bond, S.W., see Wilson, C.G. **38**, 221
Bondi, J.N., see Radebaugh, G.W. **45**, 39; **57**, 95
Bondi, J.V., see Matuszewska, B. **136**, 147
Bondi, M., see Iannuccelli, V. **143**, 187
Bonferoni, M.C., G. Mellerio, P. Giunchedi, C. Caramella, U. Conte, Photostability evaluation of nicardipine HCl solutions **80**, 109
Bonferoni, M.C., S. Rossi, F. Ferrari, M. Bertoni, C. Caramella, Influence of medium on dissolution-erosion behaviour of Na carboxymethylcellulose and on viscoelastic properties of gels **117**, 41
Bonferoni, M.C., see Ferrari, F. **117**, 49; **121**, 11; **136**, 71
Bonfils, A., see Fortier, M. **52**, 27
Bonfils, A., see Heyman, M. **37**, 33
Bonhomme, L., D. Benhamou, T. Beugre, Slow-release effect of pH-adjusted bupivacaine: In vitro demonstration **84**, 33
Bonhomme, L., B. Duleba, T. Beugre, G. Fredj, HPLC determination of the stability of retinoic acid in gel formulation **65**, R9
Bonhomme, L., M.C. Mathieu, C. Gosse, E. Depraetere, J. Renaux, G. Fredj, M. Seiller, Charcoal tattooing of mammary carcinoma in mice **80**, 271
Bonhomme, L., see Leomy, C. **115**, 235
Bonhomme-Faivre, L., B. Paule, S. Urien, E. Rudant, L. Bottius, D. Pradel, D. Marrot, Pharmacokinetics of all-trans retinoic acid (ATRA) in pants with renal cancer concomitantly treated with interferon-α 2A (IFN) **134**, 99
Bonhomme-Faivre, L., see Bonhomme-Faivre, L. **134**, 99
Bonina, F., M. Lanza, L. Montenegro, C. Puglisi, A. Tomaino, D. Trombetta, F. Castelli, A. Saija, Flavonoids as potential protective agents against photo-oxidative skin damage **145**, 87
Bonina, F., see Montenegro, L. **138**, 191
Bonina, F., see Saija, A. **124**, 1
Bonina, F.P., F. Castelli, L. Montenegro, G. Mazzone, Partitioning and differential scanning calorimetry studies of *N*-

alkyllactame ester dermal prodrugs of indomethacin **106**, 115
Bonina, F.P., V. Carelli, Colo G. Di, L. Montenegro, E. Nannipieri, Vehicle effects on in vitro skin permeation of and stratum corneum affinity for model drugs caffeine and testosterone **100**, 41
Bonina, F.P., L. Montenegro, Penetration enhancer effects on in vitro percutaneous absorption of heparin sodium salt **82**, 171
Vehicle effects on in vitro heparin release and skin penetration from different gels **102**, 19
Effects of some non-toxic penetration enhancers on in vitro heparin skin permeation from gel vehicles **111**, 191
Bonina, F.P., L. Montenegro, Capraris P De, E. Bousquet, S. Tirendi, 1-Alkylazacycloalkan-2-one esters as prodrugs of indomethacin for improved delivery through human skin **77**, 21
Bonina, F.P., L. Montenegro, F. Guerrera, Naproxen 1-alkylazacycloalkan-2-one esters as dermal prodrugs: In vitro evaluation **100**, 99
Bonina, F.P., L. Montenegro, G. Micali, D.P. West, P. Palicharla, R.L. Koch, In vitro percutaneous absorption evaluation of phenobarbital through hairless mouse, adult and premature human skin **98**, 93
Bonina, F.P., L. Montenegro, G. Trapani, M. Franco, G. Liso, In vitro evaluation of N-acyllactam esters of indomethacin as dermal prodrugs **124**, 45
Bonina, F.P., see Bonina, F.P. **98**, 93; **106**, 115
Bonina, F.P., see Montenegro, L. **133**, 89; **140**, 51
Bonini, F., see Fawaz, F. **132**, 271; **133**, 107
Bonini, F., see Guyot, M. **123**, 53
Bonjoch, J., see Munoz, M. **139**, 249
Bonn, R., see Hadgraft, J. **64**, 187
Bonner, F.J., see Quigley, J.M. **54**, 155
Bonny, J.D., see Leuenberger, H. **52**, 91; **115**, 217
Bontinck, A.M., see Kinget, R. **3**, 65
Booth, S.W., see Craig, D.Q.M. **114**, 103
Bootsma, H.P.R., H.W. Frijlink, A. Eissens, J.H. Proost, H. Van Doorne, C.F. Lerk, β-Cyclodextrin as an excipient in solid oral dosage forms: In vitro and in vivo evaluation of spray-dried diazepam-β-cyclodextrin products **51**, 213
Boraie, N.A., see Naggar, V.F.B. **28**, 239
Borchard, G., K.L. Audus, F. Shi, J. Kreuter, Uptake of surfactant-coated poly(methyl methacrylate)-nanoparticles by bovine brain microvessel endothelial cell monolayers **110**, 29
Borchardt, R.T., see Rim, S. **32**, 79
Borchardt, R.T., see Shah, P.K. **63**, 281; **68**, 285
Borg, K., see Hjartstam, J. **61**, 101
Borga, B., see Nicklasson, M. **69**, 255
Borgstrom, L., S. Newman, Total and regional lung deposition of terbutaline sulphate inhaled via a pressurised MDI or via Turbuhaler^R **97**, 47
Bories, C., see Rodrigues, Jr. J.M. **126**, 253
Borin, M.T., J.W. Ayres, Single dose bioavailability of acetaminophen following oral administration **54**, 199
Borjesson, E., see Jerwanska, E. **149**, 131
Borka, L., E. Shefter, The polymorphism of triamcinolone diacetate **16**, 93
Borras-Blasco, J., see Lopez, A. **139**, 205
Borrell, J.H., see Montero, M.T. **138**, 113
Borsadia, S., see Seta, Y. **81**, 89
Bortolotti, F., see Esposito, E. **129**, 263
Bos, C.E., Doorne H. Van, C.F. Lerk, Microbiological stability of tablets stored under tropical conditions **55**, 175
Bos, C.E., H. Vromans, C.F. Lerk, Lubricant sensitivity in relation to bulk density for granulations based on starch or cellulose **67**, 39
Bosanquet, A.G., Instability of solutions of Niaziquone stored at negative temperatures **47**, 215
Bosanquet, A.G., see Betteridge, R.F. **56**, 37; **71**, 73
Bosch, E.H., see Van Doorne, H. **73**, 43
Boston, V., see Wolfson, A.D. **51**, 183
Botha, S.A., D.R. Flanagan, Characterization of digoxin hydrates using thermal methods **82**, 185
Non-thermal methods in characterization of anhydrous digoxin and two digoxin hydrates **82**, 195
Botha, S.A., see Van Tonder, E.C. **63**, 35
Bottari, F., V. Carelli, G. Di Colo et al., A new method for determining the diffusion coefficient of drugs in semisolid vehicles from release data **2**, 63
Bottenberg, P., D. Slop, J.P. Remon, C. de Muynck, D. Coomans, Influence of viscosity and surfactant on fissure penetration of dental fluoride gels in vitro **49**, 147
Penetration of some commercially available fluoride gels into occlusal fissures in vitro **58**, 203
Bottius, L., see Bonhomme-Faivre, L. **134**, 99
Botzolakis, J.E., L.E. Small, L.L. Augsburger, Effect of disintegrants on drug dissolution from capsules filled on a dosator-type automatic capsule-filling machine **12**, 341
Bouche, R., see Fachaux, J.M. **99**, 99
Bouckaert, S., L. Vakaet, J.P. Remon, Influence of the buccal application site of a bioadhesive slow-release tablet on salivary miconazole concentrations in irradiated pants **130**, 257
Bouckaert, S., see Ntawukulilyayo, J.D. **91**, 23; **93**, 209
Boulenc, X., T. Breul, J.C. Gautier, P. Saudemon, H. Joyeux, C. Roques, Y. Berger, G. Fabre, Sodium lauryl sulphate increases tiludronate paracellular transport using human epithelial Caco-2 monolayers **123**, 71
Boulenc, X., C. Roques, H. Joyeux, Y. Berger, G. Fabre, Bisphosphonates increase tight junction permeability in the human intestinal epithelial (Caco-2) model **123**, 13
Boulenc, X., see Caro, I. **116**, 147
Bourassa, P.A., see Shanker, R.M. **119**, 149
Bourbouze, R., see Nimier, K. **105**, 235
Bourel, D., see Martin, R.J. **39**, 23
Bourgeois, D., see Davies, M.C. **66**, 223
Bourget, P., see Rieutord, A. **119**, 57
Bourget, P., A. Lesne-Hulin, A. Sertin, A. Maillot, M. Alaya, C. Martin, Fluorescence polarization immunoassay: Does it always represent a reliable method to monitor treatment with teicoplanin? Comparison with data obtained by high-performance liquid chromatography **146**, 167

Bourget, P., V. Quinquis-Desmaris, A. Hulin, J.D. Peter, F. Jehl, Study of the protein binding of fusidic acid in cholestatic individuals and comparison with in vitro findings **128**, 223
Bourne, D.W.A., see Kwon, K. **37**, 219
Bourny, E., see Sebert, P. **106**, 103; **110**, 285; **122**, 129
Bourret, E., L. Bardet, Mechanism of gelling of agarose sols. III. Rheologic behavior of various samples and energetics of their gelling process **12**, 19
Bourrie, M., see Caro, I. **116**, 147
Bousquet, E., see Bonina, F.P. **77**, 21
Bousquet, E., see Scalia, S. **115**, 249
Bousquet, E., see Ventura, C.A. **149**, 1
Boussard, P., M.J. Devleeschouwer, J. Dony, Prevention of the adhesion of Pseudomonas aeruginosa to human buccal epithelial cells **53**, 253
In vitro modification of antimicrobial efficacy by protamine **72**, 51
Boussard, P., M.J. Devleeschouwer, M. Otten, J. Dony, Influence of chloramphenicol on the sensitivity of Pseudomonas aeruginosa to chloroxylenol and crystal violet **32**, 91
Boussard, P., see Devleeschouwer, M.J. **32**, 55
Bousselet, M., see Jacolot, A. **128**, 283
Boutell, S.L., see Bains, D. **69**, 233
Bouwstra, J.A., G.S. Gooris, J. Brussee, M.A. Salomons-de Vries, W. Bras, The influence of alkyl-azones on the ordering of the lamellae in human stratum corneum **79**, 141
Bouwstra, J.A., G.S. Gooris, M.A. Salomons-de Vries, J.A. Van der Spek, W. Bras, Structure of human stratum corneum as a function of temperature and hydration: A wide-angle X-ray diffraction study **84**, 205
Bouwstra, J.A., L.J.C. Peschier, J. Brussee, H.E. Bodde, Effect of *N*-alkyl-azocycloheptan-2-ones including azone on the thermal behaviour of human stratum corneum **52**, 47
Bouwstra, J.A., M.A. Salomons-de Vries, B.A.I. den Bergh Van, G.S. Gooris, Changes in lipid organisation of the skin barrier by *N*-alkyl-azocycloheptanones: A visualisation and X-ray diffraction study **144**, 81
Bouwstra, J.A., see Buckton, G. **83**, 115
Bouwstra, J.A., see Cornwell, P.A. **127**, 9
Bouwstra, J.A., see Lehr, C.M. **78**, 43
Bouzon, J., see Armand, J.Y. **40**, 33
Bouzon, J., see Farah, N. **36**, 81
Bouzon, J., see Laghoueg-Derriche, N. **67**, 163
Bouzon, J., see Ouriemehi, E.M. **102**, 47; **113**, 231
Bove, E.L., see Johnston, T.P. **52**, 139
Bowen, J.C., see Alpar, H.O. **88**, 335
Bower, C., C. Washington, T.S. Purewal, Fractal morphology of drug aggregates in aerosol propellant suspensions **118**, 229
Bowtle, W., T. Woodage, A. Waugh, Bromhexine: In vitro and in vivo studies of release from mono- and bi-component preparations **9**, 305
Bowtle, W.J., M.J. Heasman, A.P. Prince, D.A. Easthope, Compatibility of the cephalosporin, cefamandole nafate with injections **4**, 263
Bowtle, W.J., see Burns, S.J. **141**, 9
Boxenbaum, H., J. Barnhill, Rates of pharmacokinetic literature obsolescence **21**, 135
Boxenbaum, H., see Khor, S.P. **23**, 109; **30**, 189
Boyd, J.A., see Johnston, T.P. **52**, 139; **59**, 95
Boymond, C., see Saleh, S.I. **45**, 19
Boza, A., see Alvarez-Fuentes, J. **148**, 219
Braghiroli, L., see Fabrizio, Saettone M. **142**, 103
Bragonje, J., see Shek, E. **11**, 257
Brain, J.D., see Niven, R.W. **104**, 73
Brain, K.R., J. Hadgraft, V.J. James, V.P. Shah, K.A. Walters, A.C. Watkinson, In vitro assessment of skin permeation from a transdermal system for the delivery of oestradiol **89**, R13
Brain, K.R., J. Hadgraft, D. Lewis, G. Allan, The influence of Azone^{R} on the percutaneous absorption of methotrexate **71**, R9
Brain, K.R., see Ashton, P. **87**, 261, 265
Brain, K.R., see Bialik, W. **92**, 219
Brain, K.R., see Brain, K.R. **89**, R13
Brain, K.R., see Heard, C.M. **90**, R5
Brain, K.R., see Watkinson, A.C. **121**, 27; **123**, 303
Bramanti, G., P. Mura, A. Liguori, L. Ceccarelli, G. Santoni, G. Grossi, M.C. Torresi, Pharmaceutical availability of digoxin tablets **49**, 241
Bramanti, G., see Bramanti, G. **49**, 241
Bramanti, G., see Mura, P. **119**, 71
Brand, R.M., G. Duensing, F.G. Hamel, Iontophoretic delivery of an insulin-mimetic peroxovanadium compound **146**, 115
Brandl, M., R. Strickley, T. Bregante, L. Gu, T.W. Chan, Degradation of 4′-azidothymidine in aqueous solution **93**, 75
Brandl, M., see Bachmann, D. **91**, 69
Brandl, M., see Schneider, T. **117**, 1; **132**, 9
Brannon-Peppas, L., Recent advances on the use of biodegradable microparticles and nanoparticles in controlled drug delivery **116**, 1
Brannon-Peppas, L., N.A. Peppas, Time-dependent response of ionic polymer networks to pH and ionic strength changes **70**, 53
Braquet, P., see Dowty, M.E. **68**, 231
Bras, W., see Bouwstra, J.A. **79**, 141; **84**, 205
Brasseur, N., D. Brault, P. Couvreur, Adsorption of hematoporphyrin onto polyalkylcyanoacrylate nanoparticles: Carrier capacity and drug release **70**, 129
Brasseur, R., see Benameur, H. **89**, 157
Bratty, J.R., C.M. Smith, D.J. McEntegart, A.W. Hubbard, A double-blind study to assess the primary irritancy of flurbiprofen (local action transcutaneous) when applied to the skin of Caucasians **115**, 119
Brault, D., see Brasseur, N. **70**, 129
Braun, M.A., R. Oschmann, P.C. Schmidt, Influence of excipients and storage humidity on the deposition of disodium cromoglycate (DSCG) in the Twin Impinger **135**, 53
Brauns, U., see Muller, B.W. **26**, 77
Braxton, B.K., see Schoen, M.P. **114**, 159
Bray, D., see Rowe, R.C. **101**, 169

Breas, R., see Poelma, F.G.J. **64**, 161
Bregante, T., see Brandl, M. **93**, 75
Breheny, C., see Edwardson, P.A.D. **91**, 51
Breimer, D.D., see Noach, A.B.J. **90**, 229
Brenner, G.S., see Ip, D.P. **28**, 183
Brenner, G.S., see Zografi, G. **18**, 99
Bretal, M.J., see Remunan, C. **80**, 151
Breteau, M., see Machet, L. **72**, R1
Breul, T., see Boulenc, X. **123**, 71
Brewster, M.E., M. Deyrup, K. Seyda, N. Bodor, Synthesis, characterization and in vitro evaluation of various sulfonamide chemical delivery systems **68**, 215
Brewster, M.E., K.S. Estes, N. Bodor, An intravenous toxicity study of 2-hydroxypropyl-β-cyclodextrin, a useful drug solubilizer, in rats and monkeys **59**, 231
Brewster, M.E., T. Loftsson, S. Amselem, D. Friedmand, A. Yogev, W.R. Anderson, D.O. Helton, A. Dinculescu N. Bodor, E. Pop, Formulation development for a zidovudine chemical delivery system 1. Parenteral dosage forms **125**, 17
Brewster, M.E., T. Loftsson, J. Baldvinsdottir, N. Bodor, Stabilization of aspartame by cyclodextrins **75**, R5
Brewster, M.E., T. Loftsson, K.S. Estes, J.L. Lin, H. Fridriksdottir, N. Bodor, Effect of various cyclodextrins on solution stability and dissolution rate of doxorubicin hydrochloride **79**, 289
Brewster, M.E., see Amselem, S. **125**, 31
Brewster, M.E., see Bodor, N. **35**, 47; **53**, 195
Brewster, M.E., see Brewster, M.E. **79**, 289; **125**, 17
Brewster, M.E., see Dietzel, K. **59**, 225
Brewster, M.E., see Estes, K.S. **65**, 101
Brewster, M.E., see Pop, E. **84**, 39; **140**, 33
Brickwell, J., see Steed, K.P. **123**, 291
Bridges, J.F., J.F. Woodley, R. Duncan, J. Kopecek, Soluble *N*-(2-hydroxypropyl)methacrylamide copolymers as a potential oral, controlled-release, drug delivery system. I. Bioadhesion to the rat intestine in vitro **44**, 213
Bridges, P.A., see Leung, K.K.M. **145**, 95
Bridges, P.A., see McCallion, O.N.M. **130**, 1
Briegel, P., see Pfaff, G. **14**, 173
Briggner, L.E., G. Buckton, K. Bystrom, P. Darcy, The use of isothermal microcalorimetry in the study of changes in crystallinity induced during the processing of powders **105**, 125
Briggs, C.J., see Washington, C. **48**, 133
Brine, G.A., see Rapaka, R.S. **12**, 285
Brion, F., see Jacolot, A. **128**, 283
Brion, F., see Postaire, E. **55**, 99
Briskin, J., see Qiu, Y. **143**, 179
Brittain, H.G., see Morris, K.R. **108**, 195
Britten, A.Z., see Gonda, I. **27**, 255
Britten, J.R., M.I. Barnett, Development and validation of a capsule filling machine simulator **71**, R5
Britten, J.R., see Pilpel, N. **70**, 241
Briz, R., see Bedmar, M.C. **138**, 95
Brochart, H., P. Ehret, P. Goldbach, A. Stamm, In vitro evaluation of a new controlled release veterinary device **91**, 95
Brochart, H., see Goldbach, P. **117**, 225
Brodin, A., see Fyhr, P. **40**, 157, 193
Brodin, A., see Nicklasson, M. **15**, 87; **18**, 149; **23**, 97
Brojo, A.P., see Pina, M.E. **133**, 139; **148**, 73
Bronaugh, R.L., see Storm, J.E. **65**, 265
Brondsted, H., H.M. Nielsen, L. Hovgaard, Drug delivery studies in Caco-2 monolayers. III. Intestinal transport of various vasopressin analogues in the presence of lysophosphatidylcholine **114**, 151
Brondsted, H., see Hovgaard, L. **114**, 141
Brooke, D., see Yang, W.H. **11**, 271
Brookes, L.G., see Gilbert, J.C. **32**, 223
Brooks, J.D., see Riviere, J.E. **127**, 213
Brophy, M.R., P.B. Deasy, In vitro and in vivo studies on biodegradable polyester microparticles containing sulphamethizole **29**, 223
Application of the Higuchi model for drug release from dispersed matrices to particles of general shape **37**, 41
Brossard, D., see Arnaud, P. **129**, 279
Brossard, D., see Cohen, T. **137**, 249
Brossard, F., see Zeevi, A. **108**, 57
Brown, K.F., see Ridd, M.J. **11**, 11
Brown, M.R.W., see Alpar, H.O. **88**, 335
Brown, P.W., see Allwood, M.C. **97**, 219
Bru, N., see Roy, D. **148**, 165
Bruneaux, F., see Satou, V. **107**, 111
Bruner, M.W., see Burke, C.J. **86**, 89
Brunet, C., see Benaji, B. **105**, 83
Brunet, C., see Dine, T. **77**, 279
Brunet, C., see Faouzi, M.A. **105**, 89
Brunet, C., see Gressier, B. **104**, 151
Brunet, C., see Khalfi, F. **139**, 243
Brunmark, A., see Stjarnkvist, P. **40**, 215
Brusee, J., see Hoogstraate, A.J. **76**, 37
Brussee, J., see Bodde, H.E. **53**, 13
Brussee, J., see Bouwstra, J.A. **52**, 47; **79**, 141
Bryan, A.J., R. Kaur, G. Robinson et al., Histological and physiological studies in the intestine of the rat exposed to solutions of Myrj 52 and PEG 2000 **7**, 145
Brzokewicz, A., see Rolland, A. **76**, 217
Bubb, M.D., see Davis, S.S. **1**, 303
Buchmann, M., see Huu-Phuoc, N. **34**, 217
Buchmann, M., see Ruelle, P. **87**, 47
Buckton, G., see Ahmed, H. **130**, 195
Buckton, G., see Ashby, L.J. **51**, 245
Buckton, G., see Blackett, P.M. **125**, 133
Buckton, G., see Blair, T.C. **63**, 251; **72**, 111
Buckton, G., see Briggner, L.E. **105**, 125
Buckton, G., see Buckton, G. **83**, 115
Buckton, G., see Dove, J.W. **138**, 199
Buckton, G., see Efentakis, M. **60**, 229; **70**, 153
Buckton, G., see Forster, S. **72**, 29
Buckton, G., see Mackellar, A.J. **112**, 65, 79
Buckton, G., see Mall, S. **131**, 41
Buckton, G., see Ochoa, Machiste E. **145**, 197
Buckton, G., see Odidi, I.O. **72**, 43
Buckton, G., see Papadimitriou, E. **98**, 57

Buckton, G., see Parsons, G.E. **82**, 145; **83**, 163
Buckton, G., see Pinto, J.F. **83**, 187; **118**, 95
Buckton, G., see Rillosi, M. **117**, 75
Buckton, G., see Sheridan, P.L. **109**, 155; **125**, 141
Buckton, G., see Tapia, C. **92**, 211
Buckton, G., see Young, S.A. **60**, 235
Buckton, G., see Zajic, L. **59**, 155
Buckton, G., The assessment, and pharmaceutical importance, of the solid/liquid and the solid/vapour interface: A review with respect to powders **44**, 1
The role of compensation analysis in the study of wettability, solubility, disintegration and dissolution **66**, 175
Buckton, G., J.K. Armstrong, B.Z. Chowdhry, S. Leharne, A.E. Beezer, The use of high sensitivity differential scanning calorimetry to characterise dilute aqueous dispersion of surfactants. 2. Further studies on polyoxyethylene alkyl ethers **110**, 179
Buckton, G., A.E. Beezer, A microcalorimetric study of powder surface energetics **41**, 139
The applications of microcalorimetry in the field of physical pharmacy **72**, 181
The relationship between particle size and solubility **82**, R7
Buckton, G., A.E. Beezer, S.M. Chatham, K.K. Patel, In vitro dissolution testing of oral controlled release preparations in the presence of artificial foodstuffs. II. Probing drug/food interactions using microcalorimetry **56**, 151
Buckton, G., A.E. Beezer, S.P. Denyer, S.J. Russell, Observations on the biopharmaceutical importance of chain length in chemically related compounds **73**, 1
Buckton, G., R. Bulpett, N. Verma, Surface analysis of pharmaceutical powders: X-ray photoelectron spectroscopy (XPS) related to powder wettability **72**, 157
Buckton, G., B. Chandaria, Consideration of adhesion to modified container walls, by use of surface energy and polarity data, and Lewis acid–Lewis base interactions **94**, 223
Buckton, G., A. Choularton, A.E. Beezer, S.M. Chatham, The effect of the comminution technique on the surface energy of a powder **47**, 121
Buckton, G., B.Z. Chowdhry, J.K. Armstrong, S.A. Leharne, J.A. Bouwstra, H.E.J. Hofland, The use of high-sensitivity differential scanning calorimetry to characterise dilute aqueous dispersions of surfactants **83**, 115
Buckton, G., D. Ganderton, R. Shah, In vitro dissolution of some commercially available sustained-release theophylline preparations **42**, 35
Buckton, G., M. Efentakis, The use of thermodynamic activation parameters and compensation analysis to model drug release from hydrophobic matrices **62**, 157
Buckton, G., M. Efentakis, H. Al-Hmoud, Z. Rajan, The influence of surfactants on drug release from acrylic matrices **74**, 169
Buckton, G., P. Darcy, The influence of additives on the recrystallisation of amorphous spray dried lactose **121**, 81
The use of gravimetric studies to assess the degree of crystallinity of predominantly crystalline powders **123**, 265
Water mobility in amorphous lactose below and close to the glass transition temperature **136**, 141
Buckton, G., P. Darcy, D. Greenleaf, P. Holbrook, The use of isothermal microcalorimetry in the study of changes in crystallinity of spray-dried salbutamol sulphate **116**, 113
The use of isothermal microcalorimetry in the study of changes in crystallinity of spray-dried salbutamol sulphate (Int. J. Pharm. 116 (1995) 113–118) **122**, 130
Buckton, G., P. Darcy, A.J. Mackellar, The use of isothermal microcalorimetry in the study of small degrees of amorphous content of powders **117**, 253
Bugay, D.E., see Morris, K.R. **108**, 195
Buist, H.P.A., E. Tomlinson, J.F.M. Kinkel, Effect of α-α charge-transfer complexation on distribution of some aromatics between water and 2,2,4-trimethylpentane **15**, 347
Buitendijk, H.H., see Van, der Veen J **87**, 161
Bulpett, R., see Buckton, G. **72**, 157
Bult, A., see Bekers, O. **52**, 239; **56**, 103; **68**, 271
Bult, A., see Leeuwenkamp, O.R. **24**, 27
Bult, A., see Van der Houwen, O.A.G.J. **67**, 155; **89**, R5; **99**, 73; **105**, 249; **109**, 191; **112**, 97
Bummer, P.M., An FTIR study of the structure of human serum albumin adsorbed to polysulfone **132**, 143
Bundgaard, H., A differential kinetic method for the determination of mecillinam in the presence of its hydrolysis and epimerization products **5**, 257
Acid-catalyzed hydrolysis of fosfomycin and its implication in oral absorption of the drug **6**, 1
Bundgaard, H., A. Buur, Prodrugs as drug delivery systems. 65. Hydrolysis of α-hydroxy- and α-acyloxy-*N*-benzoylglycine derivatives and implications for the design of prodrugs of NH-acidic compounds **37**, 185
Bundgaard, H., A. Buur, S.C. Chang, V.H.L. Lee, Prodrugs of timolol for improved ocular delivery: Synthesis, hydrolysis kinetics and lipophilicity of various timolol esters **33**, 15
Bundgaard, H., A. Buur, K.T. Hansen, J.D. Larsen, J. Moss, L. Olsen, Prodrugs as drug delivery systems. 77. Phthalidyl derivatives as prodrug forms for amides, sulfonamides, carbamates and other NH-acidic compounds **45**, 47
Bundgaard, H., A. Hoelgaard, B. Mollgaard, Leaching of hydrolytic enzymes from human skin in cutaneous permeation studies as determined with metronidazole and 5-fluorouracil pro-drugs **15**, 285
Bundgaard, H., C. Larsen, Kinetics and mechanism of the sucrose-accelerated degradation of penicillins in aqueous solution **1**, 95
Piperazinedione formation from reaction of ampicillin with carbohydrates and alcohols in aqueous solution **3**, 1
Pro-drugs as drug delivery systems. XVII. Esters of 4-hydroxybutyric acids as potential pro-drug types for γ-lactones **7**, 169
The influence of carbohydrates and polyhydric alcohols on the stability of cephalosporins in aqueous solution **16**, 319
Bundgaard, H., C. Larsen, E. Arnold, Prodrugs as drug delivery systems. XXVII. Chemical stability and bioavailability of a water-soluble prodrug of metronidazole for parenteral administration **18**, 79

Bundgaard, H., C. Larsen, P. Thorbek, Prodrugs as drug delivery systems. XXVI. Preparation and enzymatic hydrolysis of various water-soluble amino acid esters of metronidazole **18**, 67

Bundgaard, H., E. Falch, Allopurinol prodrugs. I. Synthesis, stability and physicochemical properties of various N_1-acyl allopurinol derivatives **23**, 223

Allopurinol prodrugs. II. Synthesis, hydrolysis kinetics and physicochemical properties of various *N*-acyloxymethyl allopurinol derivatives **24**, 307

Allopurinol prodrugs. III. Water-soluble *N*-acyloxymethyl allopurinol derivatives for rectal or parenteral use **25**, 27

Bundgaard, H., E. Falch, C. Larsen, Pro-drugs as drug delivery systems. XI. Preparation and characterisation of a novel water-soluble pro-drug type and barbituric acids **6**, 19

Bundgaard, H., E. Falch, S.B. Pedersen, G.H. Nielsen, Allopurinol prodrugs. IV. Improved rectal and parenteral delivery of allopurinol using the prodrug approach as evaluated in rabbits **27**, 71

Bundgaard, H., G.J. Friis, Prodrugs of peptides. 16. Isocyclosporin A as a potential prodrug of cyclosporin A **82**, 85

Bundgaard, H., E. Jensen, E. Falch, S.B. Pedersen, Allopurinol prodrugs. V. Water-soluble *N*-substituted (aminomethyl)benzoyloxymethyl allopurinol derivatives for parenteral or rectal delivery **64**, 75

Bundgaard, H., J. Hansen, Studies on the stability of corticosteroids. VI. Kinetics of the rearrangement of βmethasone-17-valerate to the 21-valerate ester in aqueous solution **7**, 197

Nucleophilic phosphate-catalyzed degradation of penicillins: Demonstration of a penicilloyl phosphate intermediate and transformation of ampicillin to a piperazinedione **9**, 273

Bundgaard, H., Hansen A. Bagger, C. Larsen, Pro-drugs as drug delivery systems. III. Esters of malonuric acids as novel pro-drug types for barbituric acids **3**, 341

Bundgaard, H., S.H. Hansen, Hydrolysis and epimerization kinetics of pilocarpine in basic aqueous solution as determined by HPLC **10**, 281

Bundgaard, H., M. Johansen, Pro-drugs as drug delivery systems. VIII. Bioreversible derivatization of hydantoins by *N*-hydroxymethylation **5**, 67

Pro-drugs as drug delivery systems. XV. Bioreversible derivatization of phenytoin, acetazolamide, Chloroxazone and various other NH-acidic compounds by *N*-aminomethylation to effect enhanced dissolution rates **7**, 129

Pro-drugs as drug delivery systems. XIX. Bioreversible derivatization of aromatic amines by formation of *N*-Mannich bases with succinimide **8**, 183

Hydrolysis of N-Mannich bases and its consequences for the biological testing of such agents **9**, 7

Pro-drugs as drug delivery systems. XX. Oxazolidines as potential pro-drug types for β-aminoalcohols, aldehydes or ketones **10**, 165

Hydrolysis of *N*-(α-hydroxybenzyl)benzamide and other *N*-(α-hydroxyalkyl) amide derivatives: Implications for the design of *N*-acyloxyalkyl-type prodrugs **22**, 45

Bundgaard, H., M. Johansen, V. Stella, M. Cortese, Pro-drugs as drug delivery systems. XXI. Preparation, physicochemical properties and bioavailability of a novel water-soluble prodrug type for carbamazepine **10**, 181

Bundgaard, H., N. Mork, Kinetics of the decarboxylation of foscarnet in acidic aqueous solution and its implication in its oral absorption **63**, 213

Bundgaard, H., N. Mork, A. Hoelgaard, Enhanced delivery of nalidixic acid through human skin via acyloxymethyl ester progrugs **55**, 91

Bundgaard, H., N.M. Nielsen, Glycolamide esters as a novel biolabile prodrug type for non-steroidal anti-inflammatory carboxylic acid drugs **43**, 101

Bundgaard, H., N.M. Nielsen, A. Buur, Aspirin prodrugs: Synthesis and hydrolysis of 2-acetoxybenzoate esters of various *N*-(hydroxyalkyl)amides **44**, 151

Bundgaard, H., T. Norgaard, N.M. Nielsen, Photodegradation and hydrolysis of furosemide and furosemide esters in aqueous solutions **42**, 217

Bundgaard, H., U. Klixbull, Hydrolysis of pivampicillin in buffer and plasma solutions. Formation of a 4-imidazolidinone from ampicillin and formaldehyde **27**, 175

Bundgaard, H., U. Klixbull, E. Falch, Prodrugs as drug delivery systems. 43. *O*-Acyloxymethyl salicylamide *N*-Mannich bases as double prodrug forms for amines **29**, 19

Prodrugs as drug delivery systems. 44. *O*-Acyloxymethyl, *O*-acyl and *N*-acyl salicylamide derivatives as possible prodrugs for salicylamide **30**, 111

Bundgaard, B., see Anderson, F.M. **20**, 155

Bundgaard, H., see Andersen, F.M. **19**, 189; **21**, 51

Bundgaard, H., see Buur, A. **18**, 325; **21**, 349; **23**, 209; **24**, 43; **36**, 41; **42**, 51

Bundgaard, H., see Drustrup, J. **47**, 103; **71**, 105

Bundgaard, H., see Duchateau, G.S.M.J.E. **39**, 87

Bundgaard, H., see Fredholt Thomsen, K. **91**, 39

Bundgaard, H., see Friis, G.J. **82**, 79

Bundgaard, H., see Hansen, J. **8**, 121; **81**, 253

Bundgaard, H., see Hansen, K.T. **79**, 205

Bundgaard, H., see Hansen, L.B. **84**, 253; **88**, 221, 229, 237, 243

Bundgaard, H., see Illum, L. **10**, 339; **17**, 183

Bundgaard, H., see Jensen, E. **58**, 143; **70**, 137; **71**, 117

Bundgaard, H., see Johansen, M. **7**, 119; **13**, 89

Bundgaard, H., see Kahns, A.H. **62**, 193; **71**, 31; **76**, 99; **77**, 65; **78**, 199

Bundgaard, H., see Klixbull, U. **20**, 273; **23**, 163

Bundgaard, H., see Kurono, Y. **80**, 135; **81**, 179

Bundgaard, H., see Larsen, J.D. **37**, 87; **51**, 27

Bundgaard, H., see Lenaerts, V. **39**, 121

Bundgaard, H., see Lokind, K.B. **76**, 177

Bundgaard, H., see Lundin, S. **76**, R1

Bundgaard, H., see Mollgaard, B. **12**, 153

Bundgaard, H., see Mork, N. **60**, 163

Bundgaard, H., see Moss, J. **52**, 255; **66**, 39, 183; **74**, 67; **82**, 91

Bundgaard, H., see Nielsen, L.S. **102**, 231

Bundgaard, H., see Nielsen, N.M. **29**, 9

Bundgaard, H., see Rasmussen, G.J. **71**, 45; **76**, 113
Bundgaard, H., see Steffansen, B. **56**, 159
Bundgaard, H., see Wassermann, K. **14**, 73
Bunge, A.L., see Watkinson, A.C. **83**, 171; **87**, 175
Bunjes, H., K. Westesen, M.H.J. Koch, Crystallization tendency and polymorphic transitions in triglyceride nanoparticles **129**, 159
Bunjes, H., see Westesen, K. **115**, 129
Bunt, C.R., D.S. Jones, I.G. Tucker, The effects of pH, ionic strength and organic phase on the bacterial adhesion to hydrocarbons (BATH) test **99**, 93
The effects of pH, ionic strength and polyvalent ions on the cell surface hydrophobicity of *Escherichia coli* evaluated by the BATH and HIC methods **113**, 257
Buonaguidi, M., V. Carelli, Colo G. Di, E. Nannipieri, M.F. Serafini, Evaluation of a pH-sensitive semi-interpenetrating polymer network for control of GI drug delivery **147**, 1
Burgalassi, S., see Saettone, M.F. **51**, 203
Burgalassi, S., L. Panichi, M.F. Saettone, J. Jacobsen, M.R. Rassing, Development and in vitro/in vivo testing of mucoadhesive buccal patches releasing benzydamine and lidocaine **133**, 1
Burger, A., see Griesser, U.J. **120**, 83
Burger, J.J., E. Tomlinson, E.M.A. Mulder, J.G. McVie, Albumin microspheres for intra-arterial tumour targeting. I. Pharmaceutical aspects **23**, 333
Burgess, D.J., J.E. Carless, Manufacture of gelatin/gelatin coacervate microcapsules **27**, 61
Microelectrophoretic behaviour of gelatin and acacia complex coacervates and indomethacin microcapsules **32**, 207
Burgess, D.J., C. Triqueneaux, E. Tomlinson, Potential use of albumin microspheres as a drug delivery system. I. Preparation and in vitro release of steroids **39**, 129
Burgot, G., see Cognez, P. **123**, 137
Burgot, G., M. Bona, M.O. Christen, J.L. Burgot, First relationships between detoxication properties of some 1,2-dithiole-3-thiones and their log *P* **129**, 295
Burgot, G., P. Serrand, J.L. Burgot, Thermodynamics of partitioning in the n-octanol/water system of some β-blockers **63**, 73
Burgot, G., A first estimation of the thermodynamic parameters of water/n-octanol partitioning of some tricyclic antidepressants: Similarities and differences with the values of other psychotropic drugs **94**, 135
Burgot, J.L., see Burgot, G. **63**, 73; **129**, 295
Burgot, J.L., see Cognez, P. **123**, 137
Burguillo, F.J., see Pedraz, J.L. **25**, 147
Buri, P., see Cornaz, A.L. **129**, 175
Buri, P., see Ganem-Quintanar, A. **147**, 165
Buri, P., see Ibrahim, H. **77**, 211; **87**, 239
Buri, P., see Meseguer, G. **95**, 229
Buri, P., see Nook, T. **43**, 119
Buri, P., see Ranga, Rao K.V. **52**, 265
Buri, P., see Rouge, N. **136**, 117
Buri, P.A., see Peppas, N.A. **20**, 107
Burke, C.J., B.L. Steadman, D.B. Volkin, P.K. Tsai, M.W. Bruner, C.R. Middaugh, The adsorption of proteins to pharmaceutical container surfaces **86**, 89
Burkman, A.M., see Sokoloski, T.D. **6**, 63
Burkman, A.M., see Wu, C.C. **8**, 323
Burnette, R.R., A Monte-Carlo model for the passive diffusion of drugs through the stratum corneum **22**, 89
Burnham, W.R., P.K. Hansrani, C.E. Knott et al., Stability of a fat emulsion based intravenous feeding mixture **13**, 9
Burnouf, T., see Flament, M.P. **148**, 93
Burns, S.J., D. Corness, G. Hay, S. Higginbottom, I. Whelan, D. Attwood, S.G. Barnwell, Development and validation of an in vitro dissolution method for a floating dosage form with biphasic release characteristics **121**, 37
An in vitro assessment of liquid-filled Capill^{R} potato starch capsules with biphasic release characteristics **134**, 223
Burns, S.J., S. Higginbottom, D. Corness, G. Hay, I. Whelan, D. Attwood, S.G. Barnwell, A study of enteric-coated liquid-filled hard gelatin capsules with biphasic release characteristics **110**, 291
Burns, S.J., S. Higginbottom, I. Whelan, W.J. Bowtle, E. Rosenberg, D. Corness, G. Hay, D. Attwood, S.G. Barnwell, Formulation strategies designed to maintain the biphasic release characteristics of liquid-filled capsules **141**, 9
Burns, S.J., see Barnwell, S.G. **128**, 145
Burns, S.J., see Higginbottom, S. **109**, 173
Burrell, L.S., see Betageri, G.V. **98**, 149
Burrows, R., J.H. Collett, D. Attwood, The release of drugs from monoglyceride-water liquid crystalline phases **111**, 283
Burt, C.D., see Moore, D.E. **14**, 133
Burt, H.M., A.G. Mitchell, Dissolution anisotropy in nickel sulfate α hexahydrate crystals **3**, 261
Burt, H.M., see Dordunoo, S.K. **133**, 191
Burt, H.M., see Zhang, X. **132**, 195; **137**, 199
Burton, P.S., see Cho, M.J. **52**, 79
Bury, M., J. Gerhards, W. Erni, Monitoring sedimentation processes by conductivity measurements **76**, 207
Bury, M., J. Gerhards, W. Erni, A. Stamm, Application of a new method based on conductivity measurements to determine the creaming stability of o/w emulsions **124**, 183
Bush, R.K., see Welling, P.G. **4**, 135
Bushong, J., see Small, D. **149**, 203
Busker, R.W., G.M.J. Beijersbergen Van Henegouwen, B.M.C. Kwee, J.H.M. Winkins, Photobinding of flunitrazepam and its major photo-decomposition product *N*-desmethylflunitrazepam **36**, 113
Busquets, M.A., see Alsina, M.A. **70**, 111
Busquets, M.A., see Colome, C. **90**, 59
Busquets, M.A., see Mestres, C. **76**, 145
Busquets, M.A., see Reig, F. **44**, 257
Busson, R., see Naidong, W. **96**, 13
Bustamante, P., M. Jimenez-Duran, B. Escalera, The E and C model for predicting the solubility of drugs in pure solvents **129**, 283
Bustamante, P., see Molpeceres, J. **130**, 75
Butula, I., see Kalcic, I. **136**, 31
Butula, I., see Maysinger, D. **63**, 149
Butula, I., see Zorc, B. **123**, 65
Buur, A., see Bundgaard, H. **33**, 15; **37**, 185; **44**, 151; **45**, 47

Buur, A., see Moss, J. **66**, 183
Buur, A., see Steffansen, B. **132**, 243
Buur, A., N. Larsen Chr., M. Rassing, K. Hermansen, Bioavailability of progesterone from rectal dosage forms in rabbits **16**, 41
Buur, A., H. Bundgaard, Prodrugs as drug delivery systems. XXVIII. Structural factors influencing the rate of hydrolysis of oxazolidines—a potential prodrug type **18**, 325
Prodrugs of 5-fluorouracil. I. Hydrolysis kinetics and physicochemical properties of various *N*-acyl derivatives of 5-fluorouracil **21**, 349
Prodrugs of 5-fluorouracil. III. Hydrolysis kinetics in aqueous solution and biological media, lipophilicity and solubility of various 1-carbamoyl derivatives of 5-fluorouracil **23**, 209
Prodrugs of 5-fluorouracil. VIII. Improved rectal and oral delivery of 5-fluorouracil via various prodrugs. Structure-rectal absorption relationships **36**, 41
Buur, A., H. Bundgaard, E. Falch, Prodrugs of 5-fluorouracil. IV. Hydrolysis kinetics, bioactivation and physicochemical properties of various *N*-acyloxymethyl derivatives of 5-fluorouracil **24**, 43
Buur, A., H. Bundgaard, V.H.L. Lee, Prodrugs of propranolol: Hydrolysis and intramolecular aminolysis of various propranolol esters and oxazolidin-2-one derivative **42**, 51
Buur, A., L. Trier, C. Magnusson, P. Artursson, Permeability of 5-fluorouracil and prodrugs in Caco-2 cell monolayers **129**, 223
Buxeraud, J., see Jambut-Absil, A.C. **35**, 129
Buxton, P.C., I.R. Lynch, J.M. Roe, Solid-state forms of paroxetine hydrochloride **42**, 135
Buyuktimkin, N., see Buyuktimkin, S. **118**, 113; **127**, 245
Buyuktimkin, N., see Marjukka, Turunen T. **92**, 89
Buyuktimkin, S., N. Buyuktimkin, J.H. Rytting, Preparation and-enhancing effect of 1-(*N*,*N*-dimethylamino)-2-propanol dodecanoate (DAIPD) on the transepidermal penetration of clonidine, hydrocortisone, and indomethacin through shed snake skin **118**, 113
Interaction of indomethacin with a new penetration enhancer, dodecyl 2-(*N*,*N*-dimethylamino)propionate (DDAIP): Its effect on transdermal delivery **127**, 245
Buyuktimkin, S., see Marjukka, Turunen T. **92**, 89
Bye, A., see Gilbert, J.C. **32**, 223
Bye, A., see Watkinson, A.C. **74**, 229
Byrn, S.R., see Stephenson, G.A. **146**, 93
Byron, P.R., R.E. Notari, M.Y. Huang, Pharmacokinetic predictions of optimum drug delivery rates from prodrugs designed for maximum duration **1**, 219
Byron, P.R., M.J. Rathbone, Prediction of interfacial transfer kinetics. I. Relative importance of diffusional resistance in aqueous and organic boundary layers in two-phase transfer cell **21**, 107
Prediction of interfacial transfer kinetics. II. Solute ionization and aqueous phase ionic strength effects in two-phase transfer and rotating diffusion cells **29**, 103
Byron, P.R., see Hickey, A.J. **42**, 267
Byron, P.R., see Hindle, M. **116**, 169; **134**, 137
Byron, P.R., see Jashnani, R.N. **113**, 123; **130**, 13
Byron, P.R., see Notari, R.E. **1**, 233
Byron, P.R., see Phillips, E.M. **110**, 9
Bystrom, K., see Briggner, L.E. **105**, 125
Byun, S.Y., see Shin, S.C. **137**, 95

C

Cabana, B.E., see Prasad, V.K. **11**, 81
Cabanes, A., see Fonseca, M.J. **133**, 265
Cabarga, M.M., see Sanchez, B.M. **106**, 229
Cabral Marques, H., J. Hadgraft, I.W. Kellaway, Studies of cyclodextrin inclusion complexes. I. The salbutamol–cyclodextrin complex as studied by phase solubility and DSC **63**, 259
Cabral Marques, H., J. Hadgraft, I.W. Kellaway, G. Taylor, Studies of cyclodextrin inclusion complexes. III. The pulmonary absorption of β-, DM-β- and HP-β-cyclodextrins in rabbits **77**, 297
Studies of cyclodextrin inclusion complexes. IV. The pulmonary absorption of salbutamol from a complex with 2-hydroxypropyl-β-cyclodextrin in rabbits **77**, 303
Cabral, Marques H., J. Hadgraft, I.W. Kellaway, W.J. Pugh, Studies of cyclodextrin inclusion complexes. II. Molecular modelling and ^{1}H NMR evidence for the salbutamol-β-cyclodextrin complex **63**, 267
Caccialanza, G., see Maggi, L. **136**, 43
Cachet, T., see Vinckier, C. **55**, 67
Cachet, T., G. Van den Mooter, R. Hauchecorne, C. Vinckier, J. Hoogmartens, Decomposition kinetics of erythromycin A in acidic aqueous solutions **55**, 59
Cachon, M., see Gehenot, A. **45**, 13
Cacini, W., see Ritschel, W.A. **13**, 353
Cadorniga, R., see Frances, C. **77**, 193
Cadorniga, R., see Torrado, S. **86**, 147; **133**, 65; **140**, 45; **140**, 247
Cagiano, R., see Altomare, C. **48**, 91
Cainey, J.M., see Meakin, B.J. **119**, 91; **119**, 103
Caira, M.R., see Van Tonder, E.C. **63**, 35
Cairncross, C., C. Ackermann, Permeation of digoxin through skin in vitro **27**, 361
Cajkovac, M., see Skalko, N. **85**, 97
Cal, S., G. Iglesias, C. Souto, A. Concheiro, J.L. Gomez-Amoza, R. Martinez-Pacheco, Effects of hydration on the properties of a roller-dried β-lactose for direct compression **129**, 253
Cal, S., B. Rodriguez-Puente, C. Souto, A. Concheiro, J.L. Gomez-Amoza, R. Martinez-Pacheco, Comparison of a spray-dried α-lactose monohydrate with a fully hydrated roller-dried β-lactose **136**, 13
Calabrese, G., see Dal, Pozzo A. **70**, 219
Caldwell, L.J., see Itoh, T. **33**, 265
Caldwell, R.W., M.A. Chryssanthis, V.O. Davis, Systemic delivery of sympathomimetic amines by transdermal iontophoresis **123**, 181
Caldwell, R.W., see Loftsson, T. **22**, 345
Callant, D., see Morgan, S.M. **122**, 121

Callegan, M.C., J.A. Hobden, R.J. O'Callaghan, J.M. Hill, Ocular drug delivery: A comparison of transcorneal iontophoresis to corneal collagen shields **123**, 173
Callegaro, L., see Ghezzo, E. **87**, 21
Calpena, A.C., J. Lauroba, M. Suriol, R. Obach, Berrozpe J. Domenech, Effect of D-limonene on the transdermal permeation of nifedipine and domperidone **103**, 179
Calvert, R.T., see Hunter, E. **4**, 175
Calvo, M.B., J.L. Pedraz, M.T. Vicente, A. Dominguez-Gil, Pharmacokinetics of cefazoline and dibenzylamine administered in a sustained drug delivery system to healthy volunteers **67**, 259
Calvo, M.B., see Hernandez, R.M. **139**, 45
Calvo, M.V., Gil A. Dominguez, Binding of naproxen to human albumin. Interaction with palmitic acid **16**, 215
Calvo, M.V., J.M. Lanao, A. Dominguez-Gil, Bioavailability of rectally administered naproxen **38**, 117
Calvo, P., C. Thomas, M.J. Alonso, J.L. Vila-Jato, J.R. Robinson, Study of the mechanism of interaction of poly(ϵ-caprolactone) nanocapsules with the cornea by confocal laser scanning microscopy **103**, 283
Calvo, P., see Alonso, M.J. **68**, 69
Camber, O., P. Edman, Factors influencing the corneal permeability of prostaglandin $F_{2\alpha}$ and its isopropyl ester in vitro **37**, 27
Camber, O., P. Edman, L.I. Olsson et al., Permeability of prostaglandin $F_{2\alpha}$ and prostaglandin $F_{2\alpha}$ esters across cornea in vitro **29**, 259
Camber, O., U. Muller, Influence of some preservatives on the corneal permeability of pilocarpine and dexamethasone, in vitro **39**, 229
Camber, O., see Bjerre, C. **144**, 217
Camber, O., see Gren, T. **141**, 53
Camber, O., see Prisell, P.T. **85**, 51
Camenisch, G., G. Folkers, H. Van de Waterbeemd, Comparison of passive drug transport through Caco-2 cells and artificial membranes **147**, 61
Cameroni, R., see Iannuccelli, V. **143**, 195
Cameroni, R., see Vandelli, M.A. **100**, 107; **100**, 115
Campigli, V., see Saettone, M.F. **43**, 67
Cannon, J., see Gupta, P. **140**, 119
Cannon, J.B., N.A. Williams, K.J. Papp, Reduction of pain on intravenous infusion with bile salt formulations for a macrolide antibiotic **114**, 65
Cannon, J.B., see Lovell, M.W. **109**, 45
Canton, E., Esteban M. Jesus, F. Rius, Factors affecting the stability of ceftriaxone sodium in solution on storage **92**, 47
Capone, G., see Carli, F. **21**, 317
Cappel, M.J., J. Kreuter, Effect of nonionic surfactants on transdermal drug delivery. I. Polysorbates **69**, 143
Effect of nonionic surfactants on transdermal drug delivery. II. Poloxamer and poloxamine surfactants **69**, 155
Cappel, M.J., see Tsai, J.C. **68**, 127
Caputo, O., see Cavalli, R. **89**, R9; **148**, 47
Caraballo, I., M. Fernandez-Arevalo, M.A. Holgado, A.M. Rabasco, Percolation theory: Application to the study of the release behaviour from inert matrix systems **96**, 175
Caraballo, I., M. Fernandez-Arevalo, M.A. Holgado, A.M. Rabasco, H. Leuenberger, Study of the release mechanism of carteolol inert matrix tablets on the basis of percolation theory **109**, 229
Caraballo, I., M. Fernandez-Arevalo, M. Millan, A.M. Rabasco, H. Leuenberger, Study of percolation thresholds in ternary tablets **139**, 177
Caraballo, I., A.M. Rabasco, M. Fernandez-Arevalo, Study of thimerosal degradation mechanism **89**, 213
Caraballo, I., see Alvarez-Fuentes, J. **139**, 237
Caraballo, I., see Gonzalez-Rodriguez, M.L. **146**, 175
Caraballo, I., see Holgado, M.A. **114**, 13; **118**, 151
Carabello, I., see Alvarez-Fuentes, J. **148**, 219
Caram-Lelham, N., L.O. Sundelof, Some aspects on characterization and properties of charged polysaccharides. An investigation of the system carrageenan/amitriptyline/water with relation to amphiphile adsorption and charge density **115**, 103
Caram-Lelham, N., see Sjokvist, E. **79**, 123
Caramella, C., P. Colombo, U. Conte, F. Ferrari, A. Gazzaniga, A. LaManna, N.A. Peppas, A physical analysis of the phenomenon of tablet disintegration **44**, 177
Caramella, C., see Bonferoni, M.C. **80**, 109; **117**, 41
Caramella, C., see Ferrari, F. **109**, 115; **112**, 29; **117**, 49; **121**, 11; **136**, 71
Caramella, C., see Gazzaniga, A. **91**, 167
Caramella, C., see Giunchedi, P. **77**, 177
Caramella, C., see Hernandez, R.M. **139**, 45
Caramella, C., see Pena Romero, A. **73**, 239
Caramella, C., see Peppas, N.A. **51**, 77
Caramella, C., see Ruizk, S. **112**, 125
Cardile, V., see Panico, A.M. **98**, 19
Cardillo, P., see Dobetti, L. **128**, 277
Cardot, J.M., see Aaiche, J.M. **55**, 147
Carelli, V., Colo G. Di, C. Guerrini, E. Nannipieri, Drug release from silicone elastomer through controlled polymer cracking: An extension to macromolecular drugs **50**, 181
Carelli, V., Colo G. Di, E. Nannipieri, Effect of water-soluble additives on drug release from silicone rubber matrices. III. A study of release mechanism by differential scanning calorimetry **30**, 9
Factors in zero-order release of clonidine hydrochloride from monolithic polydimethylsiloxane matrices **35**, 21
Carelli, V., Colo G. Di, E. Nannipieri, M.F. Serafini, Evaluation of the solution impregnation method for loading drugs into suspension-type polymer matrices: A study of factors determining the patterns of solid drug distribution in matrix and drug release from matrix **55**, 199
Enhancement effects in the permeation of Alprazolam through hairless mouse skin **88**, 89
Bile acids as enhancers of steroid penetration through excised hairless mouse skin **89**, 81
A study of controlled-release systems for progesterone based on crosslinked poly(ethylene oxides) **94**, 103
Carelli, V., see Bilia, A. **130**, 83
Carelli, V., see Bonina, F.P. **100**, 41

Carelli, V., see Bottari, F. **2**, 63
Carelli, V., see Buonaguidi, M. **147**, 1
Carelli, V., see Di, Colo G. **30**, 1
Cariola, C.M., see Sheen, P.C. **118**, 221
Carless, J.E., see Burgess, D.J. **27**, 61; **32**, 207
Carless, J.E., see Rowe, J.S. **13**, 313
Carlfors, J., see Edsman, K. **137**, 233
Carlfors, J., see Ghaderi, R. **141**, 205
Carlfors, J., see Sturesson, C. **89**, 235
Carli, F., G. Capone, I. Colombo et al., Surface and transport properties of acrylic polymers influencing drug release from porous matrices **21**, 317
Carli, F., I. Colombo, L. Magarotto et al., Influence of polymer characteristics on drug loading into crospovidone **33**, 115
Carlin, A.S., see Storm, J.E. **65**, 265
Carlisi, B., see Castelli, F. **59**, 19
Carlisi, B., see Giammona, G. **57**, 55; **64**, 239
Carlisle, M.R., M.L. Chicoine, M.B. Wygant, A stability-indicating high-performance liquid chromatographic assay for nicotine in transdermal patches **80**, 227
Carlotti, M.E., see Cavalli, R. **148**, 47
Carlotti, M.E., see Gasco, M.R. **18**, 235
Carlson, G.T., see Sinko, C.M. **114**, 85
Carlucci, G., A. Colanzi, P. Mazzeo, M.G. Quaglia, Determination of 2-aminopyridine in piroxicam by derivative UV-spectrophotometry **53**, 257
Carlucci, G., Giuseppe E Di, P. Mazzeo, P. Stanley, Simultaneous determination of enalapril maleate and hydrochlorothiazide in tablets by derivative UV spectrophotometry and high-performance liquid chromatography **93**, 245
Carlucci, G., P. Mazzeo, M.G. Quaglia, C. Vetuschi, Determination of 5-methoxy-2-methylindole-3-acetic acid in indomethacin by derivative UV-spectrophotometry **49**, 79
Carlucci, G., P. Mazzco, C. Vetuschi, Giuseppe E Di, Reversed-phase high-performance liquid chromatographic and derivative UV spectrophotometric determination of famotidine in pharmaceutical preparations **102**, 271
Carlucci, G., see Veronico, M. **119**, 109
Carman-Meakin, B., see Farr, S.J. **51**, 39
Carney, R.F., see Maurin, M.B. **144**, 99
Caro, I., X. Boulenc, M. Rousset, V. Meunier, M. Bourrie, B. Julian, H. JoyeuxRoques, C.Berger, Y.Zweibaum, A.G. Fabre, Characterisation of a newly isolated Caco-2 clone (TC-7), as a model of transport processes and biotransformation of drugs **116**, 147
Caroff, E., see Jacolot, A. **128**, 283
Caroff, E., see Valiere, C. **138**, 125
Carola, K.V., see Shanker, R.M. **119**, 149
Carotta, V., see Esposito, E. **142**, 9
Carotti, A., see Altomare, C. **48**, 91; **56**, 273
Carpenter, B.G., see Mortazavi, S.A. **83**, 221; **94**, 195
Carpenter, J.F., see Foster, T.M. **134**, 193
Carpignano, R., see Gasco, M.R. **18**, 235
Carpignano, R., see Pattarino, F. **91**, 157
Carr, L., see Patel, J.P. **65**, 195
Carreno-Gomez, B., R. Duncan, Evaluation Of the biological properties of soluble chitosan and chitosan microspheres **148**, 231
Carret, L., see Lafforgue, C. **147**, 243
Carrio, I., see Roca, M. **69**, 189
Carrupt, P.A., see Rothlisberger, T. **44**, 141; **51**, 73
Carrupt, P.A., see Tsai, R.S. **80**, 39
Carruthers, K.M., see Smith, A. **59**, 115
Carson, J.H., see Lamson, M.J. **105**, 259
Carson, L.W., see Lee, V.H.L. **29**, 43
Carstensen, H., B.W. Muller, R.H. Muller, Adsorption of ethoxylated surfactants on nanoparticles. I. Characterization by hydrophobic interaction chromatography **67**, 29
Carstensen, J., see Bechgaard, E. **89**, 147
Carstensen, J.T., A. Li Wan Po, The state of water in drug decomposition in the moist solid state: Description and modelling **83**, 87
Carstensen, J.T., F. Puisieux, A. Mehta, M.A. Zoglio, Milling kinetics of granules **1**, 65
Carstensen, J.T., see Acree, Jr. W.E. **8**, 69
Carstensen, J.T., see Bamba, M. **2**, 307
Carstensen, J.T., see Ertel, K.D. **34**, 179; **42**, 171; **61**, 9
Carstensen, J.T., see Fessi, H. **1**, 265; **30**, 209
Carstensen, J.T., see Franchini, M.K. **111**, 153
Carstensen, J.T., see Hou, X.P. **25**, 207
Carstensen, J.T., see Lai, T.Y.F. **1**, 33
Carstensen, J.T., see Mouazen, F. **1**, 275
Carstensen, J.T., see Van Scoik, K.G. **58**, 185
Carstensen, J.T., see Vaution, C. **1**, 349
Carstensen, J.T., see Wesoluch, F. **2**, 343
Carter, J.E., see Skelly, J.P. **63**, 83
Carter, K.C., J. O'Grady, T.F. Dolan, A.J. Baillie, J. Alexander, J. Keys, A direct comparison of sodium stibogluconate treatment in two animal models of human visceral leishmaniasis, mouse and hamster **53**, 129
Carter, K.C., see Baillie, A.J. **57**, 23
Carter, K.C., see Carter, K.C. **53**, 129
Carter, K.C., see Collins, M. **83**, 251
Cartilier, L.H., see Moussa, I.S. **149**, 139
Cartwright-Shamoon, J.M., J.C. McElnay, P.F. D'Arcy, Examination of sorption and photodegradation of amsacrine during storage in intravenous burette administration sets **42**, 41
Cartwright-Shamoon, J., see McElnay, J.C. **47**, 239
Carvajal, M.T., see Shah, N.H. **106**, 15
Casabo, V.G., see Collado, E.F. **44**, 187
Casabo, V.G., see Diez-Sales, O. **77**, 1
Casabo, V.G., see Martin-Algarra, V. **122**, 1
Casabo, V.G., see Perez-Buendia, M.D. **55**, 165
Casabo, V.G., see Pla-Delfina, J.M. **37**, 49
Casadebaig, J., see Bassani, V. **63**, 57
Casadebaig, J., see Doko, M.B. **76**, 199
Casalderrey, M., see Landin, M. **107**, 247; **123**, 143
Casals, E., M. Gallardo, J. Estelrich, Factors influencing the encapsulation of thioguanine in DRV liposomes **143**, 171
Cashman, J.R., see Gumbleton, M. **71**, 175
Castano, A., see Navarro, A.S. **37**, 113

Castellano, I., I. Goni, M. Gurruchaga, M.V. Velasco, A. Munoz, M.R. Jimenez-Castellanos, Contribution to the study of new graft copolymer matrices for drug delivery systems. Technological study **146**, 71
Castellano, I., see Ferrero, M.C. **149**, 233
Castellano, I., see Velasco, M.V. **136**, 107
Castelli, F., B. Conti, G. Puglisi, U. Conte, G. Mazzone, Calorimetric studies on tolmetin release from poly-DL-lactide microspheres to lipid model membrane **103**, 217
Castelli, F., G. Giammona, G. Puglisi, B. Carlisi, S. Gurrieri, Interaction of macromolecular pro-drugs with lipid model membrane: Calorimetric study of 4-biphenylacetic aciid linked to α,β-poly(*N*-hydroxyethyl)-DL-aspartamide interacting with phosphatidylcholine vesicles **59**, 19
Castelli, F., G. Giammona, A. Raudino, G. Puglisi, Macromolecular prodrugs interaction with mixed lipid membrane. A calorimetric study of naproxen linked to polyaspartamide interacting with phosphatidylcholine and phosphatidylcholine-phosphatidic acid vesicles **70**, 43
Castelli, F., G. Puglisi, G. Giammona, C.A. Ventura, Effect of the complexation of some nonsteroidal anti-inflammatory drugs with β-cyclodextrin on the interaction with phosphatidylcholine liposomes **88**, 1
Castelli, F., G. Puglisi, R. Pignatello, S. Gurrieri, Calorimetric studies of the interaction of 4-biphenylacetic acid and its β-cyclodextrin inclusion compound with lipid model membrane **52**, 115
Castelli, F., see Bonina, F. **145**, 87
Castelli, F., see Bonina, F.P. **106**, 115
Castelli, F., see Saija, A. **124**, 1
Castensson, S., see Angberg, M. **61**, 67; **73**, 209; **77**, 269; **81**, 153; **83**, 11; **90**, 19
Casu, B., see Ahmed, S.M. **109**, 99
Catellani, P.L., P. Predella, A. Bellotti, P. Colombo, Tablet water uptake and disintegration force measurements **51**, 63
Catellani, P.L., P. Santi, E. Gasperini, S. Ciceri, G. Dondi, P. Colombo, Centrifugal die filling system in a new rotary tablet machine **88**, 285
Catellani, P.L., see Catellani, P.L. **88**, 285
Catellani, P.L., see Colombo, P. **88**, 99
Catellani, P.L., see Santi, P. **68**, 43; **92**, 23
Catherpermal, S., see Jorgensen, L. **113**, 83
Catz, P., D.R. Friend, Alkyl esters as skin permeation enhancers for indomethacin **55**, 17
Transdermal delivery of levonorgestrel. VIII. Effect of enhancers on rat skin, hairless mouse skin, hairless guinea pig skin, and human skin **58**, 93
Catz, P., see Nolen, III H.W. **107**, 169
Cavailles, L., see Doko, M.B. **76**, 199
Cavallaro, G., M. Fresta, G. Giammona, G. Puglisi, A. Villari, Entrapment of β-lactams antibiotics in polyethylcyanoacrylate nanoparticles: Studies on the possible in vivo application of this colloidal delivery system **111**, 31
Cavallaro, G., G. Giammona, Manna G. La, S. Palazzo, G. Pitarresi, Liveri V. Turco, Viscosimetric investigation of the interaction between sodium dodecylsulfate micelles and a polymer drug carrier **90**, 195
Cavallaro, G., see Cavallaro, G. **90**, 195
Cavallaro, G., see Fresta, M. **99**, 145
Cavallaro, G., see Giammona, G. **64**, 239; **105**, 57
Cavallaro, G., see Turco, Liveri V. **99**, 285
Cavalli, R., O. Caputo, M.E. Carlotti, M. Trotta, C. Scarnecchia, M.R. Gasco, Sterilization and freeze-drying of drug-free and drug-loaded solid lipid nanoparticles **148**, 47
Cavalli, R., O. Caputo, M.R. Gasco, Solid lipospheres of doxorubicin and idarubicin **89**, R9
Cavalli, R., S. Morel, M.R. Gasco, P. Chetoni, M.F. Saettone, Preparation and evaluation in vitro of colloidal lipospheres containing pilocarpine as ion pair **117**, 243
Cavalli, R., see Cavalli, R. **148**, 47
Cavalli, R., see Morel, S. **105**, 189; **119**, 126; **132**, 259
Cavalli, R., see Trotta, M. **143**, 67
Cave, G., see Rabaron, A. **99**, 29
Cave, G., see Santos Magalhaes N.S. **76**, 225
Cavrini, V., Pietra A.M Di, M.A. Raggi, High-pressure liquid chromatographic (HPLC) analysis of imidazole antifungals in commercial dosage forms **10**, 119
High-performance liquid chromatographic (HPLC) analysis of methenolone esters in pharmaceutical formulations **13**, 333
Cavrini, V., see Di, Pietra A.M. **35**, 13
Cavrini, V., see Gatti, R. **115**, 113
Cavrini, V., see Nobile, L. **40**, 85
Cazin, J.C., see Benaji, B. **105**, 83
Cazin, J.C., see Dine, T. **77**, 279
Cazin, J.C., see Faouzi, M.A. **89**, 125; **105**, 89
Cazin, J.C., see Gressier, B. **104**, 151
Cazin, J.C., see Khalfi, F. **139**, 243
Cazin, M., see Benaji, B. **105**, 83
Cazin, M., see Dine, T. **77**, 279
Cazin, M., see Faouzi, M.A. **89**, 125; **105**, 89
Cazin, M., see Khalfi, F. **139**, 243
Ceccarelli, L., see Bramanti, G. **49**, 241
Cedro, A., see Ahmed, S.M. **109**, 99
Cefali, E.A., J.C. Nolan, W.R. McConnell, D.L. Walters, Bioavailability of silicon and aluminum from Zeolite A in dogs **127**, 147
Cejudo-Ferragud, E., A. Nacher, A. Polache, T. Cercos-Fortea, M. Merino, V.G. Casabo, Evidence of competitive inhibition for the intestinal absorption of baclofen by phenylalanine **132**, 63
Celebi, N., see Acarturk, F. **85**, 1
Celebi, N., see Erden, N. **48**, 83; **137**, 57
Celebi, N., N. Erden, Interaction of naproxen with β-cyclodextrin in ground mixture **78**, 183
Celebi, N., N. Erden, A. Turkyilmaz, The preparation and evaluation of salbutamol sulphate containing poly(lactic acid-*co*-glycolic acid) microspheres with factorial design-based studies **136**, 89
Celik, M., see Maganti, L. **95**, 29; **103**, 55
Celik, M., see Mollan, M.J. **114**, 23; **144**, 1
Cellai, L., see Di, Giulio A. **144**, 27
Cellamare, S., see Altomare, C. **48**, 91; **56**, 273

Cencia-Rohan, L., S. Silvestri, Effect of solvent system on microfluidization-induced mechanical degradation **95**, 23
Ceolin, R., J. Dugue, J.C. Rouland, C. Ralambosoa, F. Lepage, Solid state studies on stiripentol: A novel anticonvulsant drug **74**, 77
Cepeda, M., see Garcia, M.J. **5**, 117
Cepelak, I., see Juretic, D. **61**, 219
Cerbai, R., see Fabrizio, Saettone M. **142**, 103
Cercos-Fortea, T., see Cejudo-Ferragud, E. **132**, 63
Cerezo, A., see Bedmar, M.C. **138**, 95
Cerezo, A., see Fernandez, M. **84**, 197; **98**, 29
Cerezo, A., see Margarit, M.V. **81**, 67; **108**, 101
Cettina, M.G., see Sclafani, J. **124**, 213
Cha, C.Y., see Lee, K.C. **59**, 27
Chabenat, C., see Ezpeleta, I. **131**, 191; **142**, 227
Chacon, M., L. Berges, J. Molpeceres, M.R. Aberturas, M. Guzman, Optimized preparation of poly D,L (lactic-glycolic) microspheres and nanoparticles for oral administration **141**, 81
Chafetz, L., see Neau, S.H. **108**, 95
Chafetz, L., see Shah, K.P. **109**, 271
Chafi, N., J.P. Montheard, J.M. Vergnaud, Release of 2-aminothiazole from polymer carriers **67**, 265
Chafi, N., J.P. Montheard, J.M. Vergnaud, S. Pinzauti, Dosage form with salicylic acid attached to a polyanhydride polymer dispersed in an Eudragit matrix **52**, 203
Chaillot, B., see Andres, C. **144**, 141
Chaing, C.H., see Law, S.L. **84**, 161
Chakrabarti, S., see Menon, A. **140**, 207
Challacombe, S.J., see O'Hagan, D.T. **108**, 133
Cham, T.M., see Chuo, W.H. **134**, 247; **144**, 241
Cham, T.M., see Lin, C.W. **118**, 169; **127**, 261
Chamart, S., M. Hanocq, G. Leclercq, N. Devleeschouwer, Adsorption of tamoxifen and 2-methyl derivatives under cell culture conditions **99**, 59
Chambers, L.A., see Walker, M. **70**, 167
Chan, H.K., I. Gonda, Development of a systematic theory of suspension inhalation aerosols. II. Aggregates of monodisperse particles nebulized in polydisperse droplets **41**, 147
Methotrexate: Existence of different types of solid **68**, 179
Chan, H.K., D.J.W. Grant, Influence of compaction on the intrinsic dissolution rate of modified acetaminophen and adipic acid crystals **57**, 117
Chan, H.K., see Rodley, G.A. **95**, 143
Chan, L.W., see Wan, L.S.C. **95**, 77; **103**, 267
Chan, S.Y., A.L.W. Po, Prodrugs for dermal delivery **55**, 1
Chan, S.Y., A. Li Wan Po, Quantitative assessment of non-steroidal anti-inflammatory topical products in nicotinate-induced erythema using tristimulus colour analysis **83**, 73
Chan, T.W., see Brandl, M. **93**, 75
Chanal, J.L., see Audran, M. **15**, 273; **20**, 129
Chanal, J.L., see Galleyrand, J.C. **42**, 225
Chandaria, B., see Buckton, G. **94**, 223
Chandler, S.G., L. Illum, N.W. Thomas, Nasal absorption in the rat. I. A method to demonstrate the histological effects of nasal formulations **70**, 19
Nasal absorption in rats. II. Effect of enhancers on insulin absorption and nasal histology **76**, 61
Chandler, S.G., N.W. Thomas, L. Illum, Nasal absorption in the rat. IV. Membrane activity of absorption enhancers **117**, 139
Chandraprakash, K.S., N. Udupa, P. Umadevi, G.K. Pillai, Pharmacokinetic evaluation of surfactant vesicle-entrapped methotrexate in tumor-bearing mice **61**, R1
Chang, C.M., R. Bodmeier, Organic solvent-free polymeric microspheres prepared from aqueous colloidal polymer dispersions by a w/o-emulsion technique **130**, 187
Binding of drugs to monoglyceride-based drug delivery systems **147**, 135
Chang, H.W., M.G. Lee, H. Lee, M.K. Park, C.K. Kim, Effects of leucovorin on the pharmacokinetics of methotrexate in rabbits **79**, 71
Chang, H.W., see Yoon, E.J. **67**, 177
Chang, L.C., see Kitamura, S. **101**, 127
Chang, P., see Law, S.L. **70**, 1; **116**, 87; **116**, 87; **128**, 139
Chang, R.K., S.I. Chang, J.R. Robinson, A study of the performance of a modified V-shaped solids mixer using segregating materials **80**, 171
Chang, R.K., G. Iturrioz, C.W. Luo, Preparation and evaluation of shellac pseudolatex as an aqueous enteric coating system for pellets **60**, 171
Chang, R.K., E.M. Rudnic, The effect of various polymeric coating systems on the dissolution and tableting properties of potassium chloride microcapsules **70**, 261
Chang, S.C., see Bundgaard, H. **33**, 15
Chang, S.I., see Chang, R.-K. **80**, 171
Chang, Y., see Chaudhuri, B. **42**, 117; **65**, 57
Chantres, J.R., see Sainz, M.C. **89**, 183; **91**, 1
Chantres, J.R., B. Elorza, M.A. Elorza, P. Rodado, Deoxycholate alters the order of acyl chains in freeze-thaw extrusion vesicles of L-α-dipalmitoyl phosphatidylcholine: Study of the 1,6-diphenyl-1,3,5-hexatriene steady-state fluorescence anisotropy **138**, 139
Chapman, C.B., see Sitaram, B.R. **118**, 181
Chapman, Jr. J.M., see Michniak, B.B. **91**, 85; **94**, 203; **110**, 231
Chapman, S.R., see Newton, J.M. **120**, 95; **120**, 101
Charman, S.A., see Bibby, D.C. **144**, 61
Charman, S.A., see Porter, C.J.H. **141**, 227
Charman, W.N., L.E. McCrossin, N.L. Pochopin, S.L.A. Munro, Study of the relative lactonization rates of pilocarpic acid and isopilocarpic acid in acidic media **88**, 397
Charman, W.N., see Bibby, D.C. **144**, 61
Charman, W.N., see Doherty, M.M. **111**, 205
Charman, W.N., see Pochopin, N.L. **105**, 169; **121**, 157
Charman, W.N., see Porter, C.J.H. **141**, 227
Charman, W.N.A., T. Noguchi, V.J. Stella, An experimental system designed to study the in situ intestinal lymphatic transport of lipophilic drugs in anesthetized rats **33**, 155
Testing potential dosage form strategies for intestinal lymphatic drug transport: Studies in the rat **33**, 173
Charman, W.N.A., V.J. Stella, Effects of lipid class and lipid vehicle volume on the intestinal lymphatic transport of DDT **33**, 165

Estimating the maximal potential for intestinal lymphatic transport of lipophilic drug molecules **34**, 175
Charman, W.N.A., see Noguchi, T. **24**, 173; **24**, 185
Chast, F., see Dumortier, G. **59**, 1
Chatham, S.M., see Buckton, G. **47**, 121; **56**, 151
Chatham, S.M., see Parsons, G.E. **82**, 145; **83**, 163
Chattaraj, S.C., I. Kanfer, Release of acyclovir from semi-solid dosage forms: A semi-automated procedure using a simple plexiglass flow-through cell **125**, 215
'The insertion cell': A novel approach to monitor drug release from semi-solid dosage forms **133**, 59
Chattaraj, S.C., J. Swarbrick, I. Kanfer, A simple diffusion cell to monitor drug release from semi-solid dosage forms **120**, 119
Chatterjee, S., see Fares, H.M. **133**, 215
Chaubal, M.G., see Vaidyanathan, R. **25**, 85
Chaudhuri, B., Y. Chang, A.H. Karara, Polyether polyurethane delivery systems. I. Evaluation of monolithic sytems for N^6-(δ^2-isopentenyl)adenosine **42**, 117
Polyether polyurethane delivery systems. II. Effect of coadministration of pentostatin with N^6-(δ^2-isopentenyl)adenosine **65**, 57
Chaudhuri, B., see Ahmed, I. **44**, 97
Chaudry, I., see Sangekar, S. **35**, 187
Chauhan, U.P.S., see Jaggi, M. **69**, 77
Chaumat, C., see Droin, A. **27**, 233
Chaumeil, J.C., see Arnaud, P. **129**, 279
Chaumeil, J.C., see Chemtob, C. **29**, 1; **29**, 83
Chaumeil, J.C., see Cohen, T. **137**, 249
Chaumeil, J.C., see Dumortier, G. **59**, 1
Chaumeil, J.C., see Fontan, J.E. **73**, 17; **82**, 67
Chaumeil, J.C., see Prudat-Christiaens, C. **141**, 109
Chaumeil, J.C., see Zuber, M. **47**, 31
Chauvet, A., see El Moussaoui, A. **106**, 109
Chauvet, A., see Lheritier, J. **123**, 273
Chawla, A., K.M.G. Taylor, J.M. Newton, M.C.R. Johnson, Production of spray dried salbutamol sulphate for use in dry powder aerosol formulation **108**, 233
Chawla, A., see Taylor, K.M.G. **85**, 203
Chawla, A., see Washington, C. **54**, 191
Chawner, J.A., P. Gilbert, Adsorption of alexidine and chlorhexidine to *Escherichia coli* and membrane components **55**, 209
Chee, S.M., see Davidson, A.G. **63**, 29
Cheema, M.S., see Henry, B.T. **73**, 81
Chemtob, C., J.C. Chaumeil, M. N'Dongo, Microencapsulation by ethylcellulose phase separation: Microcapsule characteristics **29**, 1
Tablets of metronidazole microcapsules: Release characteristics **29**, 83
Chen, C., see Cooke, N.E. **115**, 17
Chen, C.F., see Tsai, H.T. **80**, 75
Chen, C.F., see Tsai, T.H. **84**, 279
Chen, C.Y., F.A. Chen, A.B. Wu, H.C. Hsu, J.J. Kang, H.W. Cheng, Effect of hydroxypropyl-β-cyclodextrin on the solubility, photostability and in-vitro permeability of alkannin/shikonin enantiomers **141**, 171
Chen, C.Y., see Chen, C.Y. **141**, 171
Chen, C.Y., see Cheng, H.W. **120**, 137
Chen, C.Y., see Ho, H.O. **90**, 95
Chen, D., N. Ohta, T. Yotsuyanagi, K. Ikeda, Polymerized precipitation of human γ-globulin induced by *cis*-diamminedichloroplatinum(II) **106**, 249
Chen, D., see Ohta, N. **85**, 39; **118**, 85; **124**, 165
Chen, F.A., see Chen, C.Y. **141**, 171
Chen, F.A., see Cheng, H.W. **120**, 137
Chen, H., see Bodmeier, R. **51**, 1
Chen, J., P.A. Crooks, A. Hussain, Kinetics and mechanism of hydrolysis of amidals: Their relative stability compared to structurally related acetals and acylals **123**, 95
Chen, J.R., see Shah, J.C. **84**, 223; **113**, 103
Chen, K.S., see Lin, Y.Y. **124**, 53
Chen, M.L., see Pradhan, S. **138**, 207
Chen, N.R., see Truelove, J. **19**, 17
Chen, S., see Powell, M.F. **44**, 225
Chen, S.C., see Liu, C.H. **100**, 241
Chen, S.C., see Rajagopalan, N. **29**, 161
Chen, S.H., R.A. Kenley, J.S. Winterle, Sulconazole reactions with peracetic acid and hydrogen peroxide **72**, 89
Chen, S.R., see Lu, S.M. **119**, 11
Chen, T.K., D. Thornton, M.Y.K. Ho, Reduction of dimethyl sulfoxide by dihydrohalide salts of pharmaceuticals **59**, 211
Chen, W.G., G.C-C. Hwang, Adhesive and in vitro release characteristics of propranolol bioadhesive disc system **82**, 61
Chen, Y., L.E. Matheson, Quantitative structure-transportability relationship for the release of a series of substituted pyridines from a planar polydimethylsiloxane matrix **94**, 153
Chen, Y., P. Vayumhasuwan, L.E. Matheson, Prediction of flux through polydimethylsiloxane membranes using atomic charge calculations: Application to an extended data set **137**, 149
Chen, Y., W. Yang, L.E. Matheson, Prediction of flux through polydimethylsiloxane membranes using atomic charge calculations **94**, 81
Chen, Y., see Matheson, L.E. **125**, 297
Chen-Chow, P.C., S.G. Frank, The determination of lidocaine and benzocaine in isopropyl myristate **8**, 81
In vitro release of lidocaine from pluronic F-127 gels **8**, 89
Cheng, H.W., F.A. Chen, H.C. Hsu, C.Y. Chen, Photochemical decomposition of alkannin/shikonin enantiomers **120**, 137
Cheng, K.C., E. Shefter, T. Srikrishnan, Crystal structure analysis of the desolvation of the chloroform solvate of griseofulvin **2**, 81
Cheng, K.C., see Resetarits, D.E. **2**, 113
Cheng, K.C., see Shefter, E. **6**, 179
Cheng, Y.L., see Chiu, L.K. **126**, 169
Chenon, M.T., see Lecomte, S. **139**, 105
Cheron, M., see Lemesle-Lamache, V. **141**, 117
Cheskin, H., see Qiu, Y. **115**, 95; **143**, 179
Chess, E.K., see Jenke, D.R. **78**, 115; **108**, 1

Chetoni, P., P. Crotti, M.F. Saettone, Albuterol prodrugs for ocular administration: Synthesis and evaluation of the physico-chemical and IOP-depressant properties of three albuterol triesters **105**, 147
Chetoni, P., see Cavalli, R. **117**, 243
Chetoni, P., see Fabrizio, Saettone M. **142**, 103
Chetoni, P., see Gallarate, M. **100**, 219
Chetoni, P., see Saettone, M.F. **51**, 203; **72**, 131; **126**, 79
Cheung, Y.W., A. Li Wan Po, W.J. Irwin, Cutaneous biotransformation as a parameter in the modulation of the activity of topical corticosteroids **26**, 175
Resistance to enzymic hydrolysis as a parameter in drug potency **27**, 325
Chevanne, F., see Dollo, G. **131**, 219; **136**, 165
Chevanne, F., see Le Corre, P. **107**, 41
Chevanne, F., see Polard, E. **134**, 37
Chevin, J.C., P. Maincent, J.M. Mougeolle, M. Jacque, J.A. Martin, Nickel II elimination and autoradiographic distribution in normal and nickel-sensitized guinea pigs **45**, 101
Chevrier, R., see Satou, V. **107**, 111
Chi, S.C., E.S. Park, H. Kim, Effect of penetration enhancers on flurbiprofen permeation through rat skin **126**, 267
Chi, S.C., see Kim, C.K. **99**, 109
Chi, S.C., see Oh, I. **73**, 23
Chia, C.G.H., see Wan, L.S.C. **77**, 183
Chiam, S.F., see Hu, O.Y.P. **53**, 85
Chian-Jo, Chang, see Melethil, S. **31**, 83
Chiang, C., N. Weiner, Gastrointestinal uptake of liposomes. II. In vivo studies **40**, 143
Chiang, C.C., M. Longer, P. Tyle, D. Fessler, B. Shetty, Formulation development of an oral dosage form for an HIV protease inhibitor, AG1284 **117**, 197
Chiang, C.C., see Pavliv, L. **105**, 227
Chiang, C.M., G.L. Flynn, N.D. Weiner, G.J. Szpunar, Bioavailability assessment of topical delivery systems: Effect of inter-subject variability on relative in vitro deliveries of minoxidil and hydrocortisone from solution and ointment formulations **50**, 21
Bioavailability assessment of topical delivery systems: Effect of vehicle evaporation upon in vitro delivery of minoxidil from solution formulations **55**, 229
Chiang, C.M., G.L. Flynn, N.D. Weiner, W.J. Addicks, G.J. Szpunar, Bioavailability assessment of topical delivery systems: In vitro delivery of minoxidil from prototypical semisolid formulations **49**, 109
Chiang, C.M., N. Weiner, Gastrointestinal uptake of liposomes. I. In vitro and in situ studies **37**, 75
Chiang, C.M., see Ma, X. **142**, 115
Chiang, H.S., see Gu, L. **41**, 95; **41**, 105
Chiao, C.S., see Ghosh, T.K. **82**, 39
Chiappini, I., see Fardella, G. **121**, 123
Chicco, D., see Vojnovic, D. **145**, 203
Chicoine, M.L., see Carlisle, M.R. **80**, 227
Chien, Y.W., see Banga, A.K. **48**, 15
Chien, Y.W., see Chun, I.K. **121**, 217
Chien, Y.W., see Corbo, D.C. **50**, 253
Chien, Y.W., see Lelawongs, P. **56**, 13; **61**, 179
Chien, Y.W., see Liu, J.C. **25**, 265
Chien, Y.W., see Mirchandani, H. **95**, 1
Chien, Y.W., see Nair, M. **89**, 41
Chien, Y.W., see Shah, H.S. **86**, 167
Chien, Y.W., see Tan, E.L. **42**, 161
Chien, Y.W., see Toddywala, R.D. **76**, 77
Chien, Y.W., see Wearley, L.L. **59**, 87
Chien, Y.W., see Yu, K. **125**, 81
Childs, K., see Ghosh, T.K. **88**, 391
Chin, N.L.J., see Leeuwenkamp, O.R. **33**, 1
Chiorboli, V., see Gambari, R. **72**, 251
Chiori, C.O., A.A. Ghobashy, A potentiating effect of EDTA on the bactericidal activity of lower concentrations of ethanol **17**, 121
Chiossone, D.C., see Smith, P.L. **106**, 95
Chiu, L.K., W.J. Chiu, Y.L. Cheng, Effects of polymer degradation on drug release—A mechanistic study of morphology and transport properties in 50:50 poly(DL-lactide-*co*-glycolide) **126**, 169
Chiu, W.J., see Chiu, L.K. **126**, 169
Ch'ng, H.S., see Mahrag, Tur K. **148**, 63
Ch'ng, H.S., see Mumtaz, A.M. **121**, 129; **121**, 249
Cho, M.J., M.A. Allen, Quantitative assessment of the negative catalytic effects of a cationic surfactant myristyl-γ-picolinium chloride on the specific-acid catalyzed epimerization of 15(*S*)-15-methyl prostaglandin F2α **1**, 281
Cho, M.J., J.F. Scieszka, P.S. Burton, Citric acid as an adjuvant for transepithelial transport **52**, 79
Cho, M.J., see Haynes, L.C. **45**, 169
Cho, M.J., see Scieszka, J.F. **45**, 165
Chohan, R.K., J.M. Newton, Analysis of extrusion of some wet powder masses used in extrusion/spheronisation **131**, 201
Choi, H.G., see Kim, C.K. **106**, 25
Choi, J.Y., see Kim, C.K. **106**, 25
Choi, K.E., see Yoon, W.H. **102**, 249
Choi, S.H., see Gonsho, A. **65**, 183
Choi, Y.K., S.Y. Jeong, Y.H. Kim, A glucose-triggered solubilized polymer gel matrix for an insulin delivery system **80**, 9
Choi, Y.J., see Kim, C.K. **98**, 9
Chookhachizadeh-Moghaddam, M.H., see Barzegar-Jalali, M. **144**, 127
Chopineau, J., J. Barthelemy, P. Bastide, Effect of repeated formaldehyde low-temperature steam sterilization on physicomechanical properties of PVC tubing **73**, 191
Chopineau, J., see Levadoux, E. **127**, 255
Chopineau, J., see Satou, V. **107**, 111
Chopineau, J., see Sautou-Miranda, V. **130**, 251
Chou, C.K.H., R.C. Vasavada, Evaluation of propylene glycol monostearate-ethoxylated stearyl alcohol films and kinetics of cortisol release **25**, 199
Chou, H.L., see Sheu, M.T. **85**, 57
Chou, R.C., I.M. Ramzan, G. Levy, Effect of pregnancy or renal dysfunction on differences in plasma protein binding of phenytoin in peripheral blood and blood exiting the liver of rats **18**, 217

Choudhari, K.B., N.M. Sanghavi, Dissolution behaviour and characterization of diazepam-Pullulan coground mixtures **89**, 207
Choudhury, S., K.F. Nelson, Improvement of oral bioavailability of carbamazepine by inclusion in 2-hydroxypropyl-β-cyclodextrin **85**, 175
Chouinard, F., F.W.K. Kan, J.C. Leroux, C. Foucher, V. Lenaerts, Preparation and purification of polyisohexylcyanoacrylate nanocapsules **72**, 211
Chouinard, F., see Pimienta, C. **80**, 1
Choularton, A., see Buckton, G. **47**, 121
Choulis, N.H., see Al-Hmoud, H. **68**, R1
Choulis, N.H., see Athanassiou, G.C. **72**, 141; **85**, 209; **90**, 51
Choulis, N.H., see Giannakou, S.A. **125**, 7
Choulis, N.H., see Papadimitriou, E. **86**, 131
Choulis, N.H., see Vlachou, M.D. **82**, 47
Chow, A.H.L., P.K.K. Chow, W. Zhongshan, D.J.W. Grant, Modification of acetaminophen crystals: Influence of growth in aqueous solutions containing *p*-acetoxyacetanilide on crystal properties **24**, 239
Chow, A.H.L., J.D. Gordon, A. Szeitz, J.W.M. Young, Modification of phenytoin crystals. III. Influence of 3-butanoyloxymethyl-5,5-diphenylhydantoin on solution-phase crystallization and related crystal properties **126**, 11
Chow, A.H.L., D.J.W. Grant, Modification of acetaminophen crystals. II. Influence of stirring rate during solution-phase growth on crystal properties in the presence and absence of *p*-acetoxyacetanilide **41**, 29
Modification of acetaminophen crystals. III. Influence of initial supersaturation during solution-phase growth on crystal properties in the presence and absence of *p*-acetoxyacetanilide **42**, 123
Influence of crystallization conditions on the physical properties of acetaminophen crystals: Evaluation by multiple linear regression **51**, 115
Physical factors influencing the aqueos dissolution rate of acetaminophen crystals doped with *p*-acetoxyacetanilide: Evaluation by multiple linear regression **51**, 129
Chow, A.H.L., C.K. Hsia, Modification of phenytoin crystals: Influence of 3-acetoxymethyl-5,5-diphenylhydantoin on solution-phase crystallization and related crystal properties **75**, 219
Chow, A.H.L., C.K. Hsia, J.D. Gordon, J.W.M. Young, E.I. Vargha-Butler, Assessment of wettability and its relationship to the intrinsic dissolution rate of doped phenytoin crystals **126**, 21
Chow, A.H.L., see Gordon, J.D. **79**, 171
Chow, A.H.L., see Grant, D.J.W. **18**, 25
Chow, D., see Shah, J.C. **113**, 103; **84**, 223; **112**, 263; **109**, 283
Chow, D.D., A.H. Karara, Characterization, dissolution and bioavailability in rats of ibuprofen-β-cyclodextrin complex system **28**, 95
Chow, D.D., see Touitou, E. **104**, 19
Chow, K.Y., J. Go, M. Mehdizadeh, D.J.W. Grant, Modification of adipic acid crystals: Influence of growth in the presence of fatty acid additives on crystal properties **20**, 3
Chow, K.Y., J. Go, Z. Wang et al., Modification of adipic acid crystals. II. Influence of growth in the presence of oleic acid on crystal properties **25**, 41
Chow, M.Y., see Neau, S.H. **108**, 95; **131**, 47
Chow, P.K.K., see Chow, A.H.L. **24**, 239
Chow, W.S., see Rajagopalan, N. **29**, 161
Chow, Y.P., see Chowhan, Z.T. **5**, 139
Chowdhry, B.Z., see Buckton, G. **83**, 115; **110**, 179
Chowdhry, B.Z., see Mackellar, A.J. **112**, 65
Chowhan, Z.T., Y.P. Chow, Compression behavior of pharmaceutical powders **5**, 139
Chowhan, Z.T., E.E. Linn, Powder inhalation aerosol studies. II. In vitro rat lung model and its comparisons with the air sampler **3**, 117
Chowhan, Z.T., I.C. Yang, Powder flow studies. IV. Tensile strength and orifice flow rate relationships of binary mixtures **14**, 231
Chowhan, Z.T., see Gordon, M.S. **97**, 119
Chowhan, Z.T., see Ong, J.T.H. **96**, 231
Chowhan, Z.T., see Wang, L.H. **60**, 61
Christen, M.O., see Burgot, G. **129**, 295
Christensen, C.A., see Michniak, B.B. **94**, 203; **110**, 231
Christensen, F.N., see Bertelsen, F. **111**, 117
Christensen, F.N., see Jorgensen, K. **143**, 223
Christensen, J.V., see Hansen, K.T. **79**, 205
Christensen, L.N., see Pridal, L. **136**, 53
Christensen, S., see Mork, N. **60**, 163
Christodoulou, M., see Toth, I. **125**, 117
Christrup, L., see Drustrup, J. **71**, 105
Christrup, L.L., S.S. Davis, M. Frier, C.D. Melia, S.N. Rasmussen, N. Washington, I.R. Wilding, C. Andersen, Deposition of a model substance, ^{99m}Tc E-HIDA, in the oral cavity after administration of lozenges, chewing gum and sublingual tablets **66**, 169
Christrup, L.L., see Hansen, L.B. **88**, 221, 229, 237, 243
Christy, N., see Washington, C. **54**, 191
Chryssanthis, M.A., see Caldwell, R.W. **123**, 181
Chu, W.W., see Anderson, B.D. **43**, 261
Chukuka, C.S., see Okhamafe, A.O. **68**, 11
Chulia, D., see Pena, Romero A. **73**, 239
Chun, I.K., Y.W. Chien, Stabilization of methionine enkephalin in various rabbit mucosal extracts by enzyme inhibitors **121**, 217
Chun, I.K., D.S. Yun, Inclusion complexation of hydrocortisone butyrate with cyclodextrins and dimethyl-β-cyclodextrin in aqueous solution and in solid state **96**, 91
Chung, F.Y., M.D. Donovan, Nasal pre-systemic metabolism of peptide drugs: Substance P metabolism in the sheep nasal cavity **128**, 229
Chung, P.H., see Hu, O.Y.P. **53**, 85
Chung, S.M., E.J. Yoon, S.H. Kim, M.G. Lee, H. Lee, M.K. Park, C.K. Kim, Pharmacokinetics of 5-fluorouracil after intravenous infusion of 5-fluorouracil-acetic acid-human serum albumin conjugates to rabbits **68**, 61
Chungi, V.S., L.W. Dittert, R.B. Smith, Gastrointestinal sites of furosemide absorption in rats **4**, 27

Chuo, W.H., T.R. Tsai, S.H. Hsu, T.M. Cham, Development of nifedipine-loaded albumin microspheres using a statistical factorial design **134**, 247
Preparation and in-vitro evaluation of nifedipine loaded albumin microspheres cross-linked by different glutaraldehyde concentrations **144**, 241
Ciceri, S., see Catellani, P.L. **88**, 285
Ciesliga, B.L., see Johnston, T.P. **52**, 139; **59**, 95
Ciftci, K., M.J. Groves, Delivery of antitumor compounds to the rat colon: In vitro and in vivo evaluation **145**, 157
Ciftci, K., H.S. Kas, A.A. Hincal, T.M. Ercan, O. Guven, S. Ruacan, In vitro and in vivo evaluation of PLAGA (50/50) microspheres containing 5-fluorouracil prepared by a solvent evaporation method **131**, 73
Cini, M., see Saettone, M.F. **126**, 83
Civaroli, P., see Martini, A. **127**, 239
Civaroli, P., see Muggetti, L. **136**, 81
Claessen, A., see Steerenberg, P.A. **40**, 51
Clancy, M.J., J. Corish, O.I. Corrigan, A comparison of the effects of electrical current and penetration enhancers on the properties of human skin using spectroscopic (FTIR) and calorimetric (DSC) methods **105**, 47
Claramonte, M.D.C., see Morell, J.L.P. **137**, 49
Clark, A.G., see Hardy, J.G. **31**, 151
Clark, A.R., see Newman, S.P. **74**, 203; **102**, 127
Clark, A.R., The use of laser diffraction for the evaluation of the aerosol clouds generated by medical nebulizers **115**, 69
Clarke, G.M., J.M. Newton, M.B. Short, Comparative gastrointestinal transit of pellet systems of varying density **114**, 1
Gastrointestinal transit of pellets of differing size and density **100**, 81
Clarke, J.G., S.J. Farr, S.R. Wicks, Technetium-99m labelling of suspension type pressurised metered dose inhalers comprising various drug/surfactant combinations **80**, R1
Clarke, J.G., S.R. Wicks, S.J. Farr, Surfactant mediated effects in pressurized metered dose inhalers formulated as suspensions. I. Drug–surfactant interactions in a model propellant system **93**, 221
Clarke, S., see Shah, V.P. **125**, 99
Clarke, S.W., see Newman, S.P. **36**, 55; **74**, 203, 209
Clarson, S., see Hutchings, D. **104**, 203
Clas, S.D., R. Faizer, R.E. O'Connor, E.B. Vadas, Quantification of crystallinity in blends of lyophilized and crystalline MK-0591 using X-ray powder diffraction 2,2 dimethylpropionic acid **121**, 73
Clas, S.D., see Bechard, S.R. **114**, 205
Clas, S.D., see Thibert, R. **134**, 59
Claudius, J.S., S.H. Neau, Kinetic and equilibrium characterization of interactions between glycopeptide antibiotics and sodium carboxymethyl starch **144**, 71
Clegg, J.A., see Pimm, M.V. **79**, 77
Clement, G., see Valiere, C. **138**, 125
Clements, R., see Jones, H.P. **104**, 253
Clemessy, M., G. Couarraze, B. Bevan, F. Puisieux, Preservation of skin permeability during in vitro iontophoretic experiments **101**, 219
Cliff, M.J., see Landin, M. **133**, 127; **134**, 243
Climent, E., see Fabra-Campos, S. **109**, 197
Climent, E., see Garrigues, T.M. **64**, 127
Closson, S.K., see Rush, B.D. **73**, 231
Cobby, J., see Nouravarsani, F. **11**, 45
Cociancich, F., see Gianasi, E. **148**, 139
Coderch, L., M. Oliva, L. Pens, J.L. Parra, Percutaneous penetration in vivo of amino acids **111**, 7
Coderch, L., M. Oliva, M. Pons, la Maza A. De, A.M. Manich, J.L. Parra, Percutaneous penetration of liposomes using the tape stripping technique **139**, 197
Coderch, L., see Coderch, L. **139**, 197
Coenegracht, P.J.M., see Westerhuis, J.A. **143**, 151
Coffin, Beach D.P., R.G. Hollenbeck, Determination of the energy of tablet formation during compression of selected pharmaceutical powders **17**, 313
Cognez, P., D. Pape, G. Burgot, H. Allain, J.L. Burgot, Elimination of linear parameters in non-linear regressions: A fast and effective method for the determination of binding parameters **123**, 137
Cohen, E.M., see Mazuel, G. **16**, 97
Cohen, G., see Blanchon, S. **72**, 1
Cohen, T., see Cohen, T. **137**, 249
Cohen, T., H. Sauvageon-Martre, D. Brossard, F. D'Hermies, C. Bardin, F. Chast, J.C. Chaumeil, Amphotericin B eye drops as a lipidic emulsion **137**, 249
Colanzi, A., see Carlucci, G. **53**, 257
Colarte, A.I., see Heinamaki, J.T. **109**, 9
Colburn, W.A., W.L. Hayton, M. Gibaldi, Pharmacokinetic studies of the hypoglycemic effects of insulin in the treatment of diabetic ketoacidosis **1**, 121
Cole, A., see Bailey, A.J.G. **22**, 283
Cole, E.T., see Kenyon, C.J. **112**, 207
Cole, S.K., M.J. Story, D. Attwood, T. Laudanski, J. Robertson, S.G. Barnwell, Studies using a non-ionic surfactant-containing drug delivery system designed for hard gelatin capsule compatibility **88**, 211
Cole, S.K., M.J. Story, T. Laudanski, M. Dwyer, D. Attwood, J. Robertson, S.G. Barnwell, Targeting drugs to the enterohepatic circulation: A potential drug delivery system designed to enhance the bioavailability of indomethacin **80**, 63
Cole, S.K., see Cole, S.K. **80**, 63; **88**, 211
Coleman, N.J., D.Q.M. Craig, Modulated temperature differential scanning calorimetry: A novel approach to pharmaceutical thermal analysis **135**, 13
Collado, E.F., see Garrigues, T.M. **57**, 189
Collado, E.F., S. Fabra-Campos, J.E. Peris-Ribera, V.G. Casabo, A. Martin-Villodre, J.M. Pla-Delfina, Absorption-partition relationships for true homologous series of xenobiotics as a possible approach to study mechanisms of surfactants in absorption. II. Aromatic amines in rat small intestine **44**, 187
Collett, J.H., see Attwood, D. **26**, 25; **65**, 201; **70**, 147
Collett, J.H., see Bentley, M.V.L.B. **146**, 255
Collett, J.H., see Burrows, R. **111**, 283
Collett, J.H., see Martini, L. **113**, 223

Collett, J.H., see Wood, J.M. **7**, 189
Collett, J.H., see Zaniboni, H.C. **125**, 151
Collier, P.J., P. Austin, P. Gilbert, Uptake and distribution of some isothiazolone biocides into *Escherichia coli* ATCC 8739 and *Schizosaccharomyces pombe* NCYC 1354 **66**, 201
Isothiazolone biocides: Enzyme-inhibiting pro-drugs **74**, 195
Collier, P.S., The possible range of values for apparent volume of distribution at steady-state when disposition may be characterized by a tri-exponential function **18**, 345
Collier, P.S., D.W.G. Harron, K.R. Devine, Pharmacokinetic profile of oral digoxin in healthy volunteers **1**, 3
Collier, P.S., R.A. Upton, S. Riegelman, Multiple-dose bioavailability study of a sustained-release theophylline product: Choledyl S.A. tablets **26**, 99
Collier, P.S., see Grainger-Rousseau, T.J. **54**, 1
Collier, P.S., see Harron, D.W.G. **1**, 61
Collier, P.S., see McClean, E. **60**, 35
Collier, P.S., see Temple, D.J. **2**, 127
Collins, A.E.M., P.B. Deasy, D.J. MacCarthy, D.B. Shanley, Evaluation of a controlled-release compact containing tetracycline hydrochloride bonded to tooth for the treatment of periodontal disease **51**, 103
Collins, J., see Peck, C.C. **82**, 9
Collins, J.V., see Leigh, T.R. **67**, 275
Collins, M., A.J. Baillie, K.C. Carter, Visceral leishmaniasis in the BALB/c mouse: Sodium stibogluconate treatment during acute and chronic stages of infection. II. Changes in tissue drug distribution **83**, 251
Colombel, J.F., see Cortot, A. **22**, 321
Colombo, I., see Carli, F. **21**, 317; **33**, 115
Colombo, P., P.L. Catellani, N.A. Peppas, L. Maggi, U. Conte, Swelling characteristics of hydrophilic matrices for controlled release. New dimensionless number to describe the swelling and release behavior **88**, 99
Colombo, P., U. Conte, A. Gazzaniga, L. Maggi, M.E. Sangalli, N.A. Peppas, A. La Manna, Drug release modulation by physical restrictions of matrix swelling **63**, 43
Colombo, P., see am Ende, M.T. **120**, 33
Colombo, P., see Caramella, C. **44**, 177
Colombo, P., see Catellani, P.L. **51**, 63
Colombo, P., see Colombo, P. **63**, 43
Colombo, P., see Cornaz, A.L. **129**, 175
Colombo, P., see Gazzaniga, A. **91**, 167
Colombo, P., see Hariharan, D. **112**, 47
Colombo, P., see Maggi, L. **99**, 173
Colombo, P., see Peppas, N.A. **51**, 77
Colombo, P., see Santi, P. **68**, 43; **92**, 23
Colome, C., M.A. Alsina, M.A. Busquets, I. Haro, F. Reig, Interaction of aminoglycosides and colistin with model membranes: Liposomes and monolayers **90**, 59
Conard, G.J., M.J. Jernberg, F.N. Wong et al., Clinical assessment of theophylline absorption from Theolair-SR and two other sustained-release formulations relative to a conventional formulation **10**, 259
Concheiro, A., see Cal, S. **129**, 253; **136**, 13
Concheiro, A., see Doldan, C. **124**, 69
Concheiro, A., see Landin, M. **91**, 123, 133, 143; **103**, 9; **109**, 1; **107**, 247
Concheiro, A., see Martinez-Pacheco, R. **47**, 37
Concheiro, A., see Mosquera, M.J. **135**, 147
Concheiro, A., see Perez-Marcos, B. **100**, 207
Concheiro, A., see Vila-Jato, J.L. **30**, 229
Conflant, P., see Di Martino, P. **128**, 1
Conflant, P., see Fachaux, J.M. **99**, 99
Conklin, J.D., see Mason, W.D. **36**, 105
Conklin, J.D., see Wagner, D.L. **57**, 177
Conley, D.L., see Benjamin, E.J. **13**, 205
Connolly, M.A., see Washington, C. **77**, 57
Conrath, G., see Couarraze, G. **56**, 197
Constantinides, P.P., S.H. Yiv, Particle size determination of phase-inverted water-in-oil microemulsions under different dilution and storage conditions **115**, 225
Conte, U., see Bonferoni, M.C. **80**, 109
Conte, U., see Caramella, C. **44**, 177
Conte, U., see Castelli, F. **103**, 217
Conte, U., see Colombo, P. **63**, 43; **88**, 99
Conte, U., see Ferrari, F. **121**, 11
Conte, U., see Gazzaniga, A. **91**, 167
Conte, U., see Giunchedi, P. **77**, 177; **85**, 141; **94**, 15; **130**, 41
Conte, U., see Hernandez, R.M. **139**, 45
Conte, U., see Maggi, L. **99**, 173; **135**, 73; **136**, 43
Conte, U., see Ochoa, Machiste E. **126**, 65
Conte, U., see Peppas, N.A. **51**, 77
Conte, U., see Saettone, M.F. **126**, 79
Conte, U., see Sangalli, M.E. **91**, 151
Conti, B., see Castelli, F. **103**, 217
Conti, B., see Pavanetto, F. **48**, 255; **51**, 55; **54**, 143; **74**, 175; **84**, 151
Contreras Claramonte M.D., A.P. Vialard, F.G. Vilchez, An application of regular solution theory in the study of the solubility of naproxen in some solvents used in topical preparations **94**, 23
In vitro percutaneous absorption of naproxen from gels using a double-layer artificial membrane **98**, 37
Conzentino, P., see Liversidge, G.G. **125**, 309
Cook, C.E., see Shah, V.P. **82**, 1
Cook, C.S., C.L. Hauswald, A.Y. Grahn, K. Kowalski, A. Karim, R. Koch, G.L. Schoenhard, J.A. Oppermann, Suitability of the dog as an animal model for evaluating theophylline absorption and food effects from different formulations **60**, 125
Cook, J.D., see Nicholls, T.J. **138**, 175
Cooke, N.E., C. Chen, A contribution to a mathematical theory for polymer-based controlled release devices **115**, 17
Cools, A.A., L.H.M. Janssen, A quantitative approach to the determination of drug release from reverse-phase evaporation lipid vesicles. The influence of sodium ion-pair formation on warfarin partitioning and permeability **20**, 335
Coomans, D., see Bottenberg, P. **49**, 147; **58**, 203
Coombes, R., see Jacobs, G.P. **32**, 151
Cooper, C.M., J.D. Sallis, Studies on the gastrointestinal absorption of phosphocitrate, a powerful controller of hydroxyapatite formation **98**, 165

Copovi, A., see Diez-Sales, O. **77**, 1
Coppard, S., see Flinn, N. **137**, 33
Coppi, G., see Forni, F. **60**, 83
Coppi, G., see Iannuccelli, V. **143**, 187, 195
Coppi, G., see Vanelli, M.A. **100**, 115
Coran, S.A., see Vincieri, F.F. **48**, 119
Corbini, G., see Corti, P. **111**, 83
Corbo, D.C., Y.C. Huang, Y.W. Chien, Nasal delivery of progestational steroids in ovariectomized rabbits. II. Effect of penetrant hydrophilicity **50**, 253
Cordes, G., see Hadgraft, J. **64**, 187
Corish, J., see Clancy, M.J. **105**, 47
Cornaz, A.L., A. de Ascentis, P. Colombo, P. Buri, In vitro characteristics of nicotine microspheres for transnasal delivery **129**, 175
Cornelissen, P.J.G., G.M.J. Beijersbergen van Henegouwen, K.W. Gerritsma, Photochemical decomposition of 1,4-benzodiazepines. Diazepam **1**, 173
Cornelissen, P.J.G., G.M.J.B. Van Henegouwen, K.W. Gerritsma, Photochemical decomposition of 1,4-benzodiazepines. Chlordiazepoxide benzodiazepine◇ **3**, 205
Corness, D., see Barnwell, S.G. **128**, 145
Corness, D., see Burns, S.J. **110**, 291; **121**, 37; **134**, 223
Cornett, C., see Jensen, J. **88**, 177
Cornwell, P.A., B.W. Barry, The routes of penetration of ions and 5-fluorouracil across human skin and the mechanisms of action of terpene skin penetration enhancers **94**, 189
Effects of penetration enhancer treatment on the statistical distribution of human skin permeabilities **117**, 101
Cornwell, P.A., B.W. Barry, J.A. Bouwstra, G.S. Gooris, Modes of action of terpene penetration enhancers in human skin; differential scanning calorimetry, small-angle X-ray diffraction and enhancer uptake studies **127**, 9
Cornwell, P.A., see Williams, A.C. **86**, 69
Corradeschi, F., see Bocci, V. **24**, 109; **34**, 111
Corrigan, O.I., M.A. Farvar, W.I. Higuchi, Drug membrane transport enhancement using high energy drug polyvinylpyrrolidone (PVP) co-precipitates **5**, 229
Corrigan, O.I., E.M. Holohan, K. Sabra, Amorphous forms of thiazide diuretics prepared by spray-drying **18**, 195
Corrigan, O.I., C.A. Murphy, R.F. Timoney, Dissolution properties of polyethylene glycols and polyethylene glycol-drug systems **4**, 67
Corrigan, O.I., R.F. Timoney, Towards standardizing agitation conditions in the beaker dissolution method **1**, 299
Corrigan, O.I., see Clancy, M.J. **105**, 47
Corrigan, O.I., see Healy, A.M. **84**, 167; **143**, 211
Corrigan, O.I., see Ledwidge, M.T. **147**, 41
Corrigan, O.I., see McGloughlin, R.M.R. **82**, 135
Corrigan, O.I., see Millar, F.C. **92**, 97
Corrigan, O.I., see O'Reilly, J.R. **105**, 137; **109**, 147
Corrigan, O.I., see Quigley, J.M. **58**, 107
Cortese, M., see Bundgaard, H. **10**, 181
Cortesi, R., E. Esposito, E. Menegatti, R. Gambari, C. Nastruzzi, Gelatin microspheres as a new approach for the controlled delivery of synthetic oligonucleotides and PCR-generated DNA fragments **105**, 181
Effect of cationic liposome composition on in vitro cytotoxicity and protective effect on carried DNA **139**, 69
Cortesi, R., see Esposito, E. **117**, 151; **129**, 263
Corti, P., G. Corbini, P. Gratteri, S. Furlanetto, S. Pinzauti, Determination of some quinolones in tablets, human plasma and urine by differential-pulse polarography **111**, 83
Cortot, A., J.F. Colombel, Gastric emptying of OROS tablets in man **22**, 321
Coskun, T., see Onur, M.A. **88**, 313
Cossum, P.A., see Roberts, M.S. **17**, 145
Costa, M.L., see Moreira, J.N. **147**, 153
Costantino, H.R., see Lam, X.M. **142**, 85
Costantino, L., G. Rastelli, A. Albasini, Inhibitory activity of flavonols towards the xanthine oxidase enzyme **86**, 17
Costello, B., see Francoeur, M.L. **25**, 275
Cots, J., see Pujol, M. **58**, 103
Cotton, M.L., P. Lamarche, S. Motola, E.B. Vadas, The selection of an appropriate salt form and preparation of a stable oral formulation methylbutyric acid **109**, 237
Cotton, M.L., D.W. Wu, E.B. Vadas, Drug–excipient interaction study of enalapril maleate using thermal analysis and scanning electron microscopy **40**, 129
Couarraze, G., B. Leclerc, G. Conrath, F. Falson-Rieg, F. Puisieux, Diffusion of a dispersed solute in a polymeric matrix **56**, 197
Couarraze, G., see Blanchon, S. **72**, 1
Couarraze, G., see Clemessy, M. **101**, 219
Couarraze, G., see Dubernet, C. **35**, 145
Couarraze, G., see Gallardo, M. **100**, 55
Couarraze, G., see Mayorga, P. **132**, 71
Couarraze, G., see Ribardiere, A. **144**, 195
Coupe, A.J., S.S. Davis, D.F. Evans, I.R. Wilding, The effect of sleep on the gastrointestinal transit of pharmaceutical dosage forms **78**, 69
Do pellet formulations empty from the stomach with food? **92**, 167
Courage, P., see Sitaram, B.R. **118**, 181
Courteille, F., see Benoit, J.P. **29**, 95
Courtois, P., see Rochard, E. **101**, 257
Coutel-Egros, A., Y. Maitani, M. Veillard, Y. Machida, T. Nagai, Combined effects of pH, cosolvent and penetration enhancers on the in vitro buccal absorption of propranolol through excised hamster cheek pouch **84**, 117
Couvreur, P., see Blanco, Prieto M.J. **111**, 137
Couvreur, P., see Brasseur, N. **70**, 129
Couvreur, P., see Delie, F. **115**, 45
Couvreur, P., see Gallardo, M. **100**, 55
Couvreur, P., see Grislain, L. **15**, 335
Couvreur, P., see Kante, B. **7**, 45
Couvreur, P., see Lherm, C. **84**, 13
Couvreur, P., see Marchal-Heussler, L. **58**, 115
Couvreur, P., see Muller, R.H. **84**, 1
Couvreur, P., see Nemati, F. **102**, 55; **138**, 237
Couvreur, P., see Roy, D. **148**, 165
Couvreur, P., see Seijo, B. **62**, 1
Cox, D.M., see Toth, I. **73**, 259; **77**, 13

Cox, P.J., see Sujja-Areevath, J. **139**, 53
Craane, W.I.H.M., see De, Vries M. **76**, 25
Cradock, J.C., see Poochikian, G.K. **13**, 219
Craig, D.Q.M., B.S.A. Barker, D. Banning, S.W. Booth, An investigation into the mechanisms of self-emulsification using particle size analysis and low frequency dielectric spectroscopy **114**, 103
Craig, D.Q.M., H.S.R. Lievens, K.G. Pitt, D.E. Storey, An investigation into the physico-chemical properties of self-emulsifying systems using low frequency dielectric spectroscopy, surface tension measurements and particle size analysis methylphenyl)urea **96**, 147
Craig, D.Q.M., J.M. Newton, Characterisation of polyethylene glycols using differential scanning calorimetry **74**, 33
Characterisation of polyethylene glycols using solution calorimetry **74**, 43
Characterisation of polyethylene glycol solid dispersions using differential scanning calorimetry and solution calorimetry **76**, 17
The dissolution of nortriptyline HCl from polyethylene glycol solid dispersions **78**, 175
Craig, D.Q.M., K.M.G. Taylor, The use of low frequency dielectric analysis in the characterisation of metered dose inhaler formulations **114**, 124 + 128
Craig, D.Q.M., see Ah-Fat, N.M.W. **107**, 239
Craig, D.Q.M., see Coleman, N.J. **135**, 13
Craig, D.Q.M., see Sjokvist Saers, E. **83**, 211
Craig, D.Q.M., see Sutananta, W. **110**, 75; **111**, 51; **125**, 123; **132**, 1
Craig, D.Q.M., see Tamburic, S. **137**, 243
Craig, D.Q.M., see Wulff, M. **142**, 189
Crauste-Manciet, S., J.F. Huneau, M.O. Decroix, D. Tome, R. Farinotti, J.C. Chaumeil, Cefpodoxime proxetil esterase activity in rabbit small intestine: A role in the partial cefpodoxime absorption **149**, 241
Crauste-Manciet, S., see Crauste-Manciet, S. **149**, 241
Crawford, B., see McMahon, F.G. **91**, 75
Crawford, J.W., see Hansen, L.D. **135**, 31
Creekmore, J.R., see Wurster, D.E. **116**, 179
Cremers, H., see Kwon, G.S. **79**, 191
Cremers, H.F.M., L.W. Seymour, K. Lam, G. Los, Vugt M. Van, G. Kwon, Y.H. Bae, S.W. Kim, J. Feijen, Adriamycin-loaded albumin-heparin conjugate microspheres for intraperitoneal chemotherapy **110**, 117
Cremers, H.F.M., R. Verrijk, L.G. Bayon, M.M. Wesseling, J. Wondergem, G. Heuff, S. Meijer, G.S. Kwon, Y.H. Bae, S.W. Kim, J. Feijen, Improved distribution and reduced toxicity of adriamycin bound to albumin-heparin microspheres **120**, 51
Cremieux, A., see Damery, B. **49**, 205
Crison, J.R., see Yu, L.X. **140**, 111
Critchley, H., see Illum, L. **57**, 49
Crommelin, D.J.A., C.J. de Blaey, In vitro release studies on drugs suspended in non-polar media. II. The release of paracetamol and chloramphenicol from suspensions in liquid paraffin **6**, 29
Crommelin, D.J.A., Bloois L. Van, Preparation and characterization of doxorubicin-containing liposomes. II. Loading capacity, long-term stability and doxorubicin-bilayer interaction mechanism **17**, 135
Crommelin, D.J.A., J. Modderkolk, C.J. de Blaey, The pH dependence of rectal absorption of theophylline from solutions of aminophylline in situ in rats **3**, 299
Crommelin, D.J.A., N. Slaats, Bloois L. Van, Preparation and characterization of doxorubicin-containing liposomes. I. Influence of liposome charge and pH of hydration medium on loading capacity and particle size **16**, 79
Crommelin, D.J.A., see de Smidt, J.H. **77**, 255, 261
Crommelin, D.J.A., see Grijseels, H. **14**, 299, 313
Crommelin, D.J.A., see Grit, M. **50**, 1
Crommelin, D.J.A., see Janssen, M.J.H. **23**, 1
Crommelin, D.J.A., see Talsma, H. **37**, 171; **77**, 119
Crommelin, D.J.A., see Van Bommel, E.M. **22**, 299; **72**, 19
Crommelin, D.J.A., see Zuidam, N.J. **126**, 209
Crooks, P.A., see Chen, J. **123**, 95
Crooks, P.A., see Jona, J.A. **123**, 127
Crooks, P.A., see Tawfiq, F.A. **64**, 45
Cropp, J.W., L.L. Augsburger, K. Marshall, Simultaneous monitoring of tamping force and pin displacement (F-D) on an Hofliger Karg capsule filling machine **71**, 127
Crotti, P., see Chetoni, P. **105**, 147
Crundwell, E., see Jones, M.F. **4**, 1
Cruz, J.E., D.D. Maness, G.J. Yakatan, Kinetics and mechanism of hydrolysis of furosemide **2**, 275
Cruz, M.E.M., M.M. Gaspar, F. Lopes, J.S. Jorge, R. Perez-Soler, Liposomal L-asparaginase: In vitro evaluation **96**, 67
Cruz, M.E.M., see Martins, M.B.F. **142**, 75
Cruz, M.E.M., see Mimoso, I.M. **147**, 109
Cruz, T., see Zhang, X. **137**, 199
Csabai, K., T. Cserhati, J. Szejtli, Interaction of some barbituric acid derivatives with hydroxypropyl-β-cyclodextrin **91**, 15
Cserhati, T., Interaction of some anticancer drugs with carboxymethyl-β-cyclodextrin **124**, 205
Cserhati, T., E. Forgacs, J. Szejtli, Inclusion complex formation of antisense nucleotides with hydroxypropyl-β-cyclodextrin **141**, 1
Cserhati, T., J. Hollo, Interaction of taxol and other anticancer drugs with hydroxypropyl-β-cyclodextrin **108**, 69
Cserhati, T., S. Olajos, M. Szogyi, Physicochemical interaction of some benzodiazepine derivatives with amino acids and phospholipids **61**, 189
Cserhati, T., see Csabai, K. **91**, 15
Cucala, J., see Fabregas, J.L. **52**, 173
Cui, F., see Kawashima, Y. **119**, 139
Culank, L.S., R.H. Leach, Estimates of digoxin bioavailability from short-term plasma profiles and cumulative urinary excretion **2**, 173
Culverwell, A.L., see Betteridge, R.F. **56**, 37
Cumming, K.I., A.J. Winfield, In vitro evaluation of a series of sodium carboxylates as dermal penetration enhancers **108**, 141

Cuna, M., see Mosquera, M.J. **135**, 147
Cundy, K.C., see Liversidge, G.G. **125**, 91
Curl, R., see Addicks, W.J. **56**, 243
Curry, J.T., see Lambert, W.J. **95**, 181
Curry, S.H., see Aburawi, S. **22**, 327
Cutler, D.J., see Firmer, S.J. **48**, 231
Cutt, T., J.T. Fell, P.J. Rue, M.S. Spring, Granulation and compaction of a model system. I. Granule properties **33**, 81
Granulation and compaction of a model system. III. Compaction properties of the granules **49**, 157

D

Da Silva, C.P., C. De Oliveira, M.C.P. De Lima, In vitro assay of the effects of anticancer drugs on leukemic T-cells by flow microcalorimetry **111**, 25
Da Silva, H., see Gregoriadis, G. **65**, 235
Dabbagh, M.A., J.L. Ford, M.H. Rubinstein, J.E. Hogan, Effects of polymer particle size, compaction pressure and hydrophilic polymers on drug release from matrices containing ethylcellulose **140**, 85
Daca, J., see Srcic, S. **99**, 21
Dahl, G., L.E. Dahlinder, G. Ekenved et al., The effect of buffering of acetylsalicylic acid on dissolution, absorption, gastric pH and faecal blood loss **10**, 143
Dahlin, M., see Pereswetoff-Morath, L. **128**, 9
Dahlinder, L.E., see Dahl, G. **10**, 143
Daicoviciu, D., see Leucuta, S.E. **41**, 213
Dais, M., see Small, D. **149**, 195, 203
Dakas, C.J., see Takla, P.G. **43**, 225
Dakkuri, A., see Muhi-Eldeen, Z. **26**, 15
Dal Pozzo, A., M. Acquasaliente, New highly water-soluble phenytoin prodrugs **81**, 263
Dal Pozzo, A., G. Donzelli, L. Rodriquez, A. Tajana, 'In vitro' model for the evaluation of drug distribution and plasma protein-binding relationships **50**, 97
Dal Pozzo, A., E. Liggeri, C. Delucca, G. Calabrese, Prediction of skin permeation of highly lipophilic compounds: In vitro model with a modified receptor phase **70**, 219
Dal Zotto, M., see Realdon, N. **148**, 155
Dalal, P.S., M.M. Narurkar, In vitro and in vivo evaluation of sustained release suspensions of ibuprofen **73**, 157
Dalby, R.N., see Hickey, A.J. **42**, 267
Dalby, R.N., see Jashnani, R.N. **113**, 123
Dalla Fini, see Realdon, N. **148**, 155
Dallas, P.P., see Giannakou, S.A. **125**, 7
Dallas, P.P., see Vlachou, M.D. **82**, 47
Daly, P.B., S.S. Davis, M. Frier et al., Scintigraphic assessment of the in vivo dissolution rate of a sustained release tablet **10**, 17
Daly, P.B., S.S. Davis, J.W. Kennerley, The effect of anionic surfactants on the release of chlorpheniramine from a polymer matrix tablet **18**, 201
Dam, A.M., L.G. Gazso, S. Kaewpila, I. Maschek, Radiation sterilization dose calculation for heparin and aprotinin based on ISO Method 1 **121**, 245
Damery, B., A. Cremieux, Virucidal activity against Herpes and Vaccinia virus of eight antiseptic formulations **49**, 205
Damge, C., M. Aprahamian, G. Balboni et al., Polyalkylcyanoacrylate nanocapsules increase the intestinal absorption of a lipophilic drug **36**, 121
Danckwerts, M.P., Van der Watt J., The effect of processing variables on the compression properties of controlled release core-in-cup compressed tablets from a new adjustable punch **123**, 85
Danckwerts, M.P., Development of a zero-order release oral compressed tablet with potential for commercial tabletting production **112**, 37
Danhof, M., see Verhoeven, J. **45**, 65
Daniel, P.T., see Muller, C.E. **57**, 41
Daniels, R., see Weiss, G. **124**, 87, 97
Danjo, K., see Takeeda, T. **41**, 21
Dansereau, R., R. Dansereau, Development of a new coated-bead dosage form of sodium iodide I-131 **49**, 115
Dansereau, R.J., D.B. Worthen, Letter to the editor **103**, 97
Danti, A.G., see Jafari, M.R. **48**, 207
D'Antona, P., see Esposito, E. **142**, 9
Dany, S., see Touitou, E. **43**, 9
Darbord, J.C., see Guyomard, S. **40**, 173
Darcy, P., see Briggner, L.E. **105**, 125
Darcy, P., see Buckton, G. **116**, 113; **117**, 253; **121**, 81; **122**, 130; **123**, 265; **136**, 141
D'Arcy, P.F., see Cartwright-Shamoon, J.M. **42**, 41
D'Arcy, P.F., see Harron, D.W.G. **1**, 61
D'Arcy, P.F., see McElany, J.C. **5**, 167; **6**, 205; **7**, 83; **26**, 267; **31**, 107; **33**, 261; **36**, 199
D'Arcy, P.F., see Yahya, A.M. **25**, 217; **31**, 65; **34**, 137
Darrington, R.T., T.X. Xiang, B.D. Anderson, Inclusion complexes of purine nucleosides with cyclodextrins. I. Complexation and stabilization of a dideoxypurine nucleoside with 2-hydroxypropyl-β-cyclodextrin **59**, 35
Darrouzet, H., see Amdidouche, D. **54**, 175
Darwish, I.A., see El-Massik, M.A. **140**, 69
Darwish, I.A., M.A. El-Massik, E.E. Hassan, L.K. El-Khordagui, Assessment of a hydroalcoholic surfactant solution as a medium for the dissolution testing of phenytoin **140**, 25
Darwish, R.M., S.F. Bloomfield, The effect of co-solvents on the antibacterial activity of paraben preservatives **119**, 183
Effect of ethanol, propylene glycol and glycerol on the interaction of methyl and propyl *p*-hydroxybenzoate with *Staphylococcus aureus* and *Pseudomonas aeruginosa* **147**, 51
Das, N.G., see Duddu, S.P. **114**, 247
Dauphin, J.F., see Abi Khalil, F. **140**, 1
Dauphin, J.F., see Valiere, C. **138**, 125
Dave, S.C., see Muhammad, N.A. **21**, 295
Davidson, A.G., S.M. Chee, F.M. Millar, D. Watson, Isolation and identification of an alkali-catalyzed hydrolysis product of nitrazepam **63**, 29
Davidson, A.G., G.A. Smail, Isolation and characterisation of an acid-catalyzed intermediate hydrolysis product of nitrazepam **69**, 1

Davidson, J.R., see Forbes, R.T. **126**, 199
Davidson, R., R. Ek, G. Ragnarsson, Determination of the expansion characteristics of porous cellulose matrices by image analysis **112**, 153
Davidson, R., H. Nyqvist, G. Ragnarsson, Porous cellulose matrices—A novel excipient for the formulation of solid dosage forms **100**, 49
Davidson, R., see Dennis, A.B. **65**, 85
Davidson, R., see Ek, R. **122**, 49
Davidsson, B., see Edman, P. **34**, 225
Davies, M.C., J.S. Binns, C.D. Melia, P.J. Hendra, D. Bourgeois, S.P. Church, P.J. Stephenson, FT Raman spectroscopy of drug in polymers **66**, 223
Davies, M.C., I.R. Wilding, R.D. Short, M.A. Khan, J.F. Watts, C.D. Melia, An analysis of the surface chemical structure of polymethacrylate (Eudragit) film coating polymers by XPS **57**, 183
Davies, M.C., I.R. Wilding, R.D. Short, C.D. Melia, R.C. Rowe, The in-situ chemical analysis of polymer film coatings using static secondary ion mass spectrometry (SSIMS) **62**, 97
Davies, M.C., see Davies, M.C. **57**, 183; **66**, 223
Davies, M.C., see Gilbert, J.C. **40**, 93
Davies, M.C., see Porter, C.J.H. **83**, 273
Davies, M.C., see Watts, P.J. **76**, 55; **87**, 215; **98**, 63, 75; **102**, 101
Davies, N.M., S.J. Farr, I.W. Kellaway, G. Taylor, M. Thomas, A comparison of the gastric retention of alginate containing tablet formulations with and without the inclusion of excipient calcium ions **105**, 97
Davies, N.M., see Durrani, A.M. **88**, 409
Davignon, J.P., see Poochikian, G.K. **13**, 219
Davis, A.F., J. Hadgraft, Effect of supersaturation on membrane transport. 1. Hydrocortisone acetate **76**, 1
Davis, A.F., see Pellett, M.A. **111**, 1
Davis, G.W., see Jordan, C.G.M. **141**, 125
Davis, M., I. Ichikawa, E.J. Williams, G.S. Banker, Comparison and evaluation of enteric polymer properties in aqueous solutions **28**, 157
Davis, S.S., Physico-chemical studies on aerosol solutions for drug delivery. I. Water-propylene glycol systems **1**, 71
Davis, S.S., M.D. Bubb, Physico-chemical studies on aerosol solutions for drug delivery. III. The effect of relative humidity on the particle size of inhalation aerosols **1**, 303
Davis, S.S., G. Elson, J. Whitmore, Physico-chemical studies on aerosol solutions for drug delivery. II. Water-propylene glycol-ethanol systems **1**, 85
Davis, S.S., P. Hansrani, The influence of emulsifying agents on the phagocytosis of lipid emulsions by macrophages **23**, 69
Davis, S.S., J.G. Hardy, M.J. Taylor et al., A comparative study of the gastrointestinal transit of a pellet and tablet formulation **21**, 167
The effect of food on the gastrointestinal transit of pellets and an osmotic device (Osmet) **21**, 331
Davis, S.S., J.G. Hardy, C.G. Wilson et al., Gastrointestinal transit of a controlled release naproxen tablet formulation **32**, 85
Davis, S.S., L. Illum, I.M. Triccas, K.N. Winchcomb, The simultaneous degradation and sorption of diltiazem in aqueous solution **30**, 29
Davis, S.S., L. Illum, I.M. Walker, The in vivo evaluation of emulsion formulations administered intramuscularly **38**, 133
Davis, S.S., L. Illum, C. Washington, G. Harper, Studies on the interaction of charge-reversed emulsions with the reticuloendothelial system **82**, 99
Davis, S.S., R. Khosla, C.G. Wilson, N. Washington, Gastrointestinal transit of a controlled-release pellet formulation of tiaprofenic acid and the effect of food **35**, 253
Davis, S.S., R. Khosla, C.G. Wilson, N. Washington, S.T. Leslie, S. Malkowska, The gastrointestinal transit of a controlled release formulation of indomethacin **60**, 191
Davis, S.S., G.D. Parr, L. Feely, S.T. Leslie, S. Malkowska, G.F. Lockwood, The gastrointestinal transit investigation of a controlled release aminophylline formulation **49**, 183
Davis, S.S., C. Robertson, I.R. Wilding, Gastrointestinal transit of a multiparticulate tablet formulation in patients with active ulcerative colitis **68**, 199
Davis, S.S., I. Walker, Measurement of the yield of multiple emulsion droplets by a fluorescent tracer technique **17**, 203
Davis, S.S., E.A. Wilding, I.R. Wilding, Gastrointestinal transit of a matrix tablet formulation: Comparison of canine and human data **94**, 235
Davis, S.S., see Al-Khamis, K. **10**, 25; **40**, 111
Davis, S.S., see Aspden, T.J. **122**, 69
Davis, S.S., see Baveja, S.K. **39**, 39
Davis, S.S., see Christrup, L.L. **66**, 169
Davis, S.S., see Coupe, A.J. **78**, 69; **92**, 167
Davis, S.S., see Daly, P.B. **10**, 17; **18**, 201
Davis, S.S., see Douglas, S.J. **34**, 145
Davis, S.S., see Farraj, N.F. **43**, 83, 93; **52**, 11, 19
Davis, S.S., see Feely, L.C. **41**, 83; **44**, 131
Davis, S.S., see Hardy, J.G. **38**, 103; **48**, 79
Davis, S.S., see Henry, B.T. **73**, 81
Davis, S.S., see Illum, L. **11**, 323; **12**, 135; **17**, 183; **26**, 113; **29**, 53; **52**, 221; **54**, 41; **57**, 49; **63**, 207
Davis, S.S., see Jeffery, H. **77**, 169
Davis, S.S., see Jenkins, P.G. **102**, 261
Davis, S.S., see Johnson, O.L. **53**, 237; **59**, 131; **63**, 65
Davis, S.S., see Khosla, R. **52**, 1
Davis, S.S., see Kojima, I. **20**, 203, 247
Davis, S.S., see Kreuter, J. **16**, 105
Davis, S.S., see Moghimi, S.M. **68**, 121
Davis, S.S., see Nankervis, R. **119**, 173; **130**, 57
Davis, S.S., see O'Hagan, D.T. **103**, 37
Davis, S.S., see Olejnik, O. **30**, 101
Davis, S.S., see Porter, C.J.H. **83**, 273
Davis, S.S., see Price, J.M.C. **76**, 123
Davis, S.S., see Repond, C. **38**, 47
Davis, S.S., see Sastry, C.S.P. **39**, 137
Davis, S.S., see Saunders, J. **68**, 265
Davis, S.S., see Torrado, J.J. **51**, 85
Davis, S.S., see Washington, C. **44**, 169; **54**, 191

Davis, S.S., see Washington, N. **27**, 279; **28**, 139; **29**, 253; **34**, 105
Davis, S.S., see Wilding, I.R. **76**, 133; **83**, 155; **84**, 59; **101**, 263; **111**, 99; **132**, 81
Davis, V.O., see Caldwell, R.W. **123**, 181
Dawes, W.H., M.J. Groves, The effect of electrolytes on phospholipid-stabilized soyabean oil emulsions **1**, 141
Day, N.H., see Nankervis, R. **101**, 249; **119**, 173; **130**, 57
Dayan, N., see Touitou, E. **103**, 131
Dayter, L., see Randall, C.S. **120**, 235
De Ascentis, A, see Cornaz, A.L. **129**, 175
De Backer, P., see Inghelbrecht, S. **148**, 103
De Baets, P., see Inghelbrecht, S. **148**, 103
De Beukelaer Van Ooteghem, M., Interaction of certain pharmaceutical products with polysorbate 80 in solution **16**, 345
De Blaey, C.J., see Crommelin, D.J.A. **3**, 299; **6**, 29
De Blaey, C.J., see Fokkens, J.G. **14**, 79; **18**, 127; **19**, 177
De Blaey, C.J., see Grijseels, H. **9**, 337; **14**, 299, 313; **16**, 295
De Blaey, C.J., see Rutten-Kingma, J.J. **3**, 39, 179, 187
De Blaey, C.J., see Schurgers, N. **19**, 283
De Boer, A.H., D. Gjaltema, P. Hagedoorn, Inhalation characteristics and their effects on in vitro drug delivery front dry powder inhalers. Part 2. Effect of peak flow rate (PIFR) and inspiration time on the in vitro drug release from three different types of commercial dry powder inhalers **138**, 45
De Boer, A.H., H.M.I. Winter, C.F. Lerk, Inhalation characteristics and their effects on in vitro drug delivery from dry powder inhalers. Part 1. Inhalation characteristics, work of breathing and volunteers' preference in dependence of the inhaler resistance **130**, 231
De Boer, A.G., see Akiyama, Y. **136**, 155; **138**, 13
De Boer, A.G., see Noach, A.B.J. **90**, 229
De Boer, A.H., see Riepma, K.A. **66**, 47; **76**, 9; **85**, 121
De Bolos, J., C. Galindo, M. Gallardo, Pico A. Rodriguez, Prediction of stability of cefazolin sodium in perfusion fluids **40**, 175
De Bolos, J., see Pujol, M. **146**, 263
De Bolos, J., see Torres, F. **129**, 275
De Bruijn, W.C, see de Haan, F.H.N. **56**, 75
De Caprariis, see Palagiano, F. **149**, 171
De Capraris, P., see Bonina, F.P. **77**, 21
De Gand, G., see Benameur, H. **89**, 157
De Greef, H., see Kinget, R. **110**, 65
De Groot, G., see Steerenberg, P.A. **40**, 51
De Groot, J.S., see Roy, S.D. **110**, 137
De Haan, F.H.N., H.E. Bodde, W.C. de Bruijn, L.A. Ginsel, H.E. Junginger, Visualizing drug transport across stratum corneum: cryotechniques, vapour fixation, autoradiography **56**, 75
De Haan, F.H.N., A.C.A. Jansen, The hydrodynamics in a simple transport vessel in two-phase systems of organic solvent-water **18**, 311
The influences of solvent properties on interfacial transfer kinetics in stirred two-phase systems of organic solvent-water **29**, 177
De Haan, F.H.N., T. De Vringer, H. Van de Waterbeemd, A.C.A. Jansen, Transport rate constants and transport rate parameters in various organic solvent-water systems **13**, 75
De Haan, F.H.N., see Blok, D. **73**, 171
De Haan, P., C.F. Lerk, The megaloporous system: A novel principle for zero-order drug delivery. I. In vitro and in vivo performance **31**, 15
The megaloporous system: A novel principle for zero-order drug delivery. II. A model for the mechanism of drug delivery **34**, 57
De Haan, P., see Van der Voet, H. **14**, 291
De Haan, P., see Westerhuis, J.A. **143**, 151
De la Maza, A., see Coderch, L. **139**, 197
De la Torre, C, see Halbaut, L. **147**, 31
De Lima, M.C.P, see Da, Silva C.P. **111**, 25
De Lorenzi, E., see Maggi, L. **136**, 43
De Meere, A.L., P.J. Taylor, H.J.M. Grunbauer, E. Tomlinson, Local composition models in pharmaceutical chemistry. II. Differentiation of hydrophobic fragmental constants **31**, 209
De Meere, A.L., E. Tomlinson, Theoretical and experimental studies on the origin of pH-absorption shifts **17**, 331
Physicochemical description of the absorption rate of a solute between water and 2,2,4-trimethylpentane **22**, 177
De Mol, N.J., J.P.A.C.M. Maanders, Non-covalent binding of some phenothiazine drugs to DNA **16**, 153
De Meere, A.L., see Grunbauer, H.J.M. **32**, 187
De Meere, A.L., see Leahy, D.E. **50**, 117
De Morais, S.M, A.E. Beezer, L.J. Ashby, R. Bolton, Biologically based QSARs: Study of cardanol derivatives on interaction with *Saccharomyces cerevisiae* **66**, 107
De Muynck, C., R.A. Lefebvre, J.P. Remon, The influence of formulation on the relative bioavailability of indomethacin suppositories in dogs and rabbits **104**, 1
Study of the bioavilability of four indomethacin suppository formulations in healthy volunteers **104**, 87
De Muynck, C., J.P. Remon, Influence of fat composition on the melting behaviour and on the in vitro release of indomethacin suppositories **85**, 103
De Muynck, C., see Bottenberg, P. **49**, 147; **58**, 203
De Muynck, C., see de Vroe, C. **65**, 49
De Muynck, C., see Vandenbossche, G.M.R. **67**, 195
De Oliveira, C, see Da Silva, C.P. **111**, 25
De Pablos, Pon, see Gonzalez, A.G. **67**, R1
De Ponti, R., see Martini, A. **75**, 141; **127**, 239
De Ponti, R., see Muggetti, L. **136**, 81
De Ponti, R., see Torricelli, C. **71**, 19; **75**, 147
De Ronde, H.A., see Van Amerongen, I.A. **86**, 9
De Schmidt, J., see Grit, M. **50**, 1
De Smedt, S.C., see Ntawukulilyayo, J.D. **128**, 73
De Smidt, J.H., D.J.A. Crommelin, Viscosity measurement in aqueous polymer solutions by dynamic light scattering **77**, 261
De Smidt, J.H., J.C.A. Offringa, D.J.A. Crommelin, Dissolution kinetics of theophylline in aqueous polymer solutions **77**, 255
De Verdiere, A., see Nemati, F. **102**, 55; **138**, 237

De Vilder, J., see Herman, J. **56**, 51
De Villiers, C., see Munday, D.L. **69**, 123; **73**, 89
De Villiers, M., Influence of agglomeration of cohesive particles on the dissolution behaviour of furosemide powder **136**, 175
De Villiers, M., J. Van der Watt, A.P. Lotter, Kinetic study of the solid-state photolytic degradation of two polymorphic forms of furosemide **88**, 275
De Vincentis, see Wesdyk, R. **93**, 101
De Vos, A., L. Vervoort, R. Kinget, Solubilization and stability of indomethacin in a transparent oil–water gel **92**, 191
De Vries-Nijb, see Schoonen, A.J.M. **47**, 249; **49**, 261
De Vries, H., G.M.J. Beijersbergen Van Henegouwen, F.A. Huf, Photochemical decomposition of chloramphenicol in a 0.25% eyedrop and in a therapeutic intraocular concentration **20**, 265
De Vries, H., see Beijersbergen Van Henegouwen, G.M.C. **35**, 259
De Vries, J.D., J. Winkelhorst, W.J.M. Underberg, R.E.C. Henrar, J.H. Beijnen, A systematic study on the chemical stability of the novel indoloquinone antitumour agent EO9 indoledione◇ **100**, 181
De Vries, M., H.E. Bodde, J.C. Verhoef, M. Ponec, W.I.H.M. Craane, H.E. Junginger, Localization of the permeability barrier inside porcine buccal mucosa: A combined in vitro study of drug permeability, electrical resistance and tissue morphology **76**, 25
De Vries, M., see De, Vries M. **76**, 25
De Vries, M.E., see Le Brun, P.P.H. **49**, 141
De Vringer, T., see de Haan, F.H.N. **13**, 75
De Vroe, C., C. de Muynck, J.P. Remon, M. Samsom, A study on the stability of three antineoplastic drugs and on their sorption by i.v. delivery systems and end-line filters **65**, 49
De Zeeuw, D., see Franssen, E.J.F. **80**, R15
De Zeeuw, R.A., see Jonkman, J.H.G. **8**, 153; **10**, 177
De Zeeuw, R.A., see Swart, P.J. **87**, 67; **88**, 165
De Zeeuw, R.A., see Wiechers, J.W. **47**, 43; **65**, 77; **66**, 53
Dean, A.M., see Belloto, Jr. R.J. **23**, 195
Deasy, P.B., M.P. Finan, P.R. Klatt, T. Hornykiewytsch, Design and evaluation of a biodegradable implant for improved delivery of oestradiol-17β to steers **89**, 251
Deasy, P.B., M.P. Gouldson, In vitro evaluation of pellets containing enteric coprecipitates of nifedipine formed by non-aqueous spheronization **132**, 131
Deasy, P.B., M.F.L. Law, Use of extrusion-spheronization to develop an improved oral dosage form of indomethacin **148**, 201
Deasy, P.B., K.J. Quigley, Rheological evaluation of deacetylated gellan gum (Gelrite) for pharmaceutical use **73**, 117
Deasy, P.B., see Berney, B.M. **3**, 73
Deasy, P.B., see Brophy, M.R. **29**, 223; **37**, 41
Deasy, P.B., see Collins, A.E.M. **51**, 103
Deasy, P.B., see Hilton, A.K. **86**, 79
Deasy, P.B., see Law, M.F.L. **146**, 1
Deasy, P.B., see McDaid, D.M. **133**, 71
Deasy, P.B., see O'Neill, C.T. **48**, 247
Dechesne, J.P., A new enteric tablet of acetylsalicylic acid. I. Technological aspects **37**, 203
Dechesne, J.P., L. Delattre, A new enteric tablet of acetylsalicylic acid. II. Biopharmaceutical aspects **34**, 259
Decroix, M.O., see Benkhelifa, S. **145**, 115
Decroix, M.O., see Crauste-Manciet, S. **149**, 241
Decroix, M.O., see Penaud, J.F. **142**, 1
Deering, A., see Harron, D.W.G. **57**, 9
DeGrazia, F., see Huselton, C.A. **134**, 157
Deguchi, Y., see Ichimura, F. **27**, 45
Deguchi, Y., see Terasaki, T. **81**, 143
Deharo, E., see Mbela, T.K.M.N. **110**, 189
Dehpour, A.R., M.E. Zolfaghari, T. Samadian, F. Kobarfard, M. Faizi, M. Assari, Antiulcer activities of liquorice and its derivatives in experimental gastric lesion induced by ibuprofen in rats **119**, 133
Dekker, B.G., see Riepma, K.A. **87**, 149; **90**, R1
Del Cerro, J., see Fernandez-Hervas, M.J. **113**, 39
Del Estal, J.L., A.I. Alvarez, C. Villaverde, A. Justel, J.G. Prieto, Increased systemic bioavailability of albendazol when administered with surfactants in rats **102**, 257
Del Estal, J.L., A.I. Alvarez, C. Villaverde, J.G. Prieto, Comparative effects of anionic, natural bile acid surfactants and mixed micelles on the intestinal absorption of the anthelmintic albendazole **91**, 105
Del Olmo, E., I. Toth, A.N. Fonteh, W.A. Gibbons, Lipid methyl transferase inhibitory activity of novel α-amino alkylic acid derivatives **81**, 267
Del Olmo Fernandez, E., see Toth, I. **79**, 39; **83**, 131
Del Pozo, A., see Halbaut, L. **130**, 203
Del Rosario Aberturas, M., see Molpeceres, J. **130**, 75
Delalonde, M., B. Bataille, G. Baylac, J. Maurice, R. Sabatier, Definition of indices for the mechanical design of wet powders: Application to the study of a natural polymer, microcrystalline cellulose **146**, 159
Delalonde, M., G. Baylac, B. Bataille, M. Jacob, A. Puech, The rheology of wet powders: A measuring instrument, the compresso-rheometer **130**, 147
Delatour, P., see Walchshofer, N. **77**, 265
Delattre, J., see Sponton, E. **23**, 299
Delattre, J., see Weingarten, C. **26**, 251
Delattre, L., see Dechesne, J.P. **34**, 259
Delgado, A., C. Evora, M. Llabres, Optimization of 7-day release (in vitro) from DL-PLA methadone microspheres **134**, 203
Degradation of DL-PLA-methadone microspheres during in vitro release **140**, 219
Delgado, A., see Gallardo, V. **71**, 223
Delgado, A., see Munguia, O. **86**, 107
Delgado-Charro, B., see Garcia-Gonzalez, N. **100**, 25, 65; **104**, 107
Delgado-Charro, M.B., R.H. Guy, Iontophoretic delivery of nafarelin across the skin **117**, 165
Delgado-Charro, M.B, Jato J.L. Vila, In vivo study of sustained-release formulations containing amoxicillin and Gelucire 64-02 **78**, 35
Delie, F., see Blanco, Prieto M.J. **111**, 137

Delie, F., Y. Letourneux, D. Nisato, F. Puisieux, P. Couvreur, Oral administration of peptides: Study of a glycerolipidic prodrug **115**, 45
Dellerman, K.M., see Pikal, M.J. **50**, 233
Delmonte, G., see Saettone, M.F. **43**, 67
DeLuca, P.P., see Lee, K.C. **106**, 167; **114**, 215
DeLuca, P.P., see Tsai, T. **127**, 31, 43
Delucca, C., see Dal, Pozzo A. **70**, 219
D'Emanuele, A., R. Dinarvand, Preparation, characterisation, and drug release from thermoresponsive microspheres **118**, 237
Erratum: Preparation, characterisation, and drug release from thermoresponsive microspheres (Int. J. Pharm. 118 (1995) 237) **130**, 261
D'Emanuele, A., J.N. Staniforth, Effects of polymer formulation on electrophoretic drug delivery Iontophoretic transport of verapamil and melatonin. I. Cellophane membrane as a barrier **92**, 225
D'Emanuele, A., see Martini, L. **113**, 223
Demeester, J., see Jurima-Romet, M. **63**, 227
Demeester, J., see Kiekens, A. **146**, 239
Demeester, J., see Ntawukulilyayo, J.D. **128**, 73
Demuynck, C., see Ntawukulilyayo, J.D. **121**, 205
Denizot, B., see Gallardo, M. **100**, 55
Denman, S., see Billany, M.R. **124**, 279
Dennis, A.B., S.J. Farr, I.W. Kellaway, G. Taylor, R. Davidson, In vivo evaluation of rapid release and sustained release Gelucire capsule formulations **65**, 85
DeNoto, A.R., see Thombre, A.G. **111**, 181
Denyer, S.P., R.H. Guy, J. Hadgraft, W.B. Hugo, The microbial degradation of topically applied drugs **26**, 89
Denyer, S.P., W.B. Hugo, V.D. Harding, Synergy in preservative combinations **25**, 245
The biochemical basis of synergy between the antibacterial agents, chlorocresol and 2-phenylethanol **29**, 29
Denyer, S.P., see Buckton, G. **73**, 1
Denyer, S.P., see Washington, C. **39**, 33
Depraetere, E., see Bonhomme, L. **80**, 271
Derendorf, H., G. Drehsen, P. Rohdewald, In vivo-in vitro correlations of salicylate saliva levels and continuous flow cell dissolution rates **15**, 167
Derendorf, H., see Dietzel, K. **59**, 225
Derendorf, H., see Schreier, H. **87**, 183
Desager, J.P., M. Sclavons, H. Meunier, C. Harvengt, Pharmacokinetics and disposition of nifedipine and acebutolol as a fixed combination or given separately after single and repeated dose in healthy subjects **31**, 179
Desai, D.S., M.A. Abdelnasser, B.A. Rubitski, S.A. Varia, Photostabilization of uncoated tablets of sorivudine and nifedipine by incorporation of synthetic iron oxides **103**, 69
Desai, D.S., S.A. Ranadive, R. Lozano, S.A. Varia, Dissolution instability of encapsulated marketed tablets **144**, 153
Desai, D.S., B.A. Rubitski, J.S. Bergum, S.A. Varia, Effects of various formulation factors on dissolution stability of aztreonam, hydrochlorothiazide, and sorivudine capsules **110**, 249
Effects of different types of lactose and disintegrant on dissolution stability of hydrochlorothiazide capsule formulations **110**, 257
Desai, D.S., B.A. Rubitski, S.A. Varia, A.W. Newman, Physical interactions of magnesium stearate with starch-derived disintegrants and their effects on capsule and tablet dissolution **91**, 217
Desai, D.S., B.A. Rubitski, S.A. Varia, M.H. Huang, Effect of formaldehyde formation on dissolution stability of hydrochlorothiazide bead formulations **107**, 141
Desai, D.S., B.A. Rubitski, S.A. Varia, N.B. Jain, Povidone- and poloxamer-mediated degradation of hydrochlorothiazide in an antihypertensive combination tablet product **142**, 61
Deshmukh, A.A., see Yuen, K.H. **97**, 61
Desjardins, R.E., see Peck, C.C. **82**, 9
Deutsch, J., see Touitou, E. **103**, 199
Devarajan, P.V., see Mandlekar, S.V. **148**, 117
Deverre, J.R., see Ramaldes, A. **143**, 1
Devi, K.P., see Baveja, S.K. **39**, 39; **47**, 133
Devine, K.R., see Collier, P.S. **1**, 3
Devine, S., see Harron, D.W.G. **57**, 9
Devissaguet, J.P., see Fessi, H. **55**, R1
Devissaguet, J.P., see Gursoy, A. **52**, 101
Devissaguet, J.P., see Guterres, S.S. **113**, 57
Devissaguet, J.P., see Masson, V. **139**, 113
Devissaguet, J.P., see Rodrigues, Jr. J.M. **126**, 253
Devissaguet, J.P., see Tabosa, de Egito E. **145**, 17
Devissaguet, J.P., see Thioune, O. **146**, 233
Devissaguet, J.P., see Yu, W. **89**, 139
Devissauguet, J.P., see Belbella, A. **129**, 95
Devleeschouwer, M.J., P. Boussard, P. Momin, J. Dony, Difference of *Pseudomonas aeruginosa* sensitivity to chloroxylenol according to the culture medium **32**, 55
Devleeschouwer, M.J., J. Dony, An in vitro test for the evaluation of the efficacy of disinfectants **9**, 49
Devleeschouwer, M.J., see Boussard, P. **32**, 91; **53**, 253; **72**, 51
Devleeschouwer, M.J., see Dony, J. **19**, 239
Devleeschouwer, M.J., see Ibwala, N. **59**, 205
Devleeschouwer, M.J., see Vincke, B.J. **21**, 265; **32**, 61
Devleeschouwer, N., see Chamart, S. **99**, 59
Dewsbury, N.J., C.J. Kenyon, S.P. Newman, The effect of handling techniques on electrostatic charge on spacer devices: A correlation with in vitro particle size analysis **137**, 261
Dey, M., R. Enever, M. Marino, J. Michelucci, D. Smith, R. Warner, R. Weierstall, Sustained-release etodolac bioavailability and dose proprotionally: Correlation between in vivo and in vitro performance **49**, 121
Dey, M.J., see Irwin, W.J. **113**, 25
Deyrup, M., see Brewster, M.E. **68**, 215
Dezentje, R.F.R., see Van, Bommel E.M. **72**, 19
D'Hermies, F., see Cohen, T. **137**, 249
Dhillon, S., see Podczeck, F. **142**, 235
Dhodi, N., see Menon, A. **140**, 207

Di Colo, G., C. Giannessi, E. Nannipieri, M.F. Serafini, D. Vitale, Influence of drug–surfactant and skin–surfactant interactions on percutaneous absorption of two model compounds from ointment bases in vitro **50**, 27
Di Colo, G., V. Carelli, E. Nannipieri, Effect of water-soluble additives on drug release from silicone rubber matrices. II. Sustained release of prednisolone from non-swelling devices **30**, 1
Di Colo, G., see Bilia, A. **130**, 83
Di Colo, G., see Bonina, F.P. **100**, 41
Di Colo, G., see Bottari, F. **2**, 63
Di Colo, G., see Buonaguidi, M. **147**, 1
Di Colo, G., see Carelli, V. **30**, 9; **35**, 21; **50**, 181; **55**, 199; **88**, 89; **89**, 81; **94**, 103
Di Giulio, A., A. Barracchini, L. Cellai, C. Martuccio, G. Amicosante, A. Oratore, G. Pantaleoni, Rifamycins as inhibitors of collagenase activity: Their possible pharmacological role in collagen degradative diseases **144**, 27
Di Giulio, A., G. Maurizi, P. Odoardi, M.A. Saletti, G. Amicosante, A. Oratore, Encapsulation of ampicillin in reverse-phase evaporation liposomes: A direct evaluation by derivative spectrophotometry **74**, 183
Di Giulio, A., M.A. Saletti, A. Impagnatiello, M. Lucarelli, R. Strom, A. Oratore, Encapsulation and efflux of lactone and hydroxy acid forms of simvastatin in reverse-phase evaporation vesicles **89**, 133
Di Giuseppe, E., see Carlucci, G. **93**, 245; **102**, 271
Di Martino, P., A.M. Guyot-Hermann, P. Conflant, M. Drache, J.C. Guyot, A new pure paracetamol for direct compression: The orthorhombic form **128**, 1
Di Pietra, A.M., V. Cavrini, M.A. Raggi, Determination of benzaldehyde traces in benzyl alcohol by liquid chromatography (HPLC) and derivative UV spectrophotometry **35**, 13
Di Pietra, A.M., see Cavrini, V. **10**, 119; **13**, 333
Di Pietra, A.M., see Gilbert, J.C. **40**, 93
Diallo, B., see Beten, D.B. **88**, 31
Dick, C.F., see Antal, E.J. **54**, 33
Dickinson, P.A., I.W. Kellaway, G. Taylor, D. Mohr, K. Nagels, H.M. Wolff, In vitro and in vivo release of estradiol from an intra-muscular microsphere formulation **148**, 55
Dickinson, P.A., G. Taylor, Pulmonary and blood enzyme kinetics as predictors of pulmonary metabolism of esters **116**, 231
Diepold, R., J. Kreuter, P. Guggenbuhl, J.R. Robinson, Distribution of poly-hexyl-2-cyano-[3-^{14}C]acrylate nanoparticles in healthy and chronically inflamed rabbit eyes **54**, 149
Dietzel, K., K.S. Estes, M.E. Brewster, N.S. Bodor, H. Derendorf, The use of 2-hydroxypropyl-β-cyclodextrin as a vehicle for intravenous administration of dexamethasone in dogs **59**, 225
Diez-Sales, O., A. Copovi, V.G. Casabo, M. Herraez, A modelistic approach showing the importance of the stagnant aqueous layers in in vitro diffusion studies, and in vitro–in vivo correlations **77**, 1
Diez-Sales, O., E. Perez-Sayas, A. Martin-Villodre, M. Herraez-Dominguez, The prediction of percutaneous absorption. I. Influence of the dermis on in vitro permeation models **100**, 1
Diez-Sales, O., A.C. Watkinson, M. Herraez-Dominguez, C. Javaloyes, J. Hadgraft, A mechanistic investigation of the in vitro human skin permeation enhancing effect of Azone^{R} **129**, 33
Diez-Sales, O., see Lopez, A. **139**, 205
Digenis, G.A., see Fell, J.T. **22**, 1
Dighe, S., see Shah, V.P. **82**, 1
Dimitriou, A., see Malamataris, S. **68**, 51
Dimitrova, E., S. Bogdanova, E. Minkov, N. Manolova, I. Raschkov, M. Koleva, S. Mihailova, High-molecular weight polyoxyethelene as an additive in ophthalmic solutions **93**, 21
Dimitrovska, A., B. Andonovski, K. Stojanoski, Spectrophotometric study of copper(II) ion complexes with cefaclor **134**, 213
Dimitrovska, A., K. Stojanoski, K. Dorevski, Kinetics of degradation of cefaclor. I. Effects of temperature, phosphate buffer, pH and copper(II) ion on the rate of degradation **115**, 175
Dinarvand, R., see D'Emanuele, A. **118**, 237; **130**, 261
Dinculescu, A., see Brewster, M.E. **125**, 17
Dine, T., M. Luyckx, J.C. Cazin, C. Brunet, M. Cazin, F. Goudaliez, M.L. Mallevais, Stability and compatibility studies of vinblastine, vincristine, vindesine and vinorelbine with PCV infusion bags **77**, 279
Dine, T., see Benaji, B. **105**, 83
Dine, T., see Faouzi, M.A. **89**, 125; **105**, 89
Dine, T., see Gressier, B. **104**, 151
Dine, T., see Khalfi, F. **139**, 243
DiNenno, B.K., see Randall, C.S. **120**, 235
Dinesh Babu, K.V., see Anto, R.J. **131**, 1
DiPiro, J.T., see Bayoumi, S.M. **30**, 57
Dirix, C.A.M.C., see Ijntema, K. **112**, 215
Dittert, L., see Tawfiq, F.A. **64**, 45
Dittert, L.W., see Chungi, V.S. **4**, 27
Dittert, L.W., see Jona, J.A. **123**, 127
Dittgen, M., see Lukowski, G. **84**, 23
Dittrich, M., see Hampl, J. **144**, 107
Djordjevic, N.M., G. Rohr, M. Hinterleitner, B. Schreiber, Adsorption of water on cyclosporin A, from zero to finite surface coverage **81**, 21
Djuric, M., M. Jovanovic, Z. Djuric, Influence of drug amount and dioctyl sodium sulfosuccinate on dissolution of propyphenazone tablets **37**, 179
Djuric, Z., see Jovanovic, M. **75**, 155
Djuric, Z., see Rakic, R. **39**, 47
D'Mello, A., see D'Souza, M. **31**, 165
Dobetti, L., N. Mazzei, A. Grassano, R. Forster, P. Cardillo, Explosive properties of a nitroderivative compound (ITF 296), in bulk and mixed with anhydrous dibasic calcium phosphate: Involving a formulation approach **128**, 277
Dobner, B., see Schneider, I.M. **145**, 187
Dobrilovic, L., see Jacobs, G.P. **32**, 151
Dodge, J., see Singh, S.K. **125**, 243
Doelker, E., see Allemann, E. **87**, 247

Doelker, E., see Bitz, C. **131**, 171
Doelker, E., see Gander, B. **27**, 117; **58**, 63
Doelker, E., see Ibrahim, H. **77**, 211; **87**, 239
Doelker, E., see Korsmeyer, R.W. **15**, 25
Doelker, E., see Nook, T. **43**, 119
Doelker, E., see Quintanar-Guerrero, D. **143**, 133
Doelker, E., see Rouge, N. **136**, 117
Doherty, C., P. York, Mechanisms of dissolution of frusemide/PVP solid dispersions **34**, 197
Frusemide crystal forms; solid state and physicochemical analyses **47**, 141
Microenvironmental pH control of drug dissolution **50**, 222
Doherty, C., see Dove, J.W. **138**, 199
Doherty, M.M., P.J. Hughes, S.R. Kim, D.E. Mainwaring, W.N. Charman, Effect of lyophilization on the physical characteristics of medium molecular mass hyaluronates **111**, 205
Doko, M.B., V. Bassani, J. Casadebaig, L. Cavailles, M. Jacob, Preparation of proteolytic enzyme extracts from *Ananas comosus* L., Merr. fruit juice using semipermeable membrane, ammonium sulfate extraction, centrifugation and freeze-drying processes **76**, 199
Dolan, T.F., see Baillie, A.J. **57**, 23
Dolan, T.F., see Carter, K.C. **53**, 129
Doldan, C., C. Souto, A. Concheiro, R. Martinez-Pacheco, J.L. Gomez-Amoza, Dicalcium phosphate dihydrate and anhydrous dicalcium phosphate for direct compression: A comparative study **124**, 69
Dollo, G., P. Le Corre, F. Chevanne, R. Le Verge, Inclusion complexation of amide-typed local anaesthetics with 13-cyclodextrin and its derivatives. I. Physicochemical characterization **131**, 219
Inclusion complexation of amide-typed local anaesthetics with β-cyclodextrin and its derivatives. II. Evaluation of affinity constants and in vitro transfer rate constants **136**, 165
Domb, A.J., Long acting injectable oxytetracycline-liposphere formulations **124**, 271
Domenech, Berrozpe J, see Calpena, A.C. **103**, 179
Dominguez-Gil, A., see Alonso, I.G. **19**, 345
Dominguez-Gil, A., see Andersen, F.M. **19**, 189
Dominguez-Gil, A., see Calvo, M.B. **67**, 259
Dominguez-Gil, A., see Calvo, M.V. **16**, 215; **38**, 117
Dominguez-Gil, A., see Garcia, M.J. **5**, 117
Dominguez-Gil, A., see Lanao, J.M. **86**, 123
Dominguez-Gil, A., see Marino, E.L. **8**, 25; **9**, 263; **12**, 209; **16**, 23
Dominguez-Gil, A., see Navarro, A.S. **37**, 113
Dominguez-Gil, A., see Pedraz, J.L. **25**, 147
Dominguez-Gil, A., see Sanchez Camazano, M. **6**, 243
Dominguez-Gil, A.A., see Garcia, M.J. **5**, 117
Dominguez-Gil, Hurle A., see Sanchez, B.M. **106**, 229
Donaldson, J.D., M.S. Roberts, A.E. Polack, Uptake of solutes by plastic infusion tubing. Mathematical solutions **88**, 351
Donato, I., see Giammona, G. **64**, 239
Donbrow, M., see Benita, S. **12**, 251
Donbrow, M., see Gross, S.T. **29**, 213
Donbrow, M., see Hoffman, A. **29**, 195
Donbrow, M., see Touitou, E. **9**, 97
Donbrow, M., see Touitou, E. **11**, 131, 355; **15**, 13
Dondeti, P., H. Zia, T.E. Needham, In vivo evaluation of spray formulations of human insulin for nasal delivery **122**, 91
Bioadhesive and formulation parameters affecting nasal absorption **127**, 115
Dondi, G., see Catellani, P.L. **88**, 285
Dongowski, G., R. Neubert, H. Haase, B. Schnorrenberger, Interactions between food components and drugs. Part 4. Influence of pectins and bile salts on propranolol absorption **144**, 233
Donnelly, A., I.W. Kellaway, S.J. Farr, G. Taylor, N. Tudball, M. Gibson, The influence of lipophilicity upon the nasal absorption of a series of hexapeptides **135**, 191
D'Onofrio, G.P., R.C. Oppenheim, N.E. Bateman, Encapsulated microcapsules **2**, 91
Donovan, M.D., M. Zhou, Drug effects on in vivo nasal clearance in rats **116**, 77
Donovan, M.D., see Bhat, P.G. **126**, 179; **134**, 15
Donovan, M.D., see Chung, F.Y. **128**, 229
Donovan, M.D., see Miller, S.C. **12**, 147
Donovan, M.D., see Zhou, M. **130**, 93; **135**, 115
Dony, J., M.J. Devleeschouwer, P. Momin, Comparison of the sensitivity of *Pseudomonas aeruginosa* to disinfectants according to the growth conditions **19**, 239
Dony, J., see Boussard, P. **32**, 91; **53**, 253; **72**, 51
Dony, J., see Devleeschouwer, M.J. **9**, 49; **32**, 55
Dony, J., see Ibwala, N. **59**, 205
Dony, J., see Vincke, B.J. **21**, 265; **32**, 61
Donzelli, G., see Dal, Pozzo A. **50**, 97
Doornbos, D.A., see Van, der Voet H. **14**, 291
Dor, P., see Benita, S. **33**, 71
Dordunoo, S.K., H.M. Burt, Solubility and stability of taxol: Effects of buffers and cyclodextrins **133**, 191
Dorevski, K., see Dimitrovska, A. **115**, 175
Dorle, A.K., see Labhasetwar, V.D. **47**, 261
Dorle, A.K., see Pathak, Y.V. **24**, 351
Dorle, A.K., see Ramani, C.C. **137**, 11
Dorle, A.K., see Sheorey, D.S. **68**, 19
Dormish, J.J., see Foster, T.M. **134**, 193
Dorta, M.J., O. Munguia, M. Llabres, Effects of polymerization variables on PLGA properties: Molecular weight, composition and chain structure **100**, 9
Doucet, J., see Ferreira, L.A.M. **121**, 169
Douglas, S.J., S.S. Davis, L. Illum, Biodistribution of poly(butyl 2-cyanoacrylate) nanoparticles in rabbits **34**, 145
Dove, J.W., G. Buckton, C. Doherty, A comparison of two contact angle measurement methods and inverse gas chromatography to assess the surface energies of theophylline and caffeine **138**, 199
Down, G.R.B., see Baert, L. **107**, 219
Down, G.R.B., see Kwong, E.C. **103**, 259
Down, G.R.B., see Mitchell, A.G. **22**, 337

Dowse, R., Singh N. Rajendraprasad, I. Kanfer, Pharmacokinetics of phenylpropanolamine in humans after a single-dose study **39**, 141
Dowse, R., see Fassihi, A.R. **50**, 79; **72**, 175
Dowty, M.E., P. Braquet, Effect of extracellular pH on cytoplasmic pH and mechanism of pH regulation in cultured bovine corneal endothelium: Possible importance in drug transport studies **68**, 231
Dowty, M.E., K.E. Knuth, J.R. Robinson, Enzyme characterization studies on the rate-limiting barrier in rabbit buccal mucosa **88**, 293
Doyle, R., see Adjei, A. **61**, 135
Drabik, B.R., see Miller, S.C. **18**, 269
Drache, M., see Di Martino, P. **128**, 1
Drache, M., see Fachaux, J.M. **99**, 99
Dredan, J., I. Antal, I. Racz, Evaluation of mathematical models describing drug release from lipophilic matrices **145**, 61
Drehsen, G., see Derendorf, H. **15**, 167
Drenth, B.F.H., see Wiechers, J.W. **47**, 43; **65**, 77; **66**, 53
Drew, P., see Bateman, S.D. **49**, 209
Droin, A., C. Chaumat, M. Rollet et al., Model for matter transfers between sodium salicylate-Eudragit matrix and gastic liquid **27**, 233
Droin, A., see Eddine, A. **32**, 143
Drouin, D., see Sponton, E. **23**, 299
Drustrup, J., A. Fullerton, L. Christrup, H. Bundgaard, Utilization of prodrugs to enhance the transdermal absorption of morphine **71**, 105
Drustrup, Larsen, J., H. Bundgaard, V.H.L. Lee, Prodrug forms for the sulfonamide group. II. Water-soluble amino acid derivatives of *N*-methylsulfonamides as possible prodrugs **47**, 103
D'Silva, J.B., R.E. Notari, Kinetics and mechanisms of aqueous degradation of the anticancer agent, indicine *N*-oxide **8**, 45
D'Souza, M., R. Venkataramanan, A. D'Mello, P. Niphadkar, An alternative prodrug approach for reducing presystemic metabolism of drugs **31**, 165
Du, J., see Rao, Jasti B. **118**, 161
Du Plessis, J., C. Ramachandran, N. Weiner, D.G. Muller, The influence of particle size of liposomes on the deposition of drug into skin **103**, 277
Du Plessis, J., C. Ramachandran, N. Weiner, D.G. Muller, The influence of lipid composition and lamellarity of liposomes on the physical stability of liposomes upon storage **127**, 273
Du Plessis, J., L.R. Tiedt, Wyk C.J. Van, C. Ackermann, A new transmission electron microscope method for the determination of particle size in parenteral fat emulsions **34**, 173
Du Plessis, J., N. Weiner, D.G. Muller, The influence of in vivo treatment of skin with liposomes on the topical absorption of a hydrophilic and a hydrophobic drug in vitro **103**, R1
Du Reau, C., see Ward, A.J.I. **74**, 137
Dua, R., H. Zia, T. Needham, The influence of tonicity and viscosity on the intranasal absorption of salmon calcitonin in rabbits **147**, 233
Dubernet, C., J.P. Benoit, G. Couarraze, D. Duchene, Microencapsulation of nitrofurantoin in poly(ε-caprolactone): Tableting and in vitro release studies **35**, 145
Dubernet, C., J.C. Rouland, J.P. Benoit, Comparative study of two ethylcellulose forms (raw material and microspheres) carried out through thermal analysis **64**, 99
Dubernet, C., see Nemati, F. **102**, 55; **138**, 237
Dubois, J.G., M. Hanocq, G. Atassi, Extraction and assay of daunorubicin and its metabolites from neoplastic tissue **35**, 219
Dubois, J.L., see Bernard, E. **126**, 235
Dubois, J.L., see Ford, J.L. **28**, 11
Dubrovsky, J., see Bialer, M. **20**, 53; **23**, 25
Duchateau, G.S.M.J.E., H. Bundgaard, S.W.J. Basseleur, Influence of some surface-active agents on nasal absorption in rabbits **39**, 87
Duchateau, G.S.M.J.E., J. Zuidema, W.M. Albers, F.W.H.M. Merkus, Nasal absorption of alprenolol and metoprolol **34**, 131
Duchateau, G.S.M.J.E., J. Zuidema, W.A.J.J. Hermens, F.W.H.M. Merkus, In vitro ciliotoxicity of propranolol isomers **31**, 275
Duchateau, G.S.M.J.E., J. Zuidema, F.W.H.M. Merkus, Bile salts and intranasal drug absorption **31**, 193
Duchateau, G.S.M.J.E., see Zuidema, J. **47**, 1
Duchene, D., see Amdidouche, D. **54**, 175; **111**, 111
Duchene, D., see Benkerrour, L. **19**, 27
Duchene, D., see Dubernet, C. **35**, 145
Duchene, D., see Irache, J.M. **90**, R9
Duchene, D., see Lemesle-Lamache, V. **141**, 117
Duchene, D., see Leroy-Lechat, F. **101**, 97
Duchene, D., see Lin, S.Z. **69**, 211; **106**, 63
Duchene, D., see Ponchel, G. **38**, 65
Duchene, D., see Skiba, M. **120**, 1; **126**, 275; **129**, 113; **142**, 121; **145**, 241
DuCret, R., see Agyilirah, G.A. **75**, 241
Duda, E., see Mori, A. **131**, 57
Duddu, S.P., N.G. Das, T.P. Kelly, T.D. Sokoloski, Microcalorimetric investigation of phase transitions. I. Is water desorption from theophylline•HOH a single-step process? **114**, 247
Duddu, S.P., F.K.Y. Fung, D.J.W. Grant, Effects of crystallization in the presence of the opposite enantiomer on the crystal properties of (*SS*)-(+)-pseudoephedrinium salicylate **127**, 53
Effect of the opposite enantiomer on the physicochemical properties of (−)-ephedrinium 2-naphthalenesulfonate crystals **94**, 171
Due Larsen, B., see Taub, M.E. **146**, 205
Duensing, G., see Brand, R.M. **146**, 115
Duewell, S., see Schwendener, R.A. **49**, 249
Duffield, J.R., see Hall, S.B. **70**, 251
Duffy, T.D., see Ford, J.L. **12**, 11

Dugard, P.H., R.C. Scott, A method of predicting percutaneous absorption rates from vehicle to vehicle: An experimental assessment **28**, 219
Dugard, P.H., see Walters, K.A. **10**, 153
Dugue, J., see Ceolin, R. **74**, 77
Duleba, B., see Bonhomme, L. **65**, R9
Dumic, M., see Jaksic, P. **135**, 177
Dumont, P., see Mbela, T.K.M. **79**, 29; **92**, 71
Dumont, S., see Goldbach, P. **123**, 33
Dumortier, G., M. Zuber, F. Chast, P. Sandouk, J.C. Chaumeil, Systemic absorption of morphine after ocular administration: Evaluation of morphine salt insert in vitro and in vivo **59**, 1
Duncan, M.R., J.M. Lee, M.P. Warchol, Influence of surfactants upon protein/peptide adsorption to glass and polypropylene **120**, 179
Duncan, M.R., see Pujara, C.P. **114**, 197
Duncan, R., see Al-Shamkhani, A. **122**, 107
Duncan, R., see Bridges, J.F. **44**, 213
Duncan, R., see Carreno-Gomez, B. **148**, 231
Duncan, R., see Gianasi, E. **148**, 139
Duncan, R., see Hudson, A.J. **133**, 257
Duncan, R., see Morgan, S.M. **128**, 99
Duncan, R., see Pato, A.J. **104**, 227
Duncan-Hewitt, W.C., D.J.W. Grant, True density and thermal expansivity of pharmaceutical solids: Comparison of methods and assessment of crystallinity **28**, 75
Duncan-Hewitt, W.C., see Lin, M.C. **106**, 187
Dundalek, K., see Bernkop-Schnurch, A. **138**, 75
Durham, D.G., C.T. Hung, R.B. Taylor, Identification of some acids produced during autoclaving of D-glucose solutions using HPLC **12**, 31
Durham, D.G., see Taylor, R.B. **26**, 259
Durig, T., A.R. Fassihi, Preformulation study of moisture effect on the physical stability of pyridoxal hydrochloride **77**, 315
Identification of stabilizing and destabilizing effects of excipient–drug interactions in solid dosage form design **97**, 161
Durmaz, G., see Akbuga, J. **111**, 217
Duroux, J.L., see Basly, J.P. **139**, 219; **142**, 125, 247
Durr, M., see Grove, J. **66**, 23
Durrani, A.M., N.M. Davies, M. Thomas, I.W. Kellaway, Pilocarpine bioavailability from a mucoadhesive liposomal opthalmic drug delivery system **88**, 409
Durrani, A.M., S.J. Farr, I.W. Kellaway, Influence of molecular weight and formulation pH on the precorneal clearance rate of hyaluronic acid in the rabbit eye **118**, 243
Durrani, M.J., A. Kusai, N.F.H. Ho et al., Topical vaginal drug delivery in the guinea pig. I. Effect of estrous cycle on the vaginal membrane permeability of vidarabine **24**, 209
Durrani, M.J., see Neau, S.H. **131**, 47
Durrani, M.J., see Singh, S.K. **125**, 243
Durrer, C., see Irache, J.M. **90**, R9
Durrheim, H.H., see Bluhm, D.P. **68**, 277, 281
Duru, C., see Raffin, F. **120**, 205; **145**, 247
Duru, C., see Rizk, S. **126**, 57
Duru, C., see Ruizk, S. **112**, 125
Duval, O., see Pech, B. **128**, 179
Dvis, S.S., see Nankervis, R. **101**, 249
Dwivedi, S.K., S. Sattari, F. Jamali, A.G. Mitchell, Ibuprofen racemate and enantiomers: Phase diagram, solubility and thermodynamic studies **87**, 95
Dwivedi, S.K., see Tricehurst, M.D. **141**, 93
Dwyer, M., see Barnwell, S.G. **89**, 245
Dwyer, M., see Cole, S.K. **80**, 63
Dzurik, R., see Piotrovskij, V.K. **119**, 213

E

Easthope, D.A., see Bowtle, W.J. **4**, 263
Ebert, C., see Zhang, J. **101**, 15
Eberth, K., J. Merry, A comparative study of emulsions prepared by ultrasound and by a conventional method. Droplet size measurements by means of a Coulter counter and microscopy **14**, 349
Eberth, K., see Merry, J. **19**, 43
Ebian, A.R., see Nada, A.H. **53**, 175
Ebian, A.R., see Shihab, F.A. **4**, 13
Eboka, C.J., K.I. Adesanya, Assay of drugs in the presence of spectral interferants in tablets **51**, 161
Spectrophotometric assay of drugs in the presence of spectral interferants in suspensions and syrups **51**, 263
Eccleston, G.M., Phase transitions in ternary systems and oil-in-water emulsions containing cetrimide and fatty alcohols **27**, 311
Eccleston, G.M., see Pryce-Jones, R.H. **86**, 231
Economou, P.T., see Issopoulos, P.B. **57**, 235
Eddine, A., A. Droin, J.L. Taverdet, J.M. Vergnaud, Effect of pH on drug release between sodium salicylate-Eudragit compound and gastric liquid: Modelling of the process **32**, 143
Eddy, E.P., C. Wood, J. Miller, G. Wilson, I.J. Hidalgo, A comparison of the affinis of dipeptides and antibiotics for the di-/tripeptide transporter in Caco-2 cells **115**, 79
Edelstein, M., see Pedersen, M. **90**, 247
Edgar, B., see Wingstrand, K. **60**, 151
Edgar, P.J., see Ford, J.L. **40**, 223
Edman, P., P. Artursson, E. Bjork, B. Davidsson, Immobilized L-asparaginase-L-glutaminase from Acinetobacter glutaminasificans in microspheres: Some properties in vivo and in an extracorporeal system **34**, 225
Edman, P., see Aiache, J.M. **39**, 235
Edman, P., see Artursson, P. **52**, 183
Edman, P., see Bisrat, M. **63**, 49
Edman, P., see Bjork, E. **47**, 233; **62**, 187; **75**, 73
Edman, P., see Camber, O. **29**, 259; **37**, 27
Edman, P., see Fahlvik, A.K. **65**, 249
Edman, P., see Pereswetoff-Morath, L. **124**, 37; **128**, 9, 23; **125**, 205
Edman, P., see Ryden, L. **83**, 1
Edsman, K., J. Carlfors, K. Harju, Rheological evaluation and ocular contact time of some carbomer gels for ophthalmic use **137**, 233
Edsman, K., see Sturesson, C. **89**, 235

Edwards, D., A.B. Selkirk, R.B. Taylor, Determination of the stability of mitomycin C by high performance liquid chromatography **4**, 21
Edwards, D.M.F., see Efthymiopoulos, C. **95**, 127
Edwards, H.E., see Menzies, A.R. **37**, 65
Edwards, H.E., see Morton, A.K. **9**, 321; **18**, 39
Edwards, H.G.M., see Anigbogu, A.N.C. **125**, 265
Edwards, H.G.M., see Williams, A.C. **81**, R11
Edwards-Levy, F., M.C. Andry, M.C. Levy, Determination of free amino group content of serum albumin microcapsules using trinitrobenzenesulfonic acid: Effect of variations in polycondensation pH **96**, 85
Determination of free amino group content of serum albumin microcapsules. II. Effect of variations in reaction time and in terephthaloyl chloride concentration **103**, 253
Edwards-Levy, F., see Andry, M.C. **128**, 197
Edwardson, P.A.D., M. Walker, C. Breheny, Quantitative FT-IR determination of skin hydration following occlusion with hydrocolloid containing adhesive dressings **91**, 51
Eerikainen, S., Effects of spheronization on some properties of uncoated and coated granules containing different kinds of fillers **77**, 89
Eerikainen, S., A.S. Lindqvist, The behaviour of various fillers in spheronized uncoated and film-coated granules containing slightly water-soluble indomethacin **75**, 181
Eerikainen, S., E. Muttonen, J. Yliruusi, A study of sodium indomethacin-calcium hydrogen phosphate precipitation **80**, 259
Eerikainen, S., J. Yliruusi, R. Laakso, The behaviour of the sodium salt of indomethacin in the cores of film-coated granules containing various fillers **71**, 201
Eerikainen, S., see Laakso, R. **67**, 79
Efentakis, E., see Fell, J.T. **4**, 153
Efentakis, M., Release kinetics of flurbiprofen from hydrophobic heterogeneous matrices containing surfactants **85**, R1
Efentakis, M., H. Al-Hmoud, G. Buckton, Z. Rajan, The influence of surfactants on drug release from a hydrophobic matrix **70**, 153
Efentakis, M., G. Buckton, Modelling drug release from hydrophobic matrices by use of thermodynamic activation parameters **60**, 229
Efentakis, M., see Al-Hmoud, H. **68**, R1
Efentakis, M., see Buckton, G. **62**, 157; **74**, 169
Efentakis, M., see Papadimitriou, E. **86**, 131; **98**, 57
Efstathiou, C.E., see Pentari, J.G. **77**, 41
Efthymiopoulos, C., D.M.F. Edwards, M. Strolin-Benedetti, F. Ruff, C. Advenier, R. Bianchini, L. Musatti, Pharmacokinetics of acipimox and of its *N*-deoxy metabolite following single and repeated oral administration of a sustained release formulation to healthy volunteers **95**, 127
Egea, M.A., O. Valls, M.A. Alsina, M.L. Garcia, C. Losa, J.L. Vila-Jato, M.J. Alonso, Interaction of amikacin loaded nanoparticles with phosphatidylcholine monolayers as membrane models **67**, 103
Egermann, H., A. Krumphuber, Mass variation of powder samples of constant volume produced from binary random mixtures. I. Derivation of the equation **73**, 37
Eguchi, Y., see Nishida, K. **80**, 101
Ehret, P., see Brochart, H. **91**, 95
Ehtezazi, T., see Ghassabian, S. **130**, 49
Eickhoff, W.M., see Fong, G.W. **53**, 91
Eissens, A., see Bootsma, H.P.R. **51**, 213
Eissens, A.C., see Te Wierik, G.H. **98**, 219; **102**, 81; **134**, 27
Ek, R., G. Alderborn, C. Nystrom, Particle analysis of microcrystalline cellulose: Differentiation between individual particles and their agglomerates **111**, 43
Ek, R., T. Gren, U. Henriksson, H. Nyqvist, C. Nystrom, L. Odberg, Prediction of drug release by characterisation of the tortuosity in porous cellulose beads using a spin echo NMR technique **124**, 9
Ek, R., H. Lennholm, R. Davidson, C. Nystrom, G. Ragnarsson, Pore swelling in beads made of cellulose fibres and fibre fragments **122**, 49
Ek, R., P. Wormald, J. Ostlius, T. Iversen, C. Nystrom, Crystallinity index of microcrystalline cellulose particles compressed into tablets **125**, 257
Ek, R., see Davidson, R. **112**, 153
Ek, R., see Johansson, B. **117**, 57
Ekenved, G., see Dahl, G. **10**, 143
El Egakey, M.A., V. Bentele, J. Kreuter, Molecular weights of polycyanoacrylate nanoparticles **13**, 349
El Eini, D.I.D., see Vincke, B.J. **54**, 181
El Gindy, N.A., M.W. Samaha, Tensile strength of some pharmaceutical compacts and their relation to surface free energy **13**, 35
El Gindy, N.A., see Samaha, M.W. **19**, 63
El Khalik, A.F., see Gonda, I. **27**, 255
El Khawas, M., see Khalil, S.A.H. **18**, 157
El Khordagui, L.K., see Badwan, A.A. **13**, 67
El Khordagui, L.K., see Khalil, S.A. **16**, 271
El Khordagui, L.K., see Saleh, A.M. **17**, 115
El Moussaoui, A., A. Chauvet, J. Masse, D. Gaudy, A. Puech, Lyoavailability of nordazepam solid dispersions in relation with phase diagrams **106**, 109
El Samaligy M, P. Rohdewald, Polyacrylamide microbeads, a sustained release drug delivery system **13**, 23
El Sayed, A.A., A.J. Repta, Solubilization and stabilization of an investigational antineoplastic drug (NSC no. 278214) in an intravenous formulation using an emulsion vehicle **13**, 303
El Sayed, A.A., see Fouli, A.M. **14**, 95
El Sirag, O., see El Turabi, H. **53**, 249
El Tayar, N., see Gryllaki, M. **51**, 95
El Tayar, N., see Tsai, R.S. **80**, 39
El Taylor, N., H. Van de Waterbeemd, M. Gryllaki et al., The lipophilicity of deuterium atoms. A comparison of shake-flask and HPLC methods **19**, 271
El Turabi, H., O. El Sirag, M.M.A. Homeida, D.W.G. Harron, W.J. Leahey, Paracetamol pharmacokinetics in patients with hepatosplenic schistosomiasis **53**, 249
El Zenary, H., see Vojnovic, D. **145**, 203
El-Arini, S.K., H. Leuenberger, Modelling of drug release from polymer matrices: Effect of drug loading **121**, 141

El-Arini, S.K., G.K. Shiu, J.P. Skelley, An in vitro study of possible food–drug interactions of the controlled-release propranolol products **55**, 25
El-Bagory, I., B.D. Reid, A.G. Mitchell, The effect of γ radiation on the tableting properties of some pharmaceutical excipients **105**, 255
El-Dardari, M.M., see Hosny, E.A. **136**, 37
El-Faham, T.H., see Ibrahim, S.A. **61**, 1
El-Falaha, B.M.A., A.D. Russell, J.R. Furr, D.T. Rogers, Activity of benzalkonium chloride and chlorhexidine diacetate against wild-type and envelope mutants of *Escherichia coli* and *Pseudomonas aeruginosa* **25**, 329
El-Falaha, B.M.A., D.T. Rogers, J.R. Furr, A.D. Russell, Surface changes in *Pseudomonas aeruginosa* exposed to chlorhexidine diacetate and benzalkonium chloride **23**, 239
El-Gendy, G.A., K. Terada, K. Yamamoto, Y. Nakai, Molecular behavior, dissolution characteristics and chemical stability of aspirin in the ground mixture and in the inclusion complex with di-*O*-methyl-β-cyclodextrin **31**, 25
El-Gholmy, Z.A., see Khalil, S.A.H. **20**, 99
El-Gorashi, A.S., see Hosny, E.A. **117**, 147
El-Gorashi, A.S., see Niazy, E.M. **56**, 181
El-Hinnawi, M.A., N.M. Najib, Ibuprofen-polyvinylpyrrolidone dispersions. Proton nuclear magnetic resonance and infrared studies **37**, 175
El-Hinnawi, M.A., see Najib, N.M. **45**, 139
El-Khawas, M., see Khalil, S.A.H. **19**, 233
El-Khordagui, L.K., Hydrotrope-gelled starch: Study of some physicochemical properties **74**, 25
Effect of sodium salicylate on the solution properties of sodium dodecyl sulphate **83**, 53
El-Khordagui, L.K., S.A. Khalil, A.M. Saleh, Diazepam-sodium salicylate solution: Dilution with intravenous fluids, in vitro haemolytic activity and protein binding **7**, 111
El-Khordagui, L.K., see Darwish, I.A. **140**, 25
El-Khordagui, L.K., see El-Massik, M.A. **140**, 69
El-Khordagui, L.K., see Khalil, S.A.H. **20**, 99
El-Khordagui, L.K., see Saleh, A.M. **5**, 161; **24**, 231; **57**, 205
El-Kirsh, T.A., see El-Samaligy, M.S. **31**, 137
El-Koussi, A.A., N. Bodor, Formation of propranolol in the iris-ciliary body from its propranolol ketoxime precursor—A potential antiglaucoma drug **53**, 189
El-Mahrouk, G.M., see El-Samaligy, M.S. **31**, 137
El-Massik, M.A., I.A. Darwish, E.E. Hassan, L.K. El-Khordagui, Development of a dissolution medium for glibenclamide **140**, 69
El-Massik, M.A., see Darwish, I.A. **140**, 25
El-Nakeeb, M.A., see Ali, H.M. **9**, 233
El-Nimr, A.E.M., see Kassem, M.A. **12**, 1
El-Ridy, M., M. Akbarieh, M. Kassem, M. Sharkawi, R. Tawashi, Chemoprophylaxis of schistosomiasis using liposome-encapsulated tartar emetic **56**, 23
El-Ridy, M.S., see Ammar, H.O. **103**, 237
El-Sabbagh, H.M., see Soliman, O.A. **149**, 73
El-Samaligy, M.S., G.M. El-Mahrouk, T.A. El-Kirsh, Adsorption-desorption effect of microcystalline cellulose on ampicillin and amoxycillin **31**, 137
El-Samaligy, M.S., see Mahmound, H.A. **25**, 121
El-Sayed, Y.M., M.A. Al-Meshal, A.A. Al-Angary, S.R. Al-Balla, M.W. Gouda, Effects of oral administration of colestipol and cholestyramine on the pharmacokinetics of ketoprofen administered intramuscularly in man **109**, 107
El-Sayed, Y.M., M.A. Al-Meshal, A.A. Al-Angary, K.M. Lutfi, Gouda M. Wafik, Accelerated clearance of intravenous indomethacin by oral activated charcoal in rabbits **64**, 109
El-Sayed, Y.M., E.M. Niazy, S.H. Khidr, In vivo evaluation of sustained-release microspheres of metoclopramide hydrochloride in beagle dogs **123**, 113
El-Sayed, Y.M., K.F. Tabbara, M.W. Gouda, Effect of acetazolamide on the pharmacokinetics of cyclosporin in rabbits **121**, 181
El-Sayed, A.A., see Badawi, A.A. **6**, 55
El-Sayed, Y.M., see Al-Meshal, M.A. **64**, 155
El-Sayed, Y.M., see Hosny, E.A. **112**, 87
El-Shanawany, S.M., see Habib, F.S. **26**, 243
El-Tahir, K.E.H., see Hosny, E.A. **142**, 163
El-Yazbi, F.A., M.H. Barary, M.H. Abdel-Hay, Determination of nitrazepam and dipotassium clorazepate in the presence of their degradation products using second derivative spectrophotometry **27**, 139
El-Yazbi, F.A., see Wahbi, A.M. **92**, 15
Elamin, A.A., C. Ahlneck, G. Alderborn, C. Nystrom, Increased metastable solubility of milled griseofulvin, depending on the formation of a disordered surface structure **111**, 159
Elamin, A.A., G. Alderborn, C. Ahlneck, The effect of precompaction processing and storage conditions on powder and compaction properties of some crystalline materials **108**, 213
Elamin, A.A., T. Sebhatu, C. Ahlneck, The use of amorphous model substances to study mechanically activated materials in the solid state **119**, 25
Elbers, J.A.C., see Baert, L. **99**, 7
Elberse, P.A., see Riepma, K.A. **90**, R1
Eldsater, C., see Singh, S.K. **142**, 199, 215
Eley, J.G., G.W. Halbert, A.T. Florence, The incorporation of estramustine into low density lipoprotein and its activity in tissue culture **63**, 121
Incorporation of prednimustine into low density lipoprotein: Activity against P388 cells in tissue culture **65**, 219
Eli, M., see Torricelli, C. **75**, 147
Elkins, J., see Shah, V.P. **53**, 53
Elkinton, S.K., see Ross, D.L. **88**, 379; **90**, 179; **93**, 131
Elkoubi, C., see Arnaud, P. **80**, 129
Elliott, D.S., see McElnay, J.C. **36**, 199; **47**, 239
Elliott, P.N.C., see Ford, J.L. **71**, 95
Elliott, P.N.C., see Mitchell, K. **66**, 233; **100**, 155, 165, 175
Elliott, P.N.C., see Perez-Marcos, B. **111**, 251
Ellway, K., see Gaddipati, N. **128**, 55
Elmarakby, Z., see Owoyale, J.A. **12**, 265
Elmarakby, Z.S., see Owoyale, J.A. **50**, 219
Elorza, B., see Chantres, J.R. **138**, 139
Elorza, B., see Sainz, M.C. **89**, 183; **91**, 1

Elorza, M.A., see Chantres, J.R. **138**, 139
Elorza, M.A., see Sainz, M.C. **89**, 183; **91**, 1
Elsabee, M., R.J. Prankerd, Solid-state properties of drugs. II. Peak shape analysis and deconvolution of overlapping endotherms in differential scanning calorimetry of chiral mixtures **86**, 211
Solid-state properties of drugs. III. Differential scanning calorimetry of chiral drug mixtures existing as racemic solid solutions, racemic mixtures or racemic compounds **86**, 221
Elshabouri, S.R., S.A. Ibrahim, A.F. Youssef, Analysis of isoniazid using 2-methyl-3-nitropyridine-6-carboxaldehyde **2**, 257
ElShaboury, M.H., Physical properties and dissolution profiles of tablets directly compressed with β-cyclodextrin **63**, 95
Elson, G., see Davis, S.S. **1**, 85
Emori, H., see Yamamoto, K. **102**, 133
Endo, H., see Kohri, N. **81**, 49
Enever, R., see Dey, M. **49**, 121
Enever, R.P., see Lewis, G.A. **2**, 203; **3**, 275, 319
Enever, R.P., see Shahjahan, M. **70**, 9; **82**, 215, 223, 229; **143**, 75, 83
Engblom, J., S. Engstrom, Azone^R and the formation of reversed mono- and bicontinuous lipid–water phases **98**, 173
Engblom, J., see Engstrom, S. **86**, 137
Engel, A., G. Bendas, F. Wilhelm, M. Mannova, M. Ausborn, P. Nuhn, Freeze drying of liposomes with free and membrane-bound cryoprotectants—The background of protection and damaging processes **107**, 99
Engel, J., see Schelenz, G. **113**, 133
Engle, K.K., see Liversidge, G.G. **23**, 87; **30**, 247
Engstrom, L., see Engstrom, S. **79**, 113
Engstrom, S., L. Engstrom, Phase behaviour of the lidocaine-monoolein-water system **79**, 113
Engstrom, S., L. Lindahl, R. Wallin, J. Engblom, A study of polar lipid drug carrier systems undergoing a thermoreversible lamellar-to-cubic phase transition **86**, 137
Engstrom, S., see Engblom, J. **98**, 173
Engstrom, S., see Lindell, K. **95**, 219; **124**, 107
Engstrom, S., see Petersson, T. **132**, 231
Ensing, J.G., see Moolenaar, F. **9**, 353
Enthoven, R., see Zanen, P. **81**, 199
Erbod, A., see Lashmar, U.T. **125**, 315
Ercan, T.M., see Ciftci, K. **131**, 73
Erden, N., N. Celebi, A study of the inclusion complex of naproxen with β-cyclodextrin **48**, 83
Factors influencing release of salbutamol sulphate from poly(lactide-*co*-glycolide) microspheres prepared by water-in-oil-in-water emulsion technique **137**, 57
Erden, N., see Celebi, N. **78**, 183; **136**, 89
Eriksson, H., see Venho, V.M.K. **30**, 91
Eriksson, M., C. Nystrom, G. Alderborn, Evaluation of a permeametry technique for surface area measurements of coarse particulate materials **63**, 189
The use of air permeametry for the assessment of external surface area and sphericity of pelletized granules **99**, 197
Eriksson, M., G. Alderborn, Mechanisms for post-compaction changes in tensile strength of sodium chloride compacts prepared from particles of different dimensions **109**, 59
The effect of original particle size and tablet porosity on the increase in tensile strength during storage of sodium chloride tablets in a dry atmosphere **113**, 199
Eriksson, M., G. Alderborn, C. Nystrom, F. Podzceck, J.M. Newton, Comparison between and evaluation of some methods for the assessment of the sphericity of pellets **148**, 149
Ermis, D., N. Tarimci, Ketoprofen sustained-release suppositories containing hydroxypropylmethylcellulose phthalate in polyethylene glycol bases **113**, 65
Ermis, D., see Tarimci, N. **147**, 71
Erni, W., see Bury, M. **76**, 207; **124**, 183
Eroglu, L., see Gursoy, A. **52**, 101
Ertan, G., Y. Karasulu, T. Guneri, Degradation and gastrointestinal stability of nitrofurantoin in acidic and alkaline media **96**, 243
Ertan, G., see Ozyazici, M. **138**, 25
Ertel, K.D., J.T. Carstensen, Validity of the steady-state assumption in in vitro liposomal absorption enhancement modeling: The free solute mechanism as an example **34**, 179
An examination of the physical properties of pure magnesium stearate **42**, 171
Examination of a modified Arrhenius relationship for pharmaceutical stability prediction **61**, 9
Escalera, B., see Bustamante, P. **129**, 283
Esclusa-Diaz, M.T., M. Gayo-Otero, M.B. Perez-Marcos, J.L. Vila-Jato, J.J. Torres-Labandeira, Preparation and evaluation of ketoconazole-β-cyclodextrin multicomponent complexes **142**, 183
Esclusa-Diaz, M.T., M. Guimaraens-Mendez, M.B. Perez-Marcos, J.L. Vila-Jato, J.J. Torres-Labandeira, Characterization and in vitro dissolution behaviour of ketoconazole/β- and 2-hydroxypropyl-β-cyclodextrin inclusion compounds **143**, 203
Escobar, C., see Veiga, M.D. **89**, 119; **93**, 215
Esezobo, S., Disintegrants: Effect of interacting variables on the tensile strengths and disintegration times of sulphaguanidine tablets **56**, 207
Esezobo, S., see Pilpel, N. **70**, 241
Eshra, A.G., M.A. Etman, V.F. Naggar, Effect of milk and food on the bioavailability of ketoprofen in man **44**, 9
Eshra, A.G., see Hassan, E.E. **121**, 149
Espanol, O.M., see Frances, C. **77**, 193
Espino, C.D., see Farina, J.B. **45**, 123
Esposito, E., V. Carotta, A. Scabbia, L. Trombelli, P. D'Antona, E. Menegatti, C. Nastruzzi, Comparative analysis of tetracycline-containing dental gels: Poloxamer- and monoglyceride-based formulations **142**, 9
Esposito, E., R. Cortesi, F. Bortolotti, E. Menegatti, C. Nastruzzi, Production and characterization of biodegradable microparticles for the controlled delivery of proteinase inhibitors **129**, 263

Esposito, E., C. Pastesini, R. Cortesi, R. Gambari, E. Menegatti, C. Nastruzzi, Controlled release of 1-β-D-arabinofuranosylcytosine (ara-C) from hydrophilic gelatin microspheres: In vitro studies **117**, 151
Esposito, E., see Cortesi, R. **105**, 181; **139**, 69
Esposito, E., see Esposito, E. **117**, 151; **142**, 9
Esposito, E., see Nastruzzi, C. **90**, 43
Estelrich, J., see Casals, E. **143**, 171
Estelrich, J., see Foradada, M. **124**, 261
Estelrich, J., see Girona, V. **41**, 241
Estelrich, J., see Hernandez, J. **57**, 211
Estelrich, J., see Montero, M.T. **62**, 21
Estelrich, J., see Pons, M. **95**, 51
Estes, K., see Bodor, N. **53**, 195
Estes, K.S., M.E. Brewster, A.I. Webb, N. Bodor, A non-surfactant formulation for alfaxalone based on an amorphous cyclodextrin: Activity studies in rats and dogs **65**, 101
Estes, K.S., see Brewster, M.E. **59**, 231; **79**, 289
Estes, K.S., see Dietzel, K. **59**, 225
Estes, K.S., see Pop, E. **140**, 33
Estorch, M., see Roca, M. **69**, 189
Etman, M.A., F.A. Ismail, A.H. Nada, Effect of metoclopramide on ketoprofen pharmacokinetics in man **88**, 433
Etman, M.A., V.F. Naggar, Thermodynamics of paracetamol solubility in sugar-water cosolvent systems **58**, 177
Etman, M.A., see Eshra, A.G. **44**, 9
Etman, M.A., see Nada, A.H. **53**, 175
Eto, K., see Gomita, Y. **54**, 89
Etter, J.C., see Pannatier, A. **8**, 167
Etter, J.C., see Rothlisberger, T. **44**, 141
Evans, D.F., see Coupe, A.J. **78**, 69; **92**, 167
Evans, D.F., see Hardy, J.G. **37**, 245
Evans, J.C., see Gay, C.L. **49**, 39
Evans, N., R.H. Guy, J. Hadgraft et al., Transdermal drug delivery to neonates **24**, 259
Evitts, D.P., see Musson, D.G. **99**, 85
Evora, C., see Delgado, A. **134**, 203; **140**, 219
Evora, C., see Munguia, O. **86**, 107
Evora, C., see Soriano, I. **125**, 223; **142**, 135
Evora, C.M., see Farina, J.B. **45**, 123
Evora, C.M., see Sanchez, E. **83**, 59
Eyer, P., see Horst, Thiermann **114**, 125
Eyer, P., see Thiermann, H. **109**, 35; **137**, 167
Eynard, I., see Lafforgue, C. **121**, 89
Ezpeleta, I., J.M. Irache, S. Stainmesse, C. Chabenat, J. Guegeuen, Y. Popineau, A.M. Orecchioni, Gliadin nanoparticles for the controlled release of all-transretinoic-acid **131**, 191
Ezpeleta, I., J.M. Irache, S. Stainmesse, C. Chabenat, J. Gueguen, A.M. Orecchioni, Preparation of lectin–vicilin nanoparticle conjugates using the carbodiimide coupling technique **142**, 227
Ezpeleta, I., see Irache, J.M. **126**, 103
Ezzedeen, F.W., S.H. Majeed, F.A. Shibab, M.J. Mahmoud, D.H. Robinson, Y.H. Tahseen, S.J. Stohs, In vitro and in vivo evaluation of four co-trimoxazole oral suspensions **59**, 255
Ezzedeen, F.W., F.A. Shihab, S.J. Stohs, Release of sorbic acid from ointment bases **28**, 113

F

Fabbri, L., see Santoni, G. **71**, 1; **80**, 263
Fabin, B., E. Touitou, Localization of lipopholic molecules penetrating rat skin in vivo by quantitative autoradiography **74**, 59
Fabin, B., see Touitou, E. **43**, 9, 17; **70**, 159
Fabra-Campos, S., E. Climent, A. Sanchis-Cortes, J.M. Pla-Delfina, Compared effects of synthetic and natural bile acid surfactants on xenobiotic absorption. IV. Studies with taurocholate in the rat small intestine **109**, 197
Fabra-Campos, S., see Collado, E.F. **44**, 187
Fabra-Campos, S., see Garrigues, T.M. **57**, 189
Fabre, G., see Boulenc, X. **123**, 13, 71
Fabre, G., see Caro, I. **116**, 147
Fabregas, J.L., J. Cucala, Influence of microphysical structure on in vitro activity of new synthetic antacids **52**, 173
Fabrizio Saettone, M., P. Chetoni, R. Cerbai, G. Mazzanti, L. Braghiroli, Evaluation of ocular permeation enhancers: In vitro effects on corneal transport of four β-blockers, and in vitro/in vivo toxic activity **142**, 103
Fachaux, J.M., Hermann A. Guyot, J.C. Guyot, P. Conflant, M. Drache, J.P. Huvenne, R. Bouche, Compression ability improvement by solvation/desolvation process: Application to paracetamol for direct compression **99**, 99
Fagerholm, U., see Lennernas, H. **127**, 103
Fagerness, P.E., see Snider, B.G. **66**, 63
Fahelelbom, K.M.S., see Quigley, J.M. **58**, 107
Fahlvik, A.K., P. Artursson, P. Edman, Magnetic starch microspheres: Interactions of a microsphere MR contrast medium with macrophages in vitro **65**, 249
Fahlvik, A.K., see Sontum, P.C. **128**, 269
Fairbrother, J.E., see Grant, D.J.W. **5**, 109; **18**, 25
Fairbrother, J.E., see Hollingsbee, D.A. **77**, 199
Fairbrother, J.E., see Kadir, R. **60**, 139
Fairbrother, J.E., see Queen, D. **44**, 25
Faizer, R., see Clas, S.D. **121**, 73
Faizi, M., see Dehpour, A.R. **119**, 133
Fakes, M.G., see Morris, K.R. **105**, 209
Falch, E., see Bundgaard, H. **6**, 19; **23**, 223; **24**, 307; **25**, 27; **27**, 71; **29**, 19; **30**, 111; **64**, 75
Falch, E., see Buur, A. **24**, 43
Falch, E., see Jensen, E. **58**, 143
Falch, E., see Johansen, M. **13**, 89
Falch, E., see Mosher, G.L. **39**, 113
Fales, H., see Pitha, J. **29**, 73
Falkner, F.C., see Thombre, A.G. **111**, 181
Fallon, M.P., see Moore, D.E. **14**, 133
Falson, F., see Lafforgue, C. **121**, 89; **147**, 243
Falson-Rieg, F., see Couarraze, G. **56**, 197
Falson-Rieg, F., see Ganem-Quintanar, A. **147**, 165
Fang, C.L., see Fang, J.Y. **149**, 183
Fang, J.Y., C.L. Fang, Y.B. Huang, Y.H. Tsai, In vitro permeation study of capsaicin and its synthetic derivatives from ointment bases using various skin types **126**, 119

Fang, J.-Y., P.-C. Wu, Y.-B. Huang, Y.-H. Tsai, In vivo percutaneous absorption of capsaicin, nonivamide and sodium nonivamide acetate from ointment bases: Pharmacokinetic analysis in rabbits **128**, 169
Percutaneous absorption of capsaicin, nonivamide and sodium nonivamide acetate from gel and ointment bases: In vitro formulation evaluations in pigs and in vivo: Bioengineering methods in humans **130**, 121
In vivo percutaneous absorption of capsaicin, nonivamide and sodium, nonivamide acetate from ointment bases: Skin erythema test and non-invasive surface recovery technique in humans **131**, 143
Transdermal iontophoresis of sodium nonivamide acetate. I. Consideration of electrical and chemical factors **143**, 47
Transdermal iontophoresis of sodium nonivamide acetate. II. Optimization and evaluation on solutions and gels **145**, 175
Transdermal iontophoresis of sodium nonivamide acetate. III. Combined effect of pretreatment by penetration enhancers **149**, 183
Fang, J.Y., see Wu, P.C. **148**, 41
Faouzi, M.A., T. Dine, M. Luyckx, F. Goudaliez, M.L. Mallevais, C. Brunet, M. Cazin, B. Gressier, J.C. Cazin, Stability and compatibility studies of pefloxacin, ofloxacin and ciprofloxacin with PVC infusion bags **89**, 125
Faouzi, M.A., T. Dine, M. Luyckx, B. Gressier, F. Goudaliez, M.L. Mallevais, C. Brunet, M. Cazin, J.C. Cazin, Leaching of diethylhexyl phthalate from PVC bags into intravenous teniposide solution **105**, 89
Fara, J.W., see Swisher, D.A. **22**, 219
Farah, N., J. Bouzon, M. Rollet et al., 'Dry emulsion'—a sustained release form: Modelling of drug transfers in liquids **36**, 81
Fardella, G., P. Barbetti, I. Chiappini, G. Grandolini, Quantitative analysis of fluoroquinolones by ^{1}H- and ^{19}F-NMR spectroscopy **121**, 123
Fares, H.M., S. Chatterjee, M. Hayward, In vitro permeation and irritation of benzoyl peroxide-containing products **133**, 215
Farhadich, B., see Kararli, T.T. **102**, 177
Farina, J., see Munguia, O. **86**, 107
Farina, J.B., C.M. Evora, C.D. Espino, M. Llabres, Theoretical model for interpretation of in situ absorption studies **45**, 123
Farina, J.B., see Oliva, A. **143**, 163
Farinha, A., see Lewis, D. **148**, 177
Farinotti, R., see Crauste-Manciet, S. **149**, 241
Faroongsarng, D., see Mitrevej, A. **140**, 61
Farooq, A., see Jha, P. **63**, 143
Farouk, A., In vitro studies on cinchocaine-preservative combinations **9**, 1
Antibacterial activity of proteolytic enzymes **12**, 295
Farouk, A., see Ali, A.A. **9**, 239
Farouk, A., see Ali, H.M. **9**, 233
Farouk, A.M., see Ali, H.M. **6**, 301
Farr, S.J., I.W. Kellaway, B. Carman-Meakin, Comparison of solute partitioning and efflux in liposomes formed by a conventional and an aerosolised method **51**, 39
Farr, S.J., I.W. Kellaway, D.R. Parry-Jones, S.G. Woolfrey, ^{99m}technetium as a marker of liposomal deposition and clearance in the human lung **26**, 303
Farr, S.J., see Clarke, J.G. **80**, R1; **93**, 221
Farr, S.J., see Davies, N.M. **105**, 97
Farr, S.J., see Dennis, A.B. **65**, 85
Farr, S.J., see Donnelly, A. **135**, 191
Farr, S.J., see Durrani, A.M. **118**, 243
Farr, S.J., see Harnor, K.J. **95**, 111
Farr, S.J., see Jamshaid, M. **48**, 125
Farr, S.J., see Warren, S.J. **124**, 195
Farraj, N.F., P. Kahela, G.D. Parr, R. Withington, A new intramuscular formulation of rifamycin SV **38**, 39
Farraj, N.F., S.S. Davis, D.P. Graham, H.N.E. Stevens, Modification of the aqueous solubility and stability of progabide **52**, 11
Farraj, N.F., S.S. Davis, G.D. Parr, H.N.E. Stevens, Absorption of progabide from aqueous solutions in a modified recirculating rat intestinal perfusion system **43**, 93
Studies on the simultaneous dissolution and degradation of progabide powders **52**, 19
Investigation of progabide absorption from the gastrointestinal tract of the rabbit **43**, 83
Farraj, N.F., see Fisher, A.N. **74**, 147
Farraj, N.F., see Illum, L. **57**, 49; **63**, 207
Farraj, N.F., see Richardson, J.L. **88**, 319
Farrar, C., see Niven, R.W. **109**, 17
Farsam, H., H.H. Yahya-Saeb, A. Fawzi, Spectrophotometric determination of codeine in pharmaceutical preparations **7**, 343
Farvar, M.A., see Corrigan, O.I. **5**, 229
Fassihi, A.R., Mechanisms of disintegration and compactibility of disintegrants in a direct compression system **32**, 93
Continuous matrix formation for controlled drug release: Compression of isotropic polymeric system **34**, 169
Kinetics of drug release from solid matrices: Effect of compaction pressure **37**, 119
Consolidation behaviour of polymeric substances in non-disintegrating solid matrices **44**, 249
Racemates and enantiomers in drug development **92**, 1
Fassihi, A.R., R. Dowse, S.S.D. Robertson, Effect of dietary cellulose on the absorption and bioavailability of theophylline **50**, 79
Influence of sorbitol solution on the bioavailability of theophylline **72**, 175
Fassihi, A.R., M.S. Parker, Formulation effects on capping tendencies **31**, 271
Fassihi, A.R., P.H.R. Persicaner, Solid state interaction of bromazepam with polyvinylpyrrolidone in the presence of moisture **37**, 167
Fassihi, A.R., see Durig, T. **77**, 315; **97**, 161
Fassihi, A.R., see Munday, D.L. **52**, 109; **69**, 123; **73**, 89; **118**, 251
Fattal, E., see Blanco Prieto, M.J. **111**, 137
Fattal, E., see Ramaldes, A. **143**, 1
Fattal, E., see Seijo, B. **62**, 1

Fauconnet, A.L., J.P. Basly, M. Bernard, γ Radiation induced effects on isoproterenol **144**, 123
Faulkner, R.D., see Faulkner, R.D. **43**, 53
Faulkner, R.D., L.L. Sia, Z.M. Look, J.S. Barone, S.J. Forbes, A.I. Weiss, B.M. Silber, Bioequivalency of solid oral dosage forms of cefixime **43**, 53
Faustino, E., see Lewis, D. **148**, 177
Fawaz, F., F. Bonini, M. Guyot, J. Bildet, M. Maury, A.M. Lagueny, Bioavailability of norfloxacin from PEG 6000 solid dispersion and cyclodextrin inclusion complexes in rabbits **132**, 271
Fawaz, F., F. Bonini, M. Guyot, A.M. Lagueny, H. Fessi, J.P. Devissaguet, Disposition and protective effect against irritation after intravenous and rectal administration of indomethacin loaded nanocapsules to rabbits **133**, 107
Fawaz, F., see Guyot, M. **123**, 53; **144**, 209
Fawcett, J.P., see Stark, G. **143**, 93
Fawzi, A., see Farsam, H. **7**, 343
Fawzi, M.B., see Ghebre-Sellassie, I. **37**, 211
Fawzi, M.B., see Mahjour, M. **56**, 1; **95**, 161
Fazio, G., see Fernandez-Hervas, M.J. **136**, 101
Fazio, G., see Fini, A. **121**, 19; **126**, 95
Fedder, J., see Keipert, S. **142**, 153
Fedor, L.R., Acid-catalyzed hydrolysis of (*R*)-4-[(1-methyl-3-oxo-1-butenyl)amino]-3-isoxazolidinone, a prodrug of cycloserine **22**, 197
Fee, J.P.H., see McClean, E. **60**, 35
Feely, L., see Davis, S.S. **49**, 183
Feely, L.C., S.S. Davis, Influence of surfactants on drug release from hydroxypropylmethylcellulose matrices **41**, 83
The influence of polymeric excipients on drug release from hydroxypropylmethylcellulose matrices **44**, 131
Feely, L.C., see Hardy, J.G. **38**, 103
Feijen, J., see Cremers, H.F.M. **110**, 117; **120**, 51
Feijen, J., see Kwon, G.S. **79**, 191
Feldkamp, J.R., see Morefield, E.M. **34**, 263
Feldstein, M.M., V.N. Tohmakhchi, L.B. Malkhazov, A.E. Vasiliev, N.A. Plate, Hydrophilic polymeric matrices for enhanced transdermal drug delivery **131**, 229
Fell, A.F., see Neil, J.M. **22**, 105
Fell, J.T., G.A. Digenis, Imaging and behaviour of solid oral dosage forms in vivo **22**, 1
Fell, J.T., E. Efentakis, Contact angle determinations on pharmaceutical powders: A comparison of two methods **4**, 153
Fell, J.T., H.A.H. Mohammad, The wetting of powders by bile salt solutions and gastric juice **125**, 327
Fell, J.T., see Anie, S.J. **69**, 181; **76**, 183
Fell, J.T., see Ashford, M. **91**, 241; **95**, 193
Fell, J.T., see Cutt, T. **33**, 81; **49**, 157
Fell, J.T., see Harris, D. **56**, 97
Fell, J.T., see Hunter, E. **4**, 175; **17**, 59
Fell, J.T., see Kaus, L.C. **20**, 315; **22**, 99
Fell, J.T., see Luangtana-Anan, M. **60**, 197
Fell, J.T., see Mohammad, H.A.H. **17**, 39
Fell, J.T., see Plumpton, E.J. **30**, 237, 241
Fell, J.T., see Wakerly, Z. **129**, 73
Fell, J.T., see Zaniboni, H.C. **125**, 151
Felton, L.A., M.M. Haase, N.H. Shah, G. Zhang, M.H. Infeld, A.W. Malick, J.W. McGinity, Physical and enteric properties of soft gelatin capsules coated with Eudragit^R L 39 D-55 **113**, 17
Felton, L.A., N.H. Shah, G. Zhang, M.H. Infeld, A.W. Malick, J.W. McGinity, Physical-mechanical properties of film-coated soft gelatin capsules **127**, 203
Femi-Oyewo, M.N., Granulations with surfactants: Effects on pasting **40**, 73
Femi-Oyewo, M.N., M.S. Spring, Studies on paracetamol crystals produced by growth in aqueous solutions **112**, 17
Ferappi, M., see Altomare, C. **48**, 91; **56**, 273
Feriotto, G., see Gambari, R. **72**, 251
Fernandes de Aguair, P., see Inghelbrecht, S. **148**, 103
Fernandez, B., R. Mosquera, E. Uriarte, Structure-activity relationships in verapamil and analogues using molecular mechanics calculations **79**, 199
Fernandez, M., I.C. Rodriguez, M.V. Margarit, A. Cerezo, Characterization of solid dispersions of piroxicam/polyethylene glycol 4000 **84**, 197
Fernandez, M., M.V. Margarit, I.C. Rodriguez, A. Cerezo, Dissolution kinetics of piroxicam in solid dispersions with polyethylene glycol 4000 **98**, 29
Fernandez-Arevalo, M., M.A. Holgado-Villafuerte, J.M. Gines-Dorado, A.M. Rabasco-Alvarez, Effects of different fillers and wetting liquids on the dissolution behavior of carteolol hydrochloride controlled release inert matrix tablets **95**, 117
Fernandez-Arevalo, M., see Alvarez-Fuentes, J. **139**, 237
Fernandez-Arevalo, M., see Caraballo, I. **89**, 213; **96**, 175; **109**, 229; **139**, 177
Fernandez-Arevalo, M., see Holgado, M.A. **114**, 13; **118**, 151; **121**, 187
Fernandez-Arevalo, M., see Rabasco, A.M. **67**, 201
Fernandez-Hervas, M.J., M.A. Holgado, A.M. Rabasco, G. Fazio, A. Fini, Fractal and reactive dimension of a diclofenac salt: Effect of the experimental conditions **136**, 101
Fernandez-Hervas, M.J., M.A. Holgado, A.M. Rabasco, A. Fini, Use of fractal geometry on the characterization of particles morphology: Application to the diclofenac hydroxyethylpirrolidine salt **108**, 187
Fernandez-Hervas, M.J., M.T. Vela, A. Fini, A.M. Rabasco, Fractal and reactive dimension in inert matrix systems **130**, 115
Fernandez-Hervas, M.J., M.T. Vela, M.A. Holgado, Cerro J. Del, A.M. Rabasco, Determination of percolation thresholds in matrix-type controlled release systems: Application of a resistance analysis technique **113**, 39
Fernandez-Hervas, M.J., see Fini, A. **121**, 19
Fernandez-Hervas, M.J., see Gonzalez-Rodriguez, M.L. **146**, 175
Fernandez-Hervas, M.J., see Holgado, M.A. **118**, 151; **120**, 157; **121**, 187; **142**, 143
Feroci, G., see Fini, A. **126**, 95
Ferrari, F., M. Bertoni, M.C. Bonferoni, S. Rossi, C. Caramella, C. Nystrom, Investigation on bonding and disintegration properties of pharmaceutical materials **136**, 71

Ferrari, F., M. Bertoni, M.C. Bonferoni, S. Rossi, C. Caramella, M.J. Waring, Comparative evaluation of hydrocolloid dressings by means of water uptake and swelling force measurements: II **117**, 49
Ferrari, F., M. Bertoni, M.C. Bonferoni, S. Rossi, M. Saunders, M.E. Aulton, U. Conte, C. Caramella, Influence of probe shape on gel strength test **121**, 11
Ferrari, F., M. Bertoni, C. Caramella, A. La Manna, Description and validation of an apparatus for gel strength measurements **109**, 115
Ferrari, F., M. Bertoni, C. Caramella, M.J. Waring, Comparative evaluation of hydrocolloid dressings by means of water uptake and swelling force measurements: I **112**, 29
Ferrari, F., see Bonferoni, M.C. **117**, 41
Ferrari, F., see Caramella, C. **44**, 177
Ferrari, F., see Ferrari, F. **117**, 49; **121**, 11; **136**, 71
Ferrari, F., see Pena Romero, A. **73**, 239
Ferrari, F., see Ruizk, S. **112**, 125
Ferrari, L., see Rochira, M. **144**, 19
Ferreira, L.A.M., J. Doucet, M. Seiller, J.L. Grossiord, J.P. Marty, J. Wepierre, In vitro percutaneous absorption of metronidazole and glucose: Comparison of o/w, w/o/w and w/o systems **121**, 169
Ferreira, L.A.M., M. Seiller, J.L. Grossiord, J.P. Marty, J. Wepierre, Vehicle influence on in vitro release of metronidazole: Role of w/o/w multiple emulsion **109**, 251
Ferreira, L.A.M., see Ferreira, L.A.M. **121**, 169
Ferrero, C., N. Munoz, M.V. Velasco, A. Munoz-Ruiz, R. Jimenez-Castellanos, Disintegrating efficiency of croscarmellose sodium in a direct compression formulation **147**, 11
Ferrero, M.C., M.V. Velasco, A. Munoz, M.R. Jimenez-Castellanos, I. Castellano, M. Gurruchaga, I. Goni, Drug release from a new family of graft copolymers of methyl methacrylate. I **149**, 233
Ferrier, D., see Sarbach, Ch. **140**, 169
Ferry, J., see Tsai, J.C. **68**, 127; **72**, 227
Ferry, J.J., see Tsai, J.C. **96**, 111
Fessi, H., J.P. Marty, F. Puisieux, J.T. Carstensen, The Higuchi square root equation applied to matrices with high content of soluble drug substance **1**, 265
Equation relating to the Higuchi R-ratio of lubrication **30**, 209
Fessi, H., F. Piusieux, J.P. Devissaguet, N. Ammoury, S. Benita, Nanocapsule formation by interfacial polymer deposition following solvent displacement **55**, R1
Fessi, H., see Al Khouri Fallouh, N. **28**, 125
Fessi, H., see Belbella, A. **129**, 95
Fessi, H., see Fawaz, F. **133**, 107
Fessi, H., see Gursoy, A. **52**, 101
Fessi, H., see Guterres, S.S. **113**, 57
Fessi, H., see Masson, V. **139**, 113
Fessi, H., see Nemati, F. **138**, 237
Fessi, H., see Quintanar-Guerrero, D. **143**, 133
Fessi, H., see Regnault, C. **132**, 263
Fessi, H., see Rodrigues, Jr. J.M. **126**, 253
Fessi, H., see Tabosa de Egito, E. **145**, 17
Fessi, H., see Thioune, O. **146**, 233
Fessi, H., see Yu, W. **89**, 139
Fessler, D., see Chiang, C.C. **117**, 197
Field, W.N., M.D. Gamble, D.A. Lewis, A comparison of the treatment of thyroidectomized rats with free thyroxine and thyroxine encapsulated in erythrocytes **51**, 175
Field, W.N., see Alpar, H.O. **62**, 133
Fielden, K.E., J.M. Newton, R.C. Rowe, Movement of liquids through powder beds **79**, 47
The influence of lactose particle size on spheronization of extrudate processed by a ram extruder **81**, 205
A comparison of the extrusion and spheronization behaviour of wet powder masses processed by a ram extruder and a cylinder extruder **81**, 225
The influence of moisture content on spheronization of extrudate processed by a ram extruder **97**, 79
Fielding, R.M., see Guo, L.S.S. **75**, 45
Figueiredo, M., see Simoes, S. **127**, 283; **131**, 273; **139**, 169
Filipovic-Grcic, J., M. Becirevic-Lacan, N. Skalko, I. Jalsenjak, Chitosan microspheres of nifedipine and nifedipine-cyclodextrin inclusion complexes **135**, 183
Filipovic-Grcic, J., D. Maysinger, B. Zorc, I. Jalsenjak, Macromolecular prodrugs. IV. Alginate-chitosan microspheres of PHEA-L-dopa adduct **116**, 39
Filipovic-Grcic, J., see Zorc, B. **99**, 135
Finan, M.P., see Deasy, P.B. **89**, 251
Fini, A., G. Fazio, M.J. Fernandez-Hervas, M.A. Holgado, A.M. Rabasco, Influence of crystallization solvent and dissolution behaviour for a diclofenac salt **121**, 19
Fini, A., G. Fazio, G. Feroci, Solubility and solubilization properties of non-steroidal anti-inflammatory drugs **126**, 95
Fini, A., see Fernandez-Hervas, M.J. **108**, 187; **130**, 115; **136**, 101
Fini, A., see Holgado, M.A. **120**, 157
Finlay, W.H., K.W. Stapleton, P. Zuberbuhler, Errors in regional lung deposition predictions of nebulized salbutamol sulphate due to neglect or partial inclusion of hydroscopic effects **149**, 63
Finley, R., see Adjei, A. **61**, 135; **107**, 57
Finne, U., M. Hannus, A. Urtti, Mechanisms of timolol release from monoisopropyl PVM-MA matrices with and without a basic salt **78**, 237
Finne, U., J. Salivirta, A. Urtti, Sodium acetate improves the ocular/systemic absorption ratio of timolol applied ocularly in monoisopropyl PVM-MA matrices **75**, R1
Finne, U., A. Urtti, Pharmacokinetic simulation reveals in vivo deviations from in vitro release of timolol from polymer matrices **84**, 217
Finne, U., V. Vaisanen, A. Urtti, Modification of ocular and systemic absorption of timolol from ocular inserts by a buffering agent and a vasoconstrictor **65**, 19
Finne, U., see Urtti, A. **61**, 241
Firestone, B.A., see Benjamin, E.J. **35**, 73
Firmer, S.J., D.J. Cutler, Simulation of gastrointestinal drug absorption. I. Longitudinal transport in the small intestine **48**, 231
Fisch, A., see Vincent-Ballereau, F. **58**, 25

Fisher, A.N., N.F. Farraj, D.T. O'Hagan, I. Jabbal-Gill, B.R. Johansen, S.S. Davis, L. Illum, Effect of L-α-lysophosphatidylcholine on the nasal absorption of human growth hormone in three animal species **74**, 147
Fitzgerald, P., J. Hadgraft, J. Kreuter, C.G. Wilson, A γ-scintigraphic evaluation of microparticulate opthalmic delivery systems: liposomes and nanoparticles **40**, 81
Fitzgerald, P., C.G. Wilson, J.L. Greaves, M. Frier, D. Hollingsbee, D. Gilbert, M. Richardson, Scintigraphic assessment of the precorneal residence of a new ophthalmic delivery system (NODS) in man **83**, 177
Flament, M.P., P. Leterme, T. Burnouf, A.T. Gayot, Jet nebulisation: Influence of dynamic conditions and nebuliser on nebulisation duality. Application to the α1 protease inhibitor **148**, 93
Flanagan, D.R., see Bhat, P.G. **126**, 179; **134**, 15
Flanagan, D.R., see Botha, S.A. **82**, 185, 195
Flanders, P., see Staniforth, J.N. **31**, 277
Fleitman, J., J.H. Perrin, The binding of benoxaprofen to human serum albumin. Classification of the primary binding site **11**, 215
The effects of pH, calcium and chloride ions on the binding of benoxaprofen to human serum albumin. Circular dichroic and dialysis measurements **11**, 227
Fleming, R., see Guy, R.H. **3**, 143
Flinn, N., S. Coppard, I. Toth, Oral absorption studies of lipidic conjugates of thyrotropin releasing hormone (TRH) and luteinizing hormone-releasing hormone (LHRH) **137**, 33
Flinn, N., I. Hussain, A. Shaw, P. Artursson, W.A. Gibbons, I. Toth, Oral absorption studies of lipid–polylysine conjugates of thyrotropin releasing hormone (TRH) and luteinizing hormone releasing hormone (LHRH) **138**, 167
Erratum: Oral absorption studies of lipid–polylysine conjugates of thyrotropin releasing hormone (TRH^1) and luteinizing hormone releasing hormone ($LHRH^1$) (Int. J. Pharm. 138 (1996) (167)) **143**, 125
Flinn, N., see Toth, I. **105**, 241; **125**, 117
Florence, A.T., D. Whitehill, The formulation and stability of multiple emulsions **11**, 277
Florence, A.T., see Al-Dujaili, H. **34**, 67, 75
Florence, A.T., see Eley, J.G. **63**, 121; **65**, 219
Florence, A.T., see Gianasi, E. **148**, 139
Florence, A.T., see Gregoriadis, G. **65**, 235
Florence, A.T., see Halbert, G.W. **21**, 219
Florence, A.T., see Hillery, A.M. **132**, 123
Florence, A.T., see Jani, P.U. **84**, 245; **86**, 239; **105**, 157
Florence, A.T., see Law, T.K. **21**, 277; **33**, 65
Florence, A.T., see Mendelow, A.Y. **49**, 29
Florence, A.T., see Omotosho, J.A. **61**, 51
Florence, A.T., see Ping, Q. **61**, 79
Florence, A.T., see Saleh, A.M. **57**, 205
Florence, A.T., see Tomer, R. **99**, R5
Florence, A.T., see Vezin, W.R. **3**, 231
Florence, A.T., see Walters, K.A. **10**, 153
Florence, A.T., see Wesoluch, F. **2**, 343
Florence, A.T., see Wood, D.A. **8**, 35
Florence, A.T., see Yoshioka, A.T. **105**, 1
Florence, A.T., see Yoshioks, T. **108**, 117
Flynn, G., see Addicks, W.J. **56**, 243
Flynn, G.L., see Ackermann, C. **36**, 61, 67
Flynn, G.L., see Chiang, C.M. **49**, 109; **50**, 21; **55**, 229
Flynn, G.L., see Ganesan, M.G. **20**, 139
Flynn, G.L., see Hou, S.Y.E. **66**, 79
Flynn, G.L., see Neau, S.H. **49**, 223
Flynn, G.L., see Shah, V.P. **74**, 1; **82**, 21
Flynn, G.L., see Tsai, J.C. **68**, 127; **72**, 227; **96**, 111
Focher, B., see Ahmed, S.M. **77**, 247
Fokkens, J.G., C.J. de Blaey, Drug release from non-aqueous suspensions. II. The release of methylxanthines from paraffin suspensions **18**, 127
Optimization of the in vitro release of zomepirac from suspensions and suppositories **19**, 177
Fokkens, J.G., C.J. de Amelsfoort Van Blaey et al., A thermodynamic study of the solubility of theophylline and its hydrate **14**, 79
Fokkens, J.G., see Van Bommel, E.M. **72**, 19
Fokkens, R.H., see Beijnen, J.H. **32**, 111
Folayan, J.O., see Akojie, F.O.B. **42**, 261
Folkers, G., see Camenisch, G. **147**, 61
Fong, G.W., W.M. Eickhoff, Liquid chromatographic determinations of amines in complex cough-cold formulations **53**, 91
Fonseca, M.J., A. Cabanes, Alsina M. Asuncion, F. Reig, Design of a new formulation for sustained release of gentamicin: A carbopol hydrogel **133**, 265
Fontan, J.E., P. Arnaud, J.C. Chaumeil, Enhancing properties of surfactants on the release of carbamazepine from suppositories **73**, 17
Effects of polysorbate 80 on the pharmacokinetics on a carbamazepine suppository in the rabbit **82**, 67
Fonteh, A.N., see Del, Olmo E. **81**, 267
Foradada, M., J. Estelrich, Encapsulation of thioguanine in liposomes **124**, 261
Foradada, M., see Pons, M. **95**, 51
Forbes, R.T., P. York, J.R. Davidson, Dissolution kinetics and solubilis of *p*-aminosalicylic acid and its salts **126**, 199
Forbes, S.J., see Faulkner, R.D. **43**, 53
Ford, J., see Ali, H.M. **42**, 155
Ford, J.L., K. Mitchell, D. Sawh, S. Ramdour, D.J. Armstrong, P.N.C. Elliott, C. Rostron, J.E. Hogan, Hydroxypropylmethylcellulose matrix tablets containing propranolol hydrochloride and sodium dodecyl sulphate **71**, 213
Ford, J.L., K. Mitchell, P. Rowe, D.J. Armstrong, P.N.C. Elliott, C. Rostron, J.E. Hogan, Mathematical modelling of drug release from hydroxypropylmethylcellulose matrices: Effect of temperature **71**, 95
Ford, J.L., M.H. Rubinstein, Preparation, properties and ageing of tablets prepared from the chlorpropamide-urea solid dispersion **8**, 311
Ford, J.L., M.H. Rubinstein, T.D. Duffy, D.S. Ireland, A comparison of the physical properties of some sulfacetamide eye ointments commercially available in the UK **12**, 11

Ford, J.L., M.H. Rubinstein, J.E. Hogan, Formulation of sustained release promethazine hydrochloride tablets using hydroxypropylmethylcellulose matrices **24**, 327
Propranolol hydrochloride and aminophylline release from matrix tablets containing hydroxypropylmethylcellulose **24**, 339
Ford, J.L., M.H. Rubinstein, F. McCaul, J.E. Hogan, P.J. Edgar, Importance of drug type, tablet shape and added diluents on drug release kinetics from hydroxypropylmethylcellulose matrix tablets **40**, 223
Ford, J.L., A.F. Stewart, J.L. Dubois, The properties of solid dispersions of indomethacin or phenylbutazone in polyethylene glycol **28**, 11
Ford, J.L., see Dabbagh, M.A. **140**, 85
Ford, J.L., see Ford, J.L. **71**, 95, 213
Ford, J.L., see Francomb, M.M. **103**, 87
Ford, J.L., see Mitchell, K. **66**, 233; **100**, 143, 155, 165, 175
Ford, J.L., see Nokhodchi, A. **126**, 189; **129**, 21
Ford, J.L., see Perez-Marcos, B. **111**, 251
Ford, J.L., see Vulovic, N. **55**, 123
Forgacs, E., see Cserhati, T. **141**, 1
Forni, F., G. Coppi, M.A. Vandelli, M.T. Bernabei, An interpretation of the diffusion-type mechanism of drug release from microcapsules **60**, 83
Forse, S.F., see Rowe, R.C. **17**, 347
Forster, R., see Dobetti, L. **128**, 277
Forster, S., G. Buckton, A.E. Beezer, The importance of chain length on the wettability and solubility of organic homologs **72**, 29
Forsyth, A., see Mendelow, A.Y. **49**, 29
Fort, J.J., A.K. Mitra, Physicochemical properties and chromatographic behavior of a homologous series of methotrexate-α,γ-dialkyl ester prodrugs **36**, 7
Solubility and stability characteristics of a series of methotrexate dialkyl esters **59**, 271
Effects of epidermal/dermal separation methods and ester chain configuration on the bioconversion of a homologous series of methotrexate dialkyl esters in dermal and epidermal homogenates of hairless mouse skin **102**, 241
Development of a physical model for simultaneous diffusion/bioconversion of methotrexate dialkyl esters: A potential parallel/sequential metabolic scheme **107**, 9
Fort, J.J., Z. Shao, A.K. Mitra, Transport and degradation characteristics of methotrexate dialkyl ester prodrugs across tape-stripped hairless mouse skin **100**, 233
Fort, J.J., see Trivedi, J.S. **123**, 217
Fortier, M., A. Bonfils, R. Zalisz, M. Heyman, P. Smets, Pharmacokinetic profile of RU 41740, a bacterial immunomodulator, in mice, rats, and monkeys **52**, 27
Fortier, M., see Heyman, M. **37**, 33
Fortman, D., see Shah, V.P. **125**, 99
Foster, T.M., J.J. Dormish, U. Narahari, J.D. Meyer, M. Vrkljan, J. Henkin, W.R. Porter, H. Staack, J.F. Carpenter, M.C. Manning, Thermal stability of low molecular weight urokinase during heat treatment. III. Effect of salts, sugars and Tween 80 **134**, 193
Foucher, C., see Chouinard, F. **72**, 211
Fouli, A.M., Sayed A.A. El, A.A. Badawi, Release of drugs from microcapsules of methacrylate polymers **14**, 95
Fouli, A.M., see Badawi, A.A. **6**, 55
Fournier, C., M. Hamon, M. Hamon, J. Wannebroucq, S. Petiprez, J.P. Pruvo, B. Hecquet, Preparation and preclinical evaluation of bioresorbable hydroxyethylstarch microspheres for transient arterial embolization **106**, 41
Fournier, J., see Galleyrand, J.C. **42**, 225
Fowler, S., D.S. Jones, Modified adherence of *Candida albicans* to human buccal epithelial cells in vitro following treatment with cationic, non-antibiotic antimicrobial agents **84**, 77
Modified adherence of *Candida albicans* to human buccal epithelial cells in vitro following treatment with cationic, non-antibiotic antimicrobial agents **86**, 193
Fowler, S., see Jones, D.S. **105**, 71
Fox, G.G., see Beezer, A.E. **45**, 153
Fox, J., see Masada, T. **49**, 57
Fox, J.L., C.D. Yu, W.I. Higuchi, N.F.H. Ho, General physical model for simultaneous diffusion and metabolism in biological membranes. The computational approach for the steady state case **2**, 41
Fox, J.L., see Kanaya, Y. **16**, 171
Fox, J.L., see Lamson, M.L. **21**, 143
Fox, J.L., see Muhammad, N.A. **21**, 295
Fox, J.L., see Spooner, P.J. **15**, 177
Fox, P.L.A., see Le Brun, P.P.H. **49**, 141
Fox, R., see Antonin, K.H. **130**, 33
Fox, R., see Hastewell, J. **101**, 115; **126**, 245
Frances, C., M.D. Veiga, O.M. Espanol, R. Cadorniga, Preparation, characterization and dissolution of ciprofloxacin/PEG 6000 binary systems **77**, 193
Franchini, M.K., J.T. Carstensen, Failure of high temperature extrapolation of oxidative reactions in solution **111**, 153
Francisco, A.P.G., see Mimoso, I.M. **147**, 109
Franco, M., see Bonina, F.P. **124**, 45
Franco, M., see Trapani, G. **116**, 95; **139**, 215
Francoeur, M., I. Ahmed, S. Sitek, T.F. Patton, Age-related differences in ophthalmic drug disposition. III. Corneal permeability of pilocarpine in rabbits **16**, 203
Francoeur, M., T.F. Patton, Kinetics of corneal drug uptake studied by corneal perfusion in situ. I. Evaluation of system and uptake of ethyl *p*-aminobenzoate in rabbits **2**, 337
Francoeur, M.L., S.J. Sitek, B. Costello, T.F. Patton, Kinetic disposition and distribution of timolol in the rabbit eye. A physiologically based ocular model **25**, 275
Francoeur, M.L., see Harper, Bellantone **30**, 63
Francoeur, M.L., see Riviere, J.E. **127**, 213
Francomb, M.M., J.L. Ford, M.G. Lee, Adsorption of vincristine, vinblastine, doxorubicin and mitozantrone to in-line intravenous filters **103**, 87
Frank, S.G., see Chen-Chow, P.C. **8**, 81, 89
Frank, S.G., see Mueller-Goymann, C.C. **29**, 147
Franke, J.P., see Jonkman, J.H.G. **10**, 177
Franklin Smyth, W., see Yarnitzky, C. **75**, 161
Franssen, E.J.F., F. Moolenaar, D. de Zeeuw, D.K.F. Meijer, Renal specific delivery of sulfamethoxazole in the rat by

coupling to the low molecular weight protein lysozyme via an acid-sensitive linker **80**, R15
Franz, J., see Hampl, J. **144**, 107
Franz, J.M., J.P. Vonderscher, R. Voges, Contribution to the intestinal absorption of ergot peptide alkaloids **7**, 19
Franz, R., see Johnson, K. **73**, 197
Fratini, L., E.E.S. Schapoval, Ciprofloxacin determination by visible light spectrophotometry using iron(III)nitrate **127**, 279
Fratis, A., see Gordon, M.S. **115**, 29
Fredholt, K., N. Mork, M. Begtrup, Hemiesters of aliphatic dicarboxylic acids as cyclization-activated prodrug forms for protecting phenols against first-pass metabolism **123**, 209
Fredholt Thomsen, K., H. Bundgaard, Cyclization-activated phenyl carbamate prodrug forms for protecting phenols against first-pass metabolism **91**, 39
Fredj, G., see Bonhomme, L. **65**, R9
Fredj, G., see Leomy, C. **115**, 235
Freebern, K., see Pavliv, L. **105**, 227
Freeman, D.J., N.V. Sheth, S.L. Spruance, Preclinical studies of iododeoxyuridine in dimethyl sulfoxide for the topical treatment of cutaneous herpes simplex virus infections **48**, 141
Freeman, D.J., see Sheth, N.V. **28**, 201
Freeman, M., see Stead, J.A. **14**, 59
Freeman, S., see Walker, I. **104**, 157; **108**, 225
Freisleben, H.J., see Michel, C. **98**, 131
Freixas, J., see Montero, M.T. **149**, 161
Freney, J., see Lafforgue, C. **147**, 243
Fresta, M., G. Puglisi, Biological effects of CDP-choline loaded long circulating liposomes on rat cerebral post-ischemic reperfusion **134**, 89
Fresta, M., A. Villari, G. Puglisi, G. Cavallaro, 5-Fluorouracil: Various kinds of loaded liposomes: Encapsulation efficiency, storage stability and fusogenic properties **99**, 145
Fresta, M., see Cavallaro, G. **111**, 31
Fresta, M., see Puglisi, G. **118**, 65; **125**, 283
Fresta, M., see Villari, A. **127**, 185
Fricke, S., see Huttenrauch, R. **3**, 289
Fricker, G., see Haeberlin, B. **137**, 103
Fricker, G., see Oechslein, C.R. **139**, 25
Fridriksdottir, H., see Brewster, M.E. **79**, 289
Fridriksdottir, H., see Loftsson, T. **62**, 243; **104**, 181; **127**, 293
Friedman, D., see Amselem, S. **125**, 31
Friedman, D., see Benita, S. **30**, 47
Friedman, D., see Touitou, E. **19**, 323
Friedman, M., D. Harrari, A. Rimer, A. Stabholz, Inhibition of plaque formation by a sustained release delivery system for cetylpyridinium chloride **44**, 243
Friedman, M., see Bialer, M. **20**, 53
Friedman, M., see Hussein, Z. **37**, 97
Friedman, M., see Shani, S. **131**, 33
Friedmand, D., see Brewster, M.E. **125**, 17
Friend, D.R., S.I. Smedley, Solvent drug in ethanol/ethyl acetate enhanced skin permeation of D-norgestrel **97**, 39
Friend, D.R., see Catz, P. **55**, 17; **58**, 93
Friend, D.R., see Leopold, C.S. **126**, 139
Friend, D.R., see McLeod, A.D. **92**, 105
Friend, D.R., see Nolen, III H.W. **107**, 169
Frier, M., see Christrup, L.L. **66**, 169
Frier, M., see Daly, P.B. **10**, 17
Frier, M., see Fitzgerald, P. **83**, 177
Frier, M., see Hardy, J.G. **142**, 129
Friis, G.J., A. Bak, B.D. Larsen, S. Frokjaer, Prodrugs of peptides obtained by derivatization of the C-terminal peptide bond in order to effect protection against degradation by carboxypeptidases **136**, 61
Friis, G.J., H. Bundgaard, Kinetics of degradation of cyclosporin A in acidic aqueous solution and its implication in its oral absorption **82**, 79
Friis, G.J., see Bundgaard, H. **82**, 85
Frijlink, E., see Moolenaar, F. **114**, 117
Frijlink, H.W., A.J.M. Schoonen, C.F. Lerk, The effects of cyclodextrins on drugs absorption. I. In vitro observations **49**, 91
Frijlink, H.W., A.C. Wissens, A.J.M. Schoonen, C.F. Lerk, The effects of cyclodextrins on drug absorption. II. In vivo observations **64**, 195
Frijlink, H.W., see Bootsma, H.P.R. **51**, 213
Frioriksdottir, H., see Loftsson, T. **110**, 169
Frokjaer, S., see Friis, G.J. **136**, 61
Frokjaer, S., see Taub, M.E. **146**, 205
Frontini, R., J.B. Mielck, Formation of formaldehyde in polyebylenegycol and in poloaxmer under stress conditions **114**, 121
Fructus, A., see Tokgoz, N.S. **141**, 27
Fu, Lu M.F., see Powers, M.E. **108**, 49
Fubini, B., M.R. Gasco, M. Gallarate, Microcalorimetric study of microemulsions as potential drug delivery systems. I. Evaluation of enthalpy in the absence of any drug **42**, 19
Microcalorimetric study of microemulsions as potential drug delivery systems. II. Evaluation of enthalpy in the presence of drugs **50**, 213
Fuchs, J., see Michel, C. **98**, 131
Fuentes, A., see Lu, M.F. **71**, 25
Fuhrman, L.C., see Michniak, B.B. **94**, 203
Fuhrman, L.C., see Michniak, B.B. **110**, 231
Fuijihara, Y., see Kaneo, Y. **57**, 263
Fuji, K., see Aoki, H. **115**, 183; **149**, 15
Fujihara, Y., see Kaneo, Y. **44**, 265
Fujii, M., K. Shiozawa, T. Henmi, S. Yamanouchi, H. Suzuki, N. Yamashita, M. Matsumoto, Skin permeation of indomethacin from gel formed by fatty-acid ester and phospholipid **137**, 117
Fujimori, J., Y. Machida, S. Tanaka, T. Nagai, Effect of magnetically controlled gastric residence of sustained release tablets on bioavailability of acetaminophen **119**, 47
Fujimoto, S., see Morimoto, Y. **54**, 27
Fujimoto, T., see Nishihata, T. **42**, 251
Fujimoto, Y., see Nakano, N.I. **7**, 335
Fujinaga, T., see Uekama, K. **10**, 1
Fujioka, Y., see Motozono, H. **108**, 39

Fujita, K., see Kaneo, Y. **57**, 263
Fujita, T., T. Fujita, K. Morikawa, H. Tanaka, O. Iemura, A. Yamamoto, S. Muranishi, Improvement of intestinal absorption of human calcitonin by chemical modification with fatty acids: Synergistic effects of acylation and absorption enhancers **134**, 47
Fujita, T., see Nishikawa, M. **85**, 75
Fujita, Y., see Katsu, T. **53**, 61
Fujiwara, S., see Akimoto, K. **3**, 109
Fukuda, T., Y. Gomita, K. Furuno, Y. Araki, Determination of anpileptic drug concentrations in animal blood by the SLFIA method **36**, 1
Fukui, H., M. Murakami, K. Takada, S. Muranishi, Combinative promotion effect of Azone and fusogenic fatty acid on the large intestinal absorption in rat **31**, 239
Fukuoka, E., see Yamamura, S. **133**, 117
Fukuyasu, H., see Hodoshima, N. **146**, 81
Fukuzaki, H., see Asano, M. **67**, 67
Fukuzaki, H., see Imasaka, K. **68**, 87; **81**, 31
Fullerton, A., see Drustrup, J. **71**, 105
Fullerton, A., see Hansen, L.B. **84**, 253
Funaki, T., S. Furuta, N. Kaneniwa, Discontinuous absorption property of cimetidine **31**, 119
Funaki, T., N. Watari, S. Furuta, N. Kaneniwa, Analysis of the absorption characteristics of cimetidine with the use of the multi-segment absorption model **43**, 59
Funaki, T., see Kaneniwa, N. **26**, 157; **31**, 33; **34**, 231
Funaki, T., see Watari, N. **21**, 85
Fung, F.K.Y., see Duddu, S.P. **94**, 171; **127**, 53
Fung, H.L., see Maier, G.A. **4**, 75
Fung, H.L., see Ogata, H. **5**, 335
Furitsu, H., see Takakura, Y. **105**, 19
Furlanetto, S., see Corti, P. **111**, 83
Furlanetto, S., see Mura, P. **119**, 71
Furlanetto, S., see Santoni, G. **93**, 239
Furlani, P., see Wilding, I.R. **111**, 99
Furneri, P.M., see Puglisi, G. **118**, 65
Furono, K., see Gomita, Y. **54**, 89
Furr, J.R., see El-Falaha, B.M.A. **23**, 239; **25**, 329
Furr, J.R., see Russell, A.D. **27**, 163; **34**, 115; **35**, 227; **36**, 191
Furr, J.R., see Shaker, L.A. **34**, 51
Furst, W., R. Neubert, T. Jurkschat, L. Lucke, Prodrug approach of orotic acid using an absorption model **61**, 43
Furst, W., see Neubert, R. **75**, 89
Furuno, K., see Fukuda, T. **36**, 1
Furuseth, S., see Bjerga, Bjaen A.K. **92**, 183
Furuta, S., see Funaki, T. **31**, 119; **43**, 59
Furuta, S., see Kaneniwa, N. **31**, 33
Furuya, A., see Akimoto, M. **100**, 133; **117**, 31
Furuya, A., see Yamada, K. **119**, 1
Furuya, Y., see Takada, K. **44**, 107
Futo, T., see Ohta, N. **118**, 85
Fuwa, T., see Murakami, T. **54**, 259
Fyhr, P., A. Brodin, A preformulation study on the kinetics of pralidoxime chloride in cncentrated acidic solution **40**, 157
Fyhr, P., M. Nicklasson, K. Gunnvald, A. Brodin, A preformulation study on the kinetics of HI-6 in concentrated solution **40**, 193

G

Gaber, M., see Al-Shora, H.I. **47**, 209
Gaber, M., see Moustafa, M.A. **34**, 207
Gaber, M., see Turkistani, A.A.A. **59**, R1
Gabr, K.E., see Meshali, M.M. **89**, 177
Gabriel, D.L., see Kearney, A.S. **104**, 169
Gabrielson, G., see Silvestri, S. **71**, 65; **73**, 163
Gabrijelcic, V., M. Sentjurc, Influence of hydrogels on liposome stability and on the transport of liposome entrapped substances into the skin **118**, 207
Gabrijelcic, V., M. Sentjurc, J. Kristl, Evaluation of liposomes as drug carriers into the skin by one-dimensional EPR imaging **62**, 75
Gabrijelcic, V., M. Sentjure, M. Schara, The measurement of liposome entrapped molecules' penetration into the skin: A 1D-EPR and EPR kinetic imaging study **102**, 151
Gadalla, M.A.F., F. Touchard, M.W. Samaha, The effect of the composition of binary systems on the solubility and solubility parameter estimation of nalidixic and salicylic acids **38**, 71
Gaddipati, N., M. Graziosi, K. Ellway, M. Ganesan, H. Schreier, A novel aerosol inhalation device for pressurized metered dose inhalation aerosols: Gammascintigraphic evaluation of pulmonary deposition profiles and comparison with commercial inhalation devices **128**, 55
Gajan, V., see Le Corre, P. **107**, 41
Gajdos, M., see Piotrovskij, V.K. **119**, 213
Galindo, C., see de Bolos, J. **40**, 175
Galinsky, R.E., see Anderson, B.D. **43**, 261
Galinsky, R.E., G. Levy, Effect of cimetidine on acetaminophen pharmacokinetics in rats **10**, 301
Gallarate, M., M.R. Gasco, M. Trotta, P. Chetoni, M.F. Saettone, Preparation and evaluation in vitro of solutions and o/w microemulsions containing levobunolol as ion-pair **100**, 219
Gallarate, M., see Fubini, B. **42**, 19; **50**, 213
Gallarate, M., see Gasco, M.R. **69**, 193
Gallardo, M., G. Couarraze, B. Denizot, L. Treupel, P. Couvreur, F. Puisieux, Study of the mechanisms of formation of nanoparticles and nanocapsules of polyisobutyl-2-cyanoacrylate **100**, 55
Gallardo, M., see Casals, E. **143**, 171
Gallardo, M., see de Bolos, J. **40**, 175
Gallardo, M., see Gallardo, M. **100**, 55
Gallardo, V., J. Salcedo, A. Parera, A. Delgado, Effect of the preservatives antipyrin, benzoic acid and sodium metabisulfite on properties of the nitrofurantoin/solution interface **71**, 223
Galleyrand, J.C., J. Fournier, J.L. Chanal, C. Puozzo, Comparative study of binding of an antiallergic substance to serum albumin and erythrocytes in man, in the dog and in the rat **42**, 225
Gallo, J.M., C.T. Hung, D.G. Perrier, Analysis of albumin microsphere preparation **22**, 63

Gambari, R., V. Chiorboli, G. Feriotto, C. Nastruzzi, TAPP (tetra-*p*-amidinophenoxyneopentane) inhibits the binding of nuclear factors to target DNA sequences **72**, 251
Gambari, R., see Cortesi, R. **105**, 181; **139**, 69
Gambari, R., see Esposito, E. **117**, 151
Gambari, R., see Nastruzzi, C. **90**, 43
Gamble, M.D., see Field, W.N. **51**, 175
Gamlen, M.J., see Jones, H.P. **104**, 253
Gander, B., V. Beltrami, R. Gurny, E. Doelker, Effects of the method of drug incorporation and the size of the monolith on drug release from cross-linked polymers **58**, 63
Gander, B., P. Johansen, H. Nam-Tran, H.P. Merkle, Thermodynamic approach to protein microencapsulation into poly(D,L-lactide) by spray drying **129**, 51
Gander, B., K. Ventouras, R. Gurny, E. Doelker, In vitro dissolution medium with supramicellar surfactant concentration and its relevance for in vivo absorption **27**, 117
Gander, B.A., see Thomasin, C. **147**, 173
Ganderton, D., see Buckton, G. **42**, 35
Ganderton, D., see Leigh, T.R. **67**, 275
Ganderton, D., see Malamataris, S. **70**, 69
Ganderton, D., see Timsina, M.P. **101**, 1
Gandhi, R., J. Robinson, Mechanisms of penetration enhancement for transbuccal delivery of salicylic acid **85**, 129
Ganem-Quintanar, A., C. Lafforgue, F. Falson-Rieg, P. Buri, Evaluation of the transepidermal permeation of diethylene glycol monoethyl ether and skin water loss **147**, 165
Ganesan, M., see Gaddipati, N. **128**, 55
Ganesan, M.G., N.D. Weiner, G.L. Flynn, N.F.H. Ho, Influence of liposomal drug entrapment on percutaneous absorption **20**, 139
Ganesan, M.G., see Schwinke, D.L. **20**, 119
Gangal, S.V., see Jaggi, M. **69**, 77
Gangarosa, Sr. L.P., J.M. Hill, Modern iontophoresis for local drug delivery **123**, 159
Ganley, J.A., see McTaggart, C.M. **19**, 139
Ganza-Gonzalez, A., J.L. Vila-Jato, S. Anguiano-Igea, F.J. Otero-Espinar, J. Blanco-Mendez, A proton nuclear magnetic resonance study of the inclusion complex of naproxen with β-cyclodextrin **106**, 179
Gao, P., see Sung, K.C. **142**, 53
Garay, R.P., Azelastine: Well known ciliotoxic agent? [1] **136**, 181
Garcia Anton, J.M., see Reig, F. **44**, 257, 269
Garcia, B., J.P. Marty, J. Wepierre, Factors governing percutaneous absorption of alcohols from water/isopropyl myristate/polysorbate mixtures **4**, 205
Garcia, F.F., see Guzman, M. **80**, 119
Garcia, J., see Lucero, M.J. **116**, 31
Garcia, L., see Bedmar, M.C. **138**, 95
Garcia, M.J., A.A. Dominguez-Gil, M. Cepeda, A. Dominguez-Gil, Influence of experimental renal impairment in the pharmacokinetics of cefoxitin after intravenous administration to rabbits **5**, 117
Garcia, M.L., see Egea, M.A. **67**, 103
Garcia-Arias, T., see Alonso, M.J. **42**, 27
Garcia-Celma, M.J., N. Azemar, M.A. Pes, G. Solans, Solubilization of antifungal drugs in water/POE(20) sorbitan monooleate/oil systems **105**, 77
Garcia-Encina, G., see Torres, D. **59**, 9; **121**, 239
Garcia-Gonzalez, N., I.W. Kellaway, H. Blanco-Fuente, S. Anguiano-Igea, B. Delgado-Charro, F.J. Otero-Espinar, J. Blanco-Mendez, Influence of β-cyclodextrin concentration and polyacrylic acid molecular weight on swelling and release characteristics of metoclopramide-containing hydrogels **100**, 25
Design and evaluation of buccoadhesive metoclopramide hydrogels composed of poly(acrylic acid) crosslinked with sucrose **100**, 65
Influence of glycerol concentration and carbopol molecular weight on swelling and drug release characteristics of metoclopramide hydrogels **104**, 107
Garcia-Gonzalez, N., see Otero-Espinar, F.J. **75**, 37; **79**, 149
Gard, P.R., see Malhi, J.S. **43**, 77
Gardner, M.J., W.J. Jusko, Effects of oral contraceptives and tobacco use on the metabolic pathways of theophylline **33**, 55
Gareiss, J., see Blume, A. **99**, 219
Garr, J.S.M., M.H. Rubinstein, Direct compression characteristics of xylitol **64**, 223
An investigation into the capping of paracetamol at increasing speeds of compression **72**, 117
The effect of rate of force application on the properties of microcrystalline cellulose and dibasic calcium phosphate mixtures **73**, 75
The influence of moisture content on the consolidation and compaction properties of paracetamol **81**, 187
Consolidation and compaction characteristics of a three-component particulate system **82**, 71
Garren, J., see Adjei, A. **90**, 141
Garren, K.W., A.J. Repta, Incompatibility of cisplatin and reglan injectable **24**, 91
Buccal drug absorption. I. Comparative levels of esterase and peptidase activis in rat and hamster buccal and intestinal homogenates **48**, 189
Garren, K.W., R.A. Pyter, Aqueous solubility properties of a dibasic peptide-like compound 2,3-dihydroxy-5-methylhexyl)histidinamide **63**, 167
Garrigues Pelufo, T.M, A.T. Perez-Varona, M.V. Bermejo, A. Martin-Villodre, Absorption-partition relationships for true homologous series of xenobiotics as a possible approach to study mechanisms of surfactants in absorption. IV. Phenylacetic acid derivatives and anionic surfactants **79**, 135
Garrigues, T.M., E.F. Collado, S. Fabra-Campos, M.D. Perez-Buendia, A. Martin-Villodre, J.M. Pla-Delfina, Absorption-partition relationships for true homologous series of xenobiotics as a possible approach to study mechanisms of surfactants in absorption. III. Aromatic amines and cationic surfactants **57**, 189
Garrigues, T.M., A.T. Perez-Varona, E. Climent, M.V. Bermejo, A. Martin-Villodre, J.M. Pla-Delfina, Gastric absorption of acidic xenobiotics in the rat: Biophysical interpretation of an apparently atypical behaviour **64**, 127

Garrigues, T.M., M.J. Segura-Bono, M.V. Bermejo, V. Merino, J.M. Pla-Delfina, Compared effects of synthetic and natural bile acid surfactant on xenobiotic absorption. II. Studies with sodium glycocholate to confirm a hypothesis **101**, 209
Garrigues, T.M., see Perez-Buendia, M.D. **55**, 165
Garrigues, T.M., see Segura-Bono, M.J. **107**, 159
Garvin, C.P., see McGovern, J.G. **149**, 251
Garvin, K., see Sampath, S.S. **78**, 165
Gasco, M.R., M. Gallarate, F. Pattarino, In vitro permeation of azelaic acid from viscosized microemulsions **69**, 193
Gasco, M.R., F. Pattarino, F. Lattanzi, Long-acting delivery systems for peptides: Reduced plasma testosterone levels in male rats after a single injection **62**, 119
Gasco, M.R., M. Trotta, Nanoparticles from microemulsions **29**, 267
Gasco, M.R., M. Trotta, M.E. Carlotti, R. Carpignano, Contribution of carboxylic acids to the permeability of imipramine **18**, 235
Gasco, M.R., see Cavalli, R. **89**, R9; **117**, 243
Gasco, M.R., see Fubini, B. **42**, 19; **50**, 213
Gasco, M.R., see Gallarate, M. **100**, 219
Gasco, M.R., see Morel, S. **105**, 189; **119**, 126; **132**, 259
Gasco, M.R., see Pattarino, F. **91**, 157
Gasco, M.R., see Trotta, M. **143**, 67
Gascon, A.R., see Hernandez, R.M. **139**, 45
Gaskin, P.W., see Hall, S.B. **70**, 251
Gaspar, M.M., see Cruz, M.E.M. **96**, 67
Gaspar, R., V. Preat, M. Roland, Nanoparticles of polyisohexylcyanoacrylate (PIHCA) as carriers of primaquine: Formulation, physico-chemical characterization and acute toxicity **68**, 111
Gaspar, R., see Lourenco, C. **138**, 1
Gaspar, R., see Martins, M.B.F. **142**, 75
Gasperini, E., see Catellani, P.L. **88**, 285
Gasperlin, M., Smid-Korba Kristl, J. Kerc, The structure elucidation of semisolid w/o emulsion systems containing silicone surfactant **107**, 51
Gassim, A.E.H., P.G. Takla, K.C. James, Polymorphism and possible intramolecular bonding in benperidol **34**, 23
Gatlin, L.A., see Schoen, M.P. **114**, 159
Gatti, R., R. Gotti, V. Cavrini, M. Soli, A. Bertaccini, F. Carparelli, Stability study of prostaglandin E_1 (PGE_1) in physiological solutions by liquid chromatography (HPLC) **115**, 113
Gaudy, D., see El Moussaoui, A. **106**, 109
Gaudy, D., see Petrovick, P.D. **76**, 49
Gaudy, D., see Rizk, S. **126**, 57
Gaudy, D., see Ruizk, S. **112**, 125
Gautam, A., see Parimoo, P. **134**, 231
Gautier, J.C., see Boulenc, X. **123**, 71
Gautier, J.C., see Le Ray, A.M. **106**, 201
Gavach, C., see Raffin, F. **120**, 205
Gavriloff, C., see Leroy, P. **82**, 157
Gay, C.L., T.M. Murphy, J. Hadgraft, I.W. Kellaway, J.C. Evans, C.C. Rowlands, An electron spin resonance study of skin penetration enhancers **49**, 39
Gayo-Otero, M., see Esclusa-Diaz, M.T. **142**, 183
Gayot, A.T., see Flament, M.P. **148**, 93
Gazso, L.G., see Dam, A.M. **121**, 245
Gazzaniga, A., P. Iamartino, G. Maffione, M.E. Sangalli, Oral delayed-release system for colonic specific delivery **108**, 77
Gazzaniga, A., M.E. Sangalli, U. Conte, C. Caramella, P. Colombo, A. La Manna, On the release mechanism from coated swellable minimatrices **91**, 167
Gazzaniga, A., see Caramella, C. **44**, 177
Gazzaniga, A., see Colombo, P. **63**, 43
Gazzaniga, A., see Gazzaniga, A. **91**, 167
Gazzaniga, A., see Giordano, F. **62**, 153
Gazzaniga, A., see Peppas, N.A. **51**, 77
Gazzaniga, A., see Sangalli, M.E. **91**, 151
Gazzaniga, A., see Wilding, I.R. **111**, 99
Gebre-Mariam, T., see Armstrong, N.A. **34**, 125
Gehenot, A., R.C. Rao, G. Maire, M. Cachon, Values of thermal analysis in the critical evaluation of classical methods of melting point determination **45**, 13
Gelbcke, M., see Beten, D.B. **88**, 31
Genetet, B., see Rolland, A. **39**, 173
Gengenbacher, T., see Haeberlin, B. **137**, 103
Genta, I., see Pavanetto, F. **48**, 255; **51**, 55; **54**, 143; **74**, 175; **84**, 151
Georgakopoulos, P.P., see Avgoustakis, K. **79**, 67
Georgarakis, M., see Amperiadou, A. **115**, 1
Georgiacodis, M., see Macheras, P. **52**, 249
Georgieva, M., see Simeonova, M. **43**, 267
Geraldes, C.F., see Moreira, J.N. **147**, 153
Gerard, C.A., see Maurin, M.B. **94**, 11
Gerber, J.J., J. Van der Watt, A.P. Lotter, Physical characterisation of three pseudopolymorphs of cyclopenthiazide **69**, 265
Physical characterisation of solid forms of cyclopenthiazide **73**, 137
Gergich, G., see Zatz, J.L. **9**, 315
Gerhards, J., see Bury, M. **76**, 207; **124**, 183
Gerritsen, H.R., see Herremans, J.L.M. **129**, 191
Gerritsma, K.W., see Cornelissen, P.J.G. **1**, 173; **3**, 205
Gerritsma, K.W., see Van der Griend, J. **1**, 257
Getz, J.J., see Beall, H.D. **93**, 37
Getz, J.J., see Sloan, K.B. **93**, 27
Ghaderi, R., C. Sturesson, J. Carlfors, Effect of preparative parameters on the characteristics of poly (D,L-lactide-*co*-glycolide) microspheres made by the double emulsion method **141**, 205
Ghaly, G.M., see Nishihata, T. **38**, 83
Ghanem, A.H., H. Mahmoud, W.I. Higuchi, P. Liu, W.R. Good, The effects of ethanol on the transport of lipophilic and polar permeants across hairless mouse skin: Methods/validation of a novel approach **78**, 137
Ghanem, A.H., see Inamori, T. **105**, 113
Ghanem, A.H., see Kim, Y.H. **80**, 17
Ghanem, A.H., see Lee, P.H. **93**, 139
Ghanem, A.H., see Liu, P. **64**, 7; **78**, 123
Ghanem, A.H., see Peck, K.D. **98**, 141
Ghanem, A.H., see Seta, Y. **81**, 89

Gharaibeh, A.E.M., H.N. Alkaysi, M.S. Salam, K.I.A. Gharaibeh, Nil effect of cigarette smoking on ranitidine pharmacokinetics **100**, 253
Gharaibeh, A.M., see Alkaysi, H.N. **87**, 73
Gharaibeh, K.I.A., see Gharaibeh, A.E.M. **100**, 253
Gharavi, M., K.C. James, The properties of testosterone and related androgens crystallized from normal alkanols **14**, 325
Gharavi, M., K.C. James, L.M. Sanders, Solubilities of mestanolone, methandienone, methyltestosterone, nandrolone and testosterone in homologous series of alkanes and alkanols **14**, 333
Ghassabian, S., T. Ehtezazi, Forutan S. Mohsen, Mortazavi Alireza, Dexamethasone-loaded magnetic albumin microspheres: Preparation and in vitro release **130**, 49
Ghebre-Sellassie, I., R.H. Gordon, D.L. Middleton et al., A unique application and characterization of Eudragit E 30 D film coatings in sustained release formulations **31**, 43
Ghebre-Sellassie, I., R.H. Gordon, R.U. Nesbitt, M.B. Fawzi, Evaluation of acrylic-based modified-release film coatings **37**, 211
Ghebre-Sellassie, I., see Yang, S.T. **60**, 109; **86**, 247
Gheyouche, R., see Aignasse, M.F. **113**, 241
Ghezzo, E., L. Benedetti, M. Rochira, F. Biviano, L. Callegaro, Hyaluronane derivative microspheres as NGF delivery devices: Preparation methods and in vitro release characterization **87**, 21
Ghobashy, A.A., see Chiori, C.O. **17**, 121
Ghodbane, S., J.A. McCauley, Study of the polymorphism of 3-(((3-(2-(7-chloro-2-quinolinyl)-(*E*)-ethenyl)phenyl)((3-dimethylamino-3-oxopropyl)thio)methyl)thio)propanoic acid (MK571) by DSC, TG, XRPD and solubility measurements dimethylcarbamoyl)ethylthio]methylthio]propionic acid **59**, 281
Ghodbane, S., see Rivera, S.L. **108**, 31
Ghogare, A.B., see Sadana, G.S. **70**, 195
Ghorab, M., see Ammar, H.O. **103**, 237
Ghorab, M.M., see Ammar, H.O. **103**, 237
Ghorab, M.M., see Badawy, S.I.F. **128**, 45; **145**, 137
Ghosh, S., D.J.W. Grant, Determination of the solubilities of crystalline solids in solvent media that induce phase changes: Solubilities of 1,2-dialkyl-3-hydroxy-4-pyridones and their formic acid solvates in formic acid and water **114**, 185
Ghosh, T.K., C.S. Chiao, R.D. Gokhale, In vitro release and permeation of levobunolol from various transdermal formulations **82**, 39
Ghosh, T.K., M.J. Habib, K. Childs, M. Alexander, Transdermal delivery of metoprolol. I. Comparison between hairless mouse and human cadaver skin and effect of *n*-decylmethyl sulfoxide **88**, 391
Ghyczy, M., see Blume, A. **99**, 219
Giammona, G., B. Carlisi, G. Cavallaro, I. Donato, F. Pinio, V.T. Liveri, Calorimetric investigation of the interaction between α,β-poly(*N*-hydroxyethyl)-DL-aspartamide and surfactants **64**, 239
Giammona, G., G. Cavallaro, G. Pitarresi, C. Ventura, A.S. Palazzo, Synthesis and therapeutic potential of a macromolecular prodrug of diflunisal **105**, 57
Giammona, G., G. Pitarresi, V. Tomarchio, S. Spampinato, P. Govoni, T. Campana, New hydrogel matrices based on chemical crosslinked α,β-polyasparthydrazide: Synthesis, characterization and in vivo biocompatibility studies **127**, 165
Giammona, G., G. Puglisi, B. Carlisi, R. Pignatello, A. Spadaro, A. Caruso, Polymeric prodrugs: α,β-poly(*N*-hydroxyethyl)-DL-aspartamide as a macromolecular carrier for some non-steroidal anti-inflammatory agents **57**, 55
Giammona, G., see Castelli, F. **59**, 19; **70**, 43; **88**, 1
Giammona, G., see Cavallaro, G. **90**, 195; **111**, 31
Giammona, G., see Puglisi, G. **125**, 283
Giammona, G., see Turco Liveri, V. **99**, 285
Gianasi, E., F. Cociancich, I.F. Uchegbu, A.T. Florence, R. Duncan, Pharmaceutical and biological characterisation of a doxorubicin-polymer conjugate (PK1) entrapped in sorbitan monostearate Span 60 niosomes **148**, 139
Giannaccini, B., see Saettone, M.F. **20**, 187; **25**, 73; **31**, 261; **43**, 67; **51**, 203; **72**, 131; **86**, 159; **126**, 79
Giannakou, S.A., P.P. Dallas, D.M. Rekkas, N.H. Choulis, Development and in vitro evaluation of nitrendipine transdermal formulations using experimental design techniques **125**, 7
Giannessi, C., see Di, Colo G. **50**, 27
Giannini, R.P., see Shefter, E. **1**, 49
Gibaldi, M., P.J. McNamara, Steady-state concentrations of drugs with short half-lives when administered in oral sustained release formulations **2**, 167
Gibaldi, M., see Colburn, W.A. **1**, 121
Gibbins, L.B., K.C. James, Calculating quantities in dispersible powders containing antibiotics **4**, 353
Gibbons, W.A., see Del Olmo, E. **81**, 267
Gibbons, W.A., see Flinn, N. **138**, 167; **143**, 125
Gibbons, W.A., see Toth, I. **73**, 259; **77**, 13; **79**, 39; **83**, 131; **105**, 241; **125**, 117
Gibbons, W.A., see Valko, K. **83**, 123
Gibbons, W.A., see Wood, I.P. **87**, 141
Gibson, S.H.M., R.C. Rowe, E.F.T. White, Characterisation of the structure of filled polymer film coatings using ion beam etching **47**, 205
Mechanical properties of pigmented tablet coating formulations and their resistance to cracking. I. Static mechanical measurement **48**, 63
Quantitative assessment of additive-polymer interaction in pigmented hydroxypropyl methylcellulose formulations using immersion calorimetry **48**, 113
The mechanical properties of pigmented tablet coating formulations and their resistance to cracking. II. Dynamic mechanical measurement **50**, 163
Gienau, B., see Neubert, R. **75**, 89
Gierer, D.S., see Sinko, C.M. **114**, 85
Gilbert, D.L., T. Okano, T. Miyata, S.W. Kim, Macromolecular diffusion through collagen membranes **47**, 79

Gilbert, J.C., J. Hadgraft, A. Bye, L.G. Brookes, Drug release from Pluronic F-127 gels **32**, 223
Gilbert, J.C., C. Washington, M.C. Davies, Pietra A.M Di, The behaviour of Pluronic F127 in aqueous solution studied using fluorescent probes **40**, 93
Gilbert, P., see Al-Adham, I.S.I. **34**, 45
Gilbert, P., see Chawner, J.A. **55**, 209
Gilbert, P., see Collier, P.J. **66**, 201; **74**, 195
Gilbert, P., see Plumpton, E.J. **30**, 237, 241
Gilhar, D., see Touitou, E. **78**, 85
Gilligan, C.A., A. Li Wan Po, Factors affecting drug release from a pellet system coated with an aqueous colloidal dispersion **73**, 51
Oral vaccines: Design and delivery **75**, 1
Gilligan, C.A., see Al-Kassas, R.S. **94**, 59
Gilligan, C.A., see Li Wan, Po A. **66**, 111
Gilligan, C.A., see Wong, L.P. **83**, 95
Gimenez, F., E. Postaire, P. Prognon, Hoang M.D. Le, J.M. Lecomte, D. Pradeau, G. Hazebroucq, Study of thiorphan degradation **43**, 23
Gines, J.M., M.J. Arias-Blanco, J.R. Moyano, P.J. Sanchez-Soto, Thermal investigation of crystallization of polyethylene glycols in solid dispersions containing oxazepam **143**, 247
Gines, J.M., M.D. Veiga, M.J. Arias, A.M. Rabasco, Elaboration and thermal study of interactions between cinnarizine and gelucire^{R} 53/10 physical mixtures and solid dispersions **126**, 287
Gines, J.M., see Arias, M.J. **123**, 25
Gines, J.M., see Arias-Blanco, M.J. **143**, 113
Gines, J.M., see Moyano, J.R. **114**, 95
Gines, J.M., see Moyona, J.R. **148**, 211
Gines, J.M., see Rabasco, A.M. **67**, 201
Gines-Dorado, J.M., see Fernandez-Arevalo, M. **95**, 117
Gingrich, J., see Givens, R.S. **29**, 67
Ginsel, L.A., see de Haan, F.H.N. **56**, 75
Giordano, F., A. Gazzaniga, G.P. Bettinetti, A. La Manna, The influence of water content on the binding capacity of β-cyclodextrin **62**, 153
Giordano, F., see Moyona, J.R. **148**, 211
Gipps, E.M., P. Groscurth, J. Kreuter, P.P. Speiser, The effects of polyalkylcyanoacrylate nanoparticles in human normal and malignant mesenchymal cells in vitro **40**, 23
Girela Vilchez, F., see Contreras Claramonte, M.D. **98**, 37
Girona, V., J. Estelrich, M. Pujol, J. Bolos, Ampicillin polymers: Identification by gel-filtration chromatography **41**, 241
Girona, V., see Munoz, M. **139**, 249
Girona, V., see Pujol, M. **58**, 103; **146**, 263
Girona, V., see Torres, F. **129**, 275
Giunchedi, P., U. Conte, L. Maggi, A. La Manna, Hydrophilic matrices for the extended release of a model drug exhibiting pH-dependent solubility **85**, 141
Giunchedi, P., L. Maggi, U. Conte, C. Caramella, Ketoprofen pulsatile absorption from 'multiple unit' hydrophilic matrices **77**, 177
Giunchedi, P., L. Maggi, U. Conte, A. La Manna, Linear extended release of a water-insoluble drug, carbamazepine, from erodible matrices **94**, 15
Giunchedi, P., S. Scalia, L. Maggi, U. Conte, Ursodeoxycholic acid: Improvement of dissolution behaviour and its HPLC determination **130**, 41
Giunchedi, P., see Bonferoni, M.C. **80**, 109
Giunchedi, P., see Maggi, L. **99**, 173; **135**, 73
Giunchedi, P., see Ochoa, Machiste E. **126**, 65
Giunchedi, P., see Pavanetto, F. **84**, 151
Giunchedi, P., see Sangalli, M.E. **91**, 151
Givens, R.S., J. Gingrich, S. Mecklenburg, Photochemistry of flunitrazepam: A product and model study **29**, 67
Gizurarson, S., C. Marriott, G.P. Martin, E. Bechgaard, The influence of insulin and some excipients used in nasal insulin preparations on mucociliary clearance **65**, 243
Gizurarson, S., T. Thorvaldsson, P. Sigurdsson, E. Gunnarsson, Selective delivery of insulin into the brain: Intraolfactory absorption **140**, 77
Erratum: Selective delivery of insulin into the brain: Intraolfactory absorption (Int. J. Pharm. 140 (1996) 77) **146**, 133
Gizurarson, S., see Bechgaard, E. **87**, 125; **89**, 147; **128**, 287
Gjaltema, D., see De, Boer A.H. **138**, 45
Gjellan, K., C. Graffner, Influence of amount of hard fat in suppositories on the in vitro release rate and bioavailability of paracetamol. II. A comparison between three compositions and a rectal solution **104**, 215
Comparative dissolution studies of rectal formulations using the basket, the paddle and the flow-through methods. II. Ibuprofen in suppositories of both hydrophilic and lipophilic types **112**, 233
Gjellan, K., C. Graffner, H. Quiding, Influence of amount of hard fat in suppositories on the in vitro release rate and bioavailability of paracetamol and codeine. I. A comparison of three suppository compositions in vivo **102**, 71
Gjellan, K., C. Graffner, S. Strid, Exploratory studies on the in vitro release and bioavailability of dextropropoxyphene from lipophilic and hydrophilic suppositories **102**, 213
Glazer, M., see Karehill, P.G. **64**, 35
Glazer, M., see Nystrom, C. **23**, 255
Gleason, W.B., Calculated powder patterns of (*R*,*S*)-propanolol hydrochloride—A comment **127**, 109
Gleeson, J.M., see Seminoff, L.A. **54**, 251
Gnappi, G., see Redenti, E. **129**, 289
Go, J., see Chow, K.Y. **20**, 3; **25**, 41
Go, L.T., see Zanen, P. **107**, 211; **114**, 111
Gobert, J.G., see Penaud, J.F. **142**, 1
Gochoco, C.H., F.M. Ryan, J. Miller, P.L. Smith, I.J. Hidalgo, Uptake and transepithelial transport of the orally absorbed cephalosporin cephalexin, in the human intestinal cell line, Caco-2 **104**, 187
Gode, K.D., see Pandit, J.K. **21**, 129
Godwin, D.A., see Michniak, B.B. **116**, 201
Goehl, T.J., G.M. Sundaresan, V.K. Prasad, Analytical methodology applicable in dissolution testing of norethindrone-mestranol tablets **11**, 181

Studies of the dissolution characteristics of norethindrone-mestranol tablets **15**, 115
Goethals, F., see Tasset, C. **58**, 41
Gogate, U.S., A.J. Repta, *N*-(acyloxyalkoxycarbonyl) derivatives as potential prodrugs of amines. II. Esterase-catalysed release of parent amines from model prodrugs **40**, 249
Gogate, U.S., A.J. Repta, J. Alexander, *N*-(acyloxyalkoxycarbonyl) derivatives as potential prodrugs of amines. I. Kinetics and mechanism of degradation in aqueous solutions **40**, 235
Gogoi, P.J., see Vyas, S.P. **69**, 5
Goidas, P., see Malamataris, S. **68**, 51; **103**, 205
Gokhale, R.D., see Ghosh, T.K. **82**, 39
Goldbach, P., H. Brochart, P. Wehrle, A. Stamm, Sterile filtration of liposomes: Retention of encapsulated carboxyfluorescein **117**, 225
Goldbach, P., S. Dumont, R. Kessler, P. Poindron, A. Stamm, Preparation and characterization of interferon-γ-containing liposomes **123**, 33
Goldbach, P., see Brochart, H. **91**, 95
Goldberg, A.H., see Lin, S.Y. **95**, 57
Goldberg, E.P., see Jayakrishnan, A. **106**, 221
Goldberg-Cettina, M., P. Liu, J. Nightingale, T. Kurihara-Bergstrom, Enhanced transdermal delivery of estradiol in vitro using binary vehicles of isopropyl myristate and short-chain alkanols **114**, 237
Goldblum, R., see Gordon, M.S. **115**, 29
Golomb, G., see Hoffman, A. **111**, 197
Gombar, V.K., see Baveja, S.K. **41**, 55
Gomez-Amoza, J.L., see Cal, S. **129**, 253; **136**, 13
Gomez-Amoza, J.L., see Doldan, C. **124**, 69
Gomez-Amoza, J.L., see Landin, M. **91**, 123, 133, 143; **103**, 9; **107**, 247; **109**, 1; **123**, 143
Gomez-Amoza, J.L., see Perez-Marcos, B. **67**, 113; **100**, 207
Gomiscek, S., see Sher, A. **90**, 181; **99**, 119
Gomita, Y., M. Nanba, K. Furono, K. Eto, Y. Araki, Microdetermination of cimetidine in rat plasma by high-performance liquid chromatography **54**, 89
Gomita, Y., see Fukuda, T. **36**, 1
Gonda, I., Development of a systemic theory of suspension inhalation aerosols. I. A framework to study the effects of aggregation on the aerodynamic behaviour of drug particles **27**, 99
Gonda, I., A.F. El Khalik, A.Z. Britten, Hexamethylmelamine aerosols prepared in an evaporation-condensation generator **27**, 255
Gonda, I., see Chan, H.K. **41**, 147; **68**, 179
Gonda, I., see Rodley, G.A. **95**, 143
Gong, J.P., see Kim, C.K. **106**, 25
Goni, I., see Castellano, I. **146**, 71
Goni, I., see Ferrero, M.C. **149**, 233
Goni, I., see Velasco, M.V. **136**, 107
Gonsho, A., G. Imanidis, P. Vogt, E.R. Kern, H. Tsuge, M.H. Su, S.H. Choi, W.I. Higuchi, Controlled (trans) dermal delivery of an antiviral agent (acyclovir). I. An in vivo animal model for efficacy evaluation in cutaneous HSV-1 infections **65**, 183
Gonzalez, A.G., Practical digest for the evaluation of acidity constants of drugs by reversed-phase high performance liquid chromatography **91**, R1
Optimization of pharmaceutical formulations based on response-surface experimental designs **97**, 149
Gonzalez, A.G., A.G. Asuero, Application of the extended Hildebrand solubility parameter treatment for optimizing reversed-phase high-performance liquid chromatography determination of pharmaceuticals **93**, 183
Gonzalez, A.G., M.A. Herrador, A.G. Asuero, Evaluation of ionization constants of drugs in aqueous organic mixtures from reversed phase high-performance liquid chromatography **84**, R1
Solubility of theophylline in aqueous *N*,*N*-dimethylformamide mixtures **108**, 149
Hydrophobicity of α-adrenoceptor blocking agents: Study of correlations between retention in reversed-phase HPLC systems and octanol-water partition constants **120**, 215
Determination of β-adrenoceptor blocking agents in tablets using reversed-phase high-performance liquid chromatography **123**, 149
Gonzalez, A.G., F. Pablos, Evaluation of acidity constants for sparingly soluble compounds from fluorescence measurements **67**, R1
Gonzalez, A.G., F. Pablos, A.G. Asuero, Spectrophotometric determination of acidity constants of compounds with unsuitable absorption features **72**, 193
Gonzalez-Rodriguez, M.L., M.J. Fernandez-Hervas, I. Caraballo, A.M. Rabasco, Design and evaluation of a new central core matrix tablet **146**, 175
Gonzalez-Rothi, R.J., see Schreier, H. **87**, 183
Gonzalez-Rothi, R.J., see Wichert, B.V. **78**, 227
Gooch, C.A., see Beezer, A.E. **29**, 237; **45**, 153
Gooch, C.A., see Sekulovic, D. **38**, 255
Good, W.R., see Ghanem, A.H. **78**, 137
Good, W.R., see Liu, P. **64**, 7; **78**, 123
Goodfield, J., see Bailey, A.J.G. **22**, 283
Goodhart, F.W., see Nesbitt, R.U. **26**, 215
Goodman, M., B.W. Barry, Lipid-protein-partitioning (LPP) theory of skin enhancer activity: Finite dose technique **57**, 29
Goodman, M., see Gould, P.L. **19**, 149
Goodwin, S.R., see Miller, II K.J. **98**, 101
Gooris, G.S., see Bouwstra, J.A. **79**, 141; **84**, 205; **144**, 81
Gooris, G.S., see Cornwell, P.A. **127**, 9
Goossen, R.M.J., see Van der Houwen, O.A.G.J. **99**, 73
Gopal, N.G.S., S. Rajagopalan, Effect of γ radiation and ethylene oxide on neomycin sulphate **9**, 359
Gopal, N.G.S., see Bhalla, H.L. **17**, 351
Gopferich, A., G. Lee, An improved diffusion/compartmental model for transdermal drug delivery from a matrix-type device **71**, 237
Measurement of drug diffusivity in stratum corneum membranes and a polyacrylate matrix **71**, 245
Gorashi, A.S., see Ali, A.A. **19**, 297
Gordon, J.D., A.H.L. Chow, Modification of phenytoin crystals. II. Influence of 3-propanoyloxymethyl-5,5-diphenylhy-

dantoin on solution-phase crystallization and related crystal properties **79**, 171
Gordon, J.D., see Chow, A.H.L. **126**, 11, 21
Gordon, M., see Billany, M.R. **137**, 143
Gordon, M.S., A. Fratis, R. Goldblum, D. Jung, K.E. Schwartz, Z.T. Chowhan, In vivo and in vitro evaluation of four different aqueous polymeric dispersions for producing an enteric coated tablet **115**, 29
Gordon, M.S., V.S. Rudraraju, J.K. Rhie, Z.T. Chowhan, The effect of aging on the dissolution of wet granulated tablets containing super disintegrants **97**, 119
Gordon, M.S., see Gordon, M.S. **115**, 29
Gordon, R.E., D.L. Van Koevering, D.J. Reits, Utilization of differential scanning calorimetry in the compatibility screening of ibuprofen with the stearate lubricants and construction of phase diagrams **21**, 99
Gordon, R.H., see Ghebre-Sellassie, I. **31**, 43; **37**, 211
Gordon, R.H., see Nesbitt, R.U. **26**, 215
Gorecka, B.A., Y.D. Sanzgiri, D.S. Bindra, V.J. Stella, Effect of SBE4-β-CD, a sulfobutyl ether β-cyclodextrin, on the stability and solubility of O^6-benzylguanine (NSC-637037) in aqueous solutions **125**, 55
Gorgin, M.K., see Tu, Y.H. **61**, 119
Gorman, S.P., D.S. Jones, A.M. Loftus, The synergistic effect of direct and indirect ultrasonic energy and chlorhexidine gluconate on spores of *Bacillus subtilis* **65**, 127
Gorman, S.P., E.M. Scott, Potentiation and stabilization of glutaraldehyde biocidal activity utilizing surfactant-divalent cation combinations **4**, 57
Gorman, S.P., E.M. Scott, E.P. Hutchinson, The effect of sodium hypochlorite-methanol combinations on spores and spore-forms of *Bacillus subtilis* **17**, 291
Gorman, S.P., see Jones, D.S. **64**, R1; **119**, 247; **126**, 261; **131**, 83
Gorman, S.P., see McCafferty, D.F. **38**, 59
Gorman, S.P., see McGovern, J.G. **149**, 251
Gorman, S.P., see Woolfson, A.D. **48**, 167; **49**, 135; **84**, 69
Gortz, J.P., see Ntawukuliyayo, J.D. **139**, 79
Gos, T., see Sznitowska, M. **137**, 125
Goskonda, S.R., G.A. Hileman, S.M. Upadrashta, Controlled release pellets by extrusion-spheronization **111**, 89
Goskonda, S.R., see Hileman, G.A. **100**, 71
Gosse, C., see Bonhomme, L. **80**, 271
Goswami, S.K., see Vyas, S.P. **118**, 23
Goto, H., see Sugito, K. **60**, 89
Gottfries, J., J. Ahlbom, V. Harang, E. Johansson, M. Josefson, T. Morsing, A. Pettersson, A. Torstensson, Validation of an extended release tablet dissolution testing system using design and multivariate analysis **106**, 141
Gotti, R., see Gatti, R. **115**, 113
Gouda, M.W., S.A. Babhair, A.A. Al-Angary et al., Effect of dosage form, food, and an anticholinergic drug on the bioavailability of sulpiride **37**, 227
Gouda, M.W., A.H. Hikal, S.A. Babhair et al., Effect of sucralafate and antacids on the bioavailability of sulpride in humans **22**, 257
Gouda, M.W., M.A. Moustafa, A.M. Molokhia, Effect of storage conditions on erythromycin tablets marketed in Saudi Arabia **5**, 345
Gouda, M.W., see Al-Angary, A.A. **65**, R5
Gouda, M.W., see Al-Shora, H.I. **47**, 209
Gouda, M.W., see El-Sayed, Y.M. **109**, 107; **121**, 181
Gouda, M.W., see Moustafa, M.A. **30**, 225; **34**, 207
Gouda, M.W., see Turkistani, A.A.A. **59**, R1
Goudaliez, F., see Benaji, B. **105**, 83
Goudaliez, F., see Faouzi, M.A. **89**, 125; **105**, 89
Goudaliez, F., see Khalfi, F. **139**, 243
Gould, P.L., Salt selection for basic drugs **33**, 201
Gould, P.L., M. Goodman, J. Visser, Investigation of the solubility relationships of polar, semi-polar and non-polar drugs in mixed co-solvent systems **19**, 149
Gould, P.L., J.R. Howard, G.A. Oldershaw, The effect of hydrate formation on the solubility of theophylline in binary aqueous cosolvent systems **51**, 195
Gould, P.L., G.R. Sadeghnejad, B.J. Tighe, Polymers for biodegradable medical devices. IV. Hydroxybutyrate-valerate copolymers as non-disintegrating matrices for controlled-release oral dosage forms **38**, 231
Gould, P.L., see Howard, J.R. **25**, 359
Gould, P.L., see Reid, A.S. **40**, 181
Gould, P.L., see Thomas, N.W. **44**, 261
Gould, P.L., see Young, C. **40**, 187
Gouldson, M.P., see Deasy, P.B. **132**, 131
Goury, V., see Guyomard, S. **40**, 173
Govoni, P., see Giammona, G. **127**, 165
Gowan, W.G., S. Stavchansky, The effect of solvent composition upon the blood and lymph levels of phenytoin in rats after gastric administration **28**, 193
Gowan, W.G., F. Tio, K. Lamp et al., The effects of Tween 80, Myrj 52 and PEG 8000 upon the rate of pentobarbital disappearance, cumulative water flux and histological changes in the rat small intestine **29**, 169
Graf, E., see Marz, F. **58**, 155; **61**, 57
Graffner, C., see Gjellan, K. **102**, 71, 213; **104**, 215; **112**, 233
Graffner, C., see Nicklasson, M. **40**, 165
Graffner, C., see Ostberg, T. **97**, 183; **112**, 241
Graffner, C., see Ostenberg, T. **111**, 271
Graffner, C., see Stubberud, L. **114**, 55; **134**, 79
Graffner, C., see Waaler, P.J. **87**, 229
Graham, D.P., see Farraj, N.F. **52**, 11
Graham, D.T., A.R. Pomeroy, The effects of freezing on commercial insulin suspensions **1**, 315
Graham, E.T., see Kontny, M.J. **84**, 261
Grahn, A.Y., see Cook, C.S. **60**, 125
Grainger-Rousseau, T.J., J.C. McElnay, P.S. Collier, The influence of disease on plasma protein binding of drugs **54**, 1
Grandolfi, G.P., see Zografi, G. **42**, 77
Grandolini, G., see Fardella, G. **121**, 123
Grant, D.J.W., I.K.A. Abougela, Prediction of the solubility of griseofulvin in glycerides and the solvents of relatively low polarity from simple regular solution theory **17**, 77
Grant, D.J.W., H. Jacobson, J.E. Fairbrother, C.G. Patel, Phases in the paracetamol-phenazone system **5**, 109

Grant, D.J.W., G.G. Liversidge, J. Bell, Influence of physicochemical interactions on the properties of suppositories. V. The in vitro release of ketoprofen and metronidazole from various fatty suppository bases and correlations with in vivo plasma levels **14**, 251
Grant, D.J.W., M. Mehdizadeh, A.H.L. Chow, J.E. Fairbrother, Non-linear van't Hoff solubility—temperature plots and their pharmaceutical interpretation **18**, 25
Grant, D.J.W., P. York, A disruption index for quantifying the solid state disorder induced by additives or impurities. II. Evaluation from heat of solution **28**, 103
Entropy of processing: A new quantity for comparing the solid state disorder of pharmaceutical materials **30**, 161
Grant, D.J.W., see Chan, H.K. **57**, 117
Grant, D.J.W., see Chow, A.H.L. **24**, 239; **41**, 29; **42**, 123; **51**, 115, 129
Grant, D.J.W., see Chow, K.Y. **20**, 3
Grant, D.J.W., see Duddu, S.P. **94**, 171; **127**, 53
Grant, D.J.W., see Duncan-Hewitt, W.C. **28**, 75
Grant, D.J.W., see Ghosh, S. **114**, 185
Grant, D.J.W., see Khankari, R.K. **82**, 117
Grant, D.J.W., see Li, Z.J. **137**, 21
Grant, D.J.W., see Liversidge, G.G. **7**, 211; **13**, 243
Grant, D.J.W., see Vachon, M.G. **40**, 1
Grant, D.J.W., see York, P. **25**, 57
Grant, D.J.W., see Zhu, H. **135**, 151; **139**, 33
Grasmeyer, G., see Jonkman, J.H.G. **43**, 139
Grassano, A., see Dobetti, L. **128**, 277
Gratteri, P., see Corti, P. **111**, 83
Gratteri, P., see Orzalesi, G. **52**, 225
Gratteri, P., see Santoni, G. **50**, 75; **64**, 235; **71**, 1; **80**, 263; **93**, 239
Graziosi, M., see Gaddipati, N. **128**, 55
Greaves, J.L., see Fitzgerald, P. **83**, 177
Greaves, J.L., see Washington, N. **57**, 17
Greaves, J.L., see Wilson, C.G. **50**, 155; **72**, 79
Green, D.M., see Watkinson, A.C. **121**, 27; **123**, 303
Green, K.L., see Henriksen, I. **145**, 231
Green, K.L., see Nicholls, T.J. **138**, 175
Green, M., see Agyilirah, G.A. **75**, 241
Green, P.G., J. Hadgraft, Facilitated transfer of cationic drugs across a lipoidal membrane by oleic acid and lauric acid **37**, 251
Green, P.G., J. Hadgraft, M. Wolff, Physicochemical aspects of the transdermal delivery of Bupranolol enhanced delivery of nalidixic acid through human skin via acyloxymethyl ester prodrugs **55**, 265
Green, P.G., R.H. Guy, J. Hadgraft, In vitro and in vivo enhancement of skin permeation with oleic and lauric acids **48**, 103
Greenhill, J.V., see Kristensen, S. **100**, 15
Greenhill, J.V., see Tcnnesen, H.H. **87**, 79
Greenleaf, D., see Buckton, G. **116**, 113; **122**, 130
Greenslade, M.D., G. Taylor, A.D. Russell, Interactions between β-lactam and aminoglycoside antibiotics **26**, 133
Greenway, M.J., see Jones, S.P. **53**, 43
Gregoriadis, G., Silva H. Da, A.T. Florence, A procedure for the efficient entrapment of drugs in dehydration-rehydration liposomes (DRVs) **65**, 235
Gregoriadis, G., see Bachmann, D. **91**, 69
Gregoriadis, G., see Behari, J.R. **79**, 213
Gregoriadis, G., see Loukas, Y.L. **117**, 85; **144**, 225
Gregoriadis, G., see McCormack, B. **112**, 249
Greiner, G., see Sher, A. **133**, 237
Gremeau, I., see Satou, V. **107**, 111
Gren, T., C. Bjerre, O. Camber, G. Ragnarsson, In vitro drug release from porous cellulose matrices **141**, 53
Gren, T., C. Nystrom, Characterization of surface coverage of coarse particles coated with stearic acid **74**, 49
Gren, T., see Ek, R. **124**, 9
Gressier, B., S. Lebegue, C. Brunet, T. Dine, M. Luyckx, M. Cazin, J.C. Cazin, The effect of mesna in reversing, in vitro, the protease-antiprotease imbalance: Its reaction on the MPO-H_2O_2 system and on human leukocyte elastase **104**, 151
Gressier, B., see Benaji, B. **105**, 83
Gressier, B., see Faouzi, M.A. **89**, 125; **105**, 89
Gressier, B., see Gressier, B. **104**, 151
Gressier, B., see Khalfi, F. **139**, 243
Griesser, U.J., A. Burger, The effect of water vapor pressure on desolvation kinetics of caffeine 4/5-hydrate **120**, 83
Griffin, R.J., see Baker, N.D. **52**, 231; **65**, 115
Griffin, R.J., see Naik, A. **89**, 51, 65; **90**, 129
Griffith, I.P., see Toth, I. **79**, 39
Griffiths, H.A., see Armstrong, N.A. **41**, 115, 205
Grijseels, H., C.J. de Blaey, Dissolution at porous interfaces **9**, 337
Dissolution at porous interfaces. V. Pore effects in a parallel-plate dissolution cell **16**, 295
Grijseels, H., Bloois L. Van, D.J.A. Crommelin, C.J. de Blaey, Dissolution at porous interfaces. II. A study of pore effects through rotating disc experiments **14**, 299
Grijseels, H., D.J.A. Crommelin, C.J. de Blaey, Dissolution at porous interfaces. III. Pore effects in relation to the hydrodynamics at a rotating disc surface **14**, 313
Grijseels, H., see Schoonen, A.J.M. **47**, 249
Grimberg, N., see Jonkman, J.H.G. **8**, 153
Grinberg, L.N., O. Shalev, H.H. Tonnesen, E.A. Rachmilewitz, Studies on curcumin and curcuminoids. XXVI. Antioxidant effects of curcumin on the red blood cell membrane **132**, 251
Grinberg, L.N., see Tonnesen, H.H. **110**, 161
Grislain, L., P. Couvreur, V. Lenaerts et al., Pharmacokinetics and distribution of a biodegradable drug-carrier **15**, 335
Grislingaas, A.L., see Kristensen, S. **100**, 15
Grislingaas, A.L., see Tonnesen, H.H. **43**, 215; **60**, 157
Grit, M., J. De Schmidt, A. Struijke, D.J.A. Crommelin, Hydrolysis of phosphatidylcholine in aqueous liposome dispersions **50**, 1
Groening, R., Control of release and biological availability of drugs: Investigations into the renal elimination of nitrofurantoin **8**, 175
Grognet, J.M., see Ramaldes, A. **143**, 1

Groning, R., Electrophoretically controlled dermal or transdermal application systems with electronic indicators **36**, 37
Computer-controlled release of metoprolol from capsules **87**, 89
Groning, R., G. Heun, Dosage forms with controlled gastrointestinal passage—Studies on the absorption of nitrofurantoin **56**, 111
Groning, R., C. Walz, Development of experimental insulin pumps with glucose-controlled release **119**, 127
Groot, Bramel E.H, see Bekers, O. **52**, 239
Groscurth, P., see Gipps, E.M. **40**, 23
Gross, S.T., A. Hoffman, M. Donbrow, S. Benita, Fundamentals of release mechanism interpretation in multiparticulate systems: The prediction of the commonly observed release equations from statistical population models for particle ensembles **29**, 213
Gross, S.T., see Hoffman, A. **29**, 195
Grossiord, J.L., see Ferreira, L.A.M. **109**, 251; **121**, 169
Grossiord, J.L., see Thevenin, M.A. **137**, 177
Grossiord, J.L., see Tokgoz, N.S. **141**, 27
Grosvenor, M.P., J.N. Staniforth, The effect of molecular weight on the rheological and tensile properties of poly(ε-caprolactone) **135**, 103
Groth, N., see Michel, C. **98**, 131
Groueix, M.T., see Sautou-Miranda, V. **130**, 251
Grove, J., see Rozier, A. **57**, 163
Grove, J., see Thermes, F. **81**, 59
Grove, J., M. Durr, M.P. Quint, B. Plazonnet, The effect of vehicle viscosity on the ocular bioavailability of L-653,328 **66**, 23
Groves, M.J., M.H. Alkan, A note on the collapse of disintegrating tablets under laminar flow conditions **3**, 81
Groves, M.J., see Bastami, S.M. **1**, 151
Groves, M.J., see Ciftci, K. **145**, 157
Groves, M.J., see Dawes, W.H. **1**, 141
Grunbauer, H.J.M., A.L. De Meere, H.H. Van Rooij, Local composition models in pharmaceutical chemistry. III. Prediction of drug solubility in binary aqueous mixtures **32**, 187
Grunbauer, H.J.M., E. Tomlinson, Local composition models in pharmaceutical chemistry. I. Liquid-liquid distribution coefficients from COMMON UNIQUAC parameters **21**, 61
Grunbauer, H.J.M., see de Meere, A.L. **31**, 209
Grunenberg, A., J.O. Henck, H.W. Siesler, Theoretical derivation and practical application of energy/temperature diagrams as an instrument in preformulation studies of polymorphic drug substances **129**, 147
Grunenberg, A., B. Keil, J.O. Henck, Polymorphism in binary mixtures, as exemplified by nimodipine **118**, 11
Gryllaki, M., see El Taylor, N. **19**, 271
Gryllaki, M., H. Van de Waterbeemd, B. Testa, Tayar N. El, J.M. Mayer, P.A. Carrupt, Solvent-dependent lipophilicity and hydrogen-bonding capacity of 2-pyridylalkanols **51**, 95
Grynpas, R.S.O., D. Rosen, L.E. Sutton, The adsorption of urea on clays **22**, 357
Gstrein, K., see Nachtmann, F. **7**, 55
Gu, L., H.S. Chiang, A. Becker, Kinetics and mechanisms of the autoxidation of ketorolac tromethamine in aqueous solution **41**, 95
Gu, L., H.S. Chiang, D. Johnson, Light degradation of ketorolac tromethamine **41**, 105
Gu, L., O. Huynh, R.G. Strickley, L.H. Lin, G.C. Visor, Parenteral formulation development for the positive inotropic agent RS-82856. Hydrolysis and oxidation kinetics, solubility and i.v. formulation considerations **45**, 129
Gu, L., R.G. Strickley, A profound solvent effect on the diketopiperazine formation of the new dipeptide angiotensin-converting enzyme inhibitor, Moexipril 6,7 dimethoxy 3 isoquinolinecarboxylic acid **60**, 99
Gu, L., see Brandl, M. **93**, 75
Guard, P., see Barnwell, S.G. **89**, 245
Gudipati, R.M., S.A. Stavchansky, Percutaneous absorption of parachlorometaxylenol **118**, 41
Gudmundsdottir, T.K., see Loftsson, T. **127**, 293
Gudmundsson, O.S., see Kristmundsdottir, T. **137**, 159
Guegeuen, J., see Ezpeleta, I. **131**, 191; **142**, 227
Gueguen, J., see Irache, J.M. **126**, 103
Guentert, T.W., see Oie, S. **48**, 223
Guerrera, F., see Bonina, F.P. **100**, 99
Guerrini, C., see Carelli, V. **50**, 181
Guerrini, M., see Ahmed, S.M. **77**, 247; **109**, 99
Guggenbuhl, P., see Diepold, R. **54**, 149
Guiducci, A., see Saettone, M.F. **31**, 261
Guillard, I., see Arnaud, P. **129**, 279
Guillon, X., see Roy, D. **148**, 165
Guillory, J.K., see Hartauer, K.J. **85**, 163
Guillory, J.K., see Kitamura, S. **101**, 127
Guillory, J.K., see Roy, A.K. **120**, 169; **138**, 37
Guillot, T., see Jacolot, A. **128**, 283
Guillouzo, A., see Rolland, A. **53**, 67
Guimaraens-Mendez, M., see Esclusa-Diaz, M.T. **143**, 203
Guinducci, A., see Saettone, M.F. **25**, 73
Guinta, D., see McMahon, F.G. **91**, 75
Gulati, M., see Singh, S. **68**, 105
Gulik, A., see Skiba, M. **142**, 121
Gull, Lee M., see Kim, C.K. **89**, 91
Gullapalli, R.P., B.B. Sheth, Effect of methylcellulose on the stability of oil-in-water emulsions **140**, 97
Gumbleton, M., J.R. Cashman, L.Z. Benet, 1,2- and 1,3-dinitrate metabolites of nitroglycerin: Spectroscopic characterization and initial administration to man **71**, 175
Gumblteon, M., S. Maltby, G. Taylor, Urethane anaesthesia: Altered pharmacokinetics of the renally eliminated organic anion *p*-aminohippuric acid **38**, 261
Gummer, C.L., R.S. Hinz, H.I. Maibach, J. Hadgraft, The skin penetration cell: a design update **40**, 101
Gundersen, H.G., see Sontum, P.C. **128**, 269
Gundu, Rao P., see Subrahmanyam, C.V.S. **78**, 17
Guneri, T., see Ertan, G. **96**, 243
Gungor, U., see Izgu, E. **9**, 107

Gunnarsson, E., see Gizurarson, S. **140**, 77; **146**, 133
Gunnvald, K., see Fyhr, P. **40**, 193
Guo, L.S.S., R.M. Fielding, D.D. Lasic, R.L. Hamilton, D. Mufson, Novel antifungal drug delivery: Stable amphotericin B-cholesteryl sulfate discs **75**, 45
Gupta, P., J. Cannon, A. Adjei, Liposomal formulations of ABT-077: In vitro characterization studies **140**, 119
Gupta, P., Y. Qiu, A. Adjei, Pulmonary delivery of the 5-lipoxygenase inhibitor, Abbott-85761, in beagle dogs **147**, 207
Gupta, P., see Qiu, Y. **115**, 95; **143**, 179
Gupta, P.K., Magnetite as a tracer for the estimation of biodistribution of microspheres: A critical consideration **52**, 87
Gupta, P.K., C.T. Hung, Quantitative evaluation of targeted drug delivery systems **56**, 217
Albumin microspheres. V. Evaluation of parameters controlling the efficacy of magnetic microspheres in the targeted delivery of adriamycin in rats **59**, 57
Gupta, P.K., C.T. Hung, D.G. Perrier, Albumin microspheres. I. Release characteristics of adriamycin **33**, 137
Albumin microspheres. II. Effect of stabilization temperature on the release of adriamycin **33**, 147
Gupta, P.K., C.T. Hung, F.C. Lam, D.G. Perrier, Albumin microspheres. III. Synthesis and characterization of microspheres containing adriamycin and magnetite **43**, 167
Gupta, P.K., F.C. Lam, C.T. Hung, Albumin microspheres. IV. Effect of protein concentration and stabilization time on the release rate of adriamycin **51**, 253
Gupta, P.K., J.R. Robinson, Gastric empyting of liquids in the fasted dog **43**, 45
Effect of meal viscosity on gastric pH and residual volume in the fasted dog **109**, 91
Effect of volume and viscosity of coadministered fluid on gastrointestinal distribution of small particles **125**, 185
Gupta, P.K., see Lebo, D.B. **125**, 1
Gupta, P.K., see Lovell, M.W. **109**, 45; **118**, 47
Gupta, P.K., see Simon, B.H. **121**, 233
Gupta, R.L., see Singh, S. **42**, 263; **68**, 105
Gupta, S.K., see Pandit, J.K. **21**, 129
Gupta, S.L., see Krill, S.L. **132**, 89
Gupta, V., Stability of cocaine hydrochloride solutions at various pH values as determined by high-pressure liquid chromatography **10**, 249
Gurny, R., see Allemann, E. **87**, 247
Gurny, R., see Bernatchez, S.F. **106**, 161
Gurny, R., see Gander, B. **27**, 117; **58**, 63
Gurny, R., see Ibrahim, H. **77**, 211; **87**, 239
Gurny, R., see Korsmeyer, R.W. **15**, 25
Gurny, R., see Meseguer, G. **95**, 229
Gurrieri, S., see Castelli, F. **52**, 115; **59**, 19
Gurruchaga, M., see Castellano, I. **146**, 71
Gursoy, A., L. Eroglu, S. Ulutin, M. Tasyurek, H. Fessi, F. Puisieux, J.P. Devissaguet, Evaluation of indomethacin nanocapsules for their physical stability and inhibitory activity on inflammation and platelet aggregation **52**, 101
Gustavo Gonzalez, A., Drug solubility prediction by using solvatochromic parameters **75**, R13
Guterres, S.S., H. Fessi, G. Barratt, J.P. Devissaguet, F. Puisieux, Poly(DL-lactide) nanocapsules containing diclofenac. I. Formulation and stability study **113**, 57
Guterres, S.S., see Petrovick, P.D. **76**, 49
Gutierrez, C., see Perez-Marcos, B. **67**, 113
Gutierrez, F., see Veiga, M. **79**, 1
Gutierrez-Rocca, J.C., J.W. McGinity, Influence of water soluble and insoluble plasticizers on the physical and mechanical properties of acrylic resin copolymers **103**, 293
Guven, O., see Ciftci, K. **131**, 73
Guy, R.H., J. Hadgraft, A theoretical description relating skin penetration to the thickness of the applied medicament **6**, 321
Calculations of drug release rates from cylinders **8**, 159
A theoretical description of the effects of volatility and substantivity on percutaneous absorption **18**, 139
Pharmacokinetics of percutaneous absorption with concurrent metabolism **20**, 43
Transdermal drug delivery: A simplified pharmacokinetic approach **24**, 267
Interpretation and prediction of the kinetics of transdermal drug delivery: Oestradiol, hyoscine and timolol **32**, 159
On the determination of drug release rates from topical dosage forms **60**, R1
Rate control in transdermal drug delivery? **82**, R1
Guy, R.H., J. Hadgraft, H.I. Maibach, A pharmacokinetic model for percutaneous absorption **11**, 119
Guy, R.H., J. Hadgraft, H.I. Maibach, Percutaneous absorption: Multidose pharmacokinetics **17**, 23
Guy, R.H., D.H. Honda, A. Sickmueller, Solute transport resistance at the octanol–water interface **19**, 129
Guy, R.H., R. Fleming, The estimation of diffusion coefficients using the rotating diffusion cell **3**, 143
Guy, R.H., see Albery, W.J. **15**, 125
Guy, R.H., see Delgado-Charro, M.B. **117**, 165
Guy, R.H., see Denyer, S.P. **26**, 89
Guy, R.H., see Evans, N. **24**, 259
Guy, R.H., see Green, P.G. **48**, 103
Guy, R.H., see Oh, S.Y. **124**, 137
Guy, R.H., see Shah, V.P. **74**, 1
Guy, R.H., see Singh, P. **101**, R5
Guyomard, S., V. Goury, J. Laizier, J.C. Darbord, Defining of the pyrogenic assurance level (PAL) of irradiated medical devices **40**, 173
Guyot, Hermann A., see Fachaux, J.M. **99**, 99
Guyot, J.C., see Di Martino, P. **128**, 1
Guyot, J.C., see Fachaux, J.M. **99**, 99
Guyot, J.C., see Nokhodchi, A. **118**, 191; **120**, 13
Guyot, J.C., see Rizk, S. **126**, 57
Guyot, M., see Fawaz, F. **132**, 271; **133**, 107
Guyot, M., F. Fawaz, J. Bildet, F. Bonini, A.M. Langueny, Physicochemical characterization and dissolution of norfloxacin/cyclodextrin inclusion compounds and PEG solid dispersions **123**, 53

Guyot, M., F. Fawaz, M. Maury, In vitro release of theophylline from cross-linked gelatin capsules **144**, 209
Guyot-Hermann, A.M., see Di Martino, P. **128**, 1
Guzman, D., see Lopez, A. **139**, 205
Guzman, M., F.F. Garcia, J. Molpeceres, M.R. Aberturas, Polyoxyethylene-polyoxypropylene block copoymer gels as sustained release vehicles for subcutaneous drug administration **80**, 119
Guzman, M., see Chacon, M. **141**, 81
Guzman, M., see Molpeceres, J. **130**, 75

H

Ha, I.S., B.H. Woo, J.T. Lee, T.S. Kang, K.K. Tak, C.K. Oh, K.C. Lee, Degradation kinetics of growth hormone-releasing hexapeptide (GHRP-6) in aqueous solution **144**, 91
Haase, H., see Dongowski, G. **144**, 233
Haase, M.M., see Felton, L.A. **113**, 17
Habberfield, A., K. Jensen-Pippo, L. Ralph, S.W. Westwood, G.J. Russell-Jones, Vitamin B12-mediated uptake of erythropoietin and granulocyte colony stimulating factor in vitro and in vivo **145**, 1
Habib, F.S., M.A. Attia, S.M. El-Shanawany, Effect of different ointment bases on ocular disposition of ethylphenylephrine in rabbit eyes **26**, 243
Habib, F.S., see Kassem, M.A. **29**, 243; **32**, 47
Habib, M.J., J.A. Rogers, Kinetics of hydrolysis and stabilization of acetylsalicylic acid in liposome formulations **44**, 235
Habib, M.J., see Ghosh, T.K. **88**, 391
Hackemuller, D., see Lippold, B.C. **61**, 205
Hacker, R., see Su, X.Y. **123**, 47
Hadgraft, J., Calculations of drug release rates from controlled release devices. The slab **2**, 177
The epidermal reservoir: A theoretical approach **2**, 265
Percutaneous absorption: Possibilities and problems **16**, 255
Pharmaceutical aspects of transdermal nitroglycerin **135**, 1
Hadgraft, J., D. Beutner, H.M. Wolff, In vivo-in vitro comparisons in the transdermal delivery of nitroglycerin **89**, R1
Hadgraft, J., D. Lewis, D. Beutner, H.M. Wolff, In vitro assessments of transdermal devices containing nitroglycerin **73**, 125
Hadgraft, J., J.M. Haigh, Development of model membranes for percutaneous absorption measurements. I. Isopropyl myristate **39**, 149
Hadgraft, J., J. Peck, D.G. Williams, W.J. Pugh, G. Allan, Mechanisms of action of skin penetration enhancers/retarders: Azone and analogues **141**, 17
Hadgraft, J., W.J. Pugh, H.M. Wolff, Transdermal delivery of some anti-emetics **123**, 285
Hadgraft, J., G. Ridout, Development of model membranes for percutaneous absorption measurements. II. Dipalmitoyl phosphatidylcholine, linoleic acid and tetradecane **42**, 97
Hadgraft, J., K.A. Walters, P.K. Wotton, Facilitated percutaneous absorption: A comparison and evaluation of two in vitro models **32**, 257
Hadgraft, J., M. Wolff, R. Bonn, G. Cordes, Development of biphasic transdermal nitroglycerin delivery systems **64**, 187
Hadgraft, J., see Ahmed, M. **12**, 219; **13**, 227
Hadgraft, J., see Al-Khamis, K. **10**, 25; **40**, 111
Hadgraft, J., see Albery, W.J. **15**, 125
Hadgraft, J., see Arrowsmith, M. **14**, 191; **16**, 305; **17**, 91; **20**, 347
Hadgraft, J., see Ashton, P. **87**, 261, 265
Hadgraft, J., see Barker, N. **8**, 193
Hadgraft, J., see Beastall, J.C. **43**, 207
Hadgraft, J., see Bialik, W. **92**, 219
Hadgraft, J., see Brain, K.R. **71**, R9; **89**, R13
Hadgraft, J., see Cabral Marques, H. **63**, 259, 267; **77**, 297, 303
Hadgraft, J., see Davis, A.F. **76**, 1
Hadgraft, J., see Denyer, S.P. **26**, 89
Hadgraft, J., see Diez-Sales, O. **129**, 33
Hadgraft, J., see Evans, N. **24**, 259
Hadgraft, J., see Fitzgerald, P. **40**, 81
Hadgraft, J., see Gay, C.L. **49**, 39
Hadgraft, J., see Gilbert, J.C. **32**, 223
Hadgraft, J., see Green, P.G. **37**, 251; **48**, 103; **55**, 265
Hadgraft, J., see Gummer, C.L. **40**, 101
Hadgraft, J., see Guy, R.H. **6**, 321; **8**, 159; **11**, 119; **17**, 23; **18**, 139; **20**, 43; **24**, 267; **32**, 159; **60**, R1; **82**, R1
Hadgraft, J., see Heard, C.M. **90**, R5
Hadgraft, J., see Howard, J.R. **16**, 31
Hadgraft, J., see Lafforgue, C. **121**, 89
Hadgraft, J., see Lewis, D. **65**, 211
Hadgraft, J., see Martin, R.J. **39**, 23
Hadgraft, J., see Pellett, M.A. **111**, 1
Hadgraft, J., see Pugh, W.J. **103**, 163; **138**, 149
Hadgraft, J., see Rathbone, M.J. **74**, 9
Hadgraft, J., see Roberts, M.S. **126**, 219; **132**, 23
Hadgraft, J., see Walker, M. **70**, 167; **71**, R1
Hadgraft, J., see Watkinson, A.C. **74**, 229; **83**, 171; **87**, 175; **121**, 27; **123**, 303
Hadgraft, J., see Wotton, P.K. **24**, 19
Hadzija, B.W., E.K. Obeng, S.B. Ruddy, A. Noormohammadi, A.H. Beckett, Bioavailability of a controlled release indomethacin formulation in healthy subjects **67**, 185
Hadzija, B.W., see Beckett, A.H. **52**, 37
Haeberlin, B., T. Gengenbacher, A. Meinzer, G. Fricker, Cyclodextrins—Useful excipients for oral peptide administration? **137**, 103
Haemers, A., see Mbela, T.K.M. **92**, 71; **110**, 189
Hafeez, R.A., see Toth, I. **79**, 39; **83**, 131
Hafkenscheid, T.L., E. Tomlinson, Correlations between alkane/water and octan-1-ol/water distribution coefficients and isocratic reversed phase liquid chromatographic capacity factors of acids, bases and neutrals **16**, 225
Isocratic chromatographic retention data for estimating aqueous solubilities of acidic, basic and neutral drugs **17**, 1
Hydrogen-bonding properties of reversed-phase liquid chromatographic stationary phases used for measuring solute hydrophobic/lipophilic parameters **19**, 349
Hagedoorn, P., see de Boer, A.H. **138**, 45

Hageman, M.J., see Pena, L.E. **113**, 89
Hageman, M.J., see Repta, A.J. **10**, 239
Hahn, K.R., see Krill, S.L. **132**, 89
Haigh, J.M., I. Kanfer, Assessment of topical corticosteroid preparations: The human skin balancing assay **19**, 245
Haigh, J.M., see Hadgraft, J. **39**, 149
Haigh, J.M., see Meyer, E. **41**, 63
Haigh, J.M., see Smith, E.W. **27**, 185
Hailey, F.J., see Mason, W.D. **36**, 105
Hailey, F.J., see Wagner, D.L. **57**, 177
Hakasalo, M., see Monkkonen, J. **107**, 189
Halbaut, L., C. Barbe, M. Aroztegui, C. De la Torre, Oxidative stability of semi-solid excipient mixtures with corn oil and its implication in the degradation of vitamin A **147**, 31
Halbaut, L., C. Barbe, A. Del Pozo, An investigation into physical and chemical properties of semi-solid self-emulsifying systems for hard gelatin capsules **130**, 203
Halbert, G.W., J.F.B. Stuart, A.T. Florence, The incorporation of lipid-soluble antineoplastic agents into microemulsions-protein-free analogues of low density lipoprotein **21**, 219
Halbert, G.W., see Eley, J.G. **63**, 121; **65**, 219
Halbert, G.W., see McTaggart, L.E. **100**, 199
Hall, S.B., P.W. Gaskin, J.R. Duffield, D.R. Williams, An interfacial equilibria model for the electrokinetic properties of a fat emulsion **70**, 251
Halterlein, L.M., see Rubino, J.T. **26**, 165
Hamacher, E., see Stein, M. **80**, R11
Hamada, C., see Kurosaki, Y. **47**, 13
Hamaguchi, N., see Iga, K. **57**, 241
Hamaguchi, N., see Kikuchi, M. **38**, 191
Hamaguchi, N., see Yoshimura, Y. **23**, 117; **26**, 317
Hamaguchi, T., see Shinkuma, D. **9**, 17; **21**, 187
Hamamoto, K., R. Koike, Y. Machida, T. Nagai, Sustained release granules of sulfamethoxazole for chicken utilizing frictional power of gizzard **113**, 9
Hamed, M.Y., E.M. Mostafa, S.S. Tous, Antifertility effect of orally formulated melatonin tablets in mice **69**, 93
Hamel, F.G., see Brand, R.M. **146**, 115
Hamilton, R.L., see Guo, L.S.S. **75**, 45
Hammad, A., see Al-Gohary, O.M. **55**, 47
Hammad, A.L., see Al-Shora, A. **7**, 77
Hammounda, Y.E., N.A. Kasim, A.H. Nada, Formulation and in vitro evaluation of verapamil HCl suppositories **89**, 111
Hamon, M., see Fournier, C. **106**, 41
Hampl, J., M. Dittrich, J. Franz, S. Reschova, J. Stepanek, Adjuvant activity of linear aliphatic polyester and branched aliphatic oligoester microspheres **144**, 107
Han, S.K., Y.H. Park, C.K. Kim, Preparation of *N*-adamantyl n-alkanamides and evaluation of their transdermal penetration in the rabbit **126**, 35
Han, S.K., see Kim, C.K. **95**, 43; **101**, 191; **108**, 21
Han, K.S., see Lee, S.H. **90**, 81
Hanaee, J., see Barzegar-Jalali, M. **109**, 291; **144**, 127
Hanano, M., see Shim, C.K. **8**, 143
Hanano, M., see Watari, N. **6**, 155
Hanawa, T., see Watanabe, A. **117**, 23
Hanchard, C., see Jadoul, A. **120**, 221
Hancock, B.C., P. York, R.C. Rowe, Characterization of wet masses using a mixer torque rheometer. 2. Mixing kinetics **83**, 147
An assessment of substrate–binder interactions in model wet masses. 1. Mixer torque rheometry **102**, 167
The use of solubility parameters in pharmaceutical dosage form design **148**, 1
Hancock, B.C., P. York, R.C. Rowe, M.D. Parker, Characterization of wet masses using a mixer torque rheometer. 1. Effect of instrument geometry **76**, 239
Hanke, B., see Keipert, S. **142**, 153
Hanlon, G.W., see Bilbruck, J. **99**, 293
Hanlon, G.W., see Malhi, J.S. **43**, 77
Hanlon, G.W., see Wilkins, K.M. **57**, 1; **58**, 165
Hanna, M.S., see Bhatt, P.P. **50**, 197
Hanninen, K., see Arbin, A. **28**, 211
Hannula, A.M., see Marvola, M. **44**, 159
Hannus, M., see Finne, U. **78**, 237
Hanocq, M., see Abi Khalil, F. **140**, 1
Hanocq, M., see Chamart, S. **99**, 59
Hanocq, M., see Dubois, J.G. **35**, 219
Hanocq, M., see Verbiese-Genard, N. **2**, 155; **9**, 295
Hansen, E., see Sclafani, J. **124**, 213
Hansen, H.S., see Vermehren, C. **128**, 239
Hansen, J., B. Kreilgard, O. Nielsen, J. Veje, Kinetics of degradation of methotrexate in aqueous solution **16**, 141
Hansen, J., H. Bundgaard, A differential kinetic method for the determination of β-methasone-17-valerate in the presence of its degradation products **8**, 121
Hansen, J., N. Mork, H. Bundgaard, Phenyl carbamates of amino acids as prodrug forms for protecting phenols against first-pass metabolism **81**, 253
Hansen, J., see Bundgaard, H. **7**, 197; **9**, 273
Hansen, K.T., J.A. Jansen, H. Mengel, J.V. Christensen, H. Bundgaard, Enhanced bioavailability of a new class of dopamine D-1 antagonists following oral administration of their carbamic acid ester prodrugs to dogs **79**, 205
Hansen, K.T., see Bundgaard, H. **45**, 47
Hansen, L.B., L.L. Christrup, H. Bundgaard, Saliva-catalyzed hydrolysis of a ketobemidone ester prodrug: Factors influencing human salivary esterase activity **88**, 221
Ketobemidone prodrugs for buccal delivery: Prediction of the extent of saliva-catalyzed hydrolysis of various ester prodrugs under simulated in vivo conditions **88**, 229
Enhanced delivery of ketobemidone through porcine buccal mucosa in vitro via more lipophilic ester prodrugs **88**, 237
Hansen, L.B., A. Fullerton, L.L. Christrup, H. Bundgaard, Enhanced transdermal delivery of ketobemidone with prodrugs **84**, 253
Hansen, L.B., A. Jorgensen, S.N. Rasmussen, L.L. Christrup, H. Bundgaard, Buccal absorption of ketobemidone and various ester prodrugs in the rat **88**, 243
Hansen, L.D., J.W. Crawford, D.R. Keiser, R.W. Wood, Calorimetric method for rapid determination of critical wa-

ter vapor pressure and kinetics of water sorption on hygroscopic compounds **135**, 31
Hansen, L.D., M.T. Pyne, R.W. Wood, Water vapor sorption by cephalosporins and penicillins **137**, 1
Hansen, P.J., see Peppas, N.A. **20**, 107
Hansen, S.H., HPLC assay of the opiates in opium and cough mixtures using dynamically modified silica and UV absorbance, fluorescence and electrochemical detection **32**, 7
Hansen, S.H., see Bundgaard, H. **10**, 281
Hansen, S.H., see Jensen, J. **88**, 177
Hanson, P., see Suryanarayanan, R. **78**, 77
Hanson, P.A., see McTaggart, C.M. **19**, 139
Hansrani, P., see Davis, S.S. **23**, 69
Hansrani, P.K., see Burnham, W.R. **13**, 9
Happonen, I., see Heinamaki, J. **42**, 105; **48**, 51
Happonen, I., see Marvola, M. **44**, 159
Hara, M., see Maitani, Y. **64**, 139
Hara, T., H. Ishihara, Y. Aramaki, S. Tsuchiya, Specific uptake of asialofetuin-labeled liposomes by isolated hepatocytes **42**, 69
Characteristics of the binding of asialofetuin-labeled liposomes to isolated rat hepatocytes **67**, 123
Harada, M., see Tashima, T. **42**, 61
Harang, V., see Gottfries, J. **106**, 141
Harashima, H., see Uchiyama, K. **121**, 195
Harboe, E., C. Larsen, M. Johansen, H.P. Olesen, Macromolecular prodrugs. XIV. Absorption characteristics of naproxen after oral administration of a dextran T-70-naproxen ester prodrug in pigs **53**, 157
Hardie, L., see Taylor, R.B. **102**, 201
Harding, V.D., see Denyer, S.P. **25**, 245; **29**, 29
Hardy, J.G., S.S. Davis, R. Khosla, C.S. Robertson, Gastrointestinal transit of small tablets in patients with ulcerative colitis **48**, 79
Hardy, J.G., D.F. Evans, I. Zaki et al., Evaluation of an enteric coated Naproxen tablet using γ scintigraphy and pH monitoring **37**, 245
Hardy, J.G., L.C. Feely, E. Wood, S.S. Davis, The application of γ-scintigraphy for the evaluation of the relative spreading of suppository bases in rectal hard gelatin capsules **38**, 103
Hardy, J.G., A.K. Jasuja, M. Frier, A.C. Perkins, A small volume spacer for use with a breath-operated pressurised metered dose inhaler **142**, 129
Hardy, J.G., S.W. Lee, A.G. Clark, J.R. Raynolds, Enema volume and spreading **31**, 151
Hardy, J.G., see Bennett, C.E. **21**, 341
Hardy, J.G., see Davis, S.S. **21**, 167, 331; **32**, 85
Hardy, J.G., see Jenkins, J.R.F. **14**, 143
Hardy, J.G., see Moolenaar, F. **19**, 161
Hardy, J.G., see O'Reilly, S. **34**, 213
Hardy, J.G., see Pennington, A.K. **43**, 221
Hardy, J.G., see Rao, L.S. **17**, 283
Hardy, J.G., see Wilding, I.R. **76**, 133
Hardy, J.G., see Wilson, C.G. **38**, 221
Hardy, J.G., see Wood, E. **25**, 191
Hariharan, D., N.A. Peppas, R. Bettini, P. Colombo, Mathematical analysis of drug delivery from swellable systems with partial physical restrictions or impermeable coatings **112**, 47
Harju, K., see Edsman, K. **137**, 233
Harland, R., see Benson, H.A.E. **44**, 65
Harland, R., see McElnay, J.C. **40**, 105
Harmia, T., J. Kreuter, P. Speiser et al., Enhancement of the myotic response of rabbits with pilocarpine-loaded polybutylcyanoacrylate nanoparticles **33**, 187
Harmia, T., P. Speiser, J. Kreuter, Optimization of pilocarpine loading onto nanoparticles by sorption procedures **33**, 45
Harn, J., see Krill, S.L. **132**, 89
Harnor, K.J., A.C. Perkins, M. Wastie, C.G. Wilson, E.E. Sims, L.C. Feely, S.J. Farr, Effect of vapour pressure on the deposition pattern from solution phase metered dose inhalers **95**, 111
Haro, I., see Alsina, M.A. **70**, 111
Haro, I., see Colome, C. **90**, 59
Haro, I., see Mestres, C. **76**, 145
Harper Bellantone, N., S. Rim, M.L. Francoeur, B. Rasadi, Enhanced percutaneous absorption via iontophoresis. I. Evaluation of an in vitro system and transport of model compounds **30**, 63
Harper, G., see Davis, S.S. **82**, 99
Harper, M.I.A., see Taylor, R.B. **60**, 185
Harrari, D., see Friedman, M. **44**, 243
Harris, D., J.T. Fell, D.C. Taylor, J. Lynch, H.L. Sharma, Oral availability of a poorly absorbed drug, hydrochlorothiazide, from a bioadhesive formulation in the rat **56**, 97
Harris, R.J., see Barnwell, S.G. **88**, 423
Harrison, P.J., J.M. Newton, R.C. Rowe, The application of capillary rheometry to the extrusion of wet powder masses **35**, 235
Harrison, P.J., see Willmott, N. **43**, 161
Harron, D.W.G., Digoxin pharmacokinetic modelling—10 years later **53**, 181
Harron, D.W.G., D.E. Arndts, R.G. Shanks, Alinidine biotransformation in healthy Sudanese Arabs **12**, 277
Harron, D.W.G., J. Liddle, S. Devine, M.G. Scott, A. Deering, M. Burke, R.G. Shanks, Pharmacokinetics and dynamics of a new formulation of slow release oxprenolol combined with triamterene and hydrochlorothiazide in healthy man **57**, 9
Harron, D.W.G., J.C. McElnay, P.F. D'Arcy, P.S. Collier, An 'in vitro|apso| system for the correlation of drug/protein binding with variable plasma protein profile **1**, 61
Harron, D.W.G., see Collier, P.S. **1**, 3
Harron, D.W.G., see El Turabi, H. **53**, 249
Harron, D.W.G., see Harron, D.W.G. **57**, 9
Harron, D.W.G., see Tanira, M.O.M. **102**, 267
Harron, D.W.G., see Temple, D.J. **2**, 127
Hart, J.P., see Staniforth, J.N. **40**, 267
Hartauer, K.J., E.S. Miller, J.K. Guillory, Diffuse reflectance infrared Fourier transform spectroscopy for the quantitative analysis of mixtures of polymorphs **85**, 163
Harte, V.J., M.T. O'Hanrahan, R.F. Timoney, Microbial contamination in residues of ophthalmic preparations **1**, 165
Harter, J.G., see Peck, C.C. **82**, 9

Hartner, K.C., see Imanidis, G. **120**, 41
Harvengt, C., see Desager, J.P. **31**, 179
Hasan, M.M., see Tabba, H.D. **54**, 57
Hasegawa, T., R. Apichartpichean, T. Kuzuya, M. Nadai, Y. Tsunekawa, T. Horiuchi, K. Miyamoto, Protein binding characteristics of new bronchodilators, 1-methyl-3-propylxanthine (MPX) and 3-propylxanthine (enprofylline) **56**, 235
Hasegawa, T., K. Miyamoto, S. Wakusawa, K. Takagi, R. Apichartpichean, T. Kuzuya, M. Nadai, T. Horiuchi, Relationship between hydrophobicity and biological activities of xanthine derivatives in guinea pig tracheal smooth muscle **58**, 129
Hasegawa, T., see Kawaguchi, T. **58**, R1; **77**, 71
Hashida, M., S. Muranishi, H. Sezaki et al., Increased lymphatic delivery of bleomycin by microsphere in oil emulsion and its effect on lymph node metastasis **2**, 245
Hashida, M., see Masegi, M. **78**, 103
Hashida, M., see Mukai, E. **25**, 95
Hashida, M., see Nishida, K. **65**, 7; **80**, 101
Hashida, M., see Nishikawa, M. **85**, 75
Hashida, M., see Okamoto, H. **30**, 35
Hashida, M., see Sakane, T. **57**, 77
Hashida, M., see Sasaki, H. **15**, 49, 61; **36**, 147
Hashida, M., see Takakura, Y. **37**, 145
Hashida, M., see Yamamoto, A. **39**, 255; **51**, 137; **93**, 91
Hashida, M., see Yamashita, F. **89**, 199; **117**, 173
Hashida, M., see Yoshioka, T. **8**, 131
Hashim, F.M., see Soliman, O.A. **149**, 73
Hashim, H., A.L.W. Po, Improving the release characteristics of water-soluble drugs from hydrophilic sustained release matrices by in situ gas generation **35**, 201
Hasinoff, B.B., An HPLC and spectrophotometric study of the hydrolysis of ICRF-187 (dexrazoxane, (−)-1,2-bis(3,5-dioxopiperazinyl-1-yl)propane) and its one-ring opened intermediates **107**, 67
Haslam, J.L., A.E. Merfeld, G.S. Rork, Surface wetting effects in the lipid osmotic pump **56**, 227
Haslam, J.L., see Merfeld, A.E. **69**, 63
Haslam, J.L., see Vadnere, M. **22**, 207
Hassan, E.E., A.G. Eshra, A.H. Nada, Formulation of prolonged release lipid micropellets by emulsion congealing: Optimization of ketoprofen entrapment and release **121**, 149
Hassan, E.E., see Darwish, I.A. **140**, 25
Hassan, E.E., see El-Massik, M.A. **140**, 69
Hassan, H., see Sakellariou, P. **102**, 207
Hassan, I., see Nicklin, P. **83**, 197
Hassan, I.F., see Nicklin, P.L. **123**, 187
Hassan, M.A., N.M. Najib, M.S. Suleiman, Characterization of glibenclamide glassy state **67**, 131
Hassan, M.A., Salem M. Sheikh, M.S. Sueliman, N.M. Najib, Characterization of famotidine polymorphic forms **149**, 227
Hassan, M.A., M.S. Suleiman, N.M. Najib, Improvement of the in vitro dissolution characteristics of famotidine by inclusion in β-cyclodextrin **58**, 19
Hassan, M.A., see Suleiman, M.S. **54**, 65
Hastewell, J., K.H. Antonin, R. Fox, M. Mackay, The colonic absorption of human calcitonin: The effects of increasing local concentration and co-administration with a protease inhibitor **126**, 245
Hastewell, J., S. Lynch, R. Fox, I. Williamson, P. Skelton-Stroud, M. Mackay, Enhancement of human calcitonin absorption across the rat colon in vivo **101**, 115
Hata, T., Y. Shimazaki, A. Kagayama, S. Tamura, S. Ueda, Development of a novel drug delivery system, time-controlled explosion system (TES). V. Animal pharmacodynamic study and human bioavailability study **110**, 1
Hatakeyama, Y., see Takeeda, T. **41**, 21
Hatanaka, T., M. Inuma, K. Sugibayashi, Y. Morimoto, Prediction of skin permeability of drugs. II. Development of composite membrane as a skin alternative **79**, 21
Hatanaka, T., S. Morigaki, T. Aiba, K. Katayama, T. Koizumi, Effect of pH on the skin permeability of a zwitterionic drug, cephalexin **125**, 195
Hatanaka, T., see Panomsuk, S.P. **126**, 147
Hatanaka, T., see Sugibayashi, K. **113**, 189
Hatano, K., see Kurono, Y. **81**, 179
Hathaway, R., see Johnson, K. **73**, 197
Hathaway, R.D., see Jozwiakowski, M.J. **91**, 195
Hatjichristos, Th., see Malamataris, S. **141**, 101
Haubold, W., see Sher, A. **133**, 237
Hauchecorne, R., see Cachet, T. **55**, 59
Hauchecorne, R., see Vinckier, C. **55**, 67
Hauswald, C.L., see Cook, C.S. **60**, 125
Hay, G., see Barnwell, S.G. **128**, 145
Hay, G., see Burns, S.J. **110**, 291; **121**, 37; **134**, 223; **141**, 9
Hayashi, M., see Kajii, H. **33**, 253
Hayashi, T., see Murakami, T. **49**, 173
Hayashi, T., see Nakamura, H. **141**, 71
Hayashi, Y., see Yoshida, M. **115**, 61
Hayashibara, M., see Nagasako, S. **81**, 127
Hayes, G., see Wilding, I.R. **83**, 155
Haynes, L.C., M.J. Cho, Mechanism of Nile red transfer from o/w emulsions as carriers for passive drug targeting to peritoneal macrophages in vitro **45**, 169
Haynes, L.C., see Scieszka, J.F. **45**, 165
Hayton, W.L., see Colburn, W.A. **1**, 121
Hayward, D.S., D.R. Jenke, Interactions between polymer containers and parenteral solutions: Correlating equilibrium polymer–water interaction constants with solute partition coefficients **66**, 87
Hayward, D.S., R.A. Kenley, D.R. Jenke, Interactions between polymer containers and parenteral solutions: The correlation of equilibrium constants for polymer-water partitioning with octanol-water partition coefficients **59**, 245
Hayward, M., see Fares, H.M. **133**, 215
Hazebroucq, G., see Gimenez, F. **43**, 23
Healy, A.M., O.I. Corrigan, Predicting the dissolution rate of ibuprofen-acidic excipient compressed mixtures in reactive media **84**, 167
The influence of excipient particle size, solubility and acid strength on the dissolution of an acidic drug from two-component compacts **143**, 211

Heard, C.M., R. Suedee, Stereoselective adsorption and transmembrane transfer of propranolol enantiomers using cellulose derivatives **139**, 15
Heard, C.M., A.C. Watkinson, K.R. Brain, J. Hadgraft, In vitro skin penetration of propranolol enantiomers **90**, R5
Hearn, D.J., see Jones, H.P. **104**, 253
Heasman, M.J., see Bowtle, W.J. **4**, 263
Hecquet, B., see Fournier, C. **106**, 41
Hedin, L., see Lundin, S. **59**, 263
Hegde, S.G., see Lidgate, D.M. **96**, 51
Heiati, H., R. Tawashi, R.R. Shivers, N.C. Phillips, Solid lipid nanoparticles as drug carriers. I. Incorporation and retention of the lipophilic prodrug 3′-azido-3′-deoxythymidine palmitate **146**, 123
Heikkila, T., see Laine, E. **38**, 33
Heimlich, K.R., see Kwan, K.C. **6**, 225
Heinamaki, J., M. Marvola, I. Happonen, E. Westermarck, The fate of multiple-unit enteric-coated formulations in the stomach of the dog **42**, 105
Formulations releasing the drug proximal to the pylorus in the dog **48**, 51
Heinamaki, J.T., A.I. Colarte, A.J. Nordstrom, J.K. Yliruusi, Comparative evaluation of ammoniated aqueous and organic-solvent-based cellulose ester enteric coating systems: A study on free films **109**, 9
Heinamaki, J.T., V.M. Lehtola, P. Nikupaavo, J.K. Yliruusi, The mechanical and moisture permeability properties of aqueous-based hydroxypropyl methylcellulose coating systems plasticized with polyethylene glycol **112**, 191
Heinemann, S., see Muller, R.H. **101**, 175; **107**, 121
Heinonen, R.M., see Salomies, H.E.M. **110**, 197
Heiskanen, I., see Veski, P. **111**, 171
Helboe, P., M. Thomsen, Single-dose determination of suppositories containing phenylephrine, lidocaine and betamethasone valerate by reversed-phase ion-pair liquid chromatography **2**, 317
Helboe, P., see Majilat, P. **9**, 245
Hellen, L., J. Yliruusi, Process variables of instant granulator and spheroniser. III. Shape and shape distributions of pellets **96**, 217
Hellen, L., J. Yliruusi, E. Kristoffersson, Process variables of instant granulator and spheroniser. II. Size and size distributions of pellets **96**, 205
Hellen, L., J. Yliruusi, P. Merkku, E. Kristoffersson, Process variables of instant granulator and spheroniser. I. Physical properties of granules, extrudate and pellets **96**, 197
Heller, J., see Singh, M. **76**, R5
Helliwell, M., see Pritchard, K. **129**, 137
Hellmuth, E.W., see Neau, S.H. **99**, 303
Helm, V.J., B.W. Muller, Stability of the synthetic pentapeptide thymopentin in aqueous solution: Effect of pH and buffer on degradation **70**, 29
Helton, D.O., see Amselem, S. **125**, 31
Helton, D.O., see Brewster, M.E. **125**, 17
Hemgesberg, E., see Loth, H. **32**, 265
Hempenstall, J.M., W.J. Irwin, A. Li Wan Po, A.H. Andrews, Antibiotic granules for reconstitution as syrups: Product uniformity and stability dependent upon reconstitution procedure **23**, 131
Hempenstall, J.M., see Irwin, W.J. **20**, 25
Henck, J.O., see Grunenberg, A. **118**, 11; **129**, 147
Hendra, P.J., see Davies, M.C. **66**, 223
Hendriksen, B.A., Characterization of calcium fenoprofen. 1. Powder dissolution rate and degree of crystallinity **60**, 243
Characterization of calcium fenoprofen. 3. Mechanism of dissolution from rotating discs **75**, 63
Hendriksen, B.A., M.S. Preston, P. York, Processing effects on crystallinity of cephalexin: Characterisation by vacuum microbalance **118**, 1
Hendriksen, B.A., J.D. Williams, Characterization of calcium fenoprofen. 2. Dissolution from formulated tablets and compressed rotating discs **69**, 175
Hendrix, C., Z. Yongxin, Houtven C. Van, J. Thomas, E. Roets, J. Hoogmartens, Evaluation of analytical methods: Liquid chromatography of cefalexin **100**, 213
Heng, P.W.S., L.S.C. Wan, B.L. Ling, Assessment of powder cohesiveness in spheronization studies **116**, 119
Heng, P.W.S., L.S.C. Wan, Y.T.F. Tan, Relationship between aggregation of HPMC coated spheroids and tackiness/viscosity/additives of the coating formulations **138**, 57
Optimization of spheroid production by centrifugal rotary processing **143**, 107
Heng, P.W.S., see Wan, L.S.C. **73**, 111; **77**, 183; **88**, 159; **95**, 77; **96**, 59; **103**, 267; **116**, 159; **118**, 213; **141**, 161
Henmi, T., see Fujii, M. **137**, 117
Henrar, R.E.C., see De, Vries J.D. **100**, 181
Henriksen, I., K.L. Green, J.D. Smart, G. Smistad, J. Karlsen, Bioadhesion of hydrated chitosans: An in vitro and in vivo study **145**, 231
Henriksen, I., S.A. Sande, G. Smistad, T. Agren, J. Karlsen, In vitro evaluation of drug release kinetics from liposomes by fractional dialysis **119**, 231
Henriksen, I., O. Skaugrud, J. Karlsen, Use of chitosan and chitosan malate as an excipient in wet granulation of three water soluble drugs **98**, 181
Henriksen, I., G. Smistad, J. Karlsen, Interactions between liposomes and chitosan **101**, 227
Henriksen, I., S.R. Vagen, S.A. Sande, G. Smistad, J. Karlsen, Interactions between liposomes and chitosan. II. Effect of selected parameters on aggregation and leakage **146**, 193
Henriksson, U., see Ek, R. **124**, 9
Henry, B.T., M.S. Cheema, S.S. Davis, Gel permeation chromatography used to determine the stability of FITC-Dextrans in human saliva and porcine small intestinal mucus **73**, 81
Henry-Michelland, S., M.J. Alonso, A. Andremont et al., Attachment of antibiotics to nanoparticles: Preparation, drug-release and antimicrobial activity in vitro **35**, 121
Herbage, D., see Pajean, M. **77**, 31; **91**, 209
Herbert, T.O., see Atkins, P.J. **30**, 199
Herbette, L.G., see Lamson, M.J. **105**, 259
Herbort, J., see Muller, R.H. **84**, 1
Herbots, H., see Kinget, R. **3**, 65
Herbots, H., see Provost, C. **62**, 217

Herder, R.E., see Wiechers, J.W. **65**, 77
Herdewijn, P., see Van Schepdael, A. **73**, 105
Herishanu, Y., see Bialer, M. **16**, 285
Herman, C.S., see Suryanarayanan, R. **77**, 287
Herman, J., J.P. Remon, Modified starches as hydrophilic matrices for controlled oral delivery. II. In vitro drug release evaluation of thermally modified starches **56**, 65
Modified starches as hydrophilic matrices for controlled oral delivery. III. Evaluation of sustained-release theophylline formulations based on thermal modified starch matrices in dogs **63**, 201
Herman, J., J.P. Remon, J. De Vilder, Modified starches as hydrophilic matrices for controlled oral delivery. I. Production and characterisation of thermally modified starches **56**, 51
Herman, J., J.P. Remon, N. Visavarungroj, J.B. Schwartz, G.H. Klinger, Formation of theophylline monohydrate during the pelletisation of microcrystalline cellulose-anhydrous theophylline blends **42**, 15
Herman, J., N. Visavarungroj, J.P. Remon, Instability of drug release from anhydrous theophylline-microcrystalline cellulose formulations **55**, 143
Herman, J., see Visavarungroj, N. **59**, 73
Hermann, T.W., Recent research on bioavailability of drugs from suppositories **123**, 1
Hermansen, K., see Buur, A. **16**, 41
Hermecz, I., see Takacs-Novak, K. **79**, 89
Hermens, W.A.J.J., see Duchateau, G.S.M.J.E. **31**, 275
Hermens, W.A.J.J., see Schipper, N.G.M. **64**, 61
Hernandez, E., see Ramos, A.J. **128**, 129
Hernandez, J., J. Estelrich, M.T. Montero, O. Valls, Interaction between human serum albumin and liposomes: A monolayer and liposome study **57**, 211
Hernandez, R.M., A.R. Gascon, M.B. Calvo, C. Caramella, U. Conte, A. Dominguez-Gil, J.L. Pedraz, Correlation of 'in vitro' release and 'in vivo' absorption characteristics of four salbutamol sulphate formulations **139**, 45
Hernandez-Borrell, J., A new confirmation of selective action of liposomes **47**, 129
Hernandez-Borrell, J., see Montero, M.T. **96**, 157; **149**, 161
Herrador, M.A., see Asuero, A.G. **34**, 81
Herrador, M.A., see Gonzalez, A.G. **84**, R1; **108**, 149; **120**, 215; **123**, 149
Herraez, M., see Diez-Sales, O. **77**, 1
Herraez-Dominguez, M., see Diez-Sales, O. **100**, 1; **129**, 33
Herremans, J.L.M., F.W. Mijnheer, H.R. Gerritsen, R. Prevost, C. Kluft, D.J.A. Crommelin, Long-term stability of liposomes containing both tissue-type plasminogen activator and glu-plasminogen **129**, 191
Herremans, J.L.M., see Herremans, J.L.M. **129**, 191
Herrling, T., see Michel, C. **98**, 131
Herrmann, J., R. Bodmeier, Somatostatin containing biodegradable microspheres prepared by a modified solvent evaporation method based on w/o/w-multiple emulsions **126**, 129
Hersey, J.A., see Krycer, I. **12**, 113
Hessell, P.G., J.G. Lloyd-Jones, N.C. Muir, G.D. Parr, K. Sugden, A comparison of the bioavailability of prochlorperazine following i.m. buccal and oral administration **52**, 159
Hettler, D., see Levy, M.C. **69**, R1
Hettrick, L., see Matuszewska, B. **136**, 147
Heun, G., see Groning, R. **56**, 111
Heuvelsland, W.J.M., see Ijntema, K. **112**, 215
Heya, T., H. Okada, Y. Ogawa, H. Toguchi, Factors influencing the profiles of TRH release from copoly(D,L-lactic/glycolic acid) microspheres **72**, 199
Heya, T., H. Okada, Y. Tanigawara, Y. Ogawa, H. Toguchi, Effects of counteranion of TRH and loading amount on control of TRH release from copoly(DL-lactic/glycolic acid) microspheres prepared by an in-water drying method **69**, 69
Heya, T., see Okada, H. **54**, 231
Heyman, M., A. Bonfils, M. Fortier et al., Intestinal absorption of RU 41740, an immunomodulating compound extracted from *Klebsiella pneumoniae*, across duodenal epithelium and Peyer's patches of the rabbit **37**, 33
Heyman, M., see Fortier, M. **52**, 27
Hibi, T., see Kakee, A. **111**, 15
Hickey, A.J., R.N. Dalby, P.R. Byron, Effects of surfactants on aerosol powders in suspension. Implications for airborne particle size **42**, 267
Hickey, A.J., see Masinde, L.E. **100**, 123
Hickman, J.A., see Alpar, H.O. **62**, 133
Hidalgo, I.J., F.M. Ryan, G.J. Marks, P.L. Smith, pH-Dependent transepithelial transport of cephalexin in rabbit intestinal mucosa **98**, 83
Hidalgo, I.J., see Eddy, E.P. **115**, 79
Hidalgo, I.J., see Gochoco, C.H. **104**, 187
Hidalgo, I.J., see Shah, P.K. **63**, 281
Hidalgo, I.J., see Smith, P.L. **106**, 95
Hiestand, E.N., Tablet bond. I. A theoretical model **67**, 217
Hiestand, E.N., D.P. Smith, Tablet bond. II. Experimental check of model **67**, 231
Hiestand, E.N., see Jetzer, W.E. **26**, 329
Higaki, K., I. Kishimoto, H. Komatsu et al., Effect of medium-chain glycerides on the intestinal absorption and the hepatobiliary transport of Phenol red **36**, 131
Higaki, K., M. Takeuchi, M. Nakano, Estimation and enhancement of in vitro corneal transport of S-1033, a novel antiglaucoma medication **132**, 165
Higashi, Y., see Kadir, S. **33**, 235; **40**, 257
Higashi, Y., see Murakami, T. **49**, 173; **54**, 259
Higashiyama, K., see Nakamura, Y. **145**, 29
Higginbottom, S., C.B. Mallinson, S.J. Burns, D. Attwood, S.G. Barnwell, Ursodeoxycholic acid: Effects of formulation on in vitro dissolution **109**, 173
Higginbottom, S., see Barnwell, S.G. **128**, 145
Higginbottom, S., see Burns, S.J. **110**, 291; **121**, 37; **134**, 223; **141**, 9
High, A.J., see Malhi, J.S. **43**, 77
Higuchi, F., see Seta, Y. **41**, 245; 255
Higuchi, T., see Akazawa, M. **50**, 53
Higuchi, T., see Anderson, B.D. **1**, 15

Higuchi, T., see Bhatt, P.P. **50**, 197
Higuchi, T., see Itoh, T. **33**, 265
Higuchi, T., see Liversidge, G.G. **23**, 87; **30**, 247
Higuchi, T., see Nghiem, B.T. **44**, 125
Higuchi, T., see Nguyen, T.H. **25**, 1
Higuchi, T., see Nishihata, T. **21**, 239; **31**, 185
Higuchi, T., see Wong, O. **52**, 191
Higuchi, W.I., see Corrigan, O.I. **5**, 229
Higuchi, W.I., see Fox, J.L. **2**, 41
Higuchi, W.I., see Ghanem, A.H. **78**, 137
Higuchi, W.I., see Gonsho, A. **65**, 183
Higuchi, W.I., see Inamori, T. **105**, 113
Higuchi, W.I., see Kim, Y.H. **80**, 17
Higuchi, W.I., see Lamson, M.L. **21**, 143
Higuchi, W.I., see Liu, P. **64**, 7; **78**, 123
Higuchi, W.I., see Masada, T. **49**, 57
Higuchi, W.I., see Morimoto, Y. **32**, 31
Higuchi, W.I., see Muhammad, N.A. **21**, 295
Higuchi, W.I., see Peck, K.D. **98**, 141
Higuchi, W.I., see Seta, Y. **81**, 89
Higuchi, W.I., see Shah, V.P. **82**, 21
Higuchi, W.I., see Sheth, N.V. **28**, 201
Higuchi, W.I., see Sims, S.M. **69**, 109; **77**, 107
Higuchi, W.I., see Spooner, P.J. **15**, 177
Higuchi, W.I., see Srinivasan, V. **60**, 133
Hikal, A.H., Effect of polysorbate 80 on the apparent partition coefficient of drugs and on their intestinal absorption in the rat. II. Phenobarbital **7**, 205
Assay of fusidic acid in plasma by high-performance liquid chromatography **13**, 297
Hikal, A.H., see Gouda, M.W. **22**, 257
Hilbers, H.W., see Jansen, A.C.A. **75**, 193
Hileman, G.A., S.R. Goskonda, A.J. Spalitto, S.M. Upadrashta, Response surface optimization of high dose pellets by extrusion and spheronization **100**, 71
Hileman, G.A., see Goskonda, S.R. **111**, 89
Hileman, G.A., see Katikaneni, P.R. **117**, 13
Hileman, G.A., see Upadrashta, S.M. **112**, 173
Hill, D.G., A.R. Barnes, Compatibility of phenylmercuric acetate with cefuroxime and ceftazidime eye drops **147**, 127
Hill, J.M., see Callegan, M.C. **123**, 173
Hill, J.M., see Gangarosa, Sr. L.P. **123**, 159
Hill, R.M., see Sutananta, W. **125**, 123
Hillery, A., see Toth, I. **105**, 241
Hillery, A.M., A.T. Florence, The effect of adsorbed poloxamer 188 and 407 surfactants on the intestinal uptake of 60 nm polystyrene particles after oral administration in the rat **132**, 123
Hillery, A.M., see Toth, I. **102**, 223
Hilton, A.K., P.B. Deasy, In vitro and in vivo evaluation of an oral sustained-release floating dosage form of amoxycillin trihydrate **86**, 79
Hilton, J.E., M.P. Summers, The effect of wetting agents on the dissolution of indomethacin solid dispersion systems **31**, 157
The effect of polyvinylpyrrolidones on intestinal ulceration caused by indomethacin **32**, 13
A comparison between the in vivo serum levels and in vitro dissolution of indomethacin—polymer dispersion systems **33**, 219
Himmelstein, K.J., see Nguyen, T.H. **25**, 1
Hincal, A.A., see Ciftci, K. **131**, 73
Hincal, A.A., see Repta, A.J. **5**, 149
Hindle, M., P.R. Byron, Dose emissions from marketed dry powder inhalers **116**, 169
Hindle, M., P.R. Byron, N.C. Miller, Cascade impaction methods for dry powder inhalers using the high flowrate Marple–Miller impactor **134**, 137
Hino, T., see Kawashima, Y. **72**, 65; **89**, 9; **99**, 229; **119**, 139
Hino, T., see Morishima, K. **105**, 11
Hino, T., see Murakami, H. **149**, 43
Hino, T., see Niwa, T. **121**, 45
Hino, T., see Ohwaki, T. **85**, 19; **93**, 61
Hino, T., see Schade, A. **117**, 209
Hino, T., see Takeuchi, H. **86**, 25
Hinterleitner, M., see Djordjevic, N.M. **81**, 21
Hintz, R.J., K.C. Johnson, The effect of particle size distribution on dissolution rate and oral absorption **51**, 9
Hinz, R.S., see Gummer, C.L. **40**, 101
Hirai, S., T. Yashiki, T. Matsuzawa, H. Mima, Absorption of drugs from the nasal mucosa of rat **7**, 317
Hirai, S., T. Yashiki, H. Mima, Effect of surfactants on the nasal absorption of insulin in rats **9**, 165
Hirai, S., T. Yashiki, H. Mima, Mechanisms for the enhancement of the nasal absorption of insulin by surfactants **9**, 173
Hirai, S., T. Yoshioka, T. Kashihara et al., Bioavailability of 5-[4-(1-methylcyclohexylmethoxy)benzyl]-thiazolidine-2,4-dione (ADD-3878) in beagle dogs **15**, 261
Hirai, S., see Iga, K. **6**, 43
Hirakawa, Y., see Narisawa, S. **104**, 95; **148**, 85
Hirata, H., see Kitagawa, S. **98**, 203
Hirate, J., C. Zhu, I. Horikoshi, S. Nagase, Disposition of Warfarin in analbuminemic rats **65**, 149
Hirayama, F., M. Kurihara, K. Uekama, Improvement of chemical instability of prostacyclin in aqueous solution by complexation with methylated cyclodextrins **35**, 193
Hirayama, F., see Aungst, B.J. **38**, 199
Hirayama, F., see Hoshino, T. **38**, 265; **50**, 45
Hirayama, F., see Soliman, O.A. **149**, 73
Hirayama, F., see Uekama, K. **10**, 1; **13**, 253; **16**, 327
Hirayama, F., see Yamamoto, M. **49**, 163
Hirnle, P., R. Schubert, Liposomes containing blue dye for preoperative lymph node staining: Distribution and stability in dogs after endolymphatic injection **72**, 259
Hirotani, Y., see Nishihata, T. **42**, 257
Hirsch, C.A., see Pearlman, R. **18**, 53
Hirsch, R., C.C. Muller-Goymann, Fitting of diffusion coefficients in a three-compartment sustained release drug formulation using a genetic algorithm **120**, 229
Hirvonen, J., R. Sutinen, P. Paronen, A. Urtti, Transdermal penetration enhancers in rabbit pinna skin: Duration of

action, skin irritation, and in vivo/in vitro comparison **99**, 253
Hisaichi, S.I., see Kurosaki, Y. **47**, 13; **49**, 47; **51**, 47
Hiselius, J., see Prisell, P.T. **85**, 51
Hiura, M., H. Kawashima, N. Kaneniwa, Hepatic first-pass metabolism of hexobarbital in rabbits and dogs **18**, 185
Hiura, M., S. Nakagawa, H. Kawashima, N. Kaneniwa, Effect of lipid solubility on hepatic first-pass metabolism of barbiturates in rabbits **20**, 73
Hiura, M., see Kaneniwa, N. **26**, 157
Hiwatari, K.I., see Sakuma, S. **149**, 93
Hjartstam, J., K. Borg, B. Lindstedt, The effect of tensile stress on permeability of free films of ethyl cellulose containing hydroxypropyl methylcellulose **61**, 101
Hjartstam, J., see Lindstedt, B. **56**, 261; **67**, 21
Hjortkjaer, R.K., see Bechgaard, E. **128**, 287
Ho, A.H.L., S.C. Popat, J.K. Sugden, Some effects of metal ions on the photogeneration of hydroxyl radicals **104**, 247
Ho, A.H.L., A. Puri, J.K. Sugden, Effect of sweetening agents on the light stability of aqueous solutions of L-ascorbic acid **107**, 199
Ho, A.H.L., see Allman, J. **115**, 241
Ho, C.K., see Law, S.L. **128**, 139
Ho, H.O., C.Y. Chen, Diffusion characteristics of fibrin films **90**, 95
Ho, H.O., H.L. Su, T. Tsai, M.T. Sheu, The preparation and characterization of solid dispersions on pellets using a fluidized-bed system **139**, 223
Ho, M.Y.K., see Chen, T.K. **59**, 211
Ho, N.T., see Ruelle, P. **87**, 47
Ho, N.F.H., see Durrani, M.J. **24**, 209
Ho, N.F.H., see Fox, J.L. **2**, 41
Ho, N.F.H., see Ganesan, M.G. **20**, 139
Hoa, N.T., A. Michoel, R. Kinget, Dissolution testing of artemisinin solid oral dosage forms **138**, 185
Hoa, Ngo T., J. Vertommen, R. Kinget, Formulation of artemisinin tablets **146**, 271
Hoare, C., see Taylor, K.M.G. **98**, 45
Hobden, J.A., see Callegan, M.C. **123**, 173
Hodgin, L., see Suryanarayanan, R. **78**, 77
Hodoshima, N., C. Udagawa, T. Ando, H. Fukuyasu, H. Watanabe, S. Nakabayashi, Lipid nanoparticles for delivering antitumor drugs **146**, 81
Hodoshima, N., see Hodoshima, N. **146**, 81
Hodsdon, A.C., see Melia, C.D. **100**, 263
Hoelgaard, A., N. Moller, Hydrate formation of metronidazole benzoate in aqueous suspensions **15**, 213
Hoelgaard, A., B. Mollgaard, E. Baker, Vehicle effect on topical drug delivery. IV. Effect of *N*-methylpyrrolidone and polar lipids on percutaneous drug transport **43**, 233
Hoelgaard, A., see Bundgaard, H. **15**, 285; **55**, 91
Hoelgaard, A., see Mollgaard, B. **12**, 153; **15**, 185
Hoelgaard, A., see Wotton, P.K. **24**, 19
Hoffman, A., M. Donbrow, S.T. Gross et al., Fundamentals of release mechanism interpretation in multiparticulate systems: Determination of substrate release from single microcapsules and relation between individual and ensemble release kinetics **29**, 195
Hoffman, A., A. Nyska, A. Avramoff, G. Golomb, Continuous versus pulsatile administration of erythropoietin (EPO) via the uterus in anemic rats **111**, 197
Hoffman, A., see Gross, S.T. **29**, 213
Hoffman, M., see Kedzierewicz, F. **58**, 221; **117**, 247
Hoffman, M., see Marchal-Heussler, L. **58**, 115
Hoffman, M., see Zinutti, C. **133**, 97
Hoffmann, A.J., see Shao, Z. **112**, 181
Hoffmann, M., see Kedzierewicz, F. **94**, 69
Hogan, J.E., see Dabbagh, M.A. **140**, 85
Hogan, J.E., see Ford, J.L. **24**, 327, 339; **40**, 223; **71**, 95, 213
Hogan, J.E., see Mitchell, K. **100**, 143, 165, 175
Hogan, J.E., see Okutgen, E. **119**, 193
Holborow, D., see Medlicott, N.J. **84**, 85; **143**, 25
Holbrook, P., see Buckton, G. **116**, 113; **122**, 130
Holbrook, P.A., see Irwin, W.J. **113**, 25
Holbrook, W.P., see Kristmundsdottir, T. **139**, 63
Holbrook, W.P., see Sveinsson, S.J. **95**, 105
Holgado, M.A., I. Caraballo, J. Alvarez-Fuentes, M.J. Fernandez-Hervas, M. Fernandez-Arevalo, A.M. Rabasco, Influence of diluents and manufacturing method on the in vitro dissolution of carteolol hydrochloride matrix tablets **118**, 151
Holgado, M.A., M. Fernandez-Arevalo, J. Alvarez-Fuentes, I. Caraballo, J.M. Llera, A.M. Rabasco, Physical characterization of carteolol: Eudragit^{R} L binding interaction **114**, 13
Holgado, M.A., M.J. Fernandez-Hervas, J. Alvarez-Fuentes, M.T. Vela, A.M. Rabasco, A. Fini, Characterization of modified paracetamol by means of SEM and fractal analysis **142**, 143
Holgado, M.A., M.J. Fernandez-Hervas, M. Fernandez-Arevalo, A.M. Rabasco, Use of fractal dimensions in the study of excipients: Application to the characterization of modified lactoses **121**, 187
Holgado, M.A., M.J. Fernandez-Hervas, A.M. Rabasco, A. Fini, Characterization study of a diclofenac salt by means of SEM and fractal analysis **120**, 157
Holgado, M.A., see Alvarez-Fuentes, J. **139**, 237; **148**, 219
Holgado, M.A., see Caraballo, I. **96**, 175; **109**, 229
Holgado, M.A., see Fernandez-Hervas, M.J. **108**, 187; **113**, 39; **136**, 101
Holgado, M.A., see Fini, A. **121**, 19
Holgado, M.A., see Rabasco, A.M. **67**, 201
Holgado-Villafuerte, M.A., see Fernandez-Arevalo, M. **95**, 117
Holinej, J., see Johnson, J.L. **90**, 151
Holland, J.M., see Lambert, W.J. **95**, 181
Holland, S.J., J.K. Warrack, Low-temperature scanning electron microscopy of the phase inversion process in a cream formulation **65**, 225
Holland, S.J., see Srivastava, R.C. **38**, 239
Hollenbeck, R.G., Bioavailability of phenylpropanolamine HCl from tablet dosage forms containing croscarmellose sodium **47**, 89
Hollenbeck, R.G., see Coffin, Beach D.P. **17**, 313
Hollenbeck, R.G., see Mitrevej, A. **14**, 243

Hollenbeck, R.G., see Tabibi, S.E. **18**, 169
Holley, J.L., see Toth, I. **79**, 39; **83**, 131
Holley, J.L., see Wood, I.P. **87**, 141
Hollingsbee, D., see Fitzgerald, P. **83**, 177
Hollingsbee, D.A., J.E. Fairbrother, G.P. Martin, C. Marriott, L. Monger, The effect of a hydrocolloid dermatological patch (Actiderm) in potentiating the skin blanching activity of triamcinolone acetonide **77**, 199
Hollingsbee, D.A., see Kadir, R. **60**, 139
Hollingsbee, D.A., see Walker, M. **70**, 167
Hollingsbee, D.A., see Waring, M.J. **94**, 211
Hollingworth, A., see Newman, S.P. **102**, 127
Hollo, J., see Cserhati, T. **108**, 69
Holm, P., see Bertelsen, F. **111**, 117
Holman, L.E., The compressibility of pharmaceutical particulate systems. An illustration of percolation **71**, 81
The normalization of the solid fraction of particulate compacts **89**, R17
Holman, L.E., H. Leuenberger, Effect of compression speed on the relationship between normalised solid fraction and mechanical properties of compacts **57**, R1
Holmberg, D., see Seminoff, L.A. **54**, 251
Holmes, C.J., M.C. Allwood, The assessment of air-inlet filters for intravenous infusions in glass bottles **1**, 133
The growth of micro-organisms in parenteral nutrition solutions containing amino acids and sugars **2**, 325
Holmgren, J., see O'Hagan, D.T. **119**, 251
Holohan, E.M., see Corrigan, O.I. **18**, 195
Holthuis, J.J.M., see Beijnen, J.H. **41**, 169
Holthuis, J.J.M., see Vendrig, D.E.M.M. **43**, 131; **50**, 189
Holzer, A.W., J. Sjogren, Evaluation of sodium stearyl fumarate as a tablet lubricant **2**, 145
Friction coefficients of tablet masses **7**, 269
Holzer, A.W., see Ragnarsson, G. **3**, 127
Holzner, P.M., B.W. Muller, An in vitro evaluation of various spacer devices for metered-dose inhalers using the Twin Impinger **106**, 69
Particle size determination of metered dose inhalers with inertial separation methods: Apparatus A and B (BP), Four Stage Impinger and Andersen Mark II cascade impactor **116**, 11
Holzschuh, J., see Muller, C.E. **57**, 41
Homeida, M.M.A., see Ali, H.M. **42**, 155
Homeida, M.M.A., see El Turabi, H. **53**, 249
Honda, D.H., see Guy, R.H. **19**, 129
Hong, L.Z., see Kurosaki, Y. **49**, 47
Hong, M.S., see Kim, C.K. **95**, 43
Honigberg, I.L., see Vallner, J.J. **16**, 47
Hoogmartens, J., see Cachet, T. **55**, 59
Hoogmartens, J., see Naidong, W. **96**, 13
Hoogmartens, J., see Paesen, J. **113**, 215
Hoogmartens, J., see Vinckier, C. **55**, 67
Hoogstraate, A.J., J. Verhoef, J. Brusee, A.P. Ijzerman, F. Spies, H.E. Bodde, Kinetics, ultrastructural aspects and molecular modelling of transdermal peptide flux enhancement by *N*-alkylazacycloheptanones **76**, 37
Hook, I.I., Administration sets for intravenous fluids used in the Republic of Ireland **1**, 105
Hooper, G., see Steed, K.P. **112**, 199; **123**, 291
Hoppe, R., see Jachowicz, R. **99**, 321
Hori, M., see Melendres, J.L. **92**, 243
Horibe, K., see Kawashima, Y. **72**, 65
Horie, T., see Kajii, H. **33**, 253
Horikoshi, I., see Hirate, J. **65**, 149
Horikoshi, I., see Liu, H.X. **118**, 55
Horiuchi, T., see Hasegawa, T. **58**, 129
Hornebeck, W., see Toth, I. **125**, 117
Hornykiewytsch, T., see Deasy, P.B. **89**, 251
Horst, Thiermann, U. Spohrer, P. Eyer, Operational evaluation of wet/dry autoinjectors containing atropine in solution and powdered HI 6 **114**, 125
Horsthuis, G.J.B., J.A.H. Van Laarhoven, R.C. Van Rooij, H. Vromans, Studies on upscaling parameters of the Gral high shear granulation process **92**, 143
Horton, R.J., see Wilding, I.R. **84**, 59
Hoshino, T., F. Hirayama, K. Uekama, M. Yamasaki, Reduction of protriptyline-photosensitized hemolysis by β-cyclodextrin complexations **50**, 45
Hoshino, T., P.J. Nicholls, T. Irie, F. Hirayama, K. Uekama, Alleviation in protriptyline-photosensitized skin irritation by di-*O*-methyl-β-cyclodextrin complexation **38**, 265
Hoshino, T., K. Uekama, J. Pitha, Increase in temperature enhances solubility of drugs in aqueous solutions of hydroxypropylcyclodextrins **98**, 239
Hoshino, T., see Pitha, J. **80**, 243, 253
Hosny, E.A., Polycarbophil as a controlled release matrix **98**, 235
Formulation and comparative evaluation of bioadhesive containing diclofenac sodium and commercial enteric coated tablets in-vitro and in dogs **133**, 149
Hosny, E.A., A.A. Al-Angary, Bioavailability of sustained release indomethacin suppositories containing polycarbophil **113**, 209
Hosny, E.A., S.S. Abdel-Hady, K.E.H. El-Tahir, Formulation, in-vitro release and ex-vivo spasmolytic effects of mebeverine hydrochloride suppositories containing polycarbophil or polysorbate 80 **142**, 163
Hosny, E.A., Y.M. El-Sayed, M.A. Al-Meshal, A.A. Al-Angary, Effect of food on bioavailability of bioadhesive-containing indomethacin tablets in dogs **112**, 87
Hosny, E.A., E.M. Niazy, A.S. El-Gorashi, Effect of polycarbophil concentration on in vitro release and in vivo availability in beagle dogs of dihydroergotamine mesylate suppositories **117**, 147
Hosny, E.A., E.M. Niazy, M.M. El-Dardari, Effect of polycarbophil concentration on diclofenac sodium bioavailability from suppositories in beagle dogs **136**, 37
Hosoda, J.I., see Unezaki, S. **126**, 41; **144**, 11
Hosoya, K.I., see Kubo, H. **103**, 27
Hosoya, K.I., see Morimoto, Y. **32**, 31
Hossain, M., J.W. Ayres, Relative bioavailability of a novel sustained-release acetaminophen molded tablet **133**, 223
Hou, S., see Zhang, Z. **139**, 1

Hou, S.Y.E., G.L. Flynn, Phase volume and partitioning effects on drug delivery from topical emulsions **66**, 79
Hou, X.P., J.T. Carstensen, Compression characteristics of basic tricalcium phosphate ($Ca_3(PO_4)_2$, $Ca(OH)_2$) **25**, 207
Houjou, T., see Katayama, N. **110**, 155; **115**, 87
Hovgaard, L., H. Brondsted, H.M. Nielsen, Drug delivery studies in Caco-2 monolayers. II. Absorption enhancer effects of lysophosphatidylcholines **114**, 141
Hovgaard, L., H. Jacobs, N.A. Mazer, S.W. Kim, Stabilization of insulin by alkylmaltosides. A. Spectroscopic evaluation **132**, 107
Hovgaard, L., H. Jacobs, D.E. Wilson, S.W. Kim, Stabilization of insulin by alkylmaltosides. B. Oral absorption in vivo in rats **132**, 115
Hovgaard, L., see Brondsted, H. **114**, 151
Howard, D.R., see Neau, S.H. **77**, 239
Howard, J.R., P.L. Gould, The use of co-solvents in parenteral formulation of low-solubility drugs **25**, 359
Howard, J.R., J. Hadgraft, The clearance of oily vehicles following intramuscular and subcutaneous injections in rabbits **16**, 31
Howard, J.R., see Gould, P.L. **51**, 195
Howard, J.R., see Waring, M.J. **36**, 29; **40**, 15
Howard, K.A., see Jenkins, P.G. **102**, 261
Howe, K.D., see Rosen, A. **32**, 171
Howes, C., see Steward, A. **104**, 145
Hristov St., I., see Kalinkova, G.N. **115**, 245
Hruby, V., see Ugwu, S.O. **102**, 193
Hsia, C.K., see Chow, A.H.L. **75**, 219; **126**, 21
Hsu, C.C., see Lam, X.M. **142**, 85
Hsu, C.C., see Maa, Y.F. **140**, 155; **144**, 47
Hsu, H.C., see Chen, C.Y. **141**, 171
Hsu, H.C., see Cheng, H.W. **120**, 137
Hsu, L.R., Y.B. Huang, P.C. Wu, Y.H. Tsai, Percutaneous absorption of piroxicam from FAPG base through rat skin: Effects of fatty acid added to FAPG base **106**, 1
Hsu, L.R., Y.H. Tsai, Y.B. Huang, The effect of pretreatment by penetration enhancers on the in vivo percutaneous absorption of piroxicam from its gel form in rabbits **71**, 193
Hsu, L.R., see Lin, H.H. **112**, 165
Hsu, L.R., see Tsai, Y.H. **24**, 61, 101
Hsu, S.H., see Chuo, W.H. **134**, 247; **144**, 241
Hu, O.Y.P., W.S. Liou, T.J. Su, S.F. Chiam, J.S. Lai, M.L. Chu, P.H. Chung, Pharmacokinetic properties of intramuscular gentamicin in Chinese subjects **53**, 85
Hu, P., see Sinko, P.J. **109**, 125
Hua, S., see Naidong, W. **96**, 13
Huang, C.H., see Hussain, A. **21**, 233, 289
Huang, L., see Sorgi, F.L. **144**, 131
Huang, M.H., see Desai, D.S. **107**, 141
Huang, M.Y., see Byron, P.R. **1**, 219
Huang, M.Y., see Notari, R.E. **1**, 233
Huang, Y.B., P.C. Wu, H.M. Ko, Y.H. Tsai, Cardamom oil as a skin permeation enhancer for indomethacin, piroxicam and diclofenac **126**, 111
Effect of pretreatment by cardamom oil on in vitro percutaneous penetration of piroxicam gel **131**, 137
Effect of pretreatment by cardamom oil on in vivo percutaneous absorption of piroxicam gel: Pharmacokinetic analysis in rabbits **134**, 183
Huang, Y.B., see Fang, J.Y. **126**, 119; **128**, 169; **130**, 121; **131**, 143; **143**, 47; **145**, 175; **149**, 183
Huang, Y.B., see Hsu, L.R. **71**, 193; **106**, 1
Huang, Y.B., see Lin, H.H. **112**, 165
Huang, Y.B., see Wu, P.C. **143**, 119; **145**, 215; **148**, 41
Huang, Y.Y., S.M. Wu, Stability of peptides during iontophoretic transdermal delivery **131**, 19
Huang, Y.Y., S.M. Wu, C.Y. Wang, T.S. Jiang, Response surface method as an approach to optimization of iontophoretic transdermal delivery of pilocarpine **129**, 41
Huang, Y.C., see Corbo, D.C. **50**, 253
Huanga, L., see Mori, A. **131**, 57
Hubbard, A.W., see Bratty, J.R. **115**, 119
Huc, A., see Pajean, M. **77**, 31
Hudecz, F., see Pimm, M.V. **79**, 77
Hudson, A.J., W. Lee, J. Porter, J. Akhtar, R. Duncan, S. Akhtar, Stability of antisense oligonucleotides during incubation with a mixture of isolated lysosomal enzymes **133**, 257
Hudson, A.J., K.J. Lewis, V.M. Rao, S. Akhtar, Biodegradable polymer matrices for the sustained exogenous delivery of a biologically active c-*myc* hammerhead ribozyme **136**, 23
Hudson, A.J., see Hudson, A.J. **133**, 257
Huf, F.A., see De, Vries H. **20**, 265
Hugbo, P.G., An evaluation of antifungal properties of peppermint water **10**, 193
Hughes, P.J., see Doherty, M.M. **111**, 205
Hughes, R.A., see Toth, I. **68**, 191; **73**, 259; **77**, 13
Hugo, W.B., Membrane-active antimicrobial drugs—A reappraisal of their mode of action in the light of the chemiosmotic theory **1**, 127
Hugo, W.B., see Denyer, S.P. **25**, 245; **26**, 89; **29**, 29
Huguet, J., see Illum, L. **26**, 113
Huhtikangas, A., see Lindholm, T. **21**, 119
Hui, H.W., J.R. Robinson, Ocular delivery of progesterone using a bioadhesive polymer **26**, 203
Hui, H.W., see Lovell, M.W. **109**, 45
Hui, J., see Adjei, A. **107**, 57
Hui, L.K., W.H. Thomas, Rheological parameters for *Limulus* amoebocyte lysate-endotoxin reaction **10**, 133
Huikari, A., see Marvola, M. **3**, 13
Huizinga, T., see Moolenaar, F. **1**, 323; **4**, 99, 195; **5**, 127
Huizinga, T., see Schoonen, A.J.M. **4**, 141; **7**, 29
Hulin, A., see Bourget, P. **128**, 223
Hulshoff, A., see Janssen, M.J.H. **23**, 1
Humbert, P., see Makki, S. **133**, 245
Hume, L., see Kyyronen, K. **80**, 161
Hume, L.R., H.K. Lee, L. Benedetti, Y.D. Sanzgiri, E.M. Topp, V.J. Stella, Ocular sustained delivery of prednisolone using hyaluronic acid benzyl ester films **111**, 295
Huneau, J.F., see Benkhelifa, S. **145**, 115
Huneau, J.F., see Crauste-Manciet, S. **149**, 241

Hung, C.T., F.C. Lam, D.G. Perrier, A. Souter, A stability study of amphotericin B in aqueous media using factorial design **44**, 117
Hung, C.T., see Durham, D.G. **12**, 31
Hung, C.T., see Gallo, J.M. **22**, 63
Hung, C.T., see Gupta, P.K. **33**, 137, 147; **43**, 167; **51**, 253; **56**, 217; **59**, 57
Hunneyball, I.M., see Illum, L. **29**, 53
Hunneyball, I.M., see Smith, A. **30**, 215
Hunt, C.A., S. Tsang, α-Tocopherol retards autoxidation and prolongs the shelf-life of liposomes **8**, 101
Hunt, C.A., see Schreier, H. **37**, 233
Hunter, E., J.T. Fell, R.T. Calvert, H. Sharma, 'In vivo' disintegration of hard gelatin capsules in fasting and non-fasting subjects **4**, 175
Hunter, E., J.T. Fell, H. Sharma, The gastric emptying of hard gelatin capsules **17**, 59
Hunter, W.H., see Beezer, A.E. **38**, 251; **45**, 153
Huntington, J., see Wong, O. **52**, 191
Huselton, C.A., F. DeGrazia, E. Stahl, D.J. Liberato, A single dose pharmacokinetic study of 13-*cis* retinoic acid (Isotretinoin, Ro 04-3780) in the pregnant New Zealand rabbit using intravenous infusion **134**, 157
Hussain, A., R. Kimura, C.H. Huang, T. Kashihara, Nasal absorption of naloxone and buprenorphine in rats **21**, 233
Hussain, A., R. Kimura, C.H. Huang, R. Mustafa, Nasal absorption of ergotamine tartrate in rats **21**, 289
Hussain, A., see Babhair, S. **13**, 273
Hussain, A., see Chen, J. **123**, 95
Hussain, A., see Muhi-Eldeen, Z. **26**, 15
Hussain, A., see Tawfiq, F.A. **64**, 45
Hussain, A., see Truelove, J. **19**, 17
Hussain, A.A., see Jona, J.A. **123**, 127
Hussain, A.S., see Sathyan, G. **114**, 75
Hussain, I., see Flinn, N. **138**, 167; **143**, 125
Hussain, M., see Maurin, M.B. **94**, 11
Hussain, M.A., B.J. Aungst, Nasal mucosal metabolism of an LH-RH fragment and inhibition with boroleucine **105**, 7
Hussain, M.A., B.J. Aungst, A. Kearney, E. Shefter, Buccal and oral bioavailability of naloxone and naltrexone in rats **36**, 127
Hussain, M.A., L. Mersinger, M.B. Maurin, C. Kettner, In situ characterization of nasal leucine enkephalin degrading aminopeptidase. Susceptibility of the nasal enzyme to boronic acids and phosphorus-containing peptide and amino acid isosteres **117**, 181
Hussain, M.A., see Maurin, M.B. **144**, 99
Hussain, M.S.H., P. York, P. Timmins, A study of the formation of magnesium stearate film on sodium chloride using energy-dispersive X-ray analysis **42**, 89
Influence of commercial and high purity magnesium stearates on sodium chloride and paracetamol DC granules during tabletting **70**, 103
Effect of commercial and high purity magnesium stearates on in-vitro dissolution of paracetamol DC tablets **78**, 203
Hussein, Z., M. Bialer, M. Friedman, I. Raz, Comparative pharmacokinetic evaluation of sustained-release theophylline formulations in dogs and humans **37**, 97
Hussein, Z., see Bialer, M. **16**, 285
Husson, I., B. Leclerc, G. Spenlehauer, M. Veillard, F. Puisieux, G. Couarraze, Influence of size polydispersity on drug release from coated pellets **86**, 113
Husson, I., see Hyppola, R. **133**, 161
Hutchings, D., S. Clarson, A. Sakr, Studies of the mechanical properties of free films prepared using an ethylcellulose pseudolatex coating system **104**, 203
Hutchinson, E.P., see Gorman, S.P. **17**, 291
Hutchinson, L.E.F., see O'Mullane, J.E. **63**, 177
Hutt, A.J., see Avgerinos, A. **68**, 97
Huttenrauch, R., The influence of molecule-size on the state of order and on the order-dependent properties of ointments **6**, 349
Huttenrauch, R., S. Fricke, Mechano-chemical decomposition of drugs through galenic processes **3**, 289
Huttenrauch, R., J. Jacob, Mechanism of the tensid effect in tabletting **1**, 183
Huu-Phuoc, N., H. Nam-Tran, M. Buchmann, U.W. Kesselring, Experimentally optimized determination of the partial and total cohesion parameters of an insoluble polymer (microcrystalline cellulose) by gas-solid chromatography **34**, 217
Huynh, B., see Leomy, C. **115**, 235
Huynh, O., see Gu, L. **45**, 129
Hwang, G.C.C., see Chen, W.G. **82**, 61
Hwang, J.Y., see Anik, S.T. **16**, 181
Hwang, S.J., G.J. Rhee, K.M. Lee, K.H. Oh, C.K. Kim, Release characteristics of ibuprofen from excipient-loaded alginate gel beads **116**, 125
Hwang, S.J., see Jeong, Y.N. **102**, 35
Hwang, S.J., see Kim, C.K. **89**, 91; **147**, 131
Hyppola, R., I. Husson, F. Sundholm, Evaluation of physical properties of plasticized ethyl cellulose films cast from ethanol solution. Part I **133**, 161
Hyvonen-Dabek, M., see Mannermaa, J.P. **103**, 125

I

Iamartino, P., see Gazzaniga, A. **108**, 77
Iannuccelli, V., G. Coppi, M. Bondi, M. Pinelli, A. Mingione, R. Cameroni, Biodegradable intraoperative system for bone infection treatment. II. In vivo evaluation **143**, 187
Iannuccelli, V., G. Coppi, R. Cameroni, Biodegradable intraoperative system for bone infection treatment. I. The drug–polymer interaction **143**, 195
Ibrahim, H., C. Bindschaedler, E. Doelker, P. Buri, R. Gurny, Concept and development of ophthalmic pseudo-latexes triggered by pH **77**, 211
Aqueous nanodispersions prepared by a salting-out process **87**, 239
Ibrahim, H., see Sallam, E. **67**, 247
Ibrahim, H.G., E.S. Sallam, Dissolution from disintegrating tablet: Separate contributions from granules and primary drug particles considered **93**, 111
Ibrahim, K.E., E.I.A. Karim, B.M. Ahmed, I.M. Nour, Effects of tetracycline on the 2 h urinary and salivary excretion of metronidazole **98**, 231

Ibrahim, K.E., see A, Karim E.I. **76**, 261
Ibrahim, S.A., T.H. El-Faham, Tous S. Shawky, E.M. Mostafa, Formulation, release characteristics and evaluation of ibuprofen suppositories **61**, 1
Ibrahim, S.A., see Elshabouri, S.R. **2**, 257
Ibwala, N., M.J. Devleeschouwer, J. Dony, Inhibition of the adhesion of *Escherichia coli* to human urinary epithelial cells **59**, 205
Ichii, T., see Kurono, Y. **80**, 135
Ichikawa, I., see Davis, M. **28**, 157
Ichikawa, M., see Nakashima, M. **72**, R5
Ichikawa, T., see Sakiya, Y. **47**, 185
Ichimura, F., Y. Deguchi, K. Yokogawa, T. Yamana, Physiologically based parmacokinetics of valproic acid in rabbits **27**, 45
Ichimura, F., K. Yokogawa, T. Yamana et al., Physiological pharmacokinetic model for pentazocine. I. Tissue distribution and elimination in the rat **15**, 321
Physiological pharmacokinetic model for distribution and elimination of pentazocine. II. Study in rabbits and scale-up to man **19**, 75
Ichimura, F., see Yamana, T. **11**, 71
Iemura, O., see Fujita, T. **134**, 47
Iga, K., N. Hamaguchi, Y. Igari, Y. Ogawa, H. Toguchi, T. Shimamoto, Heat-specific drug release of large unilamellar vesicle as hyperthermia-mediated targeting delivery **57**, 241
Iga, K., S. Hirai, T. Yashiki, Assessment of local and systemic availability of tribenoside after rectal administration in rats **6**, 43
Iga, T., see Shim, C.K. **8**, 143
Igarashi, Y., see Sasaki, H. **93**, 49
Igari, Y., see Iga, K. **57**, 241
Igari, Y., see Okada, H. **54**, 231
Iglesias, G., see Cal, S. **129**, 253
Iguchi, S., see Kaneo, Y. **44**, 265
Igwilo, C.I., N. Pilpel, Effects of coating the powder on the tensile strength, disintegration and dissolution of lactose tablets **15**, 73
Igwilo, C.I., see Pilpel, N. **68**, 157
Ijntema, K., W.J.M. Heuvelsland, C.A.M.C. Dirix, A.P. Sam, Hydroxyapatite microcarriers for biocontrolled release of protein drugs **112**, 215
Ijzerman, A.P., see Hoogstraate, A.J. **76**, 37
Ikada, Y., see Yamaoka, T. **113**, 149
Ikeda, C., see Yamaguchi, T. **37**, 127
Ikeda, K., see Chen, D. **106**, 249
Ikeda, K., see Kurono, Y. **80**, 135; **81**, 179
Ikeda, K., see Ohta, N. **29**, 137
Ikeda, T., see Tokui, T. **110**, 277
Ikesue, K., P. Kopeckova, J. Kopecek, Degradation of proteins by guinea pig intestinal enzymes **95**, 171
Ilango, K., see Parimoo, P. **100**, 227
Ilievska, M., see Panzova, B. **70**, 187
Ilkka, J., P. Paronen, Prediction of the compression behaviour of powder mixtures by the Heckel equation **94**, 181
Ilkka, J., see Ketolainen, J. **92**, 157
Illek, B., see Piotrovskij, V.K. **119**, 213
Illum, L., Bioadhesive microspheres as a potential nasal drug delivery system **39**, 189
Illum, L., H. Bundgaard, Sorption of drugs by plastic infusion bags **10**, 339
Illum, L., H. Bundgaard, S.S. Davis, A constant partition model for examining the sorption of drugs by plastic infusion bags **17**, 183
Illum, L., S.S. Davis, Cellulose microspheres as a sustained release system for parenteral administration **11**, 323
Illum, L., S.S. Davis, C.G. Wilson et al., Blood clearance and organ deposition of intravenously administered colloidal particles. The effects of particle size, nature and shape **12**, 135
Illum, L., N.F. Farraj, H. Critchley, B.R. Johansen, S.S. Davis, Enhanced nasal absorption of insulin rats using lysophosphatidylcholine **57**, 49
Illum, L., N.F. Farraj, S.S. Davis, B.R. Johansen, D.T. O'Hagan, Investigation of the nasal absorption of biosynthetic human growth hormone in sheep—use of a bioadhesive microsphere delivery system **63**, 207
Illum, L., I.M. Hunneyball, S.S. Davis, The effect of hydrophilic coatings on the uptake of colloidal particles by the liver and by peritoneal macrophages **29**, 53
Illum, L., J. Huguet, M. Vert, S.S. Davis, Accumulation in the lung of [^{75}Se]norcholestenol administered intravenously in a globular partially quaternized poly[thio-1-(*N*,*N*-diethylaminomethyl)ethylene] carrier system **26**, 113
Illum, L., P.D.E. Jones, J. Kreuter et al., Adsorption of monoclonal antibodies to polyhexylcyanoacrylate nanoparticles and subsequent immunospecific binding to tumour cells in vitro **17**, 65
Illum, L., M.A. Khan, E. Mak, Evaluation of carrier capacity and release characteristics for poly(butyl 2-cyanoacrylate) nanoparticles **30**, 17
Illum, L., P. West, C. Washington, S.S. Davis, The effect of stabilising agents on the organ distribution of lipid emulsions **54**, 41
Illum, L., J. Wright, S.S. Davis, Targeting of microspheres to sites of inflammation **52**, 221
Illum, L., see Aspden, T.J. **122**, 69
Illum, L., see Chandler, S.G. **70**, 19; **76**, 61; **117**, 139
Illum, L., see Davis, S.S. **30**, 29; **38**, 133; **82**, 99
Illum, L., see Douglas, S.J. **34**, 145
Illum, L., see Fisher, A.N. **74**, 147
Illum, L., see Mayor, S.H. **149**, 123
Illum, L., see Moghimi, S.M. **68**, 121
Illum, L., see Porter, C.J.H. **83**, 273
Illum, L., see Richardson, J.L. **56**, 29; **77**, 75; **88**, 319
Illum, L., see Torrado, J.J. **51**, 85
Ilvessalo, P., see Laine, E. **20**, 307
Im, E.B., see Kim, C.K. **101**, 191
Imada, T., see Nagano, H. **147**, 119
Imai, K., see Asano, M. **67**, 67
Imai, T., Y. Saito, H. Matsumoto, T. Satoh, M. Otagiri, Influence of egg albumin on dissolution of several drugs **53**, 7

Imai, T., S. Shiraishi, H. Saito, M. Otagiri, Interaction of indomethacin with low molecular weight chitosan, and improvements of some pharmaceutical properties of indomethacin by low molecular weight chitosans **67**, 11
Imamura, Y., L.H. Wang, C.S. Lee et al., Acetaminophen–diphenhydramine interaction in rabbits **8**, 277
Imamura, Y., L.H. Wang, C.S. Lee, J.H. Perrin, Effects of chlorpromazine and atropine on acetaminophen absorption in rabbits **5**, 25
Imanidis, G., K.C. Hartner, N.A. Mazer, Intestinal permeation and metabolism of a model peptide (leuprolide) and mechanisms of permeation enhancement by non-ionic surfactants **120**, 41
Imanidis, G., see Gonsho, A. **65**, 183
Imasaka, K., M. Yoshida, H. Fukuzaki, M. Asano, M. Kumakura, T. Mashimo, H. Yamanaka, T. Nagai, A new biodegradable implant consisting of waxy-type poly(ε-caprolactone-*co*-▲-valerolactone) and estramustine **68**, 87
New biodegradable polymers of L-lactic acid and aromatic hydroxy acids and their applications in drug delivery systems **81**, 31
Imasaka, K., see Oribe, T. **124**, 27
Impagnatiello, A., see Di Giulio, A. **89**, 133
Inaba, A., see Kikuchi, S. **42**, 193
Inada, Y., see Yoshimura, Y. **103**, 1
Inagaki, H., see Nishihata, T. **34**, 157; **38**, 83
Inamori, T., A.H. Ghanem, W.I. Higuchi, V. Srinivasan, Macromolecule transport in and effective pore size of ethanol pretreated human epidermal membrane **105**, 113
Inamori, T., see Lee, P.H. **93**, 139
Infeld, M.H., see Felton, L.A. **113**, 17; **127**, 203
Infeld, M.H., see Shah, N.H. **106**, 15
Ingani, H.M., J. Timmermans, A.J. Moes, Conception and in vivo investigation of peroral sustained release floating dosage forms with enhanced gastrointestinal transit **35**, 157
Inghelbrecht, S., J.P. Remon, Fernandes de Aguair, B. Walczak, D.L. Massart, F. Van de Velde, P. Baets, H. DeVermeersch, P. De Backer, Instrumentation of a roll compactor and the evaluation of the parameter settings by neural networks **148**, 103
Ingkatawornwong, S., see Tantishaiyakul, V. **143**, 59
Ingvarsdottir, K., see Kristmundsdottir, T. **137**, 159
Ingvarsdottir, K., see Sveinsson, S.J. **92**, 29
Inotsume, N., M. Nakano, Reversible azomethine bond cleavage of 2′-fluoro derivatives of benzodiazepines in acidic solutions at body temperature **6**, 147
Reversible azomethine bond cleavage of nitrofurantoin in acidic solutions at body temperature **8**, 111
Hydrolytic behavior of dantrolene in acidic media at body temperature **17**, 357
Inotsume, N., see Nakano, M. **3**, 195
Inoue, N., D. Kobayashi, M. Kimura, M. Toyama, I. Sugawara, S. Itoyama, M. Ogihara, K. Sugibayashi, Y. Morimoto, Fundamental investigation of a novel drug delivery system, a transdermal delivery system with jet injection **137**, 75
Inoue, N., see Sugibayashi, K. **113**, 189
Inuma, M., see Hatanaka, T. **79**, 21
Ip, A.Y., see Niven, R.W. **101**, 81; **109**, 17; **127**, 191
Ip, D.P., G.S. Brenner, J.M. Stevenson et al., High resolution spectroscopic evidence and solution calorimetry studies on the polymorphs of enalapril maleate **28**, 183
Iqbal, M., see Irwin, W.J. **41**, 41; **75**, 211; **83**, 241
Irache, J.M., L. Bergougnoux, I. Ezpeleta, J. Gueguen, A.M. Orecchioni, Optimization and in vitro stability of legumin nanoparticles obtained by a coacervation method **126**, 103
Irache, J.M., C. Durrer, G. Ponchel, D. Duchene, Determination of particle concentration in latexes by turbidimetry **90**, R9
Irache, J.M., see Ezpeleta, I. **131**, 191; **142**, 227
Ireland, D.S., see Ford, J.L. **12**, 11
Irie, T., K. Wakamatsu, H. Arima, H. Aritomi, K. Uekama, Enhancing effects of cyclodextrins on nasal absorption of insulin in rats Chondroitin sulfate: A potential biodegradable carrier for colon-specific drug delivery **84**, 129
Irie, T., Y. Tsunenari, K. Uekama, J. Pitha, Effect of bile on the intestinal absorption of α-cyclodextrin in rats **43**, 41
Irie, T., see Abe, K. **123**, 103
Irie, T., see Arima, H. **57**, 107
Irie, T., see Hoshino, T. **38**, 265
Irie, T., see Pitha, J. **80**, 253
Irie, T., see Uekama, K. **23**, 35; **25**, 339
Irwin, B., see Nicklin, P. **83**, 197
Irwin, V.P., J.M. Quigley, R.F. Timoney, Quantitative relationships between in vitro antibacterial activities of cephalosporins and their *n*-octanol/water partition coefficients **34**, 241
1-Octanol-water partition coefficients of the anionic and zwitterionic species of diprotic zwitterionic cephalosporin antibiotics **43**, 187
Irwin, W.J., K.A. Belaid, Drug-delivery by ion-exchange. Stability of ester prodrugs of propranolol in surfactant and enzymatic systems **48**, 159
Irwin, W.J., J.M. Hempenstall, A. Li Wan Po, A.H. Andrews, Controlled-release penicillin complexes: Dissolution and stability of benzathine cloxacillin **20**, 25
Irwin, W.J., P.A. Holbrook, M.J. Dey, The stability of peptides in nasal enzymic systems **113**, 25
Irwin, W.J., M. Iqbal, Thermal stability of bropirimine **41**, 41
Solid-state stability: The effect of grinding solvated excipients **75**, 211
Bropirimine formulation: The dynamic testing of injections **83**, 241
Irwin, W.J., A. Li Wan Po, The dependence of amitriptyline partition coefficients on lipid phase **4**, 47
Irwin, W.J., Q.N. Masuda, Po A.L. Wan, Transesterification of salicylate esters used as topical analgesics **21**, 35
Irwin, W.J., F.D. Sanderson, A. Li Wan Po, Percutaneous absorption of ibuprofen: Vehicle effects on transport through rat skin **66**, 193
Percutaneous absorption of ibuprofen and naproxen: Effect of amide enhancers on transport through rat skin **66**, 243

Irwin, W.J., J.C. Smith, Extraction coefficients and facilitated transport: The effect of absorption enhancers **76**, 151
Irwin, W.J., see Baker, N.D. **52**, 231; **65**, 115
Irwin, W.J., see Cheung, Y.W. **26**, 175; **27**, 325
Irwin, W.J., see Hempenstall, J.M. **23**, 131
Irwin, W.J., see Li Wan Po, A. **16**, 115
Irwin, W.J., see Liak, T.L. **36**, 233
Irwin, W.J., see Meidan, V.M. **118**, 129
Irwin, W.J., see Naik, A. **89**, 51, 65; **90**, 129
Irwin, W.J., see Nicklin, P.L. **123**, 187; **140**, 175
Irwin, W.J., see Walker, I. **104**, 157; **108**, 225
Iseki, K., see Kohri, N. **68**, 255; **81**, 49
Ishida, E., see Tsunematsu, H. **68**, 77
Ishida, K., see Laine, E. **38**, 33
Ishida, O., see Maruyama, K. **111**, 103
Ishida, O., see Unezaki, S. **126**, 41; **144**, 11
Ishida, S., see Sakiya, Y. **47**, 185
Ishida, T., see Kokubo, H. **35**, 181
Ishigaki, K., see Maitani, Y. **146**, 11
Ishigami, M., see Tokui, T. **110**, 277
Ishihara, H., see Hara, T. **42**, 69; **67**, 123
Ishii, M., see Ando, H. **34**, 153
Ishikura, T., T. Senou, H. Ishihara, T. Kato, T. Ito, Drug delivery to the brain. DOPA prodrugs based on a ring-closure reaction to quaternary thiazolium compounds **116**, 51
Ishimaru, S., see Oribe, T. **124**, 27
Ishimitsu, T., H. Sakurai, Structure-ionization relationships of enkephalin and related fragments in aqueous solution **12**, 271
Isimer, A., see Onur, M.A. **88**, 313
Iskander, M.N., see Bibby, D.C. **144**, 61
Islasse, M., see Yamamoto, A. **39**, 255
Ismail, A.A., see Khalil, S.A. **19**, 307
Ismail, F.A., N. Khalafallah, S.A. Khalil, Adsorption of ketoprofen and bumadizone calcium on aluminium-containing antacids and its effect on ketoprofen bioavailability in man **34**, 189
Ismail, F.A., see Etman, M.A. **88**, 433
Ismail, S., D.J. Morton, Ophthalmic uptake of chloramphenicol from proprietary preparations using an in vitro method of evaluation **37**, 11
Isowa, K., see Akimoto, J. **128**, 251; **140**, 257
Isowa, K., see Quan, D. **68**, 239
Isshiki, R., see Ueda, H. **143**, 37
Issopoulos, P.B., Colorimetric determination of dihydralazine sulfate using paramolybdate anion as an oxidizing agent **61**, 261
Analytical investigations of isonicotinic acid hydrazide (isoniazid). VI. Sensitive colorimetric determination of micro amounts of isoniazid using an indirect redox method **70**, 201
Issopoulos, P.B., P.T. Economou, A sensitive colorimetric determination of microquantities of isonicotinic acid hydrazide (isoniazid) **57**, 235
Itiola, O.A., N. Pilpel, Tableting characteristics of metronidazole formulations **31**, 99
Ito, K., see Kurosawa, N. **27**, 81
Ito, R., Y. Machida, T. Sannan, T. Nagai, Magnetic granules: A novel system for specific drug delivery to esophageal mucosa in oral administration **61**, 109
Ito, R., see Watanabe, Y. **110**, 93
Ito, S., see Ohta, N. **85**, 39; **118**, 85
Ito, T., see Ishikura, T. **116**, 51
Ito, Y., see Adachi, T. **124**, 129
Itoh, M., see Nakano, M. **4**, 291
Itoh, T., T. Higuchi, L.J. Caldwell, Dynamic friction of some film coatings on rat stomach mucosa in vitro—Effect of pH **33**, 265
Itoh, T., see Akazawa, M. **50**, 53
Itoh, T., see Bhattachar, S.N. **79**, 263
Itoh, T., see Wong, O. **52**, 191
Itoh, Y., see Kawashima, Y. **75**, 25
Iturrioz, G., see Chang, R.K. **60**, 171
Iula, A.K., see Won, C.M. **79**, 183
Ivanova, T., see Simeonova, M. **43**, 267
Iversen, T., see Ek, R. **125**, 257
Iveson, S.R., see Staniforth, J.N. **31**, 145
Iwakawa, S., see Okumura, K. **88**, 63
Iwaki, M., see Ogiso, T. **98**, 113; **102**, 141
Iwakura, Y., see Morimoto, K. **81**, 119
Iwamoto, K., see Nagasako, S. **81**, 127
Iwamoto, T., see Kawashima, Y. **75**, 25; **89**, 9
Iwamoto, Y., see Takeeda, T. **41**, 21
Iwasa, K., see Kohri, N. **49**, 213
Iwasa, S., see Takada, S. **146**, 147
Iwata, M., see Nagano, H. **147**, 119
Iwatsuru, M., see Unezaki, S. **144**, 11
Iwuagwu, M.A., A. Jideonwo, Preliminary investigations into the in-vitro interaction of folic acid with magnesium trisilicate and edible clay **65**, 63
Iyer, B.V., R.C. Vasavada, In vitro skin penetration of triamcinolone acetonide from lanolin alcohol-ethyl cellulose films **3**, 247
Izgu, E., U. Gungor, The use of natural membranes for in vitro determination of absorption rates of drugs from suppository bases **9**, 107
Izutsu, K., S. Yoshioka, Y. Takeda, The effects of additives on the stability of freeze-dried β-galactosidase stored at elevated temperature **71**, 137
Izutsu, K.I., S. Yoshioka, T. Terao, Stabilization of β-galactosidase by amphiphilic additives during freeze-drying **90**, 187

J

Jabbal-Gill, I., see Fisher, A.N. **74**, 147
Jachowicz, R., Dissolution rates of partially water-soluble drugs from solid dispersion systems. I. Prednisolone **35**, 1
Dissolution rates of partially water-soluble drugs from solid dispersion systems. II. Phenytoin **35**, 7
Jachowicz, R., E. Nurnberg, R. Hoppe, Solid dispersions of oxazepam **99**, 321
Jackson, I.M., see Rubinstein, M.H. **36**, 99
Jackson, I.M., see Timmins, P. **11**, 329
Jackson, J.K., see Zhang, X. **132**, 195; **137**, 199

Jacob, J., see Huttenrauch, R. **1**, 183
Jacob, M., see Bassani, V. **63**, 57
Jacob, M., see Delalonde, M. **130**, 147
Jacob, M., see Doko, M.B. **76**, 199
Jacob, M., see Petrovick, P.D. **76**, 49
Jacob, M., see Raffin, F. **120**, 205; **145**, 247
Jacob, M., see Ruizk, S. **112**, 125
Jacob, M., see Sonanglio, D. **115**, 53
Jacobs, G.P., The effect of γ-rays on semi-synthetic penicillin powders **4**, 299
The radiation sterilization of cefoxitin sodium **7**, 279
Stability of cefazolin and other new cephalosporins following γ irradiation **17**, 29
Jacobs, G.P., L. Dobrilovic, R. Coombes, N. Raghaven, HPLC analysis of γ-irradiated β-lactam antibiotics **32**, 151
Jacobs, H., see Hovgaard, L. **132**, 107, 115
Jacobsen, J., see Burgalassi, S. **133**, 1
Jacobsen, J., B. Van Deurs, M. Pedersen, M. Romer Rassing, TR146 cells grown on filters as a model for human buccal epithelium. I. Morphology, growth, barrier properties, and permeability **125**, 165
Jacobsen, J., M. Pedersen, M. Romer Rassing, TR146 cells as a model for human buccal epithelium. II. Optimisation and use of a cellular sensitivity MTS/PMS assay **141**, 217
Jacobsen, L., see Jorgensen, K. **88**, 23
Jacobsen, L., see Nicklasson, M. **69**, 255
Jacobson, H., see Grant, D.J.W. **5**, 109
Jacobsson, S., see Arbin, A. **28**, 211
Jacobsson, S.E., see Moren, F. **3**, 335
Jacobsson, S.E., see Moren, F. **5**, 287
Jacolot, A., P. Arnaud, D. Lecompte, S. Alamercery, E. Caroff, C. Naveau, T. Guillot, M. Bousselet, F. Brion, Stability and compatibility of 2.5 mg/ml methotrexate solution in plastic syringes over 7 days **128**, 283
Jacque, M., see Chevin, J.C. **45**, 101
Jadoenath, S., see Van de Donk, H. **12**, 57
Jadoul, A., C. Hanchard, S. Thysman, V. Preat, Quantification and localization of fentanyl and TRH delivered by iontophoresis in the skin **120**, 221
Jafari, M.R., A.G. Danti, I. Ahmed, Comparison of polyethylene glycol, polyvinylpyrrolidone and urea as excipients for solid dispersion systems of miconazole nitrate **48**, 207
Jager, R.S., see Riepma, K.A. **90**, R1
Jaggi, M., R.K. Khar, U.P.S. Chauhan, S.V. Gangal, Liposomes as carriers of ^{99m}Tc glucoheptonate for liver imaging **69**, 77
Jahnchen, E., see Oie, S. **48**, 223
Jahnke, R.W.O., J. Kreuter, G. Ross, Content/container interactions. The phenomenon of haze formation on reconstitution of solids for parenteral use **77**, 47
Jain, G.K., A.K. Sharma, S.S. Agrawal, Transdermal controlled administration of verapamil-enhancement of skin permeability **130**, 169
Jain, N.B., see Desai, D.S. **142**, 61
Jain, N.B., see Wesdyk, R. **65**, 69; **93**, 101
Jain, N.K., see Vyas, S.P. **69**, 5
Jaksic, P., K. Mlinaric-Majerski, B. Zorc, M. Dumic, Macromolecular prodrugs. VI. Kinetic study of poly[α,β-(*N*-2-hydroxyethyl-DL-aspartamide)]-ketoprofen hydrolysis **135**, 177
Jakubowski, G., see Jenke, D.R. **108**, 1
Jalal, I., E. Zmaily, N. Najib, Dissolution kinetics of commercially available controlled-release theophylline preparations **52**, 63
Jalal, I., see Najib, N. **44**, 43
Jalal, I., see Shaheen, O. **38**, 123
Jalal, I.M., see Al-Nimry, S.S. **149**, 115
Jalsenjak, I., see Filipovic-Grcic, J. **116**, 39; **135**, 183
Jalsenjak, I., see Maysinger, D. **17**, 129; **63**, 149
Jalsenjak, I., see Skalko, N. **85**, 97
Jalsenjak, I., see Zorc, B. **99**, 135
Jalsenjak, V., see Juretic, D. **61**, 219
Jamali, F., T. Khazainia, Therani M. Rafiee, Absorption rate limited metabolism of salicylate in man: A consideration in bioavailability assessment **9**, 221
Jamali, F., see Dwivedi, S.K. **87**, 95
Jamali, F., see Mehvar, S.R. **7**, 293
Jambut-Absil, A.C., J. Buxeraud, J.F. Lagorce, C. Raby, Charge transfer complexes of drugs with iodine investigation by UV/visible spectroscopy **35**, 129
Jameel, S., see Billany, M.R. **124**, 279
James, K.C., Calculation of molecular surface areas and aqueous solubilities at ambient temperatures **21**, 123
James, K.C., see Armstrong, N.A. **6**, 185, 195; **34**, 125; **41**, 115, 205
James, K.C., see Gassim, A.E.H. **34**, 23
James, K.C., see Gharavi, M. **14**, 325, 333
James, K.C., see Gibbins, L.B. **4**, 353
James, K.C., see Saket, M.M. **21**, 155; **27**, 287
James, M.B., see Podczeck, F. **145**, 221; **149**, 151
James, V.J., see Brain, K.R. **89**, R13
Jamoulle, J.C., see Rolland, A. **76**, 217
Jamoulle, J.C., see Martin, B. **49**, 63
Jamshaid, M., S.J. Farr, P. Kearney, I.W. Kellaway, Poloxamer sorption on liposomes: Comparison with polystyrene latex and influence on solute efflux **48**, 125
Jang, S.H., S.H. Lee, S.H. Ryoo, S.H. Kim, M.G. Lee, Dose-independent pharmacokinetics of recombinant human interferon-αA in rabbits **84**, 273
Jang, S.H., see Lee, S.H. **90**, 81
Jang, S.H., see Yoon, W.H. **102**, 249
Jang, T.F., see Law, S.L. **103**, 81; **116**, 87; **128**, 139
Jani, P.U., A.T. Florence, D.E. McCarthy, Further histological evidence of the gastrointestinal absorption of polystyrene nanospheres in the rat **84**, 245
Jani, P.U., D.E. McCarthy, A.T. Florence, Nanosphere and microsphere uptake via Peyer's patches: Observation of the rate of uptake in the rat after a single oral dose **86**, 239
Titanium dioxide (rutile) particle uptake from the rat GI tract and translocation to systemic organs after oral administration **105**, 157
Janicki, S., see Sznitowska, M. **137**, 125

Jansen, A.C.A., H.W. Hilbers, X.R. Ni, Helden S.P Van, L.H.M. Janssen, Some physical-chemical properties of dantrolene and two of its analogues **75**, 193
Jansen, A.C.A., see de Haan, F.H.N **13**, 75; **18**, 311; **29**, 177
Jansen, J., C. Treiner, C. Vaution, F. Puisieux, Surface modification of alumina particles by nonionic surfactants: Adsorption of steroids, barbiturates and pilocarpine **103**, 19
Jansen, J.A., see Hansen, K.T. **79**, 205
Jansen, M., see Blume, A. **99**, 219
Jansen, W.T., see Westerhuis, J.A. **143**, 151
Janssen, L.H.M., see Cools, A.A. **20**, 335
Janssen, L.H.M., see Jansen, A.C.A. **75**, 193
Janssen, L.H.M., see Van der Giesen, W.F. **12**, 231
Janssen, M.J.H., D.J.A. Crommelin, G. Storm, A. Hulshoff, Doxorubicin decomposition on storage. Effect of pH, type of buffer and liposome encapsulation **23**, 1
Jantti, V., see Urtti, A. **19**, 53
Jarho, P., A. Urtti, D.W. Pate, P. Suhonen, T. Jarvinen, Increase in aqueous solubility, stability and in vitro corneal permeability of anandamide by hydroxypropyl-β-cyclodextrin **137**, 209
Jarvinen, T., P. Suhonen, S. Auriola, J. Vepsalainen, A. Urtti, P. Peura, *O,O'*-(1,4-*xylylene*)bispilocarpic acid esters a new potential double prodrugs of pilocarpine for improved ocular delivery. I. Synthesis and analysis **75**, 249
Bispilocarpic acid monoesters as prodrugs of pilocarpine. I. Preparation and identification **79**, 233
Jarvinen, T., P. Suhonen, A. Urtti, P. Peura, *O,O'*-(1,4-xylylene) bispilocarpic acid esters as new potential double prodrugs of pilocarpine for improved ocular delivery. II. Physicochemical properties, stability, solubility and enzymatic hydrolysis **75**, 259
Bispilocarpic acid monoesters as prodrugs of pilocarpine. II. Physicochemical properties and kinetics of hydrolysis in aqueous solution **79**, 243
Jarvinen, T., see Jarho, P. **137**, 209
Jarvinen, T., see Saarinen-Savolainen, P. **133**, 171
Jarvinen, T., see Suhonen, P. **74**, 221; **127**, 85
Jashnani, R.N., P.R. Byron, Dry powder aerosol generation in different environments: Performance comparisons of albuterol, albuterol sulfate, albuterol adipate and albuterol stearate **130**, 13
Jashnani, R.N., P.R. Byron, R.N. Dalby, Testing of dry powder aerosol formulations in different environmental conditions **113**, 123
Jasuja, A.K., see Hardy, J.G. **142**, 129
Jauw, T.H., see Moolenaar, F. **114**, 117
Javadi, M., see Adeyeye, C.M. **116**, 65
Javaloyes, C., see Diez-Sales, O. **129**, 33
Jawan, O.A., see Badwan, A.A. **28**, 41
Jay, M., R.M. Beihn, G.A. Snyder et al., In vitro and in vivo suppository studies with perturbed angular correlation and external scintigraphy **14**, 343
Jay, M., N. Ogawa, J.B. Bogardus et al., Anisotropy of indium-111 in cetyltrimethylammonium bromide solutions **19**, 97
Jay, M., see MacLean, D.S. **131**, 243
Jay, M., see Parr, A. **32**, 251
Jayakrishnan, A., W.A. Knepp, E.P. Goldberg, Casein microspheres: Preparation and evaluation as a carrier for controlled drug delivery **106**, 221
Jayasekera, P., see Loukas, Y.L. **117**, 85
Jefferis III, R.P., see Schoen, M.P. **114**, 159
Jeffery, H., S.S. Davis, D.T. O'Hagan, The preparation and characterisation of poly(lactide-*co*-glycolide) microparticles. I. Oil-in-water emulsion solvent evaporation **77**, 169
Jeffery, H., see O'Hagan, D.T. **103**, 37; **108**, 133
Jehl, F., see Bourget, P. **128**, 223
Jenke, D.R., E.K. Chess, G. Jakubowski, Modeling of the leachables impact on the engineering of parenteral product container systems **108**, 1
Jenke, D.R., E.K. Chess, D.C. Zietlow, B.E. Rabinow, Model for estimating the accumulation of solutes leaching from polymeric containers into parenteral solutions **78**, 115
Jenke, D.R., see Hayward, D.S. **59**, 245; **66**, 87
Jenkins, J.R.F., J.G. Hardy, C.G. Wilson, Monitoring antacid preparations in the stomach using γ scintigraphy **14**, 143
Jenkins, P.G., K.A. Howard, N.W. Blackhall, N.W. Thomas, S.S. Davis, D.T. O'Hagan, The quantitation of the absorption of microparticles into the intestinal lymph of Wistar rats **102**, 261
Jenquin, M.R., J.W. McGinity, Characterization of acrylic resin matrix films and mechanisms of drug–polymer interactions **101**, 23
Jensen, E., H. Bundgaard, Aminomethylbenzoate esters of chloramphenicol as a novel prodrug type for parenteral administration **70**, 137
Peptide esters as water-soluble prodrugs for hydroxyl containing agents: Chemical stability and enzymatic hydrolysis of benzyl esters of glycine, diglycine and triglycine **71**, 117
Jensen, E., H. Bundgaard, E. Falch, Design of a water-soluble, solution-stable and biolabile prodrug of metronidazole for parenteral administration: *N*-substituted aminomethylbenzoate esters **58**, 143
Jensen, E., see Bundgaard, H. **64**, 75
Jensen, J., C. Cornett, C.E. Olsen, J. Tjornelund, S.H. Hansen, Identification of major degradation products of 5-aminosalicylic acid formed in aqueous solutions and in pharmaceuticals **88**, 177
Jensen, M.M., see Kipp, J.E. **34**, 1
Jensen-Pippo, K., see Habberfield, A. **145**, 1
Jeong, E.J., see Kim, C.K. **115**, 193; **147**, 143
Jeong, K.H., see Lee, S.H. **90**, 81
Jeong, S.Y., see Choi, Y.K. **80**, 9
Jeong, Y.N., S.H. Lee, M.G. Lee, S.J. Hwang, C.K. Kim, Pharmacokinetics of methotrexate after intravenous and intramuscular injection of methotrexate-bearing negatively charged liposomes to rats **102**, 35
Jeong, Y.N., see Yoon, E.J. **97**, 213
Jernberg, M.J., see Conard, G.J. **10**, 259
Jerwanska, E., G. Alderborn, E. Borjesson, J.M. Newton, C. Nystrom, Effect of water content on tensile fracture force ana deformability of ram extruded cylinders **149**, 131

Jerwanska, E., G. Alderborn, J.M. Newton, C. Nystrom, The effect of water content on the porosity and liquid saturation of extruded cylinders **121**, 65
Jerzewski, R.L., N.G. Lordi, Water vapor diffusion in model tablet systems. I. Design of a diffusion apparatus **101**, 35
Water vapor diffusion in model tablet systems. II. Experiments with lactose anhydrous based tablets **101**, 45
Jesus Esteban, M., see Canton, E. **92**, 47
Jetzer, W.E., Compaction characteristics of binary mixtures **31**, 201
Jetzer, W.E., W.B. Johnson, E.N. Hiestand, Comparison of two different experimental procedures for determining compaction parameters **26**, 329
Jeyanthi, R., Rao K. Panduranga, Preparation of gelatin microspheres of bleomycin **35**, 177
Release characteristics of bleomycin mitomycin C and 5-fluorouracil from gelatin microspheres **55**, 31
Jha, P., K. Murugesan, A. Farooq, Particle size and release of norethindrone acetate through silastic tubing: An in vitro and in vivo study **63**, 143
Jian, L., A.L.W. Po, Effect of angiotensin-converting enzyme (ACE) inhibitors on ciliary activity **107**, 77
Jian, L., A. Li Wan Po, Effect of xylometazoline and antazoline on ciliary beat frequency **86**, 59
Effects of insulin and nasal absorption enhancers on ciliary activity **95**, 101
Jiang, J., see Zhan, X. **115**, 167; **126**, 29
Jiang, T.S., see Huang, Y.Y. **129**, 41
Jideonwo, A., see Iwuagwu, M.A. **65**, 63
Jie, L., see Van, Schepdael **73**, 105
Jimenez-Castellanos Ballesteros, M.R., see Velasco Antequera, M.V. **103**, 155
Jimenez-Castellanos, M.R., H. Zia, C.T. Rhodes, Assessment of an in vitro method for measuring the bioadhesiveness of tablets **89**, 223
Design and testing in vitro of a bioadhesive and floating drug delivery system for oral application **105**, 65
Jimenez-Castellanos, M.R., see Ferrero, M.C. **149**, 233
Jimenez-Castellanos, M.R., see Monedero, Perales M. **132**, 183
Jimenez-Castellanos, M.R., see Munoz-Ruiz, A. **110**, 37
Jimenez-Castellanos, M.R., see Velasco, M.V. **136**, 107
Jimenez-Castellanos, R., see Ferrero, C. **147**, 11
Jimenez-Castellanos, R., see Munoz-Ruiz, A. **144**, 147
Jimenez-Duran, M., see Bustamante, P. **129**, 283
Jing-Ping, Z., see Ogiso, T. **102**, 141
Johansen, B.R., see Fisher, A.N. **74**, 147
Johansen, B.R., see Illum, L. **57**, 49; **63**, 207
Johansen, L., see Attwood, D. **9**, 285
Johansen, M., B. Mollgaard, P.K. Wotton et al., In vitro evaluation of dermal prodrug delivery—Transport and bioconversion of a series of aliphatic esters of metronidazole **32**, 199
Johansen, M., C. Larsen, Stability and kinetics of hydrolysis of metronidazole monosuccinate in aqueous solution and in plasma **21**, 201
A comparison of the chemical stability and the enzymatic hydrolysis of a series of aliphatic and aromatic ester derivatives of metronidazole **26**, 227
Macromolecular prodrugs. II. Influence of variation in molecular weight and degree of substitution of *O*-benzoyl dextran conjugates on their physicochemical properties and stability in aqueous buffer and in plasma **27**, 219
Johansen, M., H. Bundgaard, Pro-drugs as drug delivery systems. XIII. Kinetics of decomposition of N-Mannich bases of salicylamide and assessment of their suitability as possible pro-drugs for amines **7**, 119
Johansen, M., H. Bundgaard, E. Falch, Spectrophotometric determination of the rates of hydrolysis of aldehyde-releasing pro-drugs in aqueous solution and plasma **13**, 89
Johansen, M., see Bundgaard, H. **5**, 67; **7**, 129; **8**, 183; **9**, 7; **10**, 165; 181; **22**, 45
Johansen, M., see Harboe, E. **53**, 157
Johansen, M., see Larsen, C. **27**, 205; **35**, 39; **41**, 121
Johansen, P., see Gander, B. **129**, 51
Johansson, B., G. Alderborn, Degree of pellet deformation during compaction and its relationship to the tensile strength of tablets formed of microcrystalline cellulose pellets **132**, 207
Johansson, B., M. Wikberg, R. Ek, G. Alderborn, Compression behaviour and compactability of microcrystalline cellulose pellets in relationship to their pore structure and mechanical properties **117**, 57
Johansson, B., see Arwidsson, H. **76**, 91
Johansson, E., see Gottfries, J. **106**, 141
Johansson, M.E., Granular magnesium stearate as a lubricant in tablet formulations **21**, 307
Johansson, M.O., see Ragnarsson, G. **79**, 223
John, E.G., see Stead, J.A. **14**, 59
Johno, I., K. Kawakatsu, H. Kuwata, S. Kitazawa, Segmental difference in transmucosal fluid movement and its effect on gastrointestinal drug absorption in rabbits **25**, 255
Johnson, B.A., see Zografi, G. **22**, 159
Johnson, D., see Gu, L. **41**, 105
Johnson, D.M., R.A. Pritchard, W.F. Taylor et al., Degradation of the LH-RH analog nafarelin acetate in aqueous solution **31**, 125
Johnson, E., see Adjei, A. **61**, 135
Johnson, H.W., see Lovell, M.W. **109**, 45; **118**, 47
Johnson, J.L., J. Holinej, M.D. Williams, Influence of ionic strength on matrix integrity and drug release from hydroxypropyl cellulose compacts **90**, 151
Johnson, K., R. Hathaway, P. Leung, R. Franz, Effect of triacetin and polyethylene glycol 400 on some physical properties of hydroxypropyl methylcellulose free films **73**, 197
Johnson, K.C., see Hintz, R.J. **51**, 9
Johnson, M.C.R., see Chawla, A. **108**, 233
Johnson, O.L., C. Washington, S.S. Davis, Thermal stability of fluorocarbon emulsions that transport oxygen **59**, 131
Long-term stability studies of fluorocarbon oxygen transport emulsions **63**, 65

Johnson, O.L., C. Washington, S.S. Davis, K. Schaupp, The destabilization of parenteral feeding emulsions by heparin **53**, 237
Johnson, T.S., see Monnot, E.A. **60**, 41
Johnson, W.B., see Jetzer, W.E. **26**, 329
Johnston, T.P., H. Beris, Z.G. Wout, J.L. Kennedy, Effects on splenic, hepatic, hematological, and growth parameters following high-dose poloxamer 407 administration to rats **100**, 279
Johnston, T.P., J.A. Boyd, B.L. Ciesliga, F.J. Schoen, G. Amidon, R.J. Levy, Controlled release of ethanehydroxy diphosphonate from polyurethane reservoirs to inhibit calcification of bovine pericardium used in bioprosthetic heart valves **59**, 95
Johnston, T.P., E.L. Bove, S.F. Bolling, J.A. Boyd, B.L. Ciesliga, G.L. Amidon, F.J. Schoen, R.J. Levy, Controlled release of 1-hydroxyethylidene diphosphonate: In vitro assessment and effects on bioprosthetic calcification in sheep tricuspid valve replacements **52**, 139
Johnston, T.P., see Wang, P. **113**, 73
Johnston, T.P., see Wang, P.L. **96**, 41; **114**, 177
Jona, J.A., L.W. Dittert, P.A. Crooks, S.M. Milosovich, A.A. Hussain, Design of novel prodrugs for the enhancement of the transdermal penetration of indomethacin **123**, 127
Jones, B.D., see Russell, A.D. **25**, 105
Jones, B.E., see Thomas, M. **27**, 299; **71**, 147; **73**, 185; **83**, 139
Jones, D.S., Preliminary investigations concerning the anti-adherence properties of polyhexamethylenebiguanide (Vantocil IB™) **117**, 123
Jones, D.S., C.G. Adair, W.M. Mawhinney, S.P. Gorman, Standardisation and comparison of methods employed for microbial cell surface hydrophobicity and charge determination **131**, 83
Jones, D.S., S. Fowler, A preliminary report concerning the effects of Brolene® on the adherence of *Candida albicans* to human buccal epithelial cells and on hyphal development in vitro **105**, 71
Jones, D.S., S.P. Gorman, Factors influencing the activity of taurolidine against spores of *Bacillus subtilis* (NCTC 10073) **126**, 261
Jones, D.S., A.M. Loftus, S.P. Gorman, Physical factors affecting the sporicidal activity of chlorhexidine gluconate **119**, 247
Jones, D.S., N.J. Medlicott, Casting solvent controlled release of chlorhexidine from ethylcellulose films prepared by solvent evaporation **114**, 257
Jones, D.S., D.F. McCafferty, A.D. Woolfson, S.P. Gorman, A study of the stability of taurolidine in plasma and protein-free serum **64**, R1
Jones, D.S., K.J. Pearce, An investigation of the effects of some process variables on the microencapsulation of propranolol hydrochloride by the solvent evaporation method **118**, 199
Jones, D.S., K.J. Pearce, Contribution of process variables to the entrapment efficiency of propranolol hydrochloride within ethylcellulose microspheres prepared by the solvent evaporation method as evaluated using a factorial design **131**, 25
Jones, D.S., K.N. Shaffer, The effects of tobramycin, amiloride, sodium cholated and sodium deoxycholate on the growth inhibition activity of ciprofloxacin against a respiratory tract isolate of *Pseudomonas aeruginosa* **101**, R7
Jones, D.S., L.J. Schep, I.G. Tucker, M.G. Shepherd, An explanation of the quaternary ammonium compound-Eosin Y equilibria in the presence and absence of Triton X-100 **126**, 281
Jones, D.S., see Bunt, C.R. **99**, 93; **113**, 257
Jones, D.S., see Fowler, S. **84**, 77; **86**, 193
Jones, D.S., see Gorman, S.P. **65**, 127
Jones, D.S., see McGovern, J.G. **149**, 251
Jones, D.S., see Medlicott, N.J. **84**, 85; **143**, 25
Jones, D.S., see Schep, L.J. **105**, 39; **113**, 263
Jones, D.S., see Woolfson, A.D. **48**, 167; **49**, 135
Jones, H.P., R. Clements, D.J. Hearn, M.J. Gamlen, Development and application of a pharmacokinetic simulation program for oral controlled release dosage forms—DIPS **104**, 253
Jones, M.F., E. Crundwell, P.J. Taylor, The stability of some aryloxy prostaglandin analogues **4**, 1
Jones, N.B., see Atkins, P.J. **30**, 199
Jones, P.D.E., see Illum, L. **17**, 65
Jones, S.P., M.J. Greenway, N.A. Orr, The influence of receptor fluid on in vitro percutaneous penetration **53**, 43
Jones, S.P., G.D. Parr, Polarographic detection of β-cyclodextrin inclusion complexes **33**, 105
The acetotoluides as models for studying cyclodextrin inclusion complexes **36**, 223
Jones, T.M., see Armstrong, N.A. **48**, 173
Jonkman, J.H.G., J.P. Franke, R. Schoenmaker, R.A. de Zeeuw, Influence of sample preparation and storage on theophylline concentrations in biological fluids **10**, 177
Jonkman, J.H.G., R. Schoenmaker, N. Grimberg, R.A. de Zeeuw, A new in vitro dissolution test for controlled-release theophylline tablets **8**, 153
Jonkman, J.H.G., V.W. Steinijans, W. Beier, J.P. Yska, F.A. Kerkhof, Noord O.E. De, G. Grasmeyer, Sustained release properties of the once daily theophylline capsule Euphylong as compared with the Theo-Dur tablets **43**, 139
Jonkman, J.H.G., L.E. Van Bork, J. Wijsbeek et al., Bioavailability after rectal administration of thiazinamium methylsulphate in different vehicula **3**, 55
Jonkman, J.H.G., W. Van der Boon, L.P. Balant, J.Y. LeCotonnec, No effect of food on single dose bioavailability of sustained release theophylline (Sabidal); a comparison between Sabidal and choline theophyllinate solution **25**, 113
Jonkman, J.H.G., see Wiechers, J.W. **47**, 43; **66**, 53
Jonsson, B., see Westerberg, M. **28**, 23
Jonsson, U.E., see Abrahamsson, B. **140**, 229
Joosten, J.G.H., see Bodde, H.E. **26**, 57
Jordan, C.G.M., G.W. Davis, R.F. Timoney, Studies showing the effect of enzymes on the stability of ester prodrugs of propranolol and oxprenolol in biological samples **141**, 125

Jordan, C.G.M., J.M. Quigley, R.F. Timoney, Synthesis, hydrolysis kinetics and lipophilicity of *O*-acyl esters of oxprenolol **84**, 175
Jordan, C.G.M., see Quigley, J.M. **101**, 145
Jorge, J.S., see Cruz, M.E.M. **96**, 67
Jorgensen, A., see Hansen, L.B. **88**, 243
Jorgensen, H., see Vromans, H. **39**, 201
Jorgensen, K., F.N. Christensen, Shape modelling of dissolution profiles by non-integer kinetic orders **143**, 223
Jorgensen, K., L. Jacobsen, Factorial design used for ruggedness testing of flow through cell dissolution method by means of Weibull transformed drug release profiles **88**, 23
Jorgensen, K., see Bertelsen, F. **111**, 117
Jorgensen, L., E. Bechgaard, Intranasal absorption of angiopeptin: In vitro study of absorption and enzymatic degradation **99**, 165
Intranasal permeation of thyrotropin-releasing hormone: In vitro study of permeation and enzymatic degradation **107**, 231
Jorgensen, L., P. Artursson, E. Bechgaard, Toxicological and absorption enhancing effects of glycofurol 75 and sodium glycocholate in monolayers of human intestinal epithelial (Caco-2) cells **95**, 209
Jorgensen, L., R. Larsen, S. Catherpermal, E. Bechgaard, Intranasal absorption of different aqueous formulations of angiopeptin: In vivo bioavailability study **113**, 83
Jorgensen, L., see Bechgaard, E. **87**, 125; **89**, 147; **106**, 237
Jorgensen, L., see Bindseil, E. **119**, 37
Josefson, M., see Gottfries, J. **106**, 141
Joseph, I., S. Venkataram, Indomethacin sustained release from alginate-gelatin or pectin-gelatin coacervates **126**, 161
Joshi, H.N., M.A. Kral, E.M. Topp, Microwave drying of aqueous tablet film coatings: A study on free films **51**, 19
Joshi, H.N., E.M. Topp, Hydration in hyaluronic acid and its esters using differential scanning calorimetry **80**, 213
Joshi, Y.M., see Wesdyk, R. **65**, 69; **93**, 101
Joubin, H., see Watkinson, A.C. **121**, 27; **123**, 303
Jousma, H., H. Talsma, F. Spies et al., Characterization of liposomes. The influence of extrusion of multilamellar vesicles through polycarbonate membranes on particle size, particle size distribution and number of bilayers **35**, 263
Jousma, H., see Talsma, H. **37**, 171
Jouyban-Gharamaleki, A., see Barzegar-Jalali, M. **140**, 237; **144**, 127
Jovanovic, M., R. Rakic, Z. Djuric, L. Zivanovic, Effect of surfactants on the rheological and electrical properties of carboxymethylcellulose aqueous solution **75**, 155
Jovanovic, M., see Djuric, M. **37**, 179
Jovanovic, M., see Kulvanich, P. **39**, 51
Joyeux, H., see Boulenc, X. **123**, 13, 71
Joyeux, H., see Caro, I. **116**, 147
Jozan, M., see Takacs-Novak, K. **79**, 89; **113**, 47
Jozwiak, Z., see Szwarocka, A. **135**, 167
Jozwiakowski, M.J., S.O. Williams, R.D. Hathaway, Relative physical stability of the solid forms of amiloride HCl **91**, 195
Ju, T.R., see Sung, K.C. **142**, 53
Judy, C.L., Jr. W.E. Acree, Solubility in binary solvent systems. V. Monomer and dimer models for the solubility of *p*-tolylacetic acid in systems of non-specific interactions **27**, 39
Judy, C.L., see Acree, Jr. W.E. **31**, 225
Juffermans, J., see Van der Griend, J. **1**, 257
Juhasz, J., V. Lenaerts, M. Bellemare, C. Pimienta, H. Ong, Stability and adsorption to polymeric surfaces of rat ANF in poloxamer 407 solutions **77**, 309
Julian, T.N., see Radebaugh, G.W. **45**, 39
Jun, H.W., J.W. Lai, Preparation and in vitro dissolution tests of egg albumin microcapsules of nitrofurantoin **16**, 65
Jun, H.W., see Suh, H. **129**, 13
Juneskans, O.T., see Karabit, M.S. **54**, 51; **56**, 169
Jung, B.H., see Shim, C.K. **82**, 233; **83**, 257
Jung, D., S. Oie, Decreased in vivo acetylation of sulfisoxazole in the rabbit in the presence of 4-*N*-acetyl-sulfisoxazole **7**, 179
Jung, D., see Gordon, M.S. **115**, 29
Jung, H.C., see Lopez, A.A. **28**, 167
Junginger, H.E., see Akiyama, Y. **136**, 155; **138**, 13
Junginger, H.E., see Baggerman, C. **27**, 17
Junginger, H.E., see Blok, D. **73**, 171
Junginger, H.E., see de Haan, F.H.N **56**, 75
Junginger, H.E., see Lehr, C.M. **70**, 235; **78**, 43
Junginger, H.E., see Luessen, H.L. **141**, 39
Junginger, H.E., see Tiemessen, H.L.G.M. **53**, 119; **56**, 87
Junginger, H.E., see Verhoeven, J. **45**, 65
Juni, K., see Kawaguchi, T. **58**, R1; **77**, 71
Juni, K., see Nakano, M. **4**, 291
Juni, K., see Natsume, H. **58**, 79
Juni, K., see Seki, T. **57**, 73
Juni, K., see Tamura, M. **131**, 263
Juppo, A.M., Relationship between breaking force and pore structure of lactose, glucose and mannitol tablets **127**, 95
Porosity parameters of lactose, glucose and mannitol tablets obtained by mercury porosimetry **129**, 1
Change in porosity parameters of lactose, glucose and mannitol granules caused by low compression force **130**, 149
Juppo, A.M., J. Yliruusi, L. Kervinen, P. Strom, Determination of size distribution of lactose, glucose and mannitol granules by sieve analysis and laser diffractometry **88**, 141
Juretic, D., I. Cepelak, V. Jalsenjak, T. Zanic-Grubisic, K. Lipovac, I. Jalsenjak, Effect of microencapsulated acetylsalicylic acid on glycosylation of human serum proteins in vitro **61**, 219
Jurima-Romet, M., R.F. Barber, J. Demeester, P.N. Shek, Distribution studies of liposome-encapsulated glutathione administered to the lung **63**, 227
Jurima-Romet, M., R.F. Barber, P.N. Shek, Liposomes and bronchoalveolar lavage fluid: Release of vesicle-entrapped glutathione **88**, 201
Jurjenson, H., see Sirkia, T. **107**, 179
Jurjenson, H., see Veski, P. **111**, 171
Jurkschat, T., see Furst, W. **61**, 43
Jusko, W.J., see Gardner, M.J. **33**, 55

Juslin, L., O. Antikainen, P. Merkku, J. Yliruusi, Droplet size measurement. I. Effect of three independent variables on droplet size distribution and spray angle from a pneumatic nozzle **123**, 247
Droplet size measurement. II. Effect of three independent variables on parameters describing the droplet size distribution from a pneumatic nozzle studied by multilinear stepwise regression analysis **123**, 257
Juslin, M., see Urtti, A. **25**, 165
Justel, A., see Del Estal, J.L. **102**, 257
Justo, A., see Munoz-Ruiz, A. **144**, 147

K

Kadir, F., C.B.J. Seijsener, J. Zuideman, Influence of the injection volume on the release pattern of intramuscularly administered propranolol to rats **81**, 193
Kadir, F., J. Zuidema, A. Pijpers, A. Vulto, J.H.M. Verheijden, Drug lipophilicity and release pattern of some β-blocking agents after intra-adipose injection in pigs **64**, 171
Kadir, F., see Zuidema, J. **105**, 189
Kadir, R., B.W. Barry, α-Bisabolol, a possible safe penetration enhancer for dermal and transdermal therapeutics **70**, 87
Kadir, R., B.W. Barry, J.E. Fairbrother, D.A. Hollingsbee, Delivery of triamcinolone acetonide through human epidermis: Effect of Actiderm, a new hydrocolloid dermatological patch **60**, 139
Kadir, S., T. Murakami, Y. Higashi, N. Yata, Gastrointestinal absorption of griseofulvin from liquid organic acids and esters in rats **33**, 235
Kadir, S., C. Nitta, I. Koga, T. Murakami, Y. Higashi, N. Yata, Possible factors behind the enhanced gastrointestinal absorption of griseofulvin from liquid organic acid ester solutions in rats **40**, 257
Kaewpila, S., see Dam, A.M. **121**, 245
Kagayama, A., R. Mustafa, E. Akaho et al., Mechanism of diffusion of compounds through ethylene vinyl acetate copolymers. I. Kinetics of diffusion of 1-chloro-4-nitrobenzene, 3,4-dimethylphenol and 4-hexylresorcinol **18**, 247
Kagayama, A., see Hata, T. **110**, 1
Kahela, P., R. Aaltonen, E. Lewing et al., Pharmacokinetics and dissolution of two crystalline forms of carbamazepine **14**, 103
Kahela, P., see Farraj, N.F. **38**, 39
Kahela, P., see Laine, E. **20**, 307; **43**, 241
Kahkonen, K., see Malkki-Laine, L. **117**, 189
Kahns, A.H., H. Bundgaard, Prodrugs as drug delivery systems. 107. Synthesis and chemical and enzymatic hydrolysis kinetics of various mono- and diester prodrugs of *N*-acetylcysteine **62**, 193
N-acyl derivatives as prodrug forms for amides: Chemical stability and enzymatic hydrolysis of various *N*-acyl and *N*-alkoxycarbonyl amide derivative **71**, 31
Prodrugs of peptides. 14. Bioreversible derivatization of the tyrosine phenol group to effect protection of tyrosyl peptides against α-chymotrypsin **76**, 99
Facile α-chymotrypsin-catalyzed degradation of the HIV inhibitor [D-Ala^1]-peptide T amide **77**, 65
Kahns, A.H., J. Moss, H. Bundgaard, Improved oral bioavailability of salicylamide in rabbits by a 1,3-benzoxazine-2,4-dione prodrug **78**, 199
Kaji, Y., K. Uekama, H. Yoshikawa et al., Selective transfer of 1-hexylcarbamoyl-5-fluorouracil into lymphatics by combination of β-cyclodextrin polymer complexation and absorption promoter in the rat **24**, 79
Kajii, H., T. Horie, M. Hayashi, S. Awazu, Effects of sodium salicylate and caprylate as adjuvants of drug absorption on isolated rat small intestinal epithelial cells **33**, 253
Kajimoto, H., see Wong, J. **144**, 177
Kajiyama, A., see Sako, K. **137**, 225
Kaka, I., see Shah, J.C. **109**, 283; **112**, 263
Kaka, J.S., K.I. Al-Khamis, Effect of domperidone on acetyl salicylic acid and acetaminophen absorption in rabbits **28**, 133
Kakee, A., H. Yamane, T. Nanayama, T. Hibi, K. Sugimori, T. Handa, K. Miyajima, Structure and tumor necrosis factor-inducing activity of dispersed particles of a lipid A analogue, GLA-60, and phosphatidylcholine mixture **111**, 15
Kakemi, M., see Morimoto, K. **113**, 1
Kakeya, N., see Yoshimura, Y. **17**, 47; **26**, 317
Kalala, W., see Samyn, C. **121**, 211
Kalala, W., R. Kinget, G. Van den Mooter, C. Samyn, Colonic drug-targeting: In vitro release of ibuprofen from capsules coated with poly(ether–ester) azopolymers **139**, 187
Kalcic, I., B. Zorc, I. Butula, Macromolecular prodrugs. VII. Polymer–dopamine conjugates **136**, 31
Kalcic, I., see Zorc, B. **123**, 65
Kale, N.J., Jr. L.V. Allen, Studies on microemulsions using Brij 96 as surfactant and glycerin, ethylene glycol and propylene glycol as cosurfactants **57**, 87
Kalinkova, G.N., I. Hristov St., L. Petrov, Comparative studies of different modifications of calcium valproate **115**, 245
Kalinkova, G.N., S. Stoeva, Polymorphism of azlocillin sodium **135**, 111
Kaliszan, R., M. Markuszewski, Brain/blood distribution described by a combination of partition coefficient and molecular mass **145**, 9
Kaluzny, B.D., see Snider, B.G. **66**, 63
Kamada, A., see Nishihata, T. **27**, 245; **33**, 89, 181; **38**, 83; **42**, 251
Kamali, F., see Wilson, C.G. **50**, 155
Kamani, A., see Komiya, I. **4**, 249
Kamide, S., see Nishihata, T. **42**, 251
Kaminski, J.J., N. Bodor, Determination of penicillins and cephalosporins using 3-bromo-4,4-dimethyl-2-oxazolidinone **3**, 151
Kaminski, J.J., see Bodor, N. **1**, 189
Kamiya, E., K. Morimoto, T. Takeeda et al., Rectal absorption or flurbiprofen, ketoprofen and indomethacin in polyacrylic acid equeous gel preparation in rats **17**, 273
Kamiya, E., see Morimoto, K. **14**, 149
Kamiya, H., K. Morimoto, K. Morisaka, Dissolution behavior and bioavailability of cimetidine-HCl (cimetidine monohydrochloride monohydrate) **26**, 197

Kan, F.W.K., see Chouinard, F. **72**, 211
Kanaya, Y., P. Spooner, J.L. Fox et al., Mechanistic studies on the bioavailability of calcium fluoride for remineralization of dental enamel **16**, 171
Kanaya, Y., see Spooner, P.J. **15**, 177
Kaneniwa, N., T. Funaki, S. Furuta, N. Watari, Application of the Pidgeon–Pitlick method for calculation of the absorption rate constant of drugs whose disposition can be described by the two-compartment open model **31**, 33
Kaneniwa, N., M. Hiura, T. Funaki, The absorption kinetics of barbiturates in rabbits **26**, 157
Kaneniwa, N., N. Watari, T. Funaki, Use of fluorescence HPLC for determining extrarenal creatinine clearance in rats **34**, 231
Kaneniwa, N., see Funaki, T. **31**, 119; **43**, 59
Kaneniwa, N., see Hiura, M. **18**, 185; **20**, 73
Kaneniwa, N., see Otsuka, M. **62**, 65
Kaneniwa, N., see Watari, N. **6**, 155; **7**, 307; **21**, 85
Kaneo, Y., T. Tanaka, Y. Fujihara, H. Mori, S. Iguchi, Delivery of glutathione, as a dextran conjugate, into the liver **44**, 265
Kaneo, Y., Y. Fuijihara, T. Tanaka, K. Ogawa, K. Fujita, S. Iguchi, Preparation and characterization of a soluble glutathione-dextran conjugate **57**, 263
Kanfer, I., see Chattaraj, S.C. **120**, 119; **125**, 215; **133**, 59
Kanfer, I., see Dowse, R. **39**, 141
Kanfer, I., see Haigh, J.M. **19**, 245
Kanfer, I., see Meyer, E. **41**, 63
Kanfer, I., see Skinner, M. **88**, 151
Kanfer, I., see Smith, E.W. **27**, 185
Kanfer, I., see Stubbs, C. **63**, 113
Kanfer, I., see Tandt, L.A.G.L. **104**, 11
Kanfer, I., see Terepolsky, S.A. **115**, 123
Kang, J.J., see Chen, C.Y. **141**, 171
Kang, T.S., see Ha, I.S. **144**, 91
Kaniwa, N., see Ogata, H. **23**, 277, 289; **29**, 113, 121
Kanjia, J., see Pandit, N.K. **122**, 27; **141**, 197
Kannikoski, A., see Marvola, M. **53**, 145
Kante, B., P. Couvreur, V. Lenaerts et al., Tissue distribution of [^{3}H]actinomycin D adsorbed on polybutylcyanoacrylate nanoparticles **7**, 45
Kao, C.C., see Liu, C.H. **100**, 241
Kao, C.C., see Sheu, M.T. **85**, 57
Kao, Y.H., see Lin, S.Y. **56**, 249
Kao, Y.H., see Liu, C.H. **100**, 241
Kaplan, J.E., see Schreier, H. **37**, 233
Karabit, M.S., Studies on the evaluation of preservative efficacy. V. Effect of concentration of micro-organisms on the antimicrobial activity of phenol **60**, 147
Karabit, M.S., O.T. Juneskans, P. Lundgren, Studies on the evaluation of preservative efficacy. IV. The determination of antimicrobial characteristics of some pharmaceutical compounds in aqueous solutions **54**, 51
Factorial designs in the evaluation of preservative efficacy **56**, 169
Karami, K., see Sjoberg, H. **141**, 63
Karara, A.H., see Chaudhuri, B. **42**, 117; **65**, 57
Karara, A.H., see Chow, D.D. **28**, 95
Kararli, T.T., B. Farhadich, S. Bittner, M. Babler, P.C. Yang, G.M. Walsh, Role of intestinal transport and first pass liver extraction on oral delivery of renin inhibitor compounds **102**, 177
Karasulu, Y., see Ertan, G. **96**, 243
Karau, C., M. Petszulat, P.C. Schmidt, Preparation and stability of interferon-α-containing liposomes **128**, 89
Karba, R., see Kmetec, V. **21**, 211
Karba, R., see Kristl, A. **100**, 33
Karba, R., see Mrhar, A. **62**, 15
Karba, R., see Srcic, S. **73**, 221
Karehill, P.G., M. Glazer, C. Nystrom, Studies on direct compression of tablets. XXIII. The importance of surface roughness for the compactability of some directly compressible materials with different bonding and volume reduction properties **64**, 35
Karehill, P.G., C. Nystrom, Studies on direct compression of tablets. XXI. Investigations of bonding mechanisms of some directly compressed materials by strenght characterization in media with different dielectric constants (relative permittivity) **61**, 251
Studies on direct compression of tablets. XXII. Investigation of strength increase upon ageing and bonding mechanisms for some plastically deforming materials **64**, 27
Karidas, T., see Malamataris, S. **103**, 205; **104**, 115
Karim, A., see Cook, C.S. **60**, 125
Karim, E.F.I.A., J.S. Millership, D.J. Temple, A.D. Woolfson, The influence of diet on drug metabolism studies of S-carboxymethyl-L-cysteine **52**, 155
Karim, E.I.A., see Ibrahim, K.E. **98**, 231
Karjalainen, P., see Vidgren, M.T. **37**, 239; **39**, 101, 107; **42**, 211
Karkkainen, A., see Vidgren, M.T. **37**, 239; **39**, 107; **42**, 211
Karlsen, J., see Beezer, A.E. **38**, 251
Karlsen, J., see Henriksen, I. **98**, 181; **101**, 227; **119**, 231; **145**, 231; **146**, 193
Karlsen, J., see Kristensen, S. **100**, 15
Karlsen, J., see Nord, K. **72**, 11
Karlsen, J., see Sande, S.A. **55**, 193; **73**, 147; **98**, 209
Karlsen, J., see Tonnesen, H.H. **41**, 75; **43**, 215; **90**, 221
Karlsson, J., P. Artursson, A method for the determination of cellular permeability coefficients and aqueous boundary layer thickness in monolayers of intestinal epithelial (Caco-2) cells grown in permeable filter chambers **71**, 55
Karlsson, J., A. Wikman, P. Artursson, The mucus layer as a barrier to drug absorption in monolayers of human intestinal epithelial HT29-H goblet cells **99**, 209
Karlsson, S., see Singh, S.K. **142**, 199
Karnachi, A.A., M.A. Khan, Box-behnken design for the optimization of formulation variables of indomethacin coprecipitates with polymer mixtures **131**, 9
Karnik, N.A., see Moelands, D. **86**, 263
Kas, H.S., see Ciftci, K. **131**, 73
Kasama, Y., see Terasaki, T. **81**, 143
Kashihara, T., see Hirai, S. **15**, 261
Kashihara, T., see Hussain, A. **21**, 233

Kasim, N.A., see Hammounda, Y.E. **89**, 111
Kassem, M., M. Akbarieh, R. Tawashi, Effect of fluid viscosity on the size and morphic features of salicylic acid during ball-mill grinding **47**, 179
Kassem, M., see El-Ridy, M. **56**, 23
Kassem, M.A., M.A. Attia, F.S. Habib, A.A. Mohamed, In vitro release study of β-methasone and phenylephrine hydrochloride from ophthalmic preparations **29**, 243
Activity of ophthalmic gels of β-methasone and phenylephrine hydrochloride in the rabbit's eye **32**, 47
Kassem, M.A., A.G. Mattha, A.E.M. El-Nimr, S.M. Omar, Study of the influence of sodium taurocholate (SIC) and sodium glycocholate (SGC) on the mass transfer of certain drugs. Digoxin **12**, 1
Kassem, M.A., see Attia, M.A. **47**, 21
Kassem, M.A., see Mattha, A.G. **11**, 27
Katagi, T., see Takahashi, K. **49**, 179
Katagiri, Y., see Nagasako, S. **81**, 127
Katakai, R., see Miyajima, M. **95**, 153
Katakam, M., see Banga, A.K. **116**, 211
Katayama, K., see Hatanaka, T. **125**, 195
Katayama, K., see Panomsuk, S.P. **126**, 147
Katayama, N., T. Houjou, K. Takada, Lymphatic transport of cyclosporin A from the abdominal cavity **110**, 155
Katayama, N., R. Tanaka, Y. Ohno, C. Ueda, T. Houjou, K. Takada, Implantable slow release cyclosporin A (CYA) delivery system to thoracic lymph duct **115**, 87
Katikaneni, P.R., S.M. Upadrashta, S.H. Neau, A.K. Mitra, Ethylcellulose matrix controlled release tablets of a water-soluble drug **123**, 119
Katikaneni, P.R., S.M. Upadrashta, C.E. Rowlings, S.H. Neau, G.A. Hileman, Consolidation of ethylcellulose: Effect of particle size, press speed, and lubricants **117**, 13
Katikaneni, P.R., see Upadrashta, S.M. **112**, 173
Kato, C., see Yoshikawa, H. **8**, 291
Kato, S., see Kurono, Y. **81**, 179
Kato, T., see Ishikura, T. **116**, 51
Kato, T., see Yamauchi, H. **113**, 141
Katsu, T., M. Kuroko, K. Sanchika, T. Morikawa, Y. Kurosaki, T. Nakayama, T. Kimura, Y. Fujita, Interaction of 1-dodecylazacycloheptan-2-one (Azone) with erythrocyte membrane **53**, 61
Kaufman, M.J., Rate and equilibrium constants for acid-catalyzed lactone hydrolysis of HMG-CoA reductase inhibitors **66**, 97
Kaufmann, B.G., see Moolenaar, F. **33**, 249
Kaul, D., N.T. Nguyen, S. Venkataram, Crystal habit modifications and altered tabletting characteristics **88**, 345
Kaur, R., see Bryan, A.J. **7**, 145
Kaus, L.C., J.T. Fell, H. Sharma, D.C. Taylor, Gastric emptying and intestinal transit of non-disintegrating capsules—the influence of metoclopramide **22**, 99
Kaus, L.C., J.T.H. Fell Sharma, D.C. Taylor, On the intestinal transit of a single non-disintegrating object **20**, 315
Kawabe, S., see Kim, S. **21**, 179
Kawabe, S., see Nishihata, T. **22**, 147
Kawaguchi, T., T. Hasegawa, K. Juni, T. Seki, Rectal absorption of Zidovudine **77**, 71
Kawaguchi, T., T. Hasegawa, T. Seki, K. Juni, M. Saneyoshi, Ester prodrugs of 2′,3′-dideoxy-2′,3′-didehydrothymidine (D4T) **58**, R1
Kawaguchi, T., see Seki, T. **57**, 73
Kawaguchi, T., see Tamura, M. **131**, 263
Kawahara, M., see Muhi-Eldeen, Z. **26**, 15
Kawahara, Y., see Seta, Y. **41**, 245, 255, 263
Kawaharada, U., see Asano, M. **67**, 67
Kawakami, U., see Tashima, T. **42**, 61
Kawakatsu, K., see Johno, I. **25**, 255
Kawaratani, T., see Yamamoto, A. **39**, 255; **51**, 137
Kawashima, H., see Hiura, M. **18**, 185; **20**, 73
Kawashima, H., see Naito, S.I. **17**, 109
Kawashima, S., see Murata, Y. **96**, 139
Kawashima, Y., F. Cui, H. Takeuchi, T. Niwa, T. Hino, K. Kiuchi, Parameters determining the agglomeration behaviour and the micromeritic properties of spherically agglomerated crystals prepared by the spherical crystallization-technique with miscible solvent systems **119**, 139
Kawashima, Y., H. Takeuchi, T. Hino, T. Niwa, T.L. Lin, F. Sekigawa, M. Ohya, Preparation of prolonged-release matrix tablet of acetaminophen with pulverized low-substituted hydroxypropylcellulose via wt granulation **99**, 229
Kawashima, Y., S.Y. Lin, M. Ueda, H. Takenaka, Preparation of directly compressible powders of a physical mixture and a complex of theophylline-phenobarbital using spray-drying **18**, 335
Kawashima, Y., T. Hino, H. Takeuchi, T. Niwa, K. Horibe, Rheological study of w/o/w emulsion by a cone-and-plate viscometer: Negative thixotropy and shear-induced phase inversion **72**, 65
Kawashima, Y., T. Iwamoto, T. Niwa, H. Takeuchi, T. Hino, Uniform and improved bioavailability of newly developed rapid and sustained release suspensions of ibuprofen microspheres **89**, 9
Kawashima, Y., T. Iwamoto, T. Niwa, H. Takeuchi, Y. Itoh, Preparation and characterization of a new controlled release ibuprofen suspension for improving suspendability **75**, 25
Kawashima, Y., see Morishima, K. **105**, 11
Kawashima, Y., see Murakami, H. **149**, 43
Kawashima, Y., see Niwa, T. **121**, 45
Kawashima, Y., see Ohwaki, T. **85**, 19; **93**, 61
Kawashima, Y., see Schade, A. **117**, 209
Kawashima, Y., see Takeuchi, H. **86**, 25
Kawnopparat, N., see Tantishaiyakul, V. **143**, 59
Kayano, M., see Aoki, S. **85**, 65
Kayes, J.B., see Law, S.L. **15**, 251
Kayes, J.B., see Rawlins, D.A. **13**, 145, 159, 171
Kazama, H., see Wong, J. **144**, 177
Kearney, A., see Hussain, M.A. **36**, 127
Kearney, A.S., D.L. Gabriel, S.C. Mehta, G.W. Radebaugh, Effect of polyvinylpyrrolidone on the crystallinity and dissolution rate of solid dispersions of the antiinflammatory CI-987 **104**, 169

Kearney, A.S., S.C. Mehta, C. Mehta, G.W. Radebaugh, The effect of structural changes on the intramolecular degradation of the 'dipeptoid' CI-988 **92**, 63
Kearney, A.S., S.C. Mehta, G.W. Radebaugh, The effect of cyclodextrins on the rate of intramolecular lactamization of gabapentin in aqueous solution **78**, 25
Preformulation studies to aid in the development of an injectable formulation of PD 144872, a radiosensitizing anticancer agent **102**, 63
Kearney, A.S., K. Patel, N.R. Palepu, Preformulation studies to aid in the development of a ready-to-use injectable solution of the antitumor agent, topotecan **127**, 229
Kearney, M., see Niven, R.W. **101**, 81
Kearney, P., see Jamshaid, M. **48**, 125
Kearney, P., C. Marriott, The effects of mucus glycoproteins on the bioavailability of tetracycline. I. Dissolution rate **28**, 33
The effects of mucus glycoproteins on the bioavailability of tetracycline. II. Binding **35**, 211
Kearney, P., M.J. Myers, The effects of mucus glycoproteins on the bioavailbility of tetracycline. III. Everted gut studies **38**, 211
Keates, A.C., see Nicklin, P. **128**, 155
Keating, M., see Barnwell, S.G. **88**, 423
Kedor, E.R.M., see Bentley, M.V.L.B. **146**, 255
Kedzierewicz, F., M. Hoffman, P. Maincent, Comparison of tolbutamide β-cyclodextrin inclusion compounds and solid dispersions. Physicochemical characteristics and dissolution studies **58**, 221
Kedzierewicz, F., F. Villieras, C. Zinutti, M. Hoffman, P. Maincent, A 3 year stability study of tolbutamide solid dispersions and β-cyclodextrin complex **117**, 247
Kedzierewicz, F., C. Zinutti, M. Hoffmann, P. Maincent, Bioavailability study of tolbutamide β-cyclodextrin inclusion compounds, solid dispersions and bulk powder **94**, 69
Kedzierewicz, F., see Veiga, F. **129**, 63
Kedzierewicz, F., see Zinutti, C. **133**, 97
Keigami, M., see Nishihata, T. **42**, 251
Keil, B., see Grunenberg, A. **118**, 11
Keipert, S., J. Fedder, A. Bohm, B. Hanke, Interactions between cyclodextrins and pilocarpine—as an example of a hydrophilic drug **142**, 153
Keirns, J.J., see Matzek, K.M. **28**, 85, 151
Keiser, D.R., see Hansen, L.D. **135**, 31
Kellaway, I.W., see Ahmed, M. **12**, 219; **13**, 227
Kellaway, I.W., see Arrowsmith, M. **14**, 191; **16**, 305; **17**, 91; **20**, 347
Kellaway, I.W., see Cabral-Marques H. **63**, 259, 267; **77**, 297, 303
Kellaway, I.W., see Davies, N.M. **105**, 97
Kellaway, I.W., see Dennis, A.B. **65**, 85
Kellaway, I.W., see Dickinson, P.A. **148**, 55
Kellaway, I.W., see Donnelly, A. **135**, 191
Kellaway, I.W., see Durrani, A.M. **88**, 409; **118**, 243
Kellaway, I.W., see Farr, S.J. **26**, 303; **51**, 39
Kellaway, I.W., see Garcia-Gonzalez, N. **100**, 25, 65; **104**, 107
Kellaway, I.W., see Gay, C.L. **49**, 39
Kellaway, I.W., see Jamshaid, M. **48**, 125
Kellaway, I.W., see Khan, Z.U. **55**, 129
Kellaway, I.W., see Saket, M.M. **21**, 155; **27**, 287
Kellaway, I.W., see Smart, J.D. **53**, 79
Kellaway, I.W., see Taylor, K.M.G. **58**, 49, 57
Kellaway, I.W., see Thomas, M. **27**, 299; **71**, 147; **73**, 185; **83**, 139
Kellaway, I.W., see Woolfrey, S.G. **26**, 35
Kellaway, I.W., N.M. Najib, The adsorption of hydrophilic polymers at the polystyrene–water interface **6**, 285
The effect of hydrophilic polymers on the electrophoretic mobility of suspended particles **7**, 285
Hydrophilic polymers as stabilisers and flocculants of sulphadimidine suspensions **9**, 59
Kelly, T.P., see Duddu, S.P. **114**, 247
Kempers, Y.A.G., see Bekers, O. **68**, 271
Kendall, A.T., E. Stark, J.K. Sugden, Effect of hydroxyl radicals on the stability of metronidazole in buffer solution at pH 9.2 **57**, 217
Kenley, R.A., see Chen, S.H. **72**, 89
Kenley, R.A., see Hayward, D.S. **59**, 245
Kenley, R.A., see Powell, M.F. **44**, 225
Kennedy, J.L., see Johnston, T.P. **100**, 279
Kennedy, P.E., C.M. Riley, V.J. Stella, Degradation of the antineoplastic drug, clomesone (2-chloroethyl(methylsulfonyl)methanesulfonate, NSC-338947). A kinetic and mechanistic study **48**, 179
Kennedy, R.A., see Padmadisastra, Y. **104**, R1; **112**, 55
Kennedy, T.A., see McElnay, J.C. **40**, 105
Kennerley, J., see Wilson, C.G. **40**, 119
Kennerley, J.W., see Daly, P.B. **18**, 201
Kennerley, J.W., see Wilson, C.G. **18**, 1
Kent, J.S., E.J. Mroszczak, R.L. Roe, R. Runkel, Bioavailability and in vitro dissolution of tiopinac from solution and capsule formulations **7**, 245
Kenyon, C.J., E.T. Cole, I.R. Wilding, The effect of food on the in vivo behaviour of enteric coated starch capsules **112**, 207
Kenyon, C.J., see Dewsbury, N.J. **137**, 261
Kerc, J., S. Srcic, B. Kofler, J. Smid-Korbar, Molar solubility of felodipine in different aqueous systems **81**, R1
Kerc, J., S. Srcic, M. Mohar, J. Smid-Korbar, Some physicochemical properties of glassy felodipine **68**, 25
Kerc, J., see Gasperlin, M. **107**, 51
Kerc, J., see Srcic, S. **87**, 1
Kerc, J., see Valenta, C. **148**, 131
Kerkdijk, H.G., see Beijnen, J.H. **41**, 169
Kerkhof, F.A., see Jonkman, J.H.G. **43**, 139
Kern, E.R., see Gonsho, A. **65**, 183
Kern, E.R., see Lee, P.H. **93**, 139
Kerr, B., see McElnay, J.C. **33**, 195
Kervinen, L., J. Yliruusi, Modelling S-shaped dissolution curves **92**, 115
Kervinen, L., see Juppo, A.M. **88**, 141

Kesselring, U.W., see Huu-Phuoc, N. **34**, 217
Kesselring, U.W., see Ruelle, P. **87**, 47; **104**, 125
Kessler, R., see Goldbach, P. **123**, 33
Ketolainen, J., J. Ilkka, P. Paronen, Temperature changes during tabletting measured using infrared thermoviewer **92**, 157
Ketolainen, J., M. Oksanen, J. Rantala, J. Stor-Pellinen, M. Luukkala, P. Paronen, Photoacoustic evaluation of elasticity and integrity of pharmaceutical tablets **125**, 45
Ketolainen, J., see Pesonen, T. **57**, 139
Ketolainen, J., see Pirttimaki, J. **95**, 93
Ketolainen, J., see Silvennoinen, R. **131**, 209
ketolainen, P., see Silvennoinen, R. **131**, 209
Kettner, C., see Hussain, M.A. **117**, 181
Keyhani, K., see Waldrep, J.C. **97**, 205
Khalafallah, N., see Ismail, F.A. **34**, 189
Khalfi, F., T. Dine, B. Gressier, M. Luyckx, C. Brunet, L. Ballester, F. Goudaliez, M. Cazin, J.C. Cazin, Compatibility and stability of vancomycin hydrochloride with PVC infusion material in various conditions using stability-indicating high-performance liquid chromatographic assay **139**, 243
Khalil, S.A., Khordagui El, A.M. Saleh, Interaction of caffeine with phenothiazine derivatives **16**, 271
Khalil, S.A., V.F. Naggar, I.A. Zaghloul, A.A. Ismail, In vitro anticoagulant–antacid interactions **19**, 307
Khalil, S.A., see Badwan, A.A. **13**, 67
Khalil, S.A., see El-Khordagui, L.K. **7**, 111
Khalil, S.A., see Ismail, F.A. **34**, 189
Khalil, S.A., see Saleh, A.M. **5**, 161
Khalil, S.A.H., L.K. El-Khordagui, Z.A. El-Gholmy, Effect of antacids on oral absorption of rifampicin **20**, 99
Khalil, S.A.H., L.M. Mortada, M. El-Khawas, The uptake of ampicillin and amoxycillin by some adsorbents **18**, 157
Decreased bioavailability of ampicillin and amoxycillin in presence of kaolin **19**, 233
Khalil, S.A.H., see Abd El Fattah, S. **18**, 225
Khalil, S.A.H., see Ali, H.M. **9**, 233
Khan, I., see Zahirul, M. **140**, 131
Khan, K., see Paesen, J. **113**, 215
Khan, K.A., see Sujja-Areevath, J. **139**, 53
Khan, M.A., S. Bolton, M.S. Kislalioglu, Optimization of process variables for the preparation of ibuprofen coprecipitates with Eudragit S100 **102**, 185
Khan, M.A., see Davies, M.C. **57**, 183
Khan, M.A., see Illum, L. **30**, 17
Khan, M.A., see Karnachi, A.A. **131**, 9
Khan, M.A., see Singh, S.K. **125**, 243; **141**, 179
Khan, M.Z.I., I.G. Tucker, J.P. Opdebeeck, Cholesterol and lecithin implants for sustained release of antigen: Release and erosion in vitro, and antibody response in mice **76**, 161
Evaluation of cholesterol-lecithin implants for sustained delivery of antigen: Release in vivo and single-step immunisation of mice **90**, 255
Khan, R., see Pereswetoff-Morath, L. **128**, 9
Khan, R.M.A., see Najjar, T.A. **98**, 51
Khan, Z.U., I.W. Kellaway, Differential scanning calorimetry of dimethylsulphoxide-treated human stratum corneum **55**, 129
Khankari, R.K., D. Law, D.J.W. Grant, Determination of water content in pharmaceutical hydrates by differential scanning calorimetry **82**, 117
Khanna, R., S.P. Agarwal, A. Ahuja, Preparation and evaluation of bioerodible buccal tablets containing clotrimazole **138**, 67
Khar, R.K., see Jaggi, M. **69**, 77
Khatib, K., see Billany, M.R. **137**, 143
Khazainia, T., see Jamali, F. **9**, 221
Khetarpal, V.K., see Sheen, P.C. **118**, 221
Khidr, S.H., see Al-Angary, A.A. **65**, R5; **114**, 221; **128**, 163
Khidr, S.H., see El-Sayed, Y.M. **123**, 113
Khor, S.P., H.J. Wu, H. Boxenbaum, Simultaneous correction for volume shifts and protein leakage in equilibrium dialysis protein binding experiments **23**, 109
Fluid shift and protein leakage corrections in protein binding equilibrium dialysis experiments **30**, 189
Khosla, R., S.S. Davis, Gastric emptying and small and large bowel transit of non-disintegrating tablets in fasted subjects **52**, 1
Khosla, R., see Davis, S.S. **35**, 253; **60**, 191
Khosla, R., see Hardy, J.G. **48**, 79
Khotz, M.S., see Vervaet, C. **106**, 157
Kido, M., see Nakamura, J. **87**, 59; **99**, 189
Kiekens, A. Vermeire, N. Samyn, J. Demeester, J.P. Remon, Optimisation of electrical conductance measurements for the quantification and prediction of phase separation in o/w-emulsions, containing hydroxypropylmethylcelluloses as emulsifying agents **146**, 239
Kiesvaara, J., J. Yliruusi, The use of the Washburn method in determining the contact angles of lactose powder **92**, 81
Kiesvaara, J., J. Yliruusi, E. Ahomaki, Contact angles and surface free energies of theophylline and salicylic acid powders determined by the Washburn method **97**, 101
Kikuchi, H., see Maruyama, K. **111**, 103
Kikuchi, H., see Sakuma, S. **149**, 93
Kikuchi, H., see Yamauchi, H. **90**, 73
Kikuchi, M., N. Hamaguchi, K. Uekama, Improvement of oral and rectal bioavailabilis of carmofur by methylated β-cyclodextrin complexations **38**, 191
Kikuchi, R., see Watanabe, Y. **110**, 93
Kikuchi, S., A. Inaba, M. Takahashi, Y. Aramaki, S. Tsuchiya, Decreased distribution of gentamicin in rat kidney by complexation with dextran sulfate **42**, 193
Kildsig, D.O., see Basu, K. **47**, 195
Kildsig, D.O., see Spancake, C.W. **75**, 231
Killion, Jr. R.B., V.J. Stella, The nucleophilicity of dextrose, sucrose, sorbitol, and mannitol with *p*-nitrophenyl esters in aqueous solution **66**, 149
Killion, Jr. R.B., see Stella, V.J. **71**, 157
Killip, M., see Newman, S.P. **19**, 333
Kim, B., see Yoon, E.J. **97**, 213
Kim, C.K., Y.J. Choi, S.J. Lim, M.G. Lee, S.H. Lee, S.J. Hwang, Lymph node targeting and pharmacokinetics of

[^{3}H]methotrexate-encapsulated neutral large unilamellar vesicles and immunoliposomes **98**, 9
Kim, C.K., J.Y. Choi, Y.S. Yoon, J.P. Gong, H.G. Choi, J.Y. Kong, B.J. Lee, Preparation and evaluation of a dry elixir for the enhancement of the dissolution rate of poorly water-soluble drugs **106**, 25
Kim, C.K., M.S. Hong, Y.B. Kim, S.K. Han, Effect of penetration enhancers (pyrrolidone derivatives) on multilamellar liposomes of stratum corneum lipid: A study by UV spectroscopy and differential scanning calorimetry **95**, 43
Kim, C.K., S.J. Hwang, Lee M. Gull, The organ targetability of small and large albumin microspheres containing free and HSA conjugated methotrexate **89**, 91
Kim, C.K., E.B. Im, S.J. Lim, Y.K. Oh, S.K. Han, Development of glucose-triggered pH-sensitive liposomes for a potential insulin delivery **101**, 191
Kim, C.K., E.J. Jeong, Development of dried liposome as effective immuno-adjuvant for hepatitis B surface antigen **115**, 193
Enhanced lymph node delivery and immunogenicity of hepatitis B surface antigen entrapped in galactosylated liposomes **147**, 143
Kim, C.K., J.H. Kim, K.M. Park, K.H. Oh, U. Oh, S.J. Hwang, Preparation and evaluation of a titrated extract of *Centella asiatica* injection in the form of an extemporaneous micellar solution **146**, 63
Kim, C.K., J.J. Kim, S.C. Chi, C.K. Shim, Effect of fatty acids and urea on the penetration of ketoprofen through rat skin **99**, 109
Kim, C.K., M.J. Kim, K.H. Oh, Preparation and evaluation of sustained release microspheres of terbutaline sulfate **106**, 213
Kim, C.K., S.C. Kim, H.J. Shin, K.M. Kim, K.H. Oh, Y.B. Lee, I.J. Oh, Preparation and characterization of cytarabine-loaded w/o/w multiple emulsions **124**, 61
Kim, C.K., E.J. Lee, The controlled release of blue dextran from alginate beads **79**, 11
Kim, C.K., M.K. Lee, J.H. Han, B.J. Lee, Pharmacokinetics and tissue distribution of methotrexate after intravenous injection of differently charged liposome-entrapped methotrexate to rats **108**, 21
Kim, C.K., K.H. Min, Y.K. Oh, K.M. Park, K.M. Kim, Preparation and lectin binding characteristics of *N*-stearyl lactobionamide liposomes **128**, 65
Kim, C.K., Y.K. Oh, Development of hydrophilic human serum albumin microspheres using a drug–albumin conjugate **47**, 163
Kim, C.K., S.A. Ryuu, K.M. Park, S.J. Lim, S.J. Hwang, Preparation and physicochemical characterization of phase inverted water/oil microemulsion containing cyclosporin A **147**, 131
Kim, C.K., Y.S. Yoon, J.Y. Kong, Preparation and evaluation of flurbiprofen dry elixir as a novel dosage form using a spray-drying technique **120**, 21
Kim, C.K., see Chang, H.W. **79**, 71
Kim, C.K., see Chung, S.M. **68**, 61
Kim, C.K., see Hwang, S.J. **116**, 125
Kim, C.K., see Jeong, Y.N. **102**, 35
Kim, C.K., see Lee, K.C. **114**, 215
Kim, C.K., see Han, S.K. **126**, 35
Kim, H., see Chi, S.C. **126**, 267
Kim, J.H., see Kim, C.K. **146**, 63
Kim, J.J., see Kim, C.K. **99**, 109
Kim, K.M., see Kim, C.K. **124**, 61; **128**, 65
Kim, K.M., see Lim, C.R. **132**, 175
Kim, M.J., see Kim, C.K. **106**, 213
Kim, M.M., see Yoon, E.J. **97**, 213
Kim, S., T. Nishihata, S. Kawabe et al., Effect of gelatin in the formulation on rectal insulin absorption in the presence of enamine in normal rats and depancreatized dogs **21**, 179
Kim, S., see Nishihata, T. **31**, 185
Kim, S.C., see Kim, C.K. **124**, 61
Kim, S.K., see Shim, C.K. **56**, 269
Kim, S.K., see Young, C.S. **45**, 59
Kim, S.H., see Chung, S.M. **68**, 61
Kim, S.H., see Jang, S.H. **84**, 273
Kim, S.H., see Yoon, E.J. **99**, 343
Kim, S.R., see Doherty, M.M. **111**, 205
Kim, S.W., see Cremers, H.F.M. **110**, 117; **120**, 51
Kim, S.W., see Gilbert, D.L. **47**, 79
Kim, S.W., see Hovgaard, L. **132**, 107, 115
Kim, S.W., see Kwon, G.S. **79**, 191
Kim, S.W., see Sato, S. **22**, 229
Kim, S.W., see Seminoff, L.A. **54**, 241, 251
Kim, S.W., see Yuk, S.H. **77**, 221
Kim, W.B., see Lee, K.C. **44**, 49; **59**, 27
Kim, W.B., see Shim, H.J. **103**, 147
Kim, Y.B., see Kim, C.K. **95**, 43
Kim, Y.H., A.H. Ghanem, H. Mahmoud, W.I. Higuchi, Short chain alkanols as transport enhancers for lipophilic and polar/ionic permeants in hairless mouse skin: Mechanism(s) of action **80**, 17
Kim, Y.H., see Choi, Y.K. **80**, 9
Kimura, K., see Soliman, O.A. **149**, 73
Kimura, M., see Inoue, N. **137**, 75
Kimura, M., see Sugibayashi, K. **113**, 189
Kimura, R., see Hussain, A. **21**, 233, 289
Kimura, T., see Araki, M. **81**, 39
Kimura, T., see Katsu, T. **53**, 61
Kimura, T., see Kurosaki, Y. **47**, 13; **49**, 47; **51**, 47; **82**, 165
Kimura, T., see Nakayama, T. **121**, 55
Kimura, T., see Sawamoto, T. **146**, 181
Kindberg, C.G., see Monnot, E.A. **60**, 41
Kinget, R., A.M. Bontinck, H. Herbots, Problems of dialysis techniques in the study of macromolecule binding of drugs **3**, 65
Kinget, R., H. De Greef, Absorption characteristics of novel 8-MOP semi-solid-lipid-matrix formulations: In vitro–in vivo correlation **110**, 65
Kinget, R., see de Vos, A. **92**, 191
Kinget, R., see Hoa, N.T. **138**, 185
Kinget, R., see Hoa Ngo, T. **146**, 271
Kinget, R., see Kalala, W. **139**, 187
Kinget, R., see Peeters, R. **94**, 125

Kinget, R., see Provost, C. **62**, 217
Kinget, R., see Samyn, C. **121**, 211
Kinget, R., see Talukdar, M.M. **120**, 63; **129**, 233
Kinget, R., see Ugwoke, M.I. **148**, 23
Kinget, R., see Van den Mooter, G. **87**, 37; **97**, 133; **111**, 127
Kinget, R., see Vertommen, J. **146**, 21
Kinget, R., see Vervoort, L. **129**, 185
Kinkel, J.F.M., E. Tomlinson, Drug liquid-liquid distribution based on the fundamentals of segmented flow **6**, 261
Kinkel, J.F.M., see Buist, H.P.A. **15**, 347
Kinnunen, P., see Murtoniemi, E. **110**, 101; **108**, 155
Kinsel, J.F., E.I. Melnik, S. Lindenbaum et al., A model for lasalocid-mediated electrical transport of biogenic amines through lipid bilayer membranes **12**, 97
Kinugawa, M., see Muranushi, N. **4**, 271; 281
Kipp, J.E., Nonisothermal kinetics. Comparison of two methods of data treatment **26**, 339
Kipp, J.E., M.M. Jensen, K. Kronholm, M. McHalsky, Automated liquid chromatography for non-isothermal kinetic studies **34**, 1
Kipp, J.E., W.J. Smith III, P.B. Myrdal, Determination of phosphate function group acid dissociation constants of clindamycin 2-phosphate using ^{31}P Fourier transform NMR spectrometry **74**, 215
Kirby, C.J., see Onur, M.A. **88**, 313
Kiriyama, A., see Ushirogawa, Y. **81**, 133
Kiriyama, M., see Watanabe, Y. **110**, 93
Kirk, O., see Pridal, L. **136**, 53
Kisaka, J., see Pandit, N.K. **145**, 129
Kishimoto, I., see Higaki, K. **36**, 131
Kishimoto, M., see Murakami, T. **49**, 173; **54**, 259
Kishimoto, S.I., see Takada, K. **101**, 89
Kislal, O., see Acarturk, F. **85**, 1
Kislalioglu, M.S., see Khan, M.A. **102**, 185
Kissel, T., see Kriwet, B. **127**, 135
Kissel, T., see Oechslein, C.R. **139**, 25
Kitagawa, S., M. Sawada, H. Hirata, Fluorescence analysis with diphenylhexatriene and its ionic derivatives of the fluidity of liposomes constituted from stratum corneum lipids: Contribution of each lipid component and effects of long-chain unsaturated fatty acids **98**, 203
Kitagawa, S., N. Yokochi, N. Murooka, pH-Dependence of phase transition of the lipid bilayer of liposomes of stratum corneum lipids **126**, 49
Kitajima, M., see Takakura, Y. **37**, 135
Kitamura, S., L.C. Chang, J.K. Guillory, Polymorphism of mefloquine hydrochloride **101**, 127
Kitamura, S., S. Koda, A. Miyamae, T. Yasuda, Y. Morimoto, Dehydration effect on the stability of cefixime trihydrate **59**, 217
Kitamura, S., A. Miyamae, S. Koda, Y. Morimoto, Effect of grinding on the solid-state stability of cefixime trihydrate **56**, 125
Kitano, M., see Yamashita, F. **117**, 173
Kitaura, I., see Takeeda, T. **41**, 21
Kitazawa, S., see Johno, I. **25**, 255
Kivinen, A., I. Vikholm, S. Tarpila, A film balance study of the monolayer-forming properties of dietary phospholipids and the interaction with NSAIDs on the monolayers **108**, 109
Kiwada, H., see Yamashita, C. **70**, 225
Kiyohara, G., see Lee, G. **23**, 43
Kjaervig, Kristensen, see Majilat, P. **9**, 245
Klang, S., see Zeevi, A. **108**, 57
Klang, S.H., A. Baszkin, S. Benita, The stability of piroxicam incorporated in a positively-charged submicron emulsion for ocular administration **132**, 33
Klatt, P.R., see Deasy, P.B. **89**, 251
Kleinebudde, P., Application of low substituted hydroxypropylcellulose (L-HPC) in the production of pellets using extrusion/spheronization **96**, 119
Shrinking and swelling properties of pellets containing microcrystalline cellulose and low substituted hydroxypropylcellulose. I. Shrinking properties **109**, 209
Shrinking and swelling properties of pellets containing microcrystalline cellulose and low substituted hydroxypropylcellulose. II. Swelling properties **109**, 221
Kleinebudde, P., H. Lindner, Experiments with an instrumented twin-screw extruder using a single-step granulation/extrusion process **94**, 49
Kleinebudde, P., see Bleich, J. **106**, 77
Kleinebudde, P., see Ruchatz, F. **142**, 67
Klimek, R., see Thoma, K. **67**, 169
Klimmek, R., see Thiermann, H. **109**, 35
Klinger, G.H., see Herman, J. **42**, 15
Klixbull, U., H. Bundgaard, Prodrugs as drug delivery systems. XXX. 4-Imidazolidinones as potential bioreversible derivatives for the α-aminoamide moiety in peptides **20**, 273
Kinetics of reversible reactions of ampicillin with various aldehydes and ketones with formation of 4-imidazolidinones **23**, 163
Klixbull, U., see Bundgaard, H. **27**, 175; **29**, 19; **30**, 111
Klooster, N.T.M., see Van, Amerongen **86**, 9
Kluft, C., see Herremans, J.L.M. **129**, 191
Kmetec, V., A. Mrhar, R. Karba, F. Kozjek, Evaluation of drug stability data by analog-hybrid computer: Application to lorazepam **21**, 211
Knapczyk, J., Antimycotic buccal and vaginal tablets with chitosan **88**, 9
Excipient ability of chitosan for direct tableting **89**, 1
Chitosan hydrogen as a base for semisolid drug forms **93**, 233
Knapczyk, J., A.B. Macura, B. Pawlik, Simple tests demonstrating the antimycotic effect of chitosan **80**, 33
Knaub, S.R., see Ross, D.L. **93**, 131
Knepp, W.A., see Jayakrishnan, A. **106**, 221
Knez, Z., see Sencar-Bozic, P. **148**, 123
Knight, K.S., see Shankland, K. **137**, 255
Knight, P., see Baert, L. **86**, 187
Knight, P., see Prasad, V.K. **13**, 1
Knight, P.W., see Prasad, V.K. **11**, 81
Knight, V., see Vidgren, M. **115**, 209

Knight, V., see Waldrep, J.C. **97**, 205
Knoch, A., see Weiss, G. **124**, 87, 97
Knott, C.E., see Burnham, W.R. **13**, 9
Knowles, M., see McMahon, F.G. **91**, 75
Knuth, K.E., see Dowty, M.E. **88**, 293
Ko, H.M., see Huang, Y.B. **126**, 111; **131**, 137; **134**, 183
Kobal, E., Synthesis and antibacterial activity of fluorobenzoylcephalosporins **75**, 131
Kobarfard, F., see Dehpour, A.R. **119**, 133
Kobayashi, D., see Inoue, N. **137**, 75
Kobayashi, D., see Morimoto, Y. **91**, 9
Kobayashi, D., see Sugibayashi, K. **113**, 189
Kobayashi, M., see Narisawa, S. **148**, 85
Kobayashi, M., see Sugibayashi, K. **113**, 189
Kobayashi, Y., see Watanabe, Y. **99**, 337
Kobe, J., see Kristl, A. **57**, 229
Koch, M.H.J., see Bunjes, H. **129**, 159
Koch, M.H.J., see Westesen, K. **93**, 189; **103**, 225
Koch, S.A.M., K.B. Sloan, N-Mannich base derivatives of 5-fluorocytosine: A prodrug approach to improve topical delivery **35**, 243
Koch, S.A.M., see Sloan, K.B. **21**, 251
Kochhar, S.K., M.H. Rubinstein, D. Barnes, Slugging and recompression characterisation of some blends of pharmaceutical excipients **112**, 225
The effects of slugging and recompression on pharmaceutical excipients **115**, 35
Koda, S., see Kitamura, S. **56**, 125; **59**, 217
Koerten, H.K., see Bodde, H.E. **53**, 13
Kofler, B., see Kerc, J. **81**, R1
Koga, I., see Kadir, S. **40**, 257
Kogoshi, T., see Watanabe, Y. **64**, 147
Kogure, T., see Sugito, K. **47**, 157; **60**, 89
Koh, G.L., I.G. Tucker, Variability in the phenol-sulphuric acid assay for sodium carboxymethylcellulose **34**, 183
Koh, I.B., see Lee, K.C. **44**, 49
Kohli, D.V., see Tripathi, M. **67**, 207
Kohli, R., W.I. Archer, A. Li Wan Po, Laser velocimetry for the non-invasive assessment of the percutaneous absorption of nicotinates **36**, 91
Kohli, R., W.I. Archer, J.M.C. Roberts et al., Impedance measurements for the non-invasive monitoring of skin hydration: A reassessment **26**, 275
Kohri, N., K. Iwasa, J. Kurihara, K. Miyazaki, T. Arita, Inter-subject variation in oral absorption of ketoprofen from controlled-release granules in rabbits **49**, 213
Kohri, N., N. Miyata, M. Takahashi, H. Endo, K. Iseki, K. Miyazaki, S. Takechi, A. Nomura, Evaluation of pH-independent sustained-release granules of dipyridamole by using gastric-acidity-controlled rabbits and human subjects **81**, 49
Kohri, N., H. Yatabe, K. Iseki, K. Miyazaki, A new type of a pH-independent controlled release tablet **68**, 255
Kohri, N., see Nakano, M. **3**, 195
Koike, H., see Seta, Y. **41**, 255
Koike, R., see Hamamoto, K. **113**, 9
Koizumi, N., see Adachi, T. **124**, 129
Koizumi, T., see Hatanaka, T. **125**, 195
Koizumi, T., see Panomsuk, S.P. **126**, 147
Koizumi, T., S.P. Panomsuk, Release of medicaments from spherical matrices containing drug in suspension: Theoretical aspects **116**, 45
Kojima, I., S.S. Davis, The effect of salt concentration on the distribution of phenol between aqueous sodium chloride and carbon tetrachloride **20**, 203
Thermodynamics of distribution of *para*-substituted phenols between organic solvents and sodium chloride solution **20**, 247
Kojima, M., see Sasaki, H. **44**, 15; **60**, 177
Kok, W.J.J., W.P. Vrijhof, Impairment of carbamazepine tablet disintegration in patients **86**, 259
Kokubo, H., K. Morimoto, T. Ishida et al., Bioavailability and inhibitory effect for stress ulcer of cimetidine polymorphs in rats **35**, 181
Kolb, M., see Leuenberger, H. **115**, 217
Kolli, M., J.P. Montheard, J.M. Vergnaud, Release of benzocaine, procaine, 2-aminothiazole and 4-amino-4H-1,2,4-triazole from polymer carriers **81**, 103
Kolter, K., see Backensfeld, T. **74**, 85
Komada, F., see Okumura, K. **88**, 63
Komatsu, H., see Higaki, K. **36**, 131
Komatsu, H., see Okamoto, H. **30**, 35
Komiya, I., J.Y. Park, A. Kamani, Quantitative mechanistic studies in simultaneous fluid flow and intestinal absorption using steroids as model solutes **4**, 249
Konecny, A., see McMahon, F.G. **91**, 75
Kong, J.Y., see Kim, C.K. **120**, 21
Konieczny, M., see Randall, C.S. **120**, 235
Koning, B., see Moolenaar, F. **4**, 99
Konishi, R., see Wong, O. **52**, 191
Konstantinov, C., see Atcheva, M. **103**, 303
Kontny, M.J., S. Koppenol, E.T. Graham, Use of the sorption–desorption moisture transfer model to assess the utility of a desiccant in a solid product **84**, 261
Kontny, M.J., see Zografi, G. **18**, 99; **42**, 7
Koosha, F., see Washington, C. **59**, 79
Kopecek, J., see Bridges, J.F. **44**, 213
Kopecek, J., see Ikesue, K. **95**, 171
Kopecek, J., see Rihova, B. **87**, 105
Kopeckova, P., see Ikesue, K. **95**, 171
Kopeckova, P., see Rihova, B. **87**, 105
Koppenol, S., see Kontny, M.J. **84**, 261
Kopra, T., see Marvola, M. **44**, 159
Korsmeyer, R.W., R. Gurny, E. Doelker et al., Mechanisms of solute release from porous hydrophilic polymers **15**, 25
Kostenbauder, H.B., see Adams, W.P. **25**, 293, 313
Koster, V.S., P.F.M. Kuks, R. Lange, H. Talsma, Particle size in parenteral fat emulsions, what are the true limitations? **134**, 235
Kosykh, V.A., E.A. Podres, D.K. Novikov, S.A. Volgushev, A.V. Victorov, V.S. Repin, V.N. Smirnov, Effects of khellin and timefurone on secretion and catabolism of lipoproteins by cultured rabbit and human hepatocytes **57**, 223
Kotaras, A.D., see Rowe, R.C. **22**, 57

Kotzan, J.A., see Vallner, J.J. **16**, 47; **30**, 123; **33**, 125; **36**, 73; **54**, 123
Koupparis, M., P. Macheras, C. Reppas, Application of automated flow injection analysis (FIA) to dissolution studies **20**, 325
Koupparis, M., P. Macheras, C. Tsaprounis, Automated flow-injection colorimetric determination of acetaminophen for assays and dissolution studies of multicomponent dosage forms **27**, 349
Koupparis, M., see Parissi-Poulou, M. **51**, 169
Koupparis, M.A., see Pentari, J.G. **77**, 41
Kourounakis, A., N. Bodor, J. Simpkins, Synthesis and evaluation of a brain-targeted catechol derivative as a potential NGF-inducer **141**, 239
Kouta, H.I., see Moustafa, M.A. **36**, 185
Kovanen, A., see Sutinen, R. **57**, 149
Kowalski, K., see Cook, C.S. **60**, 125
Kowalski, P., see Regosz, A. **88**, 437
Kowaluk, E.A., see Roberts, M.S. **17**, 145
Koyama, Y., see Nishikawa, M. **85**, 75
Koyama, Y., see Yamashita, F. **89**, 199; **117**, 173
Koyanagi, Y., see Unezaki, S. **144**, 11
Kozjek, F., see Kmetec, V. **21**, 211
Kozjek, F., see Kristl, A. **57**, 229; **99**, 79
Krahn, F.U., J.B. Mielck, Effect of type and extent of crystalline order on chemical and physical stability of carbamazepine **53**, 25
Kraisintu, K., R.T. Parfitt, M.G. Rowan, A high-performance liquid chromatographic method for the determination and control of the composition of gentamicin sulphate **10**, 67
Kral, M.A., see Joshi, H.N. **51**, 19
Kramer, P.A., see Lamson, M.J. **105**, 259
Kramer, W.G., see Patel, V.S. **30**, 221
Krasowska, H., The hydrolysis of indomethacin in aqueous solutions of polysorbates **4**, 89
Effect of micellar solubilization on the gastrointestinal absorption of indomethacin in the rat **7**, 137
Krause, H.J., A. Schwarz, P. Rohdewald, Polylactic acid nanoparticles, a colloidal drug delivery system for lipophilic drugs **27**, 145
Kreilgard, B., see Hansen, J. **16**, 141
Kreuter, J., Physicochemical characterization of polyacrylic nanoparticles **14**, 43
Kreuter, J., S.N. Mills, S.S. Davis, C.G. Wilson, Polybutylcyanoacrylate nanoparticles for the delivery of [^{75}Se]norcholestenol **16**, 105
Kreuter, J., U. Muller, K. Munz, Quantitative and microautoradiographic study on mouse intestinal distribution of polycyanoacrylate nanoparticles **55**, 39
Kreuter, J., see Bentele, V. **13**, 109
Kreuter, J., see Borchard, G. **110**, 29
Kreuter, J., see Cappel, M.J. **69**, 143, 155
Kreuter, J., see Diepold, R. **54**, 149
Kreuter, J., see El Egakey, M.A. **13**, 349
Kreuter, J., see Fitzgerald, P. **40**, 81
Kreuter, J., see Gipps, E.M. **40**, 23
Kreuter, J., see Harmia, T. **33**, 45, 187
Kreuter, J., see Illum, L. **17**, 65
Kreuter, J., see Jahnke, R.W.O. **77**, 47
Kreuter, J., see Langer, K. **110**, 21; **137**, 67
Kreuter, J., see Michel, C. **84**, 93
Kreuter, J., see Pirker, S. **128**, 189
Kreuter, J., see Ruland, A. **72**, 149; **85**, 7; **101**, 57; **103**, 77; **107**, 23
Kreuter, J., see Scherer, D. **101**, 165
Kreuter, J., see Troster, S.D. **45**, 91; **61**, 85
Kreuter, J., see Waser, P.G. **39**, 213
Kreuter, J., see Wood, R.W. **23**, 175
Kreuter, J., see Zimmer, A.K. **110**, 211
Krill, S.L., J. Harn, K.R. Hahn, S.L. Gupta, Activity loss on room temperature storage of Survanta^{R}, a bovine lung extract based surfactant **132**, 89
Krishnan, T.R., I. Abraham, Butler E.I Vargha, Wettability study of clofazimine and poly(vinyl methyl ether/maleic anhydride) copolymer coevaporates **80**, 277
Kristensen, A., see Alderborn, G. **37**, 155
Kristensen, S., A.L. Grislingaas, J.V. Greenhill, T. Skjetne, J. Karlsen, H.H. Tonnesen, Photochemical stability of biologically active compounds. V. Photochemical degradation of primaquine in an aqueous medium **100**, 15
Kristensen, S., see Tonnesen, H.H. **110**, 161
Kristjansson, F., L.A. Sternson, S. Lindenbaum, An investigation on possible oligomer formation in pharmaceutical formulations of cisplatin **41**, 67
Kristl, A., A. Mrhar, F. Kozjek, The ionisation properties of acyclovir and deoxyacyclovir **99**, 79
Kristl, A., A. Mrhar, F. Kozjek, J. Kobe, Lipophilicity of guanine derivatives **57**, 229
Kristl, A., S. Srcic, F. Vrecer, B. Sustar, D. Vojnovic, Polymorphism and pseudopolymorphism: Influencing the dissolution properties of the guanine derivative acyclovir **139**, 231
Kristl, A., D. Vojnovic, R. Karba, A. Mrhar, F. Rubessa, B. Zupani, Mixture design of theophylline retard formulation **100**, 33
Kristl, J., J. Smid-Korbar, E. Struc, M. Schara, H. Rupprecht, Hydrocolloids and gels of chitosan as drug carriers **99**, 13
Kristl, J., see Gabrijelcic, V. **62**, 75
Kristl Smid-Korba, J.J., see Gasperlin, M. **107**, 51
Kristmundsdottir, T., O.S. Gudmundsson, K. Ingvarsdottir, Release of diltiazem from Eudragit microparticles prepared by spray-drying **137**, 159
Kristmundsdottir, T., T. Loftsson, W.P. Holbrook, Formulation and clinical evaluation of a hydrocortisone solution for the treatment of oral disease **139**, 63
Kristmundsdottir, T., see Arnardottir, H.B. **117**, 237; **134**, 71
Kristmundsdottir, T., see Sveinsson, S.J. **82**, 129; **92**, 29
Kristoffersson, E., see Hellen, L. **96**, 197, 205
Kristoffersson, E., see Laakso, R. **19**, 35
Kriwet, B., T. Kissel, Interactions between bioadhesive poly(-acrylic acid) and calcium ions **127**, 135
Kriwet, K., C.C. Muller-Goymann, Diclofenac release from phospholipid drug systems and permeation through excised human stratum corneum **125**, 231
Kronholm, K., see Kipp, J.E. **34**, 1

Kruithof, M.A.M., see Bodde, H.E. **53**, 13
Krumphuber, A., see Egermann, H. **73**, 37
Kruse, J., see Pirker, S. **128**, 189
Krycer, I., D.G. Pope, J.A. Hersey, An evaluation of the techniques employed to investigate powder compaction behaviour **12**, 113
Krykowski, E., see Szwarocka, A. **135**, 167
Ktistis, G., A viscosity study on oil-in-water microemulsions **61**, 213
Ktistis, G., see Attwood, D. **52**, 165; **88**, 417
Ktistis, G., see Ruth, H.S. **116**, 253
Kuang, L.R., see Li, C. **94**, 143
Kubo, H., K.I. Hosoya, H. Natsume, K. Sugibayashi, Y. Morimoto, In vitro permeation of several model drugs across rabbit nasal mucosa **103**, 27
Kubo, K., see Yoshimura, Y. **103**, 1
Kubota, K., M. Sznitowska, H.I. Maibach, Percutaneous permeation of β-methasone 17-valerate from different vehicles **96**, 105
Kudla, R.J., see Lambert, W.J. **95**, 181
Kuehnhoff, J., see Wong, O. **52**, 191
Kuge, Y., see Yanai, S. **123**, 237
Kuhfeld, M.T., Jr. R.E. Stratford, In vitro measurement of drug transport using a new diffusion chamber compatible with Millicell^{R} culture supports: Performance with Caco-2 monolayers **133**, 47
Kujari, H., see Urtti, A. **19**, 53
Kuks, P.F.M., see Koster, V.S. **134**, 235
Kulvanich, P., M. Jovanovic, Correlation between total adhesion and charge decay of a model interactive system during storage **39**, 51
Kulvanich, P., P.J. Stewart, Fundamental considerations in the measurement of adhesional forces between particles using the centrifuge method **35**, 111
An evaluation of the air stream Faraday cage in the electrostatic charge measurement of interactive drug systems **36**, 243
Kumakura, M., see Asano, M. **67**, 67
Kumakura, M., see Imasaka, K. **68**, 87; **81**, 31
Kumar, R., see Baveja, S.K. **24**, 355
Kumar, V., G.S. Banker, Incompatibility of polyvinyl acetate phthalate with benzocaine: Isolation and characterization of 4-phthalimidobenzoic acid ethyl ester **79**, 61
Kumar, V., see Wei, S. **142**, 175
Kunchandy, E., M.N.A. Rao, Effect of curcumin on hydroxyl radical generation through Fenton reaction **57**, 173
Oxygen radical scavenging activity of curcumin **58**, 237
Generation of superoxide anion and hydrogen peroxide by (+)-cyanidanol-3 **65**, 261
Kurihara, J., see Kohri, N. **49**, 213
Kurihara, M., see Hirayama, F. **35**, 193
Kurihara-Bergstrom, T., see Goldberg-Cettina, M. **114**, 237
Kurihara-Bergstrom, T., see Liu, P. **64**, 7; **78**, 123; **90**, 171
Kurihara-Bergstrom, T., see Mork Nielsen, N. **39**, 75
Kurihara-Bergstrom, T., see Wilding, I.R. **132**, 81
Kuriyama, K., see Aioi, A. **93**, 1; **113**, 159
Kuroda, M., see Yamaoka, T. **113**, 149
Kurokawa, T., see Takada, S. **146**, 147
Kuroko, M., see Katsu, T. **53**, 61
Kurono, Y., S. Kato, T. Kuwayama, H. Tamaki, K. Hatano, T. Yashiro, K. Ikeda, H. Bundgaard, *cis/trans* Isomerization and proton exchange reactions of 3,11b-dimethylbenzodiazepinooxazole **81**, 179
Kurono, Y., H. Tamaki, T. Kuwayama, T. Ichii, H. Sawabe, T. Yashiro, K. Ikeda, H. Bundgaard, Kinetics and mechanism of the acid-base equilibrium of mexazolam analogues: A modification of the mechanism proposed previously for mexazolam **80**, 135
Kurosaki, Y., S.I. Hisaichi, C. Hamada, T. Nakayama, T. Kimura, Effects of surfactants on the absorption of salicylic acid from hamster cheek pouch as a model of keratinized oral mucosa **47**, 13
Kurosaki, Y., S.I. Hisaichi, L.Z. Hong, T. Nakayama, T. Kimura, Enhanced permeability of keratinized oral-mucosa to salicylic acid with 1-dodecylazacycloheptan-2-one (Azone). In vitro studies in hamster cheek pouch **49**, 47
Kurosaki, Y., S.I. Hisaichi, T. Nakayama, T. Kimura, Enhancing effect of 1-dodecylazacycloheptan-2-one (Azone) on the absorption of salicylic acid from keratinized oral mucosa and the duration of enhancement in vivo **51**, 47
Kurosaki, Y., N. Nagahara, T. Tanizawa, H. Nishimura, T. Nakayama, T. Kimura, Use of lipid disperse systems in transdermal drug delivery: Comparative study of flufenamic acid permeation among rat abdominal skin, silicon rubber membrane and stratum corneum sheet isoated from hamster cheek pouch **67**, 1
Kurosaki, Y., H. Nishimura, K. Terao, T. Nakayama, T. Kimura, Existence of a specialized absorption mechanism for cefadroxil, an aminocephalosporin antibiotic, in the human oral cavity **82**, 165
Kurosaki, Y., see Araki, M. **81**, 39
Kurosaki, Y., see Katsu, T. **53**, 61
Kurosaki, Y., see Kurosaki, Y. **67**, 1
kurosaki, Y., see Nakayama, T. **121**, 55
Kurosaki, Y., see Noach, A.B.J. **90**, 229
Kurosaki, Y., see Sawamoto, T. **146**, 181
Kurosawa, N., E. Owada, K. Ito et al., Bioavailability of nifedipine suppository in healthy subjects **27**, 81
Kurtzhals, P., see Larsen, C. **41**, 121
Kusai, A., see Durrani, M.J. **24**, 209
Kusai, A., see Wakiyama, N. **78**, 95
Kusanoi, Y., see Murakami, M. **79**, 159
Kushla, G.P., J.L. Zatz, Influence of pH on lidocaine penetration through human and hairless mouse skin in vitro **71**, 167
Kussendrager, K.D., see Riepma, K.A. **66**, 47
Kussendrager, K.D., see Van Kamp, H.V. **28**, 229
Kussendrager, K.D., see Vromans, H. **39**, 201, 207
Kuttan, G., see Anto, R.J. **131**, 1
Kuttan, R., see Anto, R.J. **131**, 1
Kuu, W.Y., Determination of residence-time distribution in i.v. tubing of in-line drug delivery system using deconvolution technique **88**, 369

Kuu, W.Y., J. McShane, J. Wong, Determination of mass transfer coefficients during freeze drying using modeling and parameter estimation techniques **124**, 241
Kuu, W.Y., M.R. Prisco, R.W. Wood, T.J. Roseman, Studies of dissolution behavior of highly soluble drugs using a rotating disk **55**, 77
Kuu, W.Y., see Wong, J. **144**, 177
Kuwata, H., see Johno, I. **25**, 255
Kuwayama, T., see Kurono, Y. **80**, 135; **81**, 179
Kuzuya, T., see Hasegawa, T. **56**, 235
Kwan, K.C., K.R. Heimlich, The role of pharmacokinetics in drug product design **6**, 225
Kwee, B.M.C., see Busker, R.W. **36**, 113
Kwon, G.S., Y.H. Bae, H. Cremers, J. Feijen, S.W. Kim, Release of macromolecules from albumin-heparin microspheres **79**, 191
Kwon, G.S., see Cremers, H.F.M. **120**, 51
Kwon, K., D.W.A. Bourne, Physiological pharmacokinetic model for the distribution and elimination of tenoxicam **37**, 219
Kwong, E., see Bechard, S.R. **87**, 133
Kwong, E.C., P.L. Lamarche, G.R.B. Down, S.A. McClintock, E.B. Vadas, M.L. Cotton, Formulation assessment of MK-886, a poorly water-soluble drug, in the beagle dog dimethylpropionic acid **103**, 259
Kynoch, D.J., see Washington, C. **64**, 217
Kyroudis, A., S.L. Markantonis, A.H. Beckett, Relationship between plasma concentration, saliva concentration and urinary excretion rate of lithium in man **43**, 145
Kyyronen, K., L. Hume, L. Benedetti, A. Urtti, E. Topp, V. Stella, Methylprednisolone esters of hyaluronic acid in ophthalmic drug delivery: In vitro and in vivo release studies **80**, 161

L

La Manna, A., see Colombo, P. **63**, 43
La Manna, A., see Ferrari, F. **109**, 115
La Manna, A., see Giordano, F. **62**, 153
La Manna, A., see Giunchedi, P. **85**, 141; **94**, 15
La Manna, G., see Cavallaro, G. **90**, 195
La Porta, E., see Santoni, G. **50**, 75
La Rotonda, M., see Palagiano, F. **149**, 171
Laakso, R., S. Eerikainen, Effects of core components on indomethacin release from film-coated granules **67**, 79
Laakso, R., E. Kristoffersson, M. Marvola, Bi-exponential first-order release kinetics of indomethacin from tablets containing polysorbate 80 **19**, 35
Laakso, R., see Eerikainen, S. **71**, 201
Laakso, T., B. Smedsrod, Cellular distribution in rat liver of intravenously administered polyacryl starch and chondroitin sulfate microparticles **36**, 253
Laakso, T., see Stjarnkvist, P. **40**, 215
Labhasetwar, V.D., A.K. Dorle, A study of the dielectric constant of salicylic acid and aspirin compacts during aging **47**, 261
Labib, A., see Pimienta, C. **80**, 1
Lacy, J.E., R.T. Parfitt, M.G. Rowan, The effects of column packing material and inorganic cations on the separation of fluorescent *o*-phthalaldehyde derivatives of gentamicin by high-performance liquid chromatography **43**, 111
Lack, L.J., see Thomas, N.W. **44**, 261
Lafforgue, C., L. Carret, F. Falson, M.E. Reverdy, J. Freney, Percutaneous absorption of a chlorhexidine digluconate solution **147**, 243
Lafforgue, C., I. Eynard, F. Falson, A.C. Watkinson, J. Hadgraft, Percutaneous absorption of methyl nicotinate **121**, 89
Lafforgue, C., see Ganem-Quintanar, A. **147**, 165
Lafleuriel, M.T., see Vincent-Ballereau, F. **50**, 87
Lagas, M., C.F. Lerk, The polymorphism of sulphathiazole **8**, 11
Laghoueg, N., J. Paulet, J.L. Taverdet, J.M. Vergnaud, Oral polymer-drug devices with a core and an erodible shell for constant drug delivery **50**, 133
Laghoueg-Derriche, N., J. Bouzon, J.M. Vergnaud, Effect of temperature of the drying process of dosage forms prepared by a humidity technique **67**, 163
Laghoueg-Derriche, N., J.M. Vergnaud, Modelling the process of drying of dosage forms made of drug dispersed in a polymer **67**, 51
Drying of dosage forms prepared by a humidity technique using a programmed temperature system **71**, 229
Lagorce, J.F., see Jambut-Absil, A.C. **35**, 129
Lagueny, A.M., see Fawaz, F. **133**, 107
Lahajnar, G., see Srcic, S. **87**, 1
Lai, J.S., see Hu, O.Y.P. **53**, 85
Lai, J.W., see Jun, H.W. **16**, 65
Lai, T.Y.F., J.T. Carstensen, Effect of shape factor on cube root dissolution behavior **1**, 33
Lai, W.F., see Wan, L.S.C. **72**, 163; **81**, 75; **94**, 39
Laicher, A., see Weiss, G. **124**, 87, 97
Laik, Ali S., High performance liquid chromatographic separation and determination of chlordiazepoxide hydrochloride and two of its decomposition products **5**, 85
Laine, E., P. Auramo, P. Kahela, On the structural behaviour of triglycerides with time **43**, 241
Laine, E., K. Ishida, R. Rajala, T. Heikkila, K. Saarnivaara, I. Piippo, Crystal forms and bioavailability of erythromycin **38**, 33
Laine, E., V. Tuominen, P. Ilvessalo, P. Kahela, Formation of dihydrate from carbamazepine anhydrate in aqueous conditions **20**, 307
Laine, E., see Niemi, L. **68**, 205
Laine, E., see Pirttimaki, J. **95**, 93
Laizier, J., see Guyomard, S. **40**, 173
Lakings, D.B., see Rush, B.D. **73**, 231
Lam, F.C., see Gupta, P.K. **43**, 167; **51**, 253
Lam, F.C., see Hung, C.T. **44**, 117
Lam, K., see Cremers, H.F.M. **110**, 117
Lam, S., see Sitaram, B.R. **118**, 181
Lam, S.Y., see Shah, V.P. **53**, 53
Lam, S.Y., see Pershing, L.K. **86**, 201

Lam, X.M., H.R. Costantino, D.E. Overcashier, T.H. Nguyen, C.C. Hsu, Replacing succinate with glycolate buffer improves the stability of lyophilized interferon- **142**, 85
Lamarche, P., see Cotton, M.L. **109**, 237
Lamarche, P.L., see Kwong, E.C. **103**, 259
Lambert, L.D., see Pershing, L.K. **86**, 201
Lambert, W.J., R.J. Kudla, J.M. Holland, J.T. Curry, A biodegradable transdermal penetration enhancer based on *N*-(2-hydroxyethyl)-2-pyrrolidone. I. Synthesis and characterization **95**, 181
Lammers, J.W.J., see Zanen, P. **107**, 211; **114**, 111
Lamont, G., see Washington, N. **108**, 125
Lamp, K., see Gowan, W.G. **29**, 169
Lampard, J.F., see Smith, A. **59**, 115
Lampard, J.F., see Wilson, C.G. **50**, 155
Lamprecht, I., see Pfaff, G. **14**, 173
Lamson, M.J., L.G. Herbette, K.R. Peters, J.H. Carson, F. Morga, D.C. Chester, P.A. Kramer, Effects of hexagonal phase induction by dolichol on phospholipid membrane permeability and morphology **105**, 259
Lamson, M.L., J.L. Fox, W.I. Higuchi, Calcium and 1-hydroxyethylidene-1,1-bisphosphonic acid: Polynuclear complex formation in the physiological range of pH **21**, 143
Lan, E.L., see Ugwu, S.O. **102**, 193
Lanao, J.M., M.T. Vicente, M.L. Sayalero, Calculation of partial components of bioavailability in slow release formulations using model-independent methods **117**, 113
Lanao, J.M., M.T. Vicente, M.L. Sayalero, A. Dominguez-Gil, A method for the calculation of bioavailability in slow release formulations in the presence of within-individual variability **86**, 123
Lanao, J.M., see Calvo, M.V. **38**, 117
Lanao, J.M., see Navarro, A.S. **37**, 113
Lancaster, J., see Adjei, A. **107**, 57
Landau, I., see Mbela, T.K.M.N. **110**, 189
Landin, M., M. Casalderry, R. Martinez-Pacheco, J.L. Gomez-Amoza, C. Souto, A. Concheiro, R.C. Rowe, Chemical stability of acetylsalicylic acid in tablets prepared with different particle size fractions of a commercial brand of dicalcium phosphate dihydrate **123**, 143
Landin, M., R. Martinez-Pacheco, J.L. Gomez-Amoza, C. Souto, A. Concheiro, R.C. Rowe, Effect of country of origin on the properties of microcrystalline cellulose **91**, 123
Effect of batch variation and source of pulp on the properties of microcrystalline cellulose **91**, 133
Influence of microcrystalline cellulose source and batch variation on the tabletting behaviour and stability of prednisone formulations **91**, 143
The effect of country of origin on the properties of dicalcium phosphate dihydrate powder **103**, 9
Dicalcium phosphate dihydrate for direct compression: Characterization and intermanufacturer variability **109**, 1
Landin, M., B. Perez-Marcos, M. Casalderrey, R. Martinez-Pacheco, J.L. Gomez-Amoza, C. Souto, A. Concheiro, R.C. Rowe, Chemical stability of acetylsalicylic acid in tablets prepared with different commercial brands of dicalcium phosphate dihydrate **107**, 247
Landin, M., R.C. Rowe, P. York, Particle size effects on the dehydration of dicalcium phosphate dihydrate powders **104**, 271
Landin, M., P. York, M.J. Cliff, R.C. Rowe, A.J. Wigmore, Scale-up of a pharmaceutical granulation in fixed bowl mixer-granulators **133**, 127
The effect of batch size on scale-up of a pharmaceutical granulation in a fixed bowl mixer granulator **134**, 243
Lang, J.E., see Pikal, M.J. **17**, 237
Lang, P.O., see Alderborn, G. **37**, 155
Lange, R., see Koster, V.S. **134**, 235
Langer, K., C. Marburger, A. Berthold, J. Kreuter, F. Stieneker, Methylmethacrylate sulfopropylmethacrylate copolymer nanoparticles for drug delivery. Part I. Preparation and physicochemical characterization **137**, 67
Langer, K., E. Seegmuller, A. Zimmer, J. Kreuter, Characterisation of polybutylcyanoacrylate nanoparticles. I. Quantification of PBCA polymer and dextrans **110**, 21
Langer, K., see Pirker, S. **128**, 189
Langguth, P., see Luessen, H.L. **141**, 39
Langueny, A.M., see Guyot, M. **123**, 53
Langworthy, M., see Park, J.Y. **2**, 215
Lansley, A.B., see Pritchard, K. **129**, 137
Lanutti, A.T., see Wall, D.A. **97**, 171
Lanza, M., see Bonina, F. **145**, 87
Lanzarotti, E., see Ahmed, S.M. **109**, 99
Larhammar, H., I. Sjoholm, B. Skoog, A. Wichman, Comparative conformational analysis of human somatotropin and biosynthetic methionyl human somatotropin by spectroscopic titrations **23**, 13
Larhrib, H., J.I. Wells, M.H. Rubinstein, Compressing polyethylene glycols: The effect of compression pressure and speed **147**, 199
Larhrib, H., J.I. Wells, M.H. Rubinstein, G. Ricart, Characterization of PEGs using matrix-assisted laser desorption/ionisation mass spectrometry and other related techniques **147**, 187
Larhrib, H., see Nokhodchi, A. **118**, 191; **120**, 13
Larsen, B.D., see Friis, G.J. **136**, 61
Larsen, C., Macromolecular prodrugs. XII. Kinetics of release of naproxen from various polysaccharide ester prodrugs in neutral and alkaline solution **51**, 233
Macromolecular prodrugs. XIII. Determination of the ionization constant of dextran by potentiometric titration and from kinetic analysis of the hydrolysis of dextran indomethacin ester conjugates **52**, 55
Larsen, C., M. Johansen, Macromolecular prodrugs. I. Kinetics and mechanism of hydrolysis of *O*-benzoyl dextran conjugates in aqueous buffer and in human plasma **27**, 205
Larsen, C., M. Johansen, Macromolecular prodrugs. IV. Kinetics of hydrolysis of metronidazole monosuccinate dextran ester conjugates in aqueous solution and in plasma—Sequential release of metronidazole from the conjugates at physiological pH **35**, 39
Larsen, C., P. Kurtzhals, M. Johansen, Kinetics of regeneration of metronidazole from hemiesters of maleic acid, succinic acid and glutaric acid in aqueous buffer, human plasma and pig liver homogenate **41**, 121

Larsen, C., see Bundgaard, H. **1**, 95; **3**, 1, 341; **6**, 19; **7**, 169; **16**, 319; **18**, 67, 79
Larsen, C., see Harboe, E. **53**, 157
Larsen, C., see Johansen, M. **21**, 201; **26**, 227; **27**, 219
Larsen, Chr. N., see Buur, A. **16**, 41
Larsen, J.D., H. Bundgaard, Prodrug forms for the sulfonamide group. I. Evaluation of *N*-acyl derivatives, *N*-sulfonylamidines, *N*-sulfonylsulfilimines and sulfonylureas as possible prodrug derivatives **37**, 87
Prodrug forms for the sulfonamide group. III. Chemical and enzymatic hydrolysis of various *N*-sulfonyl imidates—A novel prodrug form for a sulfonamide group or an ester function **51**, 27
Larsen, J.D., see Bundgaard, H. **45**, 47
Larsen, R., see Bechgaard, E. **87**, 125; **89**, 147
Larsen, R., see Bindseil, E. **119**, 37
Larsen, R., see Jorgensen, L. **113**, 83
Larsson, A., see Stubberud, L. **134**, 79
Lashmar, U.T., J. Beesley, Correlation of rheological properties of an oil in water emulsion with manufacturing procedures and stability **91**, 59
Lashmar, U.T., J. Manger, Investigation into the potential for iontophoresis facilitated transdermal delivery of acyclovir **111**, 73
Lashmar, U.T., D. Provasi, S.W. Poole, Evaluation of the delivery of suspensions from an ultrasonic nebuliser with and without the lung surfactant 'Exosurf' **103**, 119
Lashmar, U.T., J.P. Richardson, A. Erbod, Correlation of physical parameters of an oil in water emulsion with manufacturing procedures and stability **125**, 315
Lasic, D.D., see Guo, L.S.S. **75**, 45
Lastra, C.F., see Modamio, P. **107**, 243; **112**, 93; **130**, 137
Lastres, J.L., see Sanchez, L. **115**, 201
Latrofa, A., see Trapani, G. **116**, 95; **139**, 215
Lattanzi, F., see Gasco, M.R. **62**, 119
Lattin, G., see Wong, O. **76**, 171
Lau, S.W.J., J.T. Slattery, Absorption of diazepam and lorazepam following intranasal administration **54**, 171
Lau, S.W.J., see Shah, J.C. **109**, 283
Laudanski, T., see Barnwell, S.G. **88**, 423; **89**, 245
Laudanski, T., see Cole, S.K. **80**, 63; **88**, 211
Laughlin, S.M., see Rocco, W.L. **122**, 17
Launchbury, A.P., see McCafferty, D.F. **31**, 9
Launchbury, A.P., see Murphy, A. **38**, 257
Launchbury, A.P., see Pickup, M.E. **22**, 311
Launchbury, A.P., see Woolfson, A.D. **34**, 17
Laurin, P., N. Nguyen, G. L'Esperance, R. Tawashi, Effect of particle morphology on the hiding power of talc powder **28**, 177
Lauroba, J., see Calpena, A.C. **103**, 179
Law, D., see Khankari, R.K. **82**, 117
Law, M.F.L., P.B. Deasy, Use of canonical and other analyses for the optimization of an extrusion-spheronization process for indomethacin **146**, 1
Law, M.Y.L., L.K. Pershing, L.K. Roberts, Arecoline-induced skin inflammation irritant or allergic dermatitis? **88**, 251
Law, M.F.L., see Deasy, P.B. **148**, 201
Law, S.L., P. Chang, C.H. Lin, Characteristics of mitoxantrone loading on liposomes **70**, 1
Law, S.L., C.K. Ho, T.F. Jang, P. Chang, F.M. Lin, Antitumor effect of mitoxantrone-containing liposomes **128**, 139
Law, S.L., T.F. Jang, P. Chang, F.M. Lin, Stability of mitoxantrone-containing liposomes **116**, 87
Law, S.L., T.F. Jang, C.H. Lin, Release characteristics of mitoxantrone-containing liposomes **103**, 81
Law, S.L., J.B. Kayes, Adsorption of non-ionic water-soluble cellulose polymers at the solid–water interface and their effect on suspension stability **15**, 251
Law, S.L., W.Y. Lo, F.M. Lin, C.H. Chaing, Dissolution and absorption of nifedipine in polyethylene glycol solid dispersion containing phosphatidylcholine **84**, 161
Law, S.L., W.Y. Lo, S.H. Pai et al., The adsorption of bovine serum albumin by liposomes **32**, 237
Law, S.L., W.Y. Lo, S.H. Pai, G.W. Teh, The electrokinetic behavior of liposomes adsorbed with bovine serum albumin **43**, 257
Law, T.K., A.T. Florence, T.L. Whateley, Some chemically modified poloxamer hydrogels: Controlled release of prostaglandin-E_2 and testosterone **33**, 65
Law, T.K., T.L. Whateley, A.T. Florence, Some chemically modified poloxamer hydrogels: Preparation, morphology and swelling properties **21**, 277
Law, T.K., see Ping, Q. **61**, 79
Lawrence, C.B., see Aboofazeli, R. **111**, 63
Lawrence, M.J., see Aboofazeli, R. **93**, 161; **106**, 51; **111**, 63; **125**, 107
Lawter, J.R., see Touitou, E. **104**, 19
Layloff, T., see Shah, V.P. **82**, 1
Lazar, J.D., see Thombre, A.G. **111**, 181
Le Brun, P.P.H, P.L.A. Fox, M.E. De Vries, H.E. Bodde, In vitro penetration of some β-adrenoreceptor blocking drugs through porcine buccal mucosa **49**, 141
Le Corre, P., Guevello P Le, V. Gajan, F. Chevanne, Verge R. Le, Preparation and characterization of bupivacaine-loaded polylactide and polylactide-*co*-glycolide microspheres **107**, 41
Le Corre, P., see Dollo, G. **131**, 219; **136**, 165
Le Corre, P., see Polard, E. **134**, 37
Le Guevello, P, see Le Corre, P. **107**, 41
Le Hir, A., see Arnaud, P. **80**, 129
Le Hoang, M.D., see Gimenez, F. **43**, 23
Le Hoang, M.D., see Postaire, E. **55**, 99
Le Normand, Y., see Badian, M. **18**, 9
Le Ray, A.M., M. Vert, J.C. Gautier, J.P. Benoit, Fate of [^{14}C]poly(DL-lactide-*co*-glycolide) nanoparticles after intravenous and oral administration to mice **106**, 201
Le, V.H., B.C. Lippold, Influence of physicochemical properties of homologous esters of nicotinic acid on skin permeability and maximum flux **124**, 285
Le Verge, R., see Dollo, G. **131**, 219; **136**, 165
Le Verge, R., see Le Corre, P. **107**, 41
Le Verge, R., see Polard, E. **134**, 37
Le Verge, R., see Rolland, A. **39**, 173; **53**, 67
Leach, R.H., see Culank, L.S. **2**, 173

Leahey, W.J., see El Turabi, H. **53**, 249
Leahy, D.E., A.L. De Meere, A.R. Wait, P.J. Taylor, J.A. Tomenson, E. Tomlinson, A general description of water-oil partitioning rates using the rotating diffusion cell **50**, 117
Leary, J.R., S.D. Ross, Mathematical expression of tablet dissolution profiles **17**, 193
Lebegue, S., see Gressier, B. **104**, 151
Lebo, D.B., P.K. Gupta, Modulation of membrane cholesterol content in cultured LLC-PK_1 cells **125**, 1
Lechat, I., see Bassani, V. **63**, 57
Lechuga-Ballesteros, D., N. Rodriguez-Hornedo, The influence of additives on the growth kinetics and mechanism of L-alanine crystals **115**, 139
Effects of molecular structure and growth kinetics on the morphology of L-alanine crystals **115**, 151
Lechuga-Ballesteros, D., see Rodriguez-Hornedo, N. **85**, 149
Leclerc, B., see Couarraze, G. **56**, 197
Leclerc, B., see Husson, I. **86**, 113
Leclercq, G., see Chamart, S. **99**, 59
Lecompte, D., see Jacolot, A. **128**, 283
Lecomte, J.M., see Gimenez, F. **43**, 23
Lecomte, S., M.T. Chenon, NMR investigation of pefloxacin/cation/DNA interactions. Mg^{2+} and Ca^{2+} binding **139**, 105
LeCotonnec, J.Y., see Jonkman, J.H.G. **25**, 113
Ledwidge, M.T., O.I. Corrigan, Effects of environmental factors on the dehydration of diclofenac HEP dihydrate and theophyline monohydrate **147**, 41
Lee, B.J., G.H. Min, Oral controlled release of melatonin using polymer-reinforced and coated alginate beads **144**, 37
Lee, B.J., K.A. Parrott, J.W. Ayres, R.L. Sack, Design and evaluation of an oral controlled release delivery system for melatonin in human subjects **124**, 119
Lee, B.J., see Kim, C.K. **106**, 25; **108**, 21
Lee, B.L., see Pena, L.E. **113**, 89
Lee, C.S., see Imamura, Y. **5**, 25; **8**, 277
Lee, E.J., see Kim, C.K. **79**, 11
Lee, E.D., see Shim, H.J. **103**, 147
Lee, E.D., see Yoon, E.J. **109**, 181
Lee, G., J. Swarbrick, G. Kiyohara, D.W. Payling, Drug permeation through human skin. III. Effect of pH on the partitioning behavior of a chromone-2-carboxylic acid **23**, 43
Lee, G., see Gopferich, A. **71**, 237, 245
Lee, G., see Schuckler, F. **70**, 173; **80**, 81
Lee, H., see Chang, H.W. **79**, 71
Lee, H., see Chung, S.M. **68**, 61
Lee, H., see Yoon, E.J. **67**, 177
Lee, H.B., see Lim, C.R. **132**, 175
Lee, H.J., B.N. Ahn, E.J. Yoon, W.H. Paik, C.K. Shim, M.G. Lee, Pharmacokinetics and tissue distribution of adriamycin and adriamycinol after intravenous administration of adriamycin-loaded neutral proliposomes to rats **121**, 1
Lee, H.J., B.N. Ahn, W.H. Paik, C.K. Shim, M.G. Lee, Inverse targeting of reticuloendothelial system-rich organs after intravenous administration of adriamycin-loaded neutral proliposomes containing poloxamer 407 to rats **131**, 91
Lee, H.J., see Lee, H.J. **121**, 1
Lee, H.J., see Yoon, W.H. **102**, 249
Lee, H.K., see Hume, L.R. **111**, 295
Lee, H.K., see Stella, V.J. **120**, 189, 197
Lee, J.M., see Duncan, M.R. **120**, 179
Lee, J.T., see Ha, I.S. **144**, 91
Lee, K.C., The use of magnetite for estimation of microspheres in mice **47**, 265
Lee, K.C., I.B. Koh, W.B. Kim, Y.J. Lee, Size and morphological analysis of albumin microspheres in the lungs and liver of mice **44**, 49
Lee, K.C., Y.J. Lee, W.B. Kim, C.Y. Cha, Monoclonal antibody-based targeting of methotrexate-loaded microspheres **59**, 27
Lee, K.C., H.M. Song, I.J. Oh, P.P. DeLuca, Reversed-phase high-performance liquid chromatography for simultaneous determination of prostaglandins E_2, A_2 and B_2 **106**, 167
Lee, K.C., K.O. Suen, M. Yianneskis, C. Marriott, Investigation of the aerodynamic characteristics of inhaler aerosols with an inhalation simulation machine **130**, 103
Lee, K.C., J.Y. Yoon, B.H. Woo, C.K. Kim, P.P. DeLuca, Post-column fluorescence HPLC for salmon calcitonin formulations **114**, 215
Lee, K.C., see Ha, I.S. **144**, 91
Lee, K.C., see Oh, I.J. **106**, 135
Lee, K.M., see Hwang, S.J. **116**, 125
Lee, M., see Benjamin, E.J. **14**, 209
Lee, M.H., see Lee, M.J. **113**, 175
Lee, M.H., see Lee, Y.H. **76**, 71
Lee, M.H., see Young, C.S. **45**, 59
Lee, M.J., M.H. Lee, C.K. Shim, Inverse targeting of drugs to reticuloendothelial system-rich organs by lipid microemulsion emulsified with poloxamer 338 **113**, 175
Lee, M.K., see Kim, C.K. **108**, 21
Lee, M.G., see Chang, H.W. **79**, 71
Lee, M.G., see Chung, S.M. **68**, 61
Lee, M.G., see Francomb, M.M. **103**, 87
Lee, M.G., see Jang, S.H. **84**, 273
Lee, M.G., see Jeong, Y.N. **102**, 35
Lee, M.G., see Kim, C.K. **98**, 9
Lee, M.G., see Lee, H.J. **131**, 91
Lee, M.G., see Lee, S.H. **90**, 81
Lee, M.G., see Shim, H.J. **103**, 147
Lee, M.G., see Yoon, E.J. **67**, 177; **97**, 213; **99**, 343; **109**, 181
Lee, M.G., see Yoon, W.H. **102**, 249
Lee, M.H., see Shim, C.K. **56**, 269
Lee, P.H., M.H. Su, A.H. Ghanem, T. Inamori, E.R. Kern, W.I. Higuchi, An application of the C concept in predicting the topical efficacy of finite dose acyclovir in the treatment of cutaneous HSV-1 infections in hairless mice **93**, 139
Lee, R.B., see Rogers, J.D. **16**, 191
Lee, S.D., see Shim, H.J. **103**, 147
Lee, S.D., see Yoon, E.J. **109**, 181
Lee, S.H., W.H. Yoon, S.H. Jang, K.S. Han, K.H. Jeong, I.S. Kim, M.G. Lee, Pharmacokinetics of a 22 kDa variant of

unlabelled monomeric human growth hormone in rabbits **90**, 81
Lee, S.H., see Jang, S.H. **84**, 273
Lee, S.H., see Jeong, Y.N. **102**, 35
Lee, S.H., see Kim, C.K. **98**, 9
Lee, S.H., see Lee, S.H. **90**, 81
Lee, S.H., see Yoon, W.H. **102**, 249
Lee, S.J., see Yuk, S.H. **77**, 221, 231
Lee, S.W., see Hardy, J.G. **31**, 151
Lee, V.H.L., Disposition of topically applied vitamin A in the albino rabbit eye **11**, 21
Lee, V.H.L., L.W. Carson, K.A. Takemoto, Macromolecular drug absorption in the albino rabbit eye **29**, 43
Lee, V.H.L., Jr. R.E. Stratford, K.W. Morimoto, Age-related changes in esterase activity in rabbit eyes **13**, 183
Lee, V.H.L., J. Swarbrick, M.A. Redell, D.C. Yang, Vehicle influence of ocular disposition of sodium cromoglycate in the albino rabbit **16**, 163
Lee, V.H.L., J.R. Robinson, Disposition of pilocarpine in the pigmented rabbit eye **11**, 155
Lee, V.H.L., see Bundgaard, H. **33**, 15
Lee, V.H.L., see Buur, A. **42**, 51
Lee, V.H.L., see Drustrup, Larsen J. **47**, 103
Lee, V.H.L., see Redell, M.A. **17**, 299
Lee, V.H.L., see Stratford Jr., R.E. **13**, 263; **30**, 73
Lee, W., see Hudson, A.J. **133**, 257
Lee, Y.H., M.H. Lee, C.K. Shim, Pharmacokinetics of diltiazem and deacetyldiltiazem in rats **76**, 71
Lee, Y.J., see Lee, K.C. **44**, 49; **59**, 27
Lee-Amies, G., see Podczeck, F. **142**, 97
Leese, P., see McMahon, F.G. **91**, 75
Leeuwenkamp, O.R., E. Van der Mark, W. Van Bennekom, A. Bult, Investigation of the photochemical and thermal degradation of aqueous nitroprusside solutions using liquid chromatography **24**, 27
Leeuwenkamp, O.R., N.L.J. Chin, E. Van der Mark et al., In vitro degradation of nitroprusside in relation to in vivo decomposition and mechanism of action **33**, 1
Lefebvre, C., see Rambaud, J. **15**, 199
Lefebvre, R.A., see De, Muynck C. **104**, 1, 87
Legrand, P., see Tabosa, de Egito E. **145**, 17
Legros, F.J., see Benameur, H. **89**, 157
Leh, F.K.V., W. Wolf, Preparation and distribution of ^{195}Pt(*m*)-labeled bleomycin **1**, 41
Leharne, S., see Buckton, G. **110**, 179
Leharne, S.A., see Buckton, G. **83**, 115
Lehmann, K., see Beckert, T.E. **143**, 13
Lehmussaari, K., see Suhonen, P. **127**, 85
Lehner, S.J., B.W. Muller, J.K. Seydel, Interactions between *p*-hydroxybenzoic acid esters and hydroxypropyl-β-cyclodextrin and their antimicrobial effect against *Candida albicans* **93**, 201
Lehr, C.M., J.A. Bouwstra, E.H. Schacht, H.E. Junginger, In vitro evaluation of mucoadhesive properties of chitosan and some other natural polymers **78**, 43
Lehr, C.M., F.G.J. Poelma, H.E. Junginger, J.J. Tukker, An estimate of turnover time of intestinal mucus gel layer in the rat in situ loop **70**, 235
Lehr, C.M., see Schnurrer, J. **141**, 251
Lehtola, V.M., see Heinamaki, J.T. **112**, 191
Leigh, T.R., T. Nazir, J. Wiggins, D. Ganderton, J.V. Collins, Performance characteristics of the DeVilbiss Ultraneb 99 ultrasonic nebuliser with reference to use in sputum induction **67**, 275
Leike, J., see Schneider, T. **132**, 9
Leiviska, K., see Merkku, P. **102**, 117
Leiviska, K., see Murtoniemi, E. **108**, 155; **110**, 101
Lelawongs, P., J.C. Liu, Y.W. Chien, Transdermal iontophoretic delivery of arginine-vasopressin (II). Evaluation of electrical and operational factors **61**, 179
Lelawongs, P., J.C. Liu, O. Siddiqui, Y.W. Chien, Transdermal iontophoretic delivery of arginine-vasopressin (I). Physicochemical considerations **56**, 13
Lemesle-Lamache, V., D. Wouessidjewe, M. Cheron, D. Duchene, Study of β-cyclodextrin and ethylated β-cyclodextrin salbutamol complexes, in vitro evaluation of sustained-release behaviour of salbutamol **141**, 117
Lenaerts, V., H. Bundgaard, M. Quarton et al., Temperature-dependent rheological behavior of Pluronic F-127 aqueous solutions **39**, 121
Lenaerts, V., see Chouinard, F. **72**, 211
Lenaerts, V., see Grislain, L. **15**, 335
Lenaerts, V., see Juhasz, J. **77**, 309
Lenaerts, V., see Kante, B. **7**, 45
Lenaerts, V., see Pimienta, C. **80**, 1
Lencioni, E., see Bocci, V. **34**, 111
Lencioni, E., see Bocci, V. **24**, 109
Lennernas, H., K. Palm, U. Fagerholm, P. Artursson, Comparison between active and passive drug transport in human intestinal epithelial (Caco-2) cells in vitro and human jejunum in vivo **127**, 103
Lennholm, H., see Ek, R. **122**, 49
Leo, A.J., see Acree, Jr. W.E. **20**, 209
Leomy, C., L. Bonhomme, D. Amdidouche, C. Massari, B. Huynh, R. Hagipamtelli, M. Reynes, G. Fredj, J.L. Misset, G. Mathe, HPLC determination of the stability of retinoic acid in ovule formulations for the treatment of cervical dysplasia related to papillomavirus **115**, 235
Leon, M.J., see Lucero, M.J. **110**, 241; **111**, 261; **116**, 31, 125; **127**, 73; **140**, 251
Leopold, C.S., D.R. Friend, In vitro study for the assessment of poly(L-aspartic acid) as a drug carrier for colon-specific drug delivery **126**, 139
Leopold, C.S., H.I. Maibach, Effect of lipophilic vehicles on in vivo skill penetration of methyl nicotinate in different races **139**, 161
Lepage, F., see Ceolin, R. **74**, 77
Lerk, C.F., see Bos, C.E. **55**, 175; **67**, 39
Lerk, C.F., see de Boer, A.H. **130**, 231
Lerk, C.F., see de Haan, P. **31**, 15; **34**, 57
Lerk, C.F., see Frijlink, H.W. **49**, 91; **64**, 195
Lerk, C.F., see Lagas, M. **8**, 11
Lerk, C.F., see Leuenberger, H. **52**, 91
Lerk, C.F., see Piepma, K.A. **97**, 195
Lerk, C.F., see Proost, J.H. **13**, 287

Lerk, C.F., see Riepma, K.A. **66**, 47; **85**, 121; **87**, 149; **90**, R1; **97**, 29
Lerk, C.F., see Te Wierik, G.H.P. **98**, 219; **102**, 81
Lerk, C.F., see Van der Veen, J. **87**, 161
Lerk, C.F., see Van der Voort Maarschalk, K. **140**, 185
Lerk, C.F., see Van Kamp, H.V. **28**, 229
Lerk, C.F., see Vromans, H. **35**, 29; **39**, 201, 207
Lerk, C.F., see Zuurman, K. **102**, 1
Lerk, P.C., H. Sucker, Application of sucrose laurate, a new pharmaceutical excipient in peroral formulations of cyclosporin A **92**, 197
Application of sucrose laurate in topical preparations of cyclosporin A **92**, 203
Leroux, J.C., see Chouinard, F. **72**, 211
Leroy, P., C. Gavriloff, A. Nicolas, P. Archimbault, G. Ambroggi, Comparative assay of amoxicillin by high-performance liquid chromatography and microbiological methods for pharmacokinetics studies in calves **82**, 157
Leroy-Lechat, F., D. Wouessidjewe, J.P. Andreux, F. Puisieux, D. Duchene, Evaluation of the cytotoxicity of cyclodextrins and hydroxypropylated derivatives **101**, 97
Lescure, F., see Roy, D. **148**, 165
Lesko, L.J., see Pradhan, S. **138**, 207
Leslie, S.T., see Davis, S.S. **60**, 191; **49**, 183
Lesne-Hulin, A., see Bourget, P. **146**, 167
L'Esperance, G., see Laurin, P. **28**, 177
Leterme, P., see Flament, M.P. **148**, 93
Lethiecq, M., see Pelucio-Lopes, C. **96**, 249
Letourneux, Y., see Delie, F. **115**, 45
Leu, R., H. Leuenberger, The application of percolation theory to the compaction of pharmaceutical powders **90**, 213
Leucuta, S.E., The kinetics of in vitro release and the pharmacokinetics of miotic response in rabbits of gelatin and albumin microspheres with pilocarpine **54**, 71
Leucuta, S.E., R. Risca, D. Daicoviciu, D. Porutiu, Albumin microspheres as a drug delivery system for epirubicin: Pharmaceutical, pharmacokinetic and biological aspects **41**, 213
Leuenberger, H., The compressibility and compactibility of powder systems **12**, 41
Compression of binary powder mixtures and solubility parameters of solids **27**, 127
Leuenberger, H., J.D. Bonny, M. Kolb, Percolation effects in matrix-type controlled drug release systems **115**, 217
Leuenberger, H., J.D. Bonny, C.F. Lerk, H. Vromans, Relation between crushing strength and internal specific surface area of lactose compacts **52**, 91
Leuenberger, H., see Bausch, A. **101**, 63
Leuenberger, H., see Caraballo, I. **109**, 229; **139**, 177
Leuenberger, H., see El-Arini, S.K. **121**, 141
Leuenberger, H., see Holman, L.E. **57**, R1
Leuenberger, H., see Leu, R. **90**, 213
Leuenberger, H., see Mumenthaler, M. **72**, 97
Leuenberger, H., see Usteri, M. **55**, 135
Leung, K.K.M., P.A. Bridges, K.M.G. Taylor, The stability of liposomes to ultrasonic nebulisation **145**, 95
Leung, P., see Johnson, K. **73**, 197
Leuverman, A., see Moolenaar, F. **45**, 161
Levadoux, E., V. Sautou, J.E. Bazin, P. Schoeffler, J. Chopineau, Medical plastics: Compatibility of alfentanil and propofol alone or mixed. Stability of the alfentanil-propofol mixture **127**, 255
Levadoux, E., see Sautou-Miranda, V. **130**, 251
Levi-Schaffer, F., see Touitou, E. **70**, 159; **103**, 131
Levy, G., see Chou, R.C. **18**, 217
Levy, G., see Galinsky, R.E. **10**, 301
Levy, G., see Peck, C.C. **82**, 9
Levy, L., see Bechard, S.R. **114**, 205
Levy, M.C., D. Hettler, M.C. Andry, P. Rambourg, Polyhydroxamic serum albumin microcapsules: Preparation and chelating properties **69**, R1
Levy, M.C., M.C. Andry, Microcapsules prepared through interfacial cross-linking of starch derivatives **62**, 27
Levy, M.C., see Andry, M.C. **128**, 197
Levy, M.C., see Edwards-Levy, F. **96**, 85; **103**, 253
Levy, M.Y., S. Benita, Design and characterization of a submicronized o/w emulsion of diazepam for parenteral use **54**, 103
Drug release from submicronized o/w emulsion: A new in vitro kinetic evaluation model **66**, 29
Levy, M.Y., see Magenheim, B. **94**, 115
Levy, R.J., see Johnston, T.P. **52**, 139
Levy, R.J., see Sintov, A. **146**, 55
Lewing, E., see Kahela, P. **14**, 103
Lewis, D., J. Hadgraft, Mixed monolayers of dipalmitoylphosphatidylcholine with azone or oleic acid at the air–water interface **65**, 211
Lewis, D., M. Paulo, E. Faustino, A. Farinha, In vitro comparative studies of transdermal nicotine delivery systems **148**, 177
Lewis, D., see Brain, K.R. **71**, R9
Lewis, D., see Hadgraft, J. **73**, 125
Lewis, D.A., H.O. Alpar, Therapeutic possibilities of drugs encapsulated in erythrocytes **22**, 137
Lewis, D.A., see Field, W.N. **51**, 175
Lewis, G.A., Determination of dissociation constants of sparingly soluble compounds from solubility data **18**, 207
Lewis, G.A., R.P. Enever, Solution thermodynamics of some potentially long-acting norethindrone derivatives. I. Variation of enthalpies and entropies of fusion with temperature **2**, 203
Solution thermodynamics of some potentially long-acting norethindrone derivatives. II. Solutions in 2,2,4-trimethylpentane (iso-octane) **3**, 275
Solution thermodynamics of some potentially long-acting norethinedrone derivatives. III. Measurement of aqueous solubilities and the use of group free energy contributions in predicting partition coefficients **3**, 319
Lewis, K.J., see Hudson, A.J. **136**, 23
Lheritier, J., A. Chauvet, B. Abramovici, J. Masse, Improvement of the dissolution kinetics of SR 33557 by means of solid dispersions containing PEG 6000 **123**, 273
Lherm, C., R.H. Muller, F. Puisieux, P. Couvreur, Alkylcyanoacrylate drug carriers. II. Cytotoxicity of cyanoacrylate nanoparticles with different alkyl chain length **84**, 13

Lherm, C., see Muller, R.H. **84**, 1
Li, C., D.J. Yang, L.R. Kuang, S. Wallace, Polyamino acid microspheres: Preparation, characterization and distribution after intravenous injection in rats **94**, 143
Li, L.C., In vitro controlled release of theophylline from tablets containing a silicone elastomer latex **87**, 117
Li, V.H.K., J.R. Robinson, Solution viscosity effects on the ocular disposition of cromolyn sodium in the albino rabbit **53**, 219
Li, V.H.K., see Wood, R.W. **23**, 175
Li Wan Po, A., J.R. Mhando, Formulation of sustained-release products: Dissolution and diffusion-controlled release from gelatin films **20**, 87
Li Wan Po, A., P.V. Mroso, W.J. Irwin, Modelling decomposition in the solid state: Stability of salsalate in suspension in the presence of excipients **16**, 115
Li Wan Po, A., P.V. Mroso, Drug-drug incompatibility in the solid state: Kinetic interpretation, modelling and prediction **18**, 287
Li Wan Po, A., L.P. Wong, C.A. Gilligan, Characterisation of commercially available theophylline sustained- or controlled-release systems: In-vitro drug release profiles **66**, 111
Li Wan Po, A., see Al-Kassas, R.S. **94**, 59
Li Wan Po, A., see Carstensen, J.T. **83**, 87
Li Wan Po, A., see Chan, S.Y. **83**, 73
Li Wan Po, A., see Cheung, Y.W. **26**, 175; **27**, 325
Li Wan Po, A., see Gilligan, C.A. **73**, 51; **75**, 1
Li Wan Po, A., see Hempenstall, J.M. **23**, 131
Li Wan Po, A., see Irwin, W.J. **4**, 47; **20**, 25; **66**, 193, 243
Li Wan Po, A., see Jian, L. **86**, 59; **95**, 101
Li Wan Po, A., see Kohli, R. **36**, 91
Li Wan Po, A., see Liak, T.L. **36**, 233
Li Wan Po, A., see Mhando, J.R. **59**, 165; **62**, 249
Li Wan Po, A., see Su, X.Y. **123**, 47
Li Wan Po, A., see Wong, L.P. **83**, 95
Li Wan Po, A., see Zhou, X.H. **62**, 259; **69**, 29; **70**, 271; **75**, 97, 117
Li, Z.J., D.J.W. Grant, Effects of excess enantiomer on the crystal properties of a racemic compound: Ephedrinium 2-naphthalenesulfonate **137**, 21
Liak, T.L., A. Li Wan Po, W.J. Irwin, The effects of drug therapy on urinary pH-excipient effects and bioactivation of methenamine **36**, 233
Liao, G., see Zhang, Z. **139**, 1
Liao, H.M., see Yu, H.Y. **127**, 1
Liaw, J., J.R. Robinson, The effect of polyethylene glycol molecular weight on corneal transport and the related influence of penetration enhancers **88**, 125
Liaw, J., Y. Rojanasakul, J.R. Robinson, The effect of drug charge type and charge density on corneal transport **88**, 111
Liaw, J., A. Rubinstein, J.R. Robinson, Bioavailability study of Theo-dur tablets in the fasted cannulated dog **59**, 105
Liaw, J., see Rojanasakul, Y. **66**, 131
Liberato, D.J., see Huselton, C.A. **134**, 157
Lichtenberger, R., H.P. Merkle, Intrinsic diffusion coefficients of drugs in pressure-sensitive adhesive polymer masses **64**, 117
Liddle, J., see Harron, D.W.G. **57**, 9
Lidgate, D.M., S.G. Hegde, R. Maskiewicz, Conductivity measurement as a convenient technique for determination of liposome capture volume **96**, 51
Lievens, H.S.R., see Craig, D.Q.M. **96**, 147
Liew, C.V., see Wan, L.S.C. **96**, 59; **118**, 213
Liggeri, E., see Dal Pozzo, A. **70**, 219
Liguori, A., see Bramanti, G. **49**, 241
Lilley, J.C., L.H. Patterson, M.J. Taylor, Preparation and stability of liposome encapsulated doxorubicin and mitoxantrone and their internalisation by murine peritoneal macrophages **107**, 149
Lim, C.R., K.H. Oh, K.M. Kim, S.H. Yuk, H.B. Lee, C.K. Kim, The enhancement of liver targetability of [^{3}H]methotrexate-galactosylated serum albumin conjugate in mice **132**, 175
Lim, S.J., see Kim, C.K. **98**, 9; **101**, 191; **147**, 131
Lima, M.C.P., see Beezer, A.E. **38**, 251
Lin, C.W., T.M. Cham, Compression behavior and tensile strength of heat-treated polyethylene glycols **118**, 169
Effect of particle size on the available surface area of nifedipine from nifedipine-polyethylene glycol 6000 solid dispersions **127**, 261
Lin, C.H., see Law, S.L. **70**, 1; **84**, 161; **103**, 81; **116**, 87; **128**, 139
Lin, F.M., see Law, S.L. **128**, 139
Lin, H.H., see Wu, P.C. **143**, 119; **145**, 215
Lin, H.H., Y.B. Huang, L.R. Hsu, Y.H. Tsai, The influence of process-induced variations on the nature of stearic acid incorporation into an FAPG base **112**, 165
Lin, J.L., see Brewster, M.E. **79**, 289
Lin, L.H., see Gu, L. **45**, 129
Lin, L.H., see Benjamin, E.J. **35**, 73
Lin, M.C., W.C. Duncan-Hewitt, Deformation kinetics of acetaminophen crystals **106**, 187
Lin, S.Y., see Lin, Y.Y. **124**, 53
Lin, S.Y., Y.H. Kao, Solid particulates of drug-β-cyclodextrin inclusion complexes directly prepared by a spray-drying technique **56**, 249
Lin, S.Z., D. Wouessidjewe, M.C. Poelman, D. Duchene, Indomethacin and cyclodextrin complexes **69**, 211
In vivo evaluation of indomethacin/cyclodextrin complexes gastrointestinal tolerance and dermal anti-inflammatory activity **106**, 63
Lin, S.Y., see Kawashima, Y. **18**, 335
Lin, S.Y., G.L. Amidon, N.D. Weiner, A.H. Goldberg, Viscoelasticity of cellulose polymers and mucociliary transport on frog palates **95**, 57
Lin, T., see Adjei, A. **107**, 57
Lin, T., see Lu, M.F. **71**, 25
Lin, T.L., see Kawashima, Y. **99**, 229
Lin, Y.Y., K.S. Chen, S.Y. Lin, Temperature effect on the thermal characteristics and drug penetrability of the thermally on-off switching membrane **124**, 53
Lindahl, L., see Engstrom, S. **86**, 137

Lindberg, J., see Nicklasson, M. **37**, 195; **69**, 255
Lindberg, J., see Wennergren, B. **53**, 35
Lindblad, M., see O'Hagan, D.T. **119**, 251
Lindell, K., S. Engstrom, In vitro release of timolol maleate from an in situ gelling polymer system **95**, 219
Investigation of surfactant alkyl chain length and counterion effects on the thermogelling EHEC system **124**, 107
Lindenbaum, S., E.S. Rattie, G.E. Zuber et al., Polymorphism of auranofin **26**, 123
Lindenbaum, S., see Kinsel, J.F. **12**, 97
Lindenbaum, S., see Kristjansson, F. **41**, 67
Lindenbaum, S., see Oliyai, R. **73**, 33
Lindenbaum, S., see Tan, X. **74**, 127
Lindenbaum, S., see Vadnere, M. **22**, 207
Lindholm, T., A. Huhtikangas, P. Saarikivi, Organic solvent residues in free ethyl cellulose films **21**, 119
Lindley, M., see Patel, C.I. **39**, 93
Lindner, H., see Kleinebudde, P. **94**, 49
Lindqvist, A.S., see Eerikainen, S. **75**, 181
Lindqvist, A.S., see Merkku, P. **102**, 117
Lindstedt, B., G. Ragnarsson, J. Hjartstam, Osmotic pumping as a release mechanism for membrane-coated drug formulations **56**, 261
Lindstedt, B., M. Sjoberg, J. Hjartstam, Osmotic pumping release from KCl tablets coated with porous and nonporous ethylcellulose **67**, 21
Lindstedt, B., see Hjartstam, J. **61**, 101
Lindstedt, B., see Ragnarsson, G. **79**, 223
Linert, W., see Repond, C. **38**, 47
Ling, B.L., see Heng, P.W.S. **116**, 119
Ling, B.L., see Wan, L.S.C. **141**, 161
Linn, E.E., see Chowhan, Z.T. **3**, 117
Liou, W.S., see Hu, O.Y.P. **53**, 85
Lipovac, K., see Juretic, D. **61**, 219
Lippold, B.C., D. Hackemuller, The influence of skin moisturizers on drug penetration in vivo **61**, 205
Lippold, B.C., A. Ohm, Correlation between wettability and dissolution rate of pharmaceutical powders **28**, 67
Lippold, B.C., H. Schneemann, The influence of vehicles on the local bioavailability of β-methasone-17-benzoate from solution- and suspension-type ointments **22**, 31
Lippold, B.C., see Le, V.H. **124**, 285
Lippold, B.C., see Lippold, B.H. **54**, 15
Lippold, B.H., B.K. Sutter, B.C. Lippold, Parameters controlling drug release from pellets coated with aqueous ethyl cellulose dispersion **54**, 15
Lipschitz, S., see Wanwimolruk, S. **75**, 55
Liso, G., see Bonina, F.P. **124**, 45
Liso, G., see Trapani, G. **116**, 95; **139**, 215
Little, R.J., see Bodor, N. **10**, 307
Liu, C.H., S.C. Chen, Y.H. Kao, C.C. Kao, T.D. Sokoloski, M.T. Sheu, Properties of hydroxypropylmethylcellulose agranules produced by water spraying **100**, 241
Liu, C.H., see Liu, C.H. **100**, 241
Liu, C.H., see Sheu, M.T. **85**, 57
Liu, D., see Liu, F. **125**, 73
Liu, F., D. Liu, Amphipathic polyethylene glycol stabilized emulsions (o/w): Physical characterization and in vivo distribution **125**, 73
Liu, H.X., I. Adachi, I. Horikoshi, M. Ueno, Mechanism of promotion of lymphatic drug absorption by milk fat globule membrane **118**, 55
Liu, J.C., Y. Sun, O. Siddiqui, Y.W. Chien, W.M. Shi, Blood glucose control in diabetic rats by transdermal iontophoretic delivery of insulin **44**, 197
Liu, J.C., Y. Sun, K. Tojo, Y.W. Chien, Membrane permeation kinetics of nortestosterone: Effect of methyl groups on thermodynamics **25**, 265
Liu, J.C., see Lelawongs, P. **56**, 13; **61**, 179
Liu, J.C., see Tan, E.L. **42**, 161
Liu, P., W.I. Higuchi, A.H. Ghanem, T. Kurihara-Bergstrom, W.R. Good, Quantitation of simultaneous diffusion and metabolism of β-estradiol in hairless mouse skin: Enzyme distribution and intrinsic diffusion/metabolism parameters **64**, 7
Assessing the influence of ethanol on simultaneous diffusion and metabolism of β-estradiol in hairless mouse skin for the 'asymmetric' situation in vitro **78**, 123
Liu, P., A.S. Nightingale, T. Kurihara-Bergstrom, Variation of human skin permeation in vitro: Ionic vs. neutral compounds **90**, 171
Liu, P., see Ghanem, A.H. **78**, 137
Liu, P., see Goldberg-Cettina, M. **114**, 237
Liu, P., see Sclafani, J. **124**, 213
Liu, S., see Zhan, X. **115**, 167
Liversidge, G.G., P. Conzentino, Drug particle size reduction for decreasing gastric irritancy and enhancing absorption of naproxen in rats **125**, 309
Liversidge, G.G., K.C. Cundy, Particle size reduction for improvement of oral bioavailability of hydrophobic drugs. I. Absolute oral bioavailability of nanocrystalline danazol in beagle dogs **125**, 91
Liversidge, G.G., D.J.W. Grant, Influence of physicochemical interactions on the properties of suppositories. IV. Factors influencing the in vivo release of ketoprofen and metronidazole from fatty suppository bases **13**, 243
Liversidge, G.G., D.J.W. Grant, J.M. Padfield, Influence of physicochemical interactions on the properties of suppositories. I. Interactions between the constituents of fatty suppository bases **7**, 211
Liversidge, G.G., T. Nishihata, K.K. Engle, T. Higuchi, Effect of rectal suppository formulation on the release of insulin and on the gluocose plasma levels in dogs **23**, 87
Liversidge, G.G., T. Nishihata, K.K. Engle, T. Higuchi, Effect of suppository shape in the systemic availability of rectally administered insulin and sodium salicylate **30**, 247
Liversidge, G.G., see Grant, D.J.W. **14**, 251
Liversidge, G.G., see Nishihata, T. **31**, 185
Ljubic, M., see Zorc, B. **99**, 135
Llabres, M., see Delgado, A. **134**, 203; **140**, 219
Llabres, M., see Dorta, M.J. **100**, 9
Llabres, M., see Farina, J.B. **45**, 123
Llabres, M., see Munguia, O. **86**, 107

Llabres, M., see Oliva, A. **143**, 163
Llabres, M., see Sanchez, E. **83**, 59
Llabres, M., see Soriano, I. **125**, 223; **142**, 135
Llera, J.M., see Alvarez-Fuentes, J. **148**, 219
Llera, J.M., see Holgado, M.A. **114**, 13
Lloyd, A.W., G.P. Martin, S.H. Soozandehfar, Azopolymers: A means of colon specific drug delivery? **106**, 255
Lloyd, A.W., C.J. Olliff, K.J. Rutt, A comparison of carboxylate salts as liposomal cryoprotectants **131**, 257
Lloyd-Jones, J.G., see Hessell, P.G. **52**, 159
Lo, W.Y., see Law, S.L. **32**, 237; **43**, 257; **84**, 161
Loftsson, T., J. Baldvinsdottir, A.M. Sigurdardottir, The effect of cyclodextrins on the solubility and stability of medroxyprogesterone acetate and megestrol acetate in aqueous solution **98**, 225
Loftsson, T., S. Bjornsdottir, G. Palsdottir, N. Bodor, The effects of 2-hydroxypropyl-β-cyclodextrin on the solubility and stability of chlorambucil and melphalan in aqueous solution **57**, 63
Loftsson, T., H. Fridriksdottir, Degradation of lomustine (CCNU) in aqueous solutions **62**, 243
Loftsson, T., H. Fridriksdottir, T.K. Gudmundsdottir, The effect of water-soluble polymers on aqueous solubility of drugs **127**, 293
Loftsson, T., H. Frioriksdottir, A.M. Siguroardottir, H. Ueda, The effect of water-soluble polymers on drug-cyclodextrin complexation **110**, 169
Loftsson, T., H. Fridriksdottir, S. Thorisdottir, E. Stefansson, The effect of hydroxypropyl methylcellulose on the release of dexamethasone from aqueous 2-hydroxypropyl-β-cyclodextrin formulations **104**, 181
Loftsson, T., M. Matsuo, R.W. Caldwell et al., Synthesis and evaluation of novel lidocaine sulfur analogs **22**, 345
Loftsson, T., B.J. Olafsdottir, The effect of 2-hydroxypropyl-β-cyclodextrin on the simultaneous dissolution and degradation of chlorambucil **66**, 289
Loftsson, T., B.J. Olafsdottir, Cyclodextrin-accelerated degradation of β-lactam antibiotics in aqueous solutions **67**, R5
Loftsson, T., B.J. Olafsdottir, Baldvinsdottir, Estramustine: Hydrolysis, solubilization, and stabilization in aqueous solutions **79**, 107
Loftsson, T., A.M. Siguroardottir, J.H. Olafsson, Improved acitretin delivery through hairless mouse skin by cyclodextrin complexation **115**, 255
Loftsson, T., see Brewster, M.E. **75**, R5; **79**, 289; **125**, 17
Loftsson, T., see Kristmundsdottir, T. **139**, 63
Loftsson, T., see Siguroardottir, A.M. **126**, 73
Loftus, A.M., see Gorman, S.P. **65**, 127
Loftus, A.M., see Jones, D.S. **119**, 247
Lokind, K.B., F.H. Lorenzen, Oral bioavailability of 17β-estradiol and prodrugs tested in rats, pigs and dogs **127**, 155
Lokind, K.B., F.H. Lorenzen, H. Bundgaard, Oral bioavailability of 17β-estradiol and various ester prodrugs in the rat **76**, 177
Lombardo, E., see Santoni, G. **64**, 235
Long, D.F., A.J. Repta, L.A. Sternson, The reactivity of cisplatin in plasma. Implications for sample handling in pharmacokinetic studies **6**, 167
Longer, M., see Chiang, C.C. **117**, 197
Look, Z.M., see Faulkner, R.D. **43**, 53
Loos, J.A., see Baggerman, C. **27**, 17
Lootvoet, G., E. Beyssac, G.K. Shiu, J.M. Aiache, W.A. Ritschel, Study on the release of indomethacin from suppositories: In vitro–in vivo correlation **85**, 113
Looze, Y., see Vincentelli, J. **134**, 147
Lopedota, A., see Trapani, G. **139**, 215
Lopes, F., see Cruz, M.E.M. **96**, 67
Lopez, A., M.J. Morant, D. Guzman, J. Borras-Blasco, O. Diez-Sales, M. Herraez, Skin permeation model of phenylalkylcarboxylic homologous acids and their enhancer effect on percutaneous penetration of 5-Fluorouracil **139**, 205
Lopez, A., see Lopez, A. **139**, 205
Lopez, A.A., M.M. Molina, H.C. Jung et al., Bioequivalence study of nitrofurantoin tablets: In vitro–in vivo correlation **28**, 167
Lopez-Berestein, G., see Vadiei, K. **57**, 125, 133
Lordi, N.G., see Jerzewski, R.L. **101**, 35, 45
Lordi, N.G., see Masilungan, F.C. **20**, 295
Lorenzen, F.H., see Lokind, K.B. **76**, 177; **127**, 155
Lorette, G., see Machet, L. **72**, R1
Lorette, G., see Pelucio-Lopes, C. **96**, 249
Los, G., see Cremers, H.F.M. **110**, 117
Losa, C., see Alonso, M.J. **68**, 69
Losa, C., see Egea, M.A. **67**, 103
Losa, C.M., see Martinez-Pacheco, R. **47**, 37
Lostritto, R.T., see Silvestri, S.L. **50**, 141
Loth, H., Vehicular influence on transdermal drug penetration **68**, 1
Loth, H., E. Hemgesberg, Properties and dissolution of drugs micronized by crystallization from supercritical gases **32**, 265
Lott, F.D., see Niven, R.W. **101**, 81
Lotter, A.P., see de Villiers, M. **88**, 275
Lotter, A.P., see Gerber, J.J. **69**, 265; **73**, 137
Lotter, A.P., see Muller, D.G. **75**, 201; **86**, 175; **102**, 109
Lotter, A.P., see Van Tonder, E.C. **63**, 35
Louden, J.D., H.K. Patel, R.C. Rowe, A preliminary examination of the structure of gels and emulsions containing cetostearyl alcohol and cetrimide using laser Raman spectroscopy **25**, 179
Louden, J.D., R.C. Rowe, A quantitative examination of the structure of emulsions prepared using cetostearyl alcohol and cetrimide using Fourier transform infrared microscopy **63**, 219
Loukas, Y.L., P. Jayasekera, G. Gregoriadis, Novel liposome-based multicomponent systems for the protein of photolabile agents **117**, 85
Loukas, Y.L., V. Vraka, G. Gregoriadis, Use of a nonlinear least-squares model for the kinetic determination of the stability constant of cydodextrin inclusion complexes **144**, 225

Lourenco, C., M. Teixeira, S. Simoes, R. Gaspar, Steric stabilization of nanoparticles: Size and surface properties **138**, 1
Love, W.G., see Williams, A.S. **85**, 189
Lovell, M.W., H.W. Johnson, P.K. Gupta, Stability of a less-painful intravenous emulsion of clarithromycin **118**, 47
Lovell, M.W., H.W. Johnson, H.W. Hui, J.B. Cannon, P.K. Gupta, C.C. Hsu, Less-painful emulsion formulations for intravenous administration of clarithromycin **109**, 45
Lovell, M.W., S.G. Schulman, Dissociation equilibria of protonated hydroxybenzamides and alkoxybenzamides: Water as a reactant **11**, 345
Lovrecich, M., F. Nobile, F. Rubessa, G. Zingone, Effect of ageing on the release of indomethacin from solid dispersions with Eudragits **131**, 247
Lovrecich, M., F. Rubessa, Effect of loading parameters on the theophylline release from polystyrene beads **40**, 63
Low, A.S., see Taylor, R.B. **60**, 185; **102**, 201
Lowes, M.M.J., see Bluhm, D.P. **68**, 277; 281
Lowther, N., see Matthews, G.P. **50**, 111
Lozano, R., see Desai, D.S. **144**, 153
Lu, D.R., K. Abu-Izza, F. Mao, Nonlinear data fitting for controlled release devices: An integrated computer program **129**, 243
Lu, M.F., A. Fuentes, T. Lin, L. Woodward, Intranasal absorption of estropipate in the dog **71**, 25
Lu, S.M., Dimensionless presentation for drug release from a coated pure drug bead. 1. Analysis **112**, 105
Lu, S.M., S.R. Chen, Controlled release from a coated particle—Effects of initial conditions and methods of solution **119**, 11
Lu, S.M., J.Y. Yu, Dimensionless presentation for drug release from a coated pure drug bead. 2. Experiment **112**, 117
Luangtana-Anan, M., J.T. Fell, Bonding mechanisms in tabletting **60**, 197
Lubbers, S.C.M., see Van der Houwen, O.A.G.J. **112**, 97
Lucarelli, M., see Di, Giulio A. **89**, 133
Lucero, M.J., J. Garcia, J. Vigo, M.J. Leon, A theological study of semisolid preparations of Eudragit^{R} **116**, 31
Lucero, M.J., J.A. Sanchez, J. Vigo, F. Martin, A.M. Rabasco, Protection by α-tocopherol against skin ulcers induced by ionizing radiations **104**, 49
Lucero, M.J., J. Vigo, M.J. Leon, A study of shear and compression deformations on hydrophilic gels of tretinoin **106**, 125
Stability of hydrophilic gels of tretinoin **110**, 241
A study of shear and compression deformations on hydrophilic gels of α-tocopherol **111**, 261
Degradation kinetics of α-tocopherol in hydrophilic gels **140**, 251
Lucero, M.J., J. Vigo, M.J. Leon, F. Martin, J.A. Sanchez, Therapeutic efficacy of hydrophilic gels of α-tocopherol and tretinoin in skin ulcers induced by adriamycin hydrochloride **127**, 73
Lucke, L., see Furst, W. **61**, 43
Lucks, J.S., B.W. Muller, R.H. Muller, Inorganic suspensions—Interaction with salts and ionic surfactants **58**, 229
Polymeric and emulsion carriers—interaction with antiflocculants and ionic surfactants **63**, 183
Lucks, J.S., see Muller, R.H. **138**, 85
Ludden, T., see Peck, C.C. **82**, 9
Ludwig, A., M. Van Ooteghem, The evaluation of viscous ophthalmic vehicles by slit lamp fluorophotometry in humans **54**, 95
Ludwig, A., N. Unlu, M. Van Ooteghem, Evaluation of viscous ophthalmic vehicles containing carbomer by slit-lamp fluorophotometry in humans **61**, 15
Ludwig, A., see Mbela, T.K.M.N. **110**, 189
Luessen, H.L., V. Bohner, D. Perard, P. Langguth, J.C. Verhoef, A.G. De Boer, H.P. Merkle, H.E. Junginger, Mucoadhesive polymers in peroral peptide drug delivery. V. Effect of poly(acrylates) on the enzymatic degradation of peptide drugs by intestinal brush border membrane vesicles **141**, 39
Luessen, H.L., see Akiyama, Y. **136**, 155; **138**, 13
Lufti, K.M., see Al-Meshal, M.A. **64**, 155
Lugt, C.B., see Titulaer, H.A.C. **69**, 83
Luke, D.R., see Vadiei, K. **57**, 125, 133
Lukowski, G., R.H. Muller, B.W. Muller, M. Dittgen, Acrylic acid copolymer nanoparticles for drug delivery. I. Characterization of the surface properties relevant for in vivo organ distribution **84**, 23
Lund, E.M., see Ostberg, T. **112**, 241
Lundberg, B.B., Evaluation of methods for complexing prednimustine to low density lipoprotein **99**, 275
Lundberg, B.B., B.C. Mortimer, T.G. Redgrave, Submicron lipid emulsions containing amphipathic polyethylene glycol for use as drug-carriers with prolonged circulation time **134**, 119
Lundberg, B., The solubilization of lipophilic derivatives of podophyllotoxins in sub-micron sized lipid emulsions and their cytotoxic activity against cancer cells in culture **109**, 73
Lundberg, P.J., see Abrahamsson, B. **140**, 229
Lundgren, P., see Karabit, M.S. **54**, 51; **56**, 169
Lundin, S., P. Artursson, Absorption of a vasopressin analogue, 1-deamino-8-D-arginine-vasopressin (dDAVP), in a human intestinal epithelial cell line, CaCO-2 **64**, 181
Lundin, S., J. Moss, H. Bundgaard, P. Artursson, Absorption of thyrotropin-releasing hormone (TRH) and a TRH prodrug in a human intestinal cell line (Caco-2) **76**, R1
Lundin, S., N. Pantzar, L. Hedin, B.R. Westrom, Intestinal absorption enhancement by sodium taurodihydrofusidate of a peptide hormone analogue (dDAVP) and a macromolecule (BSA) in vitro and in vivo **59**, 263
Lundqvist, A.E.K., F. Podczeck, J.M. Newton, Influence of disintegrant type and proportion on the properties of tablets produced from mixtures of pellets **147**, 95
Luner, P.E., S.R. Babu, S.C. Mehta, Wettability of a hydrophobic drug by surfactant solutions **128**, 29
Lunghetti, G., see Pitcairn, G. **102**, 11
Luo, C.W., see Chang, R.K. **60**, 171

Lutfi, K.M., see El-Sayed, Y.M. **64**, 109
Luukkala, M., see Ketolainen, J. **125**, 45
Luyckx, M., see Benaji, B. **105**, 83
Luyckx, M., see Dine, T. **77**, 279
Luyckx, M., see Faouzi, M.A. **89**, 125; **105**, 89
Luyckx, M., see Gressier, B. **104**, 151
Luyckx, M., see Khalfi, F. **139**, 243
Luyt, A.S., see Siebert, J.G.H. **61**, 157; **65**, 29; **66**, 157
Lyden, M., see Alden, M. **110**, 267
Lynch, I.R., see Buxton, P.C. **42**, 135
Lynch, J., see Harris, D. **56**, 97
Lynch, S., see Hastewell, J. **101**, 115

M

Ma, B., see Zhan, X. **115**, 161; **126**, 29
Ma, L., C. Ramachandran, N.D. Weiner, Partitioning of an homologous series of alkyl *p*-aminobenzoates into multilamellar liposomes: Effect of liposome composition **70**, 209
Partitioning of a homologous series of alkyl *p*-aminobenzoates in dipalmitoylphosphatidylcholine liposomes: Effect of liposome type **77**, 127
Ma, X., J. Taw, C.M. Chiang, Control of drug crystallization in transdermal matrix system **142**, 115
Maa, Y.F., C.C. Hsu, Aggregation of recombinant human growth hormone induced by phenolic compounds **140**, 155
Maa, Y.F., P.A. Nguyen, C.C. Hsu, Spray-coating of rhDNase on lactose: Effect of system design, operational parameters and protein formulation **144**, 47
Maanders, J.P.A.C.M., see De, Mol N.J. **16**, 153
Maassen, S., see Muller, R.H. **138**, 85
Maccari, M., see Wilding, I.R. **76**, 133
Maccario, J., see Benkerrour, L. **19**, 27
MacCarthy, D.J., see Collins, A.E.M. **51**, 103
MacFarlane, C.B., see Ojile, J.E. **10**, 99
MacGregor, T.R., The mouse tail-flick method for evaluation of narcotic antagonist sustained release **38**, 171
MacGregor, T.R., see Matzek, K.M. **28**, 85, 151
Mach, H., see Thibert, R. **134**, 59
Macheras, P., Quick method for the calculation of the absorption rate constant of the linear one-compartment model using all available blood level data **19**, 339
Macheras, P., M. Koupparis, S. Antimisiaris, An in vitro model for exploring CR theophylline–milk fat interactions **54**, 123
Macheras, P., M. Koupparis, E. Apostolelli, Dissolution of 4 controlled-release theophylline formulations in milk **36**, 73
Macheras, P., M. Koupparis, C. Tsaprounis, An automated flow injection-serial dynamic dialysis technique for drug-protein binding studies **30**, 123
Drug dissolution studies in milk using the automated flow injection serial dynamic dialysis technique **33**, 125
Macheras, P., M. Symillides, M. Georgiacodis, A new approach for the in vivo evaluation of sustained release dosage forms **52**, 249
Macheras, P., C. Reppas, Dissolution and in vitro permeation behaviours of dicumarol nitrofurantoin and sulfamethizole in the presence of protein **37**, 103
Macheras, P., A. Rosen, Some considerations concerning the effect of an increased rate of oral absorption on drug levels of protein-bound drugs **6**, 175
Effect of absorption and elimination rates on the maintenance time of therapeutic drug levels. Derivation of an equation for this time and its applications **16**, 353
Macheras, P., see Koupparis, M. **20**, 325; **27**, 349
Macheras, P., see Parissi-Poulou, M. **51**, 169
Macheras, P., see Poulou, M. **34**, 29
Macheras, P., see Rosen, A. **32**, 171
Machet, L., L. Vaillant, J. Pinton, J.C. Besnard, M. Breteau, N. Cochelin, G. Lorette, Percutaneous administration of digoxin across hairless mouse skin and human skin in vitro **72**, R1
Machet, L., see Pelucio-Lopes, C. **96**, 249
Machida, R., see Ohwaki, T. **93**, 61
Machida, Y., see Coutel-Egros, A. **84**, 117
Machida, Y., see Fujimori, J. **119**, 47
Machida, Y., see Hamamoto, K. **113**, 9
Machida, Y., see Ito, R. **61**, 109
Machida, Y., see Maitani, Y. **49**, 23; **64**, 139
Machida, Y., see Morishita, I. **78**, 9; **91**, 29
Machida, Y., see Morishita, M. **78**, 1
Machida, Y., see Muramatsu, K. **107**, 1; **124**, 19
Machida, Y., see Nagano, H. **147**, 119
Machida, Y., see Nakamura, F. **134**, 173
Machida, Y., see Obata, Y. **89**, 191
Machida, Y., see Ohara, N. **105**, 31; **119**, 125
Machida, Y., see Takahashi, T. **61**, 35
Machida, Y., see Uchida, N. **74**, 95
MacKay, M., see Antonin, K.H. **130**, 33
MacKay, M., see Hastewell, J. **126**, 245
MacKay, M., see Nicklin, P. **83**, 197; **123**, 187
MacKellar, A.J., G. Buckton, J.M. Newton, B.Z. Chowdhry, C.A. Orr, The controlled crystallisation of a model powder. 1. The effects of altering the stirring rate and the supersaturation profile, and the incorporation of a surfactant (Poloxamer 188) **112**, 65
MacKellar, A.J., G. Buckton, J.M. Newton, C.A. Orr, The controlled crystallisation of a model powder. 2. Investigation into the mechanism of action of poloxamers in changing crystal properties **112**, 79
MacKellar, A.J., see Buckton, G. **117**, 253
MacLean, D.S., J.D. Robertson, E. Moody, D.J. Stalker, M. Jay, Secondary X-ray fluorescence for in vivo transdermal absorption measurements **131**, 243
Macura, A.B., see Knapczyk, J. **80**, 33
Madeira, V.M.C., see Moreira, J.N. **147**, 153
Mader, R.M., see Mader, R.M. **68**, 151
Mader, R.M., B. Rizovski, G.G. Steger, K. Moser, H. Rainer, C. Dittrich, Permeability of latex membranes to anti-cancer drugs **68**, 151
Maderich, A.B., E.T. Sugita, Absorption enhancement of dextran sulfate after enteral administration in a dispersion **137**, 85
Madhan, Kumar A.B., Rao K. Panduranga, Poly(palmitoyl-L-hydroxyproline ester) microspheres as potential oral controlled drug delivery system **149**, 107

Madu, D., see Adeyeye, C.M. **116**, 65
Maeda, T., see Murata, Y. **96**, 139
Maeda, T., see Yamahira, Y. **3**, 23
Maffione, G., see Gazzaniga, A. **108**, 77
Magai, T., see Maitani, Y. **64**, 89
Maganti, L., M. Celik, Compaction studies on pellets. I. Uncoated pellets **95**, 29
Compaction studies on pellets. II. Coated pellets **103**, 55
Magarotto, L., see Carli, F. **33**, 115
Magdassi, S., A. Siman-Tov, Formation and stabilization of perfluorocarbon emulsions **59**, 69
Magdassi, S., M. Royz, S. Shoshan, Interactions between collagen and perfluorocarbon emulsions **88**, 171
Magenheim, B., M.Y. Levy, S. Benita, A new in vitro technique for the evaluation of drug release profile from colloidal carriers—Ultrafiltration technique at low pressure **94**, 115
Maggi, L., U. Conte, P. Giunchedi, P. Colombo, Press-coated tablets for the sequential pulsed administration of two different drugs **99**, 173
Maggi, L., G. Massolini, Lorenzi E. De, U. Conte, G. Caccialanza, Evaluation of stereoselective dissolution of verapamil hydrochloride from matrix tablets press-coated with chiral excipients **136**, 43
Maggi, L., M.L. Torre, P. Giunchedi, U. Conte, Supramicellar solutions of sodium dodecyl sulphate as dissolution media to study the in vitro release characteristics of sustained-release formulations containing an insoluble drug: Nifedipine **135**, 73
Maggi, L., see Colombo, P. **63**, 43; **88**, 99
Maggi, L., see Giunchedi, P. **77**, 177; **85**, 141; **94**, 15; **130**, 41
Magill, A., see Powell, M.F. **35**, 61; **44**, 225
Magnard, F., see Armand, J.Y. **40**, 33
Magne, D., see Penaud, J.F. **142**, 1
Magnus, A.D., see Meyer, E. **41**, 63
Magnusson, A.B., see Nicklasson, M. **69**, 255
Magnusson, C., see Buur, A. **129**, 223
Magnusson, C., see Toth, I. **102**, 223
Magron, P., T.F. Patton, J.L. Taverdet, J.M. Vergnaud, Spherical oral polymer—Drug device with two polymers for constant drug delivery **38**, 91
Mahjour, .M., B.E. Mauser, M.B. Fawzi, Skin permeation enhancement effects of linoleic acid and Azone on narcotic analgesics **56**, 1
Mahjour, M., B.E. Mauser, Z.A. Rashidbaigi, M.B. Fawzi, Effects of propylene glycol diesters of caprylic and capric acids (Miglyol^{R} 840) and ethanol binary systems on in vitro skin permeation of drugs **95**, 161
Mahjour, M., see Senderoff, R.I. **83**, 65
Mahmoud, H., see Ghanem, A.H. **78**, 137
Mahmoud, H., see Kim, Y.H. **80**, 17
Mahmoud, M.J., see Ezzedeen, F.W. **59**, 255
Mahmound, H.A., M.S. El-Samaligy, Optimization of controlled drug release through interfacial polymerization **25**, 121
Mahrag Tur, K., H.S. Ch'ng, S. Baie, Use of bioadhesive polymer to improve the bioavailability of griseofulvin **148**, 63
Mahrous, G.M., see Al-Angary, A.A. **65**, R5
Maibach, H., see Nangia, A. **99**, 67
Maibach, H.I., see Gummer, C.L. **40**, 101
Maibach, H.I., see Guy, R.H. **11**, 119; **17**, 23
Maibach, H.I., see Kubota, K. **96**, 105
Maibach, H.I., see Leopold, C.S. **139**, 161
Maibach, H.I., see Melendres, J.L. **92**, 243
Maibach, H.I., see Montenegro, L. **140**, 51
Maibach, H.I., see Quan, D. **104**, 61
Maibach, H.I., see Shah, V.P. **74**, 1
Maibach, H.I., see Singh, P. **101**, R5
Maibach, H.I., see Sznitowska, M. **99**, 43; **123**, 41
Maibach, H.J., see Patil, S.M. **110**, 147
Maier, G.A., A. Poliszczuk, H.L. Fung, Stability of nitroglycerin in human and rat plasma **4**, 75
Maillot, A., see Bourget, P. **146**, 167
Mailloux, A.T., see Mitra, A.K. **32**, 133
Maincent, P., see Alonso, M.J. **42**, 27
Maincent, P., see Benoit, E. **84**, 283
Maincent, P., see Chevin, J.C. **45**, 101
Maincent, P., see Kedzierewicz, F. **58**, 221; **94**, 69; **117**, 247
Maincent, P., see Marchal-Heussler, L. **58**, 115
Maincent, P., see Veiga, F. **129**, 63
Maincent, P., see Zimmer, A.K. **110**, 211
Maincent, P., see Zinutti, C. **133**, 97
Maincent, P.D., see Benoit, E.F. **118**, 121
Mainwaring, D.E., see Doherty, M.M. **111**, 205
Maire, G., see Gehenot, A. **45**, 13
Maitani, Y., K. Ishigaki, K. Takayama, T. Nagai, In vitro nasal transport across rabbit mucosa: Effect of oxygen bubbling, pH and hypertonic pressure on permeability of lucifer yellow, diazepam and 17 β-estradiol **146**, 11
Maitani, Y., Y. Machida, T. Nagai, Influence of molecular weight and charge on nasal absorption of destran and DEAE-dextran in rabbits **49**, 23
Maitani, Y., M. Nakagaki, T. Magai, Surface potential of liposomes with entrapped insulin **64**, 89
Determination of the acid dissociation constants in ethanol-water mixtures and partition coefficients for diclofenac **74**, 105
Maitani, Y., N. Uchida, M. Nakagaki, T. Nagai, Effect of bile salts on the nasal mucosa: Membrane potential measurement **69**, 21
Maitani, Y., N. Uchida, M. Taniguchi, S. Yamazaki, M. Hara, K. Takayama, Y. Machida, T. Nagai, The inactivation process of fibroblast interferon-β in the preparation stage and in rabbit nasal absorption of mixed and freeze-dried powder dosage forms **64**, 139
Maitani, Y., T. Yamamoto, K. Takayama, T. Nagai, The effect of soybean-derived sterol and its glucoside as an enhancer of nasal absorption of insulin in rabbits in vitro and in vivo **117**, 129
Maitani, Y., T. Yamamoto, K. Takayama, N.A. Peppas, T. Nagai, A modelling analysis of drug absorption and administration from the ocular, naso-lacrimal duct, and nasal routes in rabbits **126**, 89

Maitani, Y., H. Sato, T. Nagai, Effect of ethanol on the true diffusion coefficient of diclofenac and its sodium salt in silicone membrane **113**, 165
Maitani, Y., see Coutel-Egros, A. **84**, 117
Maitani, Y., see Maitani, Y. **64**, 139
Maitani, Y., see Muramatsu, K. **107**, 1; **124**, 19
Maitani, Y., see Obata, Y. **89**, 191
Maitani, Y., see Qi, X. **114**, 33
Maitani, Y., see Qi, X.R. **146**, 31
Maitani, Y., see Uchida, N. **74**, 95
Majeed, I.A., see Al-Turk, W.A. **41**, 227
Majeed, S.H., see Ezzedeen, F.W. **59**, 255
Majilat, P., P. Helboe, Kristensen Kjaervig, Separation and quantitative determination of homatropine methylbromide and opium alkaloids in admixture in pharmaceutical preparations by gas-liquid and high-performance liquid chromatography **9**, 245
Mak, E., see Illum, L. **30**, 17
Mak, V.H.F., see Biddiscombe, M.F. **91**, 111
Makarla, K.R., see Betageri, G.V. **126**, 155
Maki, T., see Akimoto, M. **100**, 133; **117**, 31
Maki, T., see Yamada, K. **119**, 1
Makki, S., P. Muret, A.M. Said, P. Bassignot, P. Humbert, P. Agache, J. Millet, Percutaneous absorption of three psoralens commonly used in therapy: Effect of skin occlusion (in vitro study) **133**, 245
Makohon, D.J., see Barnes, A.R. **92**, 233
Malakh, A., see Najib, N.M. **32**, 229
Malamataris, S., A. Avgerinos, Controlled release indomethacin microspheres prepared by using an emulsion solvent-diffusion technique **62**, 105
Malamataris, S., D. Ganderton, Sustained release from matrix system comprising hydrophobic and hydrophilic (gel-forming) parts **70**, 69
Malamataris, S., P. Goidas, A. Dimitriou, Moisture sorption and tensile strength of some tableted direct compression excipients **68**, 51
Malamataris, S., Th. Hatjichristos, J.E. Rees, Apparent compressive elastic modulus and strength isotropy of compacts formed from binary powder mixes **141**, 101
Malamataris, S., T. Karidas, Effect of particle size and sorbed moisture on the tensile strength of some tableted hydroxypropyl methylcellulose (HPMC) polymers **104**, 115
Malamataris, S., T. Karidas, P. Goidas, Effect of particle size and sorbed moisture on the compression behaviour of some hydroxypropyl methylcellulose (HPMC) polymers **103**, 205
Malamataris, S., J.E. Rees, Viscoelastic properties of some pharmaceutical powders compared using creep compliance, extended Heckel analysis and tablet strength measurements **92**, 123
Malamataris, S., see Avgerinos, A. **63**, 77
Malamataris, S., see Pilpel, N. **68**, 157
Malhi, J.S., A.J. High, G.W. Hanlon, P.R. Gard, C. Marriott, Potentiation of oxytetracycline antimicrobial activity in vivo by concurrent administration of bromhexine **43**, 77
Malhotra, A.K., see Bhise, S.B. **24**, 297
Malick, A.W., see Felton, L.A. **127**, 203
Malick, A.W., see Shah, N.H. **106**, 15
Malkhazov, L.B., see Feldstein, M.M. **131**, 229
Malkin, J.E., see Vincent-Ballereau, F. **58**, 25
Malkki, L., S. Tammilehto, Decomposition of salbutamol in aqueous solutions. I. The effect of pH, temperature and drug concentration **63**, 17
Malkki-Laine, L., K. Purra, K. Kahkonen, S. Tammilehto, Decomposition of salbutamol in aqueous solutions. II. The effect of buffer species, pH, buffer concentration and antioxidants **117**, 189
Malkowska, S., see Davis, S.S. **49**, 183
Mall, S., G. Buckton, D.A. Rawlins, Slower dissolution rates of sulphamerazine in aqueous sodium dodecyl sulphate solutions than in water **131**, 41
Mallesch, J.L., see Moore, D.E. **76**, 187
Mallevais, M.L., see Dine, T. **77**, 279
Mallevais, M.L., see Faouzi, M.A. **89**, 125
Mallinson, C.B., see Barnwell, S.G. **88**, 423
Mallinson, C.B., see Higginbottom, S. **109**, 173
Mallon, C., see Attwood, D. **84**, R5; **88**, 417
Malmusi, L., see Vandelli, M.A. **118**, 77
Maltby, S., see Gumblteon, M. **38**, 261
Manabe, E., K. Sugibayashi, Y. Morimoto, Analysis of skin penetration enhancing effect of drugs by ethanol-water mixed systems with hydrodynamic pore theory **129**, 211
Mandal, T.K., S. Tenjarla, Preparation of biodegradable microcapsules of zidovudine using solvent evaporation: Effect of the modification of aqueous phase **137**, 187
Mandella, W., see Menon, A. **140**, 207
Manderioli, A., see Mura, P. **119**, 71
Mandlekar, S.V., S.S. Marathe, P.V. Devarajan, A novel raft-forming antacid suspension using a natural dietary fibre **148**, 117
Maness, D.D., see Cruz, J.E. **2**, 275
Manger, J., see Lashmar, U.T. **111**, 73
Manich, A.M., see Coderch, L. **139**, 197
Mannermaa, J.P., J. Raisanen, M. Hyvonen-Dabek, E. Spring, J. Yliruusi, Use of proton-induced X-ray emission (PIXE) analysis in the evaluation of large volume parenteral rubber stoppers **103**, 125
Manning, M.C., see Foster, T.M. **134**, 193
Manning, M.C., see Powers, M.E. **108**, 49
Manning, R., see Washington, C. **77**, 57
Manning, R.J., C. Washington, Chemical stability of total parenteral nutrition mixtures **81**, 1
Mannova, M., see Engel, A. **107**, 99
Manolova, N., see Dimitrova, E. **93**, 21
Mansson, P., see Olsson, B. **119**, 223
Mao, F., see Lu, D.R. **129**, 243
Marathe, S.S., see Mandlekar, S.V. **148**, 117
Marburger, C., see Langer, K. **137**, 67
Marcello, J., see Smith, P.L. **106**, 95
Marchal-Heussler, L., P. Maincent, M. Hoffman, J. Spittler, P. Couvreur, Antiglaucomatous activity of betaxolol chlorhydrate sorbed onto different isobutylcyanoacrylate nanoparticle preparations **58**, 115

Marengo, E., see Pattarino, F. **91**, 157
Margarit, M.V., I.C. Rodriguez, A. Cerezo, Myrj 51 as a suppository excipient: Influence on pharmaceutical availability and bioavailability of sodium valproate **81**, 67
Physical characteristics and dissolution kinetics of solid dispersions of ketoprofen and polyethylene glycol 6000 **108**, 101
Margarit, M.V., see Fernandez, M. **84**, 197; **98**, 29
Marignan, R., see Audran, M. **15**, 273
Marino, E.L., A. Dominguez-Gil, Determination of the macro- and micro-ionization constants of a dipolar zwitterionic cephalosporin: Cefadroxil **8**, 25
Study of accelerated inactivation of cefadroxil **12**, 209
Marino, E.L., A. Dominguez-Gil, J.M. Tabernero, F. Rubio, Pharmacokinetics of cefadroxil in patients with terminal renal impairment **9**, 263
Marino, E.L., M.T. Vicente, Gil A. Dominguez, Influence of cholestyramine on the pharmacokinetic parameters of cefadroxil after simultaneous administration **16**, 23
Marino, E.L., see Alonso, I.G. **19**, 345
Marino, E.L., see Modamio, P. **107**, 243; **112**, 93; **130**, 137
Marino, E.L., see Pedraz, J.L. **25**, 147
Marino, E.L., see Rodriguez, Barbero J. **19**, 199
Marino, M., see Dey, M. **49**, 121
Marjukka Turunen, T., S. Buyuktimkin, N. Buyuktimkin, A. Urtti, P. Paronen, J.H. Rytting, Enhanced delivery of 5-fluorouracil through shed snake skin by two new transdermal penetration enhancers **92**, 89
Markantonis, S.L., see Kyroudis, A. **43**, 145
Marks, G.J., see Hidalgo, I.J. **98**, 83
Markuszewski, M., see Kaliszan, R. **145**, 9
Marolt-Gomiscek, M., see Sher, A. **99**, 119; **148**, 191
Marolt-Gomiscek, M.M., see Sher, A. **90**, 181
Marom, G., see Benita, S. **33**, 71
Marques, M.L., see Asuero, A.G. **40**, 43
Marriott, C., see Gizurarson, S. **65**, 243
Marriott, C., see Hollingsbee, D.A. **77**, 199
Marriott, C., see Kearney, P. **28**, 33; **35**, 211
Marriott, C., see Lee, K.C. **130**, 103
Marriott, C., see Malhi, J.S. **43**, 77
Marriott, C., see Martin, G.P. **44**, 57
Marriott, C., see Pritchard, K. **129**, 137
Marriott, C., see Queen, D. **44**, 25
Marriott, C., see Timsina, M.P. **101**, 1
Marriott, C., see Waring, M.J. **94**, 211
Marriott, C., see Wilkins, K.M. **57**, 1; **58**, 165
Marriott, C., see Wilson, C.G. **38**, 221
Marriott, C., see Zeng, X.M. **107**, 205; **109**, 135; **124**, 149
Marriott, R.J., see Biddiscombe, M.F. **91**, 111
Marrot, D., see Bonhomme-Faivre, L. **134**, 99
Marsh, K., see Patel, J.P. **65**, 195
Marshall, K., see Cropp, J.W. **71**, 127
Marshall, P.V., P. York, Crystallisation solvent induced solid-state and particulate modifications of nitrofurantoin **55**, 257
The compaction properties of nitrofurantoin samples crystallised from different solvents **67**, 59
Marszall, L., E. Utsumi, Effect of choline derivatives on the cloud point of mixed ionic-non-ionic surfactants **39**, 263
Martel, P., F. Pinguet, I. Petit, S. Poujol, C. Astre, M. Fabbro, Stability of the principal cytostatic agents during storage at unusual temperatures **149**, 37
Marti, A., see Montero, M.T. **96**, 157
Martin, B., O. Watts, B. Shroot, J.C. Jamoulle, A new diffusion cell—An automated method for measuring the pharmaceutical availability of topical dosage forms **49**, 63
Martin, C., see Arellano, A. **130**, 141
Martin, C., see Santoyo, S. **117**, 219
Martin, F., see Lucero, M.J. **104**, 49; **127**, 73
Martin, F.J., see Woodle, M.C. **88**, 327
Martin, G., see Roberts, M.S. **2**, 295
Martin, G.P., A.E. Bell, C. Marriott, An in vitro method for assessing particle deposition from metered pressurised aerosols and dry powder inhalers **44**, 57
Martin, G.P., see Bilbruck, J. **99**, 293
Martin, G.P., see Gizurarson, S. **65**, 243
Martin, G.P., see Hollingsbee, D.A. **77**, 199
Martin, G.P., see Lloyd, A.W. **106**, 255
Martin, G.P., see Pritchard, K. **129**, 137
Martin, G.P., see Queen, D. **44**, 25
Martin, G.P., see Timsina, M.P. **101**, 1
Martin, G.P., see Waring, M.J. **94**, 211
Martin, G.P., see Wilkins, K.M. **58**, 165
Martin, G.P., see Wilkins, K.M. **57**, 1
Martin, G.P., see Zeng, X.M. **107**, 205; **109**, 135; **124**, 149
Martin, H., see Allwood, M.C. **127**, 65
Martin, J.A., see Chevin, J.C. **45**, 101
Martin, R.J., D. Bourel, J. Hadgraft, Skin metabolism of topically applied compounds **39**, 23
Martin-Algarra, V., R.M. Pascual-Costa, M. Merino, V.G. Casabo, Effects of polysorbate 80 on amiodarone intestinal absorption in the rat **122**, 1
Martin-Villodre, A., see Bermejo, M.V. **69**, 221
Martin-Villodre, A., see Collado, E.F. **44**, 187
Martin-Villodre, A., see Diez-Sales, O. **100**, 1
Martin-Villodre, A., see Garrigues Pelufo, T.M. **79**, 135
Martin-Villodre, A., see Garrigues, T.M. **57**, 189; **64**, 127
Martinez, D., see Postaire, E. **55**, 99
Martinez-Pacheco, R., J.L. Vila-Jato, A. Concheiro, C. Souto, C.M. Losa, T. Ramos, Effect of compression force on biopharmaceutical characteristics of Eudragit RS-based cephalexin tablets **47**, 37
Martinez-Pacheco, R., J.L. Vila-Jato, C. Souto, T. Ramos, Controlled release of cephalexin from double-layer tablets containing small proportions of acrylic resins **32**, 99
Martinez-Pacheco, R., see Doldan, C. **124**, 69
Martinez-Pacheco, R., see Landin, M. **91**, 123, 133, 143; **103**, 9; **107**, 247; **109**, 1; **123**, 143
Martinez-Pacheco, R., see Martinez-Pacheco, R. **47**, 37
Martinez-Pacheco, R., see Mosquera, M.J. **135**, 147
Martinez-Pacheco, R., see Perez-Marcos, B. **67**, 113; **100**, 207
Martini, A., R. Artico, P. Civaroli, L. Muggetti, R. de Ponti, Critical micellar concentration shifting as a simple tool for evaluating cyclodextrin–enhancer interactions **127**, 239

Martini, A., C. Torricelli, R. de Ponti, Physico-pharmaceutical characteristics of steroid/crosslinked polyvinylpyrrolidine coground systems **75**, 141
Martini, A., see Muggetti, L. **136**, 81
Martini, A., see Torricelli, C. **71**, 19; **75**, 147
Martini, L., D. Attwood, J.H. Collett, A. D'Emanuele, The bioadhesive properties of a triblock copolymer of ε-caprolactone and ethylene oxide **113**, 223
Martino, P.D., A.-M. Guyot-Hermann, P. Conflant, M. Drache, J.-C. Guyot, A new pure paracetamol for direct compression: the orthorhombic form **128**, 1
Martins, M.B.F., S.I.D. Simoes, A. Supico, M.E.M. Cruz, R. Gaspar, Enzyme-loaded PIBCA nanoparticles (SOD and L-ASNase): Optimization and characterization **142**, 75
Marttin, E., J.C. Verhoef, S.G. Romeijn, P. Zwart, F.W.H.M. Merkus, Acute histopathological effects of benzalkonium chloride and absorption enhancers on rat nasal epithelium in vivo **141**, 151
Marttin, E., see Romeijn, S.G. **135**, 137
Martuccio, C., see Di, Giulio A. **144**, 27
Marty, J.P., see Bamba, M. **2**, 307
Marty, J.P., see Fessi, H. **1**, 265; **30**, 209
Marty, J.P., see Ferreira, L.A.M. **109**, 251; **121**, 169
Marty, J.P., see Garcia, B. **4**, 205
Maruyama, K., S. Okuizumi, O. Ishida, H. Yamauchi, H. Kikuchi, M. Iwatsuru, Phosphatidyl polyglycerols prolong liposome circulation in vivo **111**, 103
Maruyama, K., see Unezaki, S. **126**, 41; **144**, 11
Maruyama, K., see Watanabe, Y. **110**, 93
Maruyama, T., M. Otagiri, S.G. Schulman, Binding characteristics of coumarin anticoagulants to human α_1-acid glycoprotein and human serum albumin **59**, 137
Maruyama, Y., see Sugito, K. **60**, 89
Marvla, M., see Sirkia, T. **107**, 179
Marvola, M., A.M. Hannula, E. Westermarck, I. Happonen, T. Kopra, Disintegration of hard gelatin capsule formulations in the dog stomach—A radiological study **44**, 159
Marvola, M., A. Kannikoski, H. Aito, S. Nykanen, The effect of food on gastrointestinal transit and drug absorption of a multiparticular sustained-release verapamil formulation **53**, 145
Marvola, M., J. Pirjola, A. Huikari, The effect of some viscosity-enhancing agents on the intestinal absorption of sulfafurazole in the rat **3**, 13
Marvola, M., see Heinamaki, J. **42**, 105; **48**, 51
Marvola, M., see Laakso, R. **19**, 35
Marvola, M., see Veski, P. **111**, 171
Marwaha, S.B., M.H. Rubinstein, Structure-lubricity evaluation of magnesium stearate **43**, 249
Marz, F., R. Scheer, E. Graf, Microbiological aspects in the production of water for injection by reverse osmosis **58**, 155
Use of a laser diffraction particle counter to monitor integrity of reverse osmosis membranes and its inefficiency to detect bacteria in product water **61**, 57
Masada, M., see Araki, M. **81**, 39
Masada, T., W.I. Higuchi, V. Srinivasan, U. Rohr, J. Fox, C. Behl, S. Pons, Examination of iontophoretic transport of ionic drugs across skin: Baseline studies with the four-electrode system **49**, 57
Masada, T., see Masada, T. **49**, 57
Masaki, K., see Akazawa, M. **50**, 53
Mascagni, P., see Toth, I. **68**, 191
Maschek, I., see Dam, A.M. **121**, 245
Masegi, M., see Masegi, M. **78**, 103
Masegi, M., N. Wakabayashi, K. Sagara, Y. Teramura, A. Yamamoto, Y. Takakura, M. Hashida, H. Sezaki, Permeability of macromolecules in cultured endothelial and tumor cell layers on microporous membranes **78**, 103
Mashimo, T., see Asano, M. **67**, 67
Masilungan, F.C., N.G. Lordi, Evaluation of film coating compositions by thermomechanical analysis. I. Penetration mode **20**, 295
Masinde, L.E., A.J. Hickey, Aerosolized aqueous suspensions of poly(L-lactic acid) microspheres **100**, 123
Maskiewicz, R., see Lidgate, D.M. **96**, 51
Mason, W.D., J.D. Conklin, F.J. Hailey, Relative bioavailability of four macrocrystalline nitrofurantoin capsules **36**, 105
Mason, W.D., see Melethil, S. **31**, 83
Massaccesi, M., see Scalia, S. **82**, 179
Massari, C., see Leomy, C. **115**, 235
Massart, D.L., see Baert, L. **96**, 225
Massart, D.L., see Inghelbrecht, S. **148**, 103
Massart, L., see Vervaet, C. **106**, 157
Masse, J., see El Moussaoui, A. **106**, 109
Masse, J., see Lheritier, J. **123**, 273
Massimo, G., see am Ende, M.T. **120**, 33
Massimo, G., see Santi, P. **92**, 23
Massolini, G., see Maggi, L. **136**, 43
Masson, V., F. Maurin, J.P. Devissaguet, H. Fessi, Stability of poly (ε-caprolacton) nanospheres in sterile aqueous media **139**, 113
Masuda, M., see Ohta, N. **124**, 165
Masuda, Q.N., see Irwin, W.J. **21**, 35
Matar, K.M., see Al-Hassan, M.I. **50**, 35; **93**, 7
Matar, S., see Touitou, E. **103**, 199
Matchet, L., J. Pinton, F. Patat, B. Arbeille, L. Pourcelot, L. Vaillant, In vitro phonophoresis of digoxin across hairless mice and human skin: Thermal effect of ultrasound **133**, 39
Mathe, G., see Leomy, C. **115**, 235
Matheson Jr., L.E., D.E. Wurster, J.A. Ostrenga, Sarin transport across excised human skin. III. The effect on solvent pretreatment on adsorption and desorption by keratin **4**, 309
Matheson, L.E., Y. Chen, A quantitative structure-transportability relationship for the release of a series of substituted benzenes and pyridines from a planar polydimethylsiloxane matrix **125**, 297
Matheson, L.E., P. Vayumhasuwan, D.M. Moeckly, The development of a predictive method for the estimation of flux through polydimethylsiloxane membranes. II. Derivation of a diffusion parameter and its application to multisubstituted benzenes **77**, 163
Matheson, L.E., see Chen, Y. **94**, 81, 153; **137**, 149
Matheson, L.E., see Moeckly, D.M. **77**, 151

Mathews, S., M.N.A. Rao, Interaction of curcumin with glutathione **76**, 257
Mathiesen, C., see Schaefer, T. **134**, 105; **139**, 125, 139
Mathieu, M.C., see Bonhomme, L. **80**, 271
Matsuda, Y., E. Tatsumi, Physicochemical characterization of furosemide modifications **60**, 11
Matsuda, Y., R. Teraoka, Improvement of the photostability of ubidecarenone microcapsules by incorporating fat-soluble vitamins **26**, 289
Matsuda, Y., R. Teraoka, I. Sugimoto, Comparative evaluation of photostability of solid-state nifedipine under ordinary and intensive light irradiation conditions **54**, 211
Matsuda, Y., see Teraoka, R. **93**, 85
Matsui, M., see Miyamoto, Y. **149**, 25
Matsukawa, Y., see Sasaki, H. **36**, 147
Matsumaru, H., see Tsuchiya, S. **14**, 279
Matsumoto, H., see Imai, T. **53**, 7
Matsumoto, K., Y. Nakai, E. Yonemochi, T. Oguchi, K. Yamamoto, Physicochemical characteristics of porous crystalline cellulose and formation of an amorphous state of ethenzamide by mixing **108**, 167
Matsumoto, K., see Oguchi, T. **113**, 97
Matsumoto, M., see Fujii, M. **137**, 117
Matsumoto, M., see Watanabe, Y. **64**, 147; **99**, 337
Matsumoto, S., see Roos, C.F. **22**, 75
Matsumoto, S., see Takakura, Y. **37**, 135
Matsumoto, Y., see Watanabe, Y. **99**, 337; **110**, 93
Matsuo, H., see Yamashita, C. **70**, 225
Matsuo, M., K. Arimori, C. Nakamura, M. Nakano, Delayed-release tablets using hydroxyethylcellulose as a gel-forming matrix **138**, 225
Matsuo, M., see Loftsson, T. **22**, 345
Matsuo, T., see Ohta, N. **124**, 165
Matsuyama, K., see Nakashima, M. **72**, R5
Matsuzawa, T., see Hirai, S. **7**, 317
Mattern, C., see Su, X.Y. **123**, 47
Mattha, A.G., S.M. Omar, M.A. Kassem, Study of the influence of sodium taurocholate (STC) and sodium glycocholate (SGC) on the mass transfer of certain drugs. Diethylstilbestrol **11**, 27
Mattha, A.G., see Kassem, M.A. **12**, 1
Matthews, G.P., N. Lowther, M.J. Shott, Crystallisation of carbamazepine in tablets stored at elevated temperatures **50**, 111
Matthews, M.L., see Mitra, A.K. **23**, 185
Matuszewska, B., L. Hettrick, J.V. Bondi, D.E. Storey, Comparative bioavailabilily of L-683,453, a 5α-reductase inhibitor, from a self-emulsifying drug delivery system in Beagle dogs **136**, 147
Matzek, K.M., T.R. MacGregor, J.J. Keirns et al., Oral pharmacokinetics of pirenzepine in man following single and multiple doses **28**, 85
Matzek, K.M., T.R. MacGregor, J.J. Keirns, M. Vinocur, Effect of food and antacids on the oral absorption of pirenzepine in man **28**, 151
Maul, K.A., P.C. Schmidt, Influence of different-shaped pigments on bisacodyl release from Eudragit L 30 D **118**, 103
Mauriac, P., see Abi, Khalil F. **140**, 1
Maurice, J., see Delalonde, M. **146**, 159
Maurin, F., see Masson, V. **139**, 113
Maurin, M.B., R.D. Vickery, R.F. Carney, M.A. Hussain, Physicochemical properties of a nonpeptide cyclic urea HIV protease inhibitor (DMP 323) **144**, 99
Maurin, M.B., R.D. Vickery, C.A. Gerard, M. Hussain, Solubility of ionization behavior of the antifungal α-(2,4-difluor ophenyl)-α-[(1-(2-(2-pyridyl)phenylethenyl)]-1H-1,2,4triazole-1-ethanol bismesylate (XD405) **94**, 11
Maurin, M.B., see Hussain, M.A. **117**, 181
Maurizi, G., see Di, Giulio A. **74**, 183
Maury, L., see Rambaud, J. **15**, 199
Maury, M., see Fawaz, F. **132**, 271
Maury, M., see Guyot, M. **144**, 209
Mauser, B.E., see Mahjour, M. **56**, 1; **95**, 161
Mawhinney, W.M., see Jones, D.S. **131**, 83
May, H.A., C.G. Wilson, R.R. Niederer, H.W. Zulliger, Monitoring radiolabelled antacid preparations in the stomach **19**, 169
Mayer, J.M., see Gryllaki, M. **51**, 95
Mayer, J.M., see McCafferty, D.F. **38**, 59
Mayer, J.M., see Rothlisberger, T. **44**, 141
Mayer, J.M., see Wernly-Chung, G.N. **63**, 129
Mayor, S.H., L. Illum, Investigation of the effect of anaesthesia on nasal absorption of insulin in rats **149**, 123
Mayorga, P., F. Puisieux, G. Couarraze, Formulation study of a transdermal delivery system of primaquine **132**, 71
Maysinger, D., I. Jalsenjak, In situ absorption and in vitro release of microencapsulated cimetidine **17**, 129
Maysinger, D., B. Zorc, I. Butula, I. Jalsenjak, Preparation and in vivo effect of microencapsulated cholinotoxin **63**, 149
Maysinger, D., see Filipovic-Grcic, J. **116**, 39
Maysinger, D., see Zorc, B. **99**, 135; **123**, 65
Mazer, N.A., see Hovgaard, L. **132**, 107
Mazer, N.A., see Imanidis, G. **120**, 41
Mazuel, C., see Rozier, A. **57**, 163
Mazuel, G., E.M. Cohen, M. Bastide, Standardization of in vitro dissolution assays **16**, 97
Mazur, J., see Nystrom, C. **10**, 209
Mazzanti, G., see Fabrizio, Saettone M. **142**, 103
Mazzco, P., see Carlucci, G. **102**, 271
Mazzei, N., see Dobetti, L. **128**, 277
Mazzeo, P., see Carlucci, G. **49**, 79; **53**, 257; **93**, 245
Mazzi, G., see Mulinacci, N. **90**, 35; **128**, 81
Mazzi, G., see Vincieri, F.F. **48**, 119
Mazzone, G., see Bonina, F.P. **106**, 115
Mazzone, G., see Castelli, F. **103**, 217
Mazzone, G., see Panico, A.M. **80**, 93; **98**, 19
Mazzone, G., see Puglisi, G. **118**, 65
Mazzone, S., see Panico, A.M. **98**, 19
Mbela, T.K.M., J.H. Poupaert, P. Dumont, Poly(diethylmethylidene malonate) nanoparticles as primaquine delivery system to liver **79**, 29

Mbela, T.K.M., A. Ludwig, I. Landau, E. Deharo, A. Haemers, Preparation, characterization and in vivo activity of mefloquine submicron emulsions **110**, 189
Mbela, T.K.M., J.H. Poupaert, P. Dumont, A. Haemers, Development of poly(dialkyl methylidenemalonate) nanoparticles as drug carriers **92**, 71
McCafferty, D.F., A.D. Woolfson, A.P. Launchbury, Stability of temazepam in parenteral formulations **31**, 9
McCafferty, D.F., J.M. Mayer, S.P. Gorman, L. Anderson, Chemical behaviour of noxythiolin in biological fluids **38**, 59
McCafferty, D.F., see Jones, D.S. **64**, R1
McCafferty, D.F., see Richardson, P.J. **78**, 189
McCafferty, D.F., see Woolfson, A.D. **34**, 17; **48**, 167; **49**, 135; **51**, 183; **62**, 9; **78**, 209; **84**, 69; **94**, 75
McCall, T.W., see Tobyn, M.J. **128**, 113
McCallion, O.N.M., M.J. Patel, Viscosity effects on nebulisation of aqueous solutions **130**, 245
McCallion, O.N.M., K.M.G. Taylor, P.A. Bridges, M. Thomas, A.J. Taylor, Jet nebulisers for pulmonary drug delivery **130**, 1
McCallion, O.N.M., K.M.G. Taylor, M. Thomas, A.J. Taylor, The influence of surface tension on aerosols produced by medical nebulisers **129**, 123
Nebulisation of monodisperse latex sphere suspensions in air-jet and ultrasonic nebulisers **133**, 203
McCallion, R., A.L.W. Po, Modelling transepidermal water loss under steady-state and non-steady-state relative humidities **105**, 103
In vivo evaluation of the effects of moisturisers on transepidermal water loss using factorial designs **113**, 247
McCarthy, D.E., see Jani, P.U. **84**, 245; **86**, 239; **105**, 157
McCarthy, T.M., see Vincke, B.J. **54**, 181
McCaul, F., see Ford, J.L. **40**, 223
McCauley, J.A., see Ghodbane, S. **59**, 281
McClean, E., P.S. Collier, J.P.H. Fee, The effect of formulation on the plasma binding and blood/plasma concentration ratio of diazepam **60**, 35
McClintock, S.A., see Kwong, E.C. **103**, 259
McColm, A.M., see Toth, I. **77**, 13
McConnell, W.R., see Cefali, E.A. **127**, 147
McCormack, B., G. Gregoriadis, Drugs-in-cyclodextrins-in liposomes: A novel concept in drug delivery **112**, 249
McCornack, J., see Bodor, N. **35**, 47
McCrossin, L.E., see Charman, W.N. **88**, 397
McCullough, B., see Shah, V.P. **125**, 99
McDaid, D.M., P.B. Deasy, Formulation development of a transdermal drug delivery system for amlodipine base **133**, 71
McDonald, C., see Shija, R. **80**, 203
McDowall, R.D., see Shah, V.P. **82**, 1
McElnay, J.C., P.F. D'Arcy, Sites and mechanisms of drug interactions. I. In vitro, intestinal and metabolic interactions **5**, 167
Sites and mechanisms of drug interactions. II. Protein binding, renal excretion and pharmacodynamic interactions **6**, 205
Aluminium content of infusion and irrigation fluids **33**, 261
McElnay, J.C., P.F. D'Arcy, O. Throne, Effect of antacid constituents, kaolin and calcium citrate on phenytoin absorption **7**, 83
McElnay, J.C., D.S. Elliott, J. Cartwright-Shamoon, P.F. D'Arcy, Stability of methotrexate and vinblastine in burette administration sets **47**, 239
McElnay, J.C., D.S. Elliott, P.F. D'Arcy, Binding of human insulin to burette administration sets **36**, 199
McElnay, J.C., T.A. Kennedy, R. Harland, The influence of ultrasound on the percutaneous absorption of fluocinolone acetonide **40**, 105
McElnay, J.C., A.C. Nicol, The comparison of a novel continuous-flow dissolution apparatus for suppositories with the rotating basket technique **19**, 89
McElany, J.C., A.M. Sidahmed, P.F. D'Arcy, Experimental modelling of drug absorption and drug absorption interactions **31**, 107
McElnay, J.C., A.M. Sidahmed, P.F. D'Arcy, R.D. McQuade, Chloroquine–digoxin interaction **26**, 267
McElnay, J.C., A.J. Taggart, B. Kerr, P. Passmore, Comparison of serum indomethacin profiles after rectal administration of two brands of indomethacin suppositories in healthy volunteers subjects **33**, 195
McElnay, J.C., see Benson, H.A.E. **44**, 65
McElnay, J.C., see Cartwright-Shamoon, J.M. **42**, 41
McElnay, J.C., see Grainger-Rousseau, T.J. **54**, 1
McElnay, J.C., see Harron, D.W.G. **1**, 61
McElnay, J.C., see Yahya, A.M. **25**, 217; **31**, 65; **34**, 137
McEntegart, D.J., see Bratty, J.R. **115**, 119
McGarron, P.A., see Woolfson, A.D. **84**, 69
McGee, J.P., see O'Hagan, D.T. **119**, 251
McGilveray, I.J., see Shah, V.P. **82**, 1
McGinity, J.W., see Bodmeier, R. **43**, 179
McGinity, J.W., see Felton, L.A. **113**, 17
McGinity, J.W., see Gutierrez-Rocca, J.C. **103**, 293
McGinity, J.W., see Jenquin, M.R. **101**, 23
McGinity, J.W., see Obara, S. **126**, 1
McGinity, J.W., see Walkow, J.C. **35**, 91, 103
McGloughlin, R.M.R., O.I. Corrigan, Dissolution characteristics of benzoic acid–Tris mixtures **82**, 135
McGovern, J.G., C.P. Garvin, D.S. Jones, A.D. Woolfson, S.P. Gorman, Modification of biomaterial surface characteristics by body fluids in vitro **149**, 251
McGowan, K.E., see Woolfson, A.D. **51**, 183; **62**, 9; **78**, 209
McGown, E., see Riviere, J.E. **127**, 213
McHalsky, M., see Kipp, J.E. **34**, 1
McKeown, R.H., see Prankerd, R.J. **62**, 37; **83**, 25, 39; **112**, 1
McKillop, A.G., see Rowe, R.C. **91**, 247; **101**, 169
McLeod, A.D., D.R. Friend, T.N. Tozer, Synthesis and chemical stability of glucocorticoid-dextran esters: Potential prodrugs for colon-specific delivery **92**, 105
McMahon, F.G., R. Vargas, P. Leese, B. Crawford, A. Konecny, K. Nichols, M. Knowles, F. Theeuwes, D. Guinta, Controlled-release dosage forms and gastrointestinal blood loss: Four clinical studies **91**, 75

McMahon, J., see Patel, H.K. **25**, 13, 237
McMahon, J., see Rowe, R.C. **31**, 281
McMullan, J., see Billany, M.R. **121**, 107
McMullen, J.N., see Alain, M. **23**, 265
McMullen, J.N., see Bechard, S. **31**, 91; **32**, 71
McNamara, P.J., see Gibaldi, M. **2**, 167
McNicol, K.J., see Schreier, H. **87**, 183
McPartlin, M., see Payne, R.S. **145**, 165
McQuade, R.D., see McElnay, J.C. **26**, 267
McShane, J., see Kuu, W.Y. **124**, 241
McTaggart, C.M., J.A. Ganley, P.A. Hanson, S.E. Walker, The evaluation of formulation and processing conditions of a melt granulation process **19**, 139
McTaggart, L.E., G.W. Halbert, Assessment of polysaccharide gels as drug delivery vehicles **100**, 199
McTiernan, R.G., see Sloan, K.B. **44**, 87
McVie, J.G., see Burger, J.J. **23**, 333
Meakin, B.J., J.M. Cainey, P.M. Woodcock, Drug delivery characteristics of Bricanyl Turbohaler(TM) dry powder inhalers **119**, 91
Simulated 'in-use' and 'mis-use' aspects of the delivery of terbutaline sulphate from Bricanyl Turbohaler(TM) dry powder inhalers **119**, 103
Mecklenburg, S., see Givens, R.S. **29**, 67
Mediratta, D.K., see Singla, A.K. **60**, 27
Medlicott, N.J., D.S. Jones, I.G. Tucker, D. Holborow, Preliminary release studies of chlorhexidine (base and diacetate) from poly(ε-caprolactone) films prepared by solvent evaporation **84**, 85
Medlicott, N.J., I.G. Tucker, M.J. Rathbone, D.W. Holborow, D.S. Jones, Chlorhexidine release from poly (ε-caprolactone) films prepared by solvent evaporation **143**, 25
Medlicott, N.J., see Jones, D.S. **114**, 257
Meenhorst, P.L., see Underberg, W.J.M. **50**, 175
Megrab, N.A., A.C. Williams, B.W. Barry, Oestradiol permeation across human skin, silastic and snake skin membranes: The effects of ethanol/water co-solvent systems **116**, 101
Mehdizadeh, M., see Chow, K.Y. **20**, 3
Mehdizadeh, M., see Grant, D.J.W. **18**, 25
Mehta, A., see Carstensen, J.T. **1**, 65
Mehta, A.M., L.L. Augsburger, A preliminary study of the effect of slug hardness on drug dissolution from hard gelatin capsules filled on an automatic capsule-filling machine **7**, 327
Mehta, C., see Kearney, A.S. **92**, 63
Mehta, R.C., see Tsai, T. **127**, 31, 43
Mehta, S.C., G.W. Radebaugh, Targeted lymphatic transport and modified systemic distribution of CI-976 a lipophilic lipid-regulator drug, via a formulation approach **108**, 85
Mehta, S.C., see Kearney, A.S. **78**, 25; **92**, 63; **102**, 63; **104**, 169
Mehta, S.C., see Luner, P.E. **128**, 29
Mehta, S.C., see Zingerman, J.P. **88**, 303
Mehvar, S.R., F. Jamali, Dipyrone-ethylbiscoumacetate interaction in man **7**, 293
Meidan, V.M., A.D. Walmsley, W.J. Irwin, Phonophoresis—Is it a reality? **118**, 129
Meijer, D.K.F., see Franssen, E.J.F. **80**, R15
Meijer, D.K.F., see Moolenaar, F. **19**, 161; **33**, 249
Meijer, S., see Cremers, H.F.M. **120**, 51
Meinzer, A., see Haeberlin, B. **137**, 103
Meisner, D., J. Pringle, M. Mezei, Liposomal ophthalmic drug delivery. III. Pharmacodynamic and biodisposition studies of atropine **55**, 105
Meisner, D.R., see Thibert, R. **134**, 59
Melani, F., see Mulinacci, N. **90**, 35; **128**, 81
Melchor, R., see Biddiscombe, M.F. **91**, 111
Melchor, R., see Yuen, K.H. **97**, 61
Melendres, J.L., A. Nangia, L. Sedik, M. Hori, H.I. Maibach, Nonane enhances propranolol hydrochloride penetration in human skin **92**, 243
Melethil, S., W.D. Mason, Chang Chian-Jo, Dose-dependent absorption and excretion of vitamin C in humans **31**, 83
Melia, C.D., A.R. Rajabi-Siahboomi, A.C. Hodsdon, J. Adler, J.R. Mitchell, Structure and behaviour of hydrophilic matrix sustained release dosage forms. 1. The origin and mechanism of formation of gas bubbles in the hydrated surface layer **100**, 263
Melia, C.D., see Christrup, L.L. **66**, 169
Melia, C.D., see Davies, M.C. **62**, 97; **66**, 223
Melia, C.D., see Watts, P.J. **76**, 55; **98**, 63, 75; **102**, 101
Mellerio, G., see Bonferoni, M.C. **80**, 109
Melnik, E.I., see Kinsel, J.F. **12**, 97
Melnik, Y., see Bialer, M. **16**, 285
Meltzer, N.M., see Repta, A.J. **18**, 277
Memoli, A., L.G. Palermiti, V. Travagli, F. Alhaique, Egg and soya phospholipids—Sonication and dialysis: A study on liposome characterization **117**, 159
Memoli, A., see Touitou, E. **78**, 85; **133**, 85
Mendelow, A.Y., A. Forsyth, A.J. Baillie, A.T. Florence, In vitro release of nickel sulphate of varying particle size from paraffin bases **49**, 29
Mendenhall, D.W., see Zografi, G. **42**, 77
Menegatti, E., see Cortesi, R. **105**, 181; **139**, 69
Menegatti, E., see Esposito, E. **117**, 151; **129**, 263
Menegatti, E., see Nastruzzi, C. **90**, 43
Mengel, H., see Hansen, K.T. **79**, 205
Mengel, H.B., see Andersen, F.M. **21**, 51
Menon, A., N. Dhodi, W. Mandella, S. Chakrabarti, Identifying fluid-bed parameters affecting product variability **140**, 207
Menon, G., see Adjei, A. **90**, 141
Menon, M.R., see Bhalla, H.L. **17**, 351
Mentrup, E., see Michel, C. **84**, 93
Menut, C., see Bassani, V. **63**, 57
Menzies, A.R., H.E. Edwards, Some of the biological properties of factor X-fractionated heparin **37**, 65
Mereish, K.A., see Repta, A.J. **18**, 277
Merfeld, A.E., J.L. Haslam, G.S. Rork, An in vitro drug release rate method for lipoidal materials **69**, 63
Merfeld, A.E., see Haslam, J.L. **56**, 227
Merino, M., see Cejudo-Ferragud, E. **132**, 63

Merino, M., see Martin-Algarra, V. **122**, 1
Merino, M., see Ramos, A.J. **128**, 129
Merino, V., see Garrigues, T.M. **101**, 209
Merino, V., see Segura-Bono, M.J. **107**, 159
Merkel, G., see Podczeck, F. **97**, 15
Merkku, P., A.S. Lindqvist, K. Leiviska, J. Yliruusi, Influence of granulation and compression process variables on flow rate of granules and on tablet properties, with special reference to weight variation **102**, 117
Merkku, P., see Hellen, L. **96**, 197
Merkku, P., see Juslin, L. **123**, 247, 257
Merkku, P., see Murtoniemi, E. **108**, 155; **110**, 101
Merkle, H.P., see Anders, R. **49**, 231
Merkle, H.P., see Gander, B. **129**, 51
Merkle, H.P., see Lichtenberger, R. **64**, 117
Merkle, H.P., see Luessen, H.L. **141**, 39
Merkle, H.P., see Thomasin, C. **147**, 173
Merkus, F.W.H.M., see Duchateau, G.S.M.J.E. **31**, 193, 275; **34**, 131
Merkus, F.W.H.M., see Marttin, E. **141**, 151
Merkus, F.W.H.M., see Romeijn, S.G. **135**, 137
Merkus, F.W.H.M., see Schipper, N.G.M. **64**, 61
Merkus, F.W.H.M., see Van de Donk, H. **12**, 57, 67
Merry, J., K. Eberth, Ultrasonic preparation of pharmaceutical emulsions. Droplet size measurements by quasi-elastic light scattering **19**, 43
Merry, J., see Eberth, K. **14**, 349
Mersinger, L., see Hussain, M.A. **117**, 181
Merville, C., see Vincent-Ballereau, F. **50**, 87
Meseguer, G., R. Gurny, P. Buri, A. Rozier, B. Plazonnet, γ-Scintigraphic study of precorneal drainage and assessment of miotic response in rabbits of various ophthalmic formulations containing pilocarpine **95**, 229
Mesens, J., see Vollmer, U. **99**, 51
Meshali, M.M., K.E. Gabr, Effect of interpolymer complex formation of chitosan with pectin or acacia on the release behaviour of chlorpromazine HCl **89**, 177
Mesiha, M., F. Plakogiannis, S. Vejosoth, Enhanced oral absorption of insulin from desolvated fatty acid-sodium glycocholate emulsions **111**, 213
Mesiha, M., M. Sidhom, Increased oral absorption enhancement of insulin by medium viscosity hydroxypropyl cellulose **114**, 137
Mestiri, M., F. Puisieux, J.P. Benoit, Preparation and characterization of cisplatin-loaded polymethyl methacrylate microspheres **89**, 229
Mestres, C., F. Reig, M.A. Busquets, I. Haro, M.A. Alsina, Interactions of naloxone with lipid monolayers **76**, 145
Mestres, C., see Alsina, M.A. **70**, 111
Mestres, C., see Reig, F. **44**, 269
Meunier, H., see Desager, J.P. **31**, 179
Meunier, V., see Caro, I. **116**, 147
Meyer, E., A.D. Magnus, J.M. Haigh, I. Kanfer, Comparison of the blanching activities of Dermovate, Betnovate and Eumovate creams and ointments **41**, 63
Meyer, J.D., see Foster, T.M. **134**, 193
Meyer, M.C., see Barry, B.W. **2**, 1, 27
Meyler, P., see Moolenaar, F. **114**, 117
Mezei, M., see Meisner, D. **55**, 105
Mezei, M., see Singh, K. **16**, 339; **19**, 263
Mhando, J.R., A. Li Wan Po, Two-stage release of benzocaine from sunflower oil/gelatin emulsion films **59**, 165
Controlled release of local anaesthetic agents from liquid–solid emulsion gels **62**, 249
Mhando, J.R., see Li Wan Po, A. **20**, 87
Miah, Y., see Podczeck, F. **144**, 187
Micali, G., see Bonina, F.P. **98**, 93
Micali, N., see Panico, A.M. **98**, 19
Micali, N., see Villari, A. **127**, 185
Michel, C., N. Groth, T. Herrling, P. Rudolph, J. Fuchs, J. Kreuter, H.J. Freisleben, Penetration of spin-labeled retinoic acid from liposomal preparations into the skin of SKH1 hairless mice. Measurement by EPR tomography **98**, 131
Michel, C., T. Purmann, E. Mentrup, E. Seiller, J. Kreuter, Effect of liposomes on percutaneous penetration of lipophilic materials **84**, 93
Michelucci, J., see Dey, M. **49**, 121
Michniak, B.B., M.R. Player, J.M. Chapman Jr., J.W. Sowell Sr., In vitro evaluation of a series of Azone analogs as dermal penetration enhancers. I **91**, 85
Michniak, B.B., M.R. Player, L.C. Fuhrman, C.A. Christensen, J.M. Chapman Jr., J.W. Sowell Jr., In vitro evaluation of a series of Azone analogs as dermal penetration enhancers. III. Acyclic amides **110**, 231
In vitro evaluation of a series of Azone analogs as dermal penetration enhancers. II (Thio)amides **94**, 203
Michniak, B.B., M.R. Player, D.A. Godwin, C.A. Phillips, Sr. J.W. Sowell, In vitro evaluation of a series of azone analogs as dermal penetration enhancers. IV. Amines **116**, 201
Michniak, B.B., see Phillips, C.A. **125**, 63
Michniak-Mikolajczak, B.B., B.W. Barry, I.L. Waller, A microbalance technique for measuring water vapour uptake and release by stratum corneum in vitro **31**, 247
Michoel, A., see Hoa, N.T. **138**, 185
Michoel, A., see Talukdar, M.M. **129**, 233
Middleton, D.L., see Ghebre-Sellassie, I. **31**, 43
Middleton, K.R., see Amidon, G.E. **45**, 79
Midha, K.K., see Shah, V.P. **82**, 1
Mielck, J.B., see Frontini, R. **114**, 121
Mielck, J.B., see Krahn, F.U. **53**, 25
Miettunen, R., see Sutinen, R. **62**, 113
Miglani, B.D., see Bhaskar, R. **28**, 59
Miglani, B.D., see Mukherji, G. **50**, 15; **65**, 1
Miglietta, M.R., see Richardson, J.L. **115**, 9
Miglietta, M.R., see Rochira, M. **144**, 19
Mihailova, S., see Dimitrova, E. **93**, 21
Miinalainen, O., see Urtti, A. **23**, 147; **25**, 165
Mijnheer, F.W., see Herremans, J.L.M. **129**, 191
Mikawa, M., see Tahara, K. **99**, 311
Mikkelson, T., see Mitra, A.K. **10**, 219
Mikkelson, T.J., see Mitra, A.K. **37**, 19
Mikkelson, T.J., see Mosher, G.L. **2**, 239
Mikos, A.G., N.A. Peppas, Measurement of the surface tension of mucin solutions **53**, 1

Milburn, P., see Russell, A.D. **25**, 105
Milecki, J., see Pitha, J. **29**, 73
Miles, R.J., see Beezer, A.E. **45**, 153
Milic, J., see Tamburic, S. **137**, 243
Millan, M., see Caraballo, I. **139**, 177
Millar, F.C., O.I. Corrigan, Dissolution mechanism of ibuprofen-casein compacts **92**, 97
Millar, F.M., see Davidson, A.G. **63**, 29
Miller, E.S., see Hartauer, K.J. **85**, 163
Miller II, K.J., Y.K. Rao, S.R. Goodwin, G.B. Westermann-Clark, D.O. Shah, Solubility and in vitro percutaneous absorption of tetracaine from solvents of propylene glycol and saline **98**, 101
Miller, J., see Eddy, E.P. **115**, 79
Miller, J., see Gochoco, C.H. **104**, 187
Miller, L.L., G.A. Smith, Iontophoretic transport of acetate and carboxylate ions through hairless mouse skin. A cation exchange membrane model **49**, 15
Miller, N.C., see Hindle, M. **134**, 137
Miller, S.C., M.D. Donovan, Effect of poloxamer 407 gel on the miotic activity of pilocarpine nitrate in rabbits **12**, 147
Miller, S.C., B.R. Drabik, Rheological properties of poloxamer vehicles **18**, 269
Miller, S.C., see Wu, H.L.S. **66**, 213
Miller, T.A., P. York, Physical and chemical characteristics of some high purity magnesium stearate and palmitate powders **23**, 55
Miller, T.A., P. York, Pharmaceutical tablet lubrication **41**, 1
Millership, J.S., M.L. Shanks, Prodrugs utilizing a reversible silyl linkage **28**, 1
Millership, J.S., see Karim, E.F.I.A. **52**, 155
Millet, J., see Makki, S. **133**, 245
Mills, S., see Al-Khamis, K. **10**, 25
Mills, S.N., see Kreuter, J. **16**, 105
Milosovich, S.M., see Jona, J.A. **123**, 127
Mima, H., see Hirai, S. **7**, 317; **9**, 165, 173
Mimoso, I.M., A.P.G. Francisco, M.E.M. Cruz, Liposomal formulation of netilmicin **147**, 109
Min, G.H., see Lee, B.J. **144**, 37
Min, K.H., see Kim, C.K. **128**, 65
Min, W., see Zhang, X. **137**, 199
Mine, T., see Sato, K. **135**, 127
Mingione, A., see Iannuccelli, V. **143**, 187
Mingxin, W., see Parkash, A.K. **92**, 151
Minhas, P.S., see Richardson, J.L. **56**, 29
Minkov, E., see Bogdanova, S. **38**, 99
Minkov, E., see Dimitrova, E. **93**, 21
Mirchandani, H., Y.W. Chien, Drug delivery approaches for anti-HIV drugs **95**, 1
Miromoto, Y., see Yamaguchi, H. **58**, 135
Mishra, B., see Pandit, J.K. **21**, 129
Misra, A., P. Upadhyay, Apparatus for preparing adhesive-dispersion transdermal patches on a laboratory scale **132**, 267
Misset, J.L., see Leomy, C. **115**, 235
Mitchell, A.G., G.R.B. Down, Recrystallization after powder compaction **22**, 337
Mitchell, A.G., see Burt, H.M. **3**, 261
Mitchell, A.G., see Dwivedi, S.K. **87**, 95
Mitchell, A.G., see El-Bagory, I. **105**, 255
Mitchell, A.G., see Suryanarayanan, R. **24**, 1; **32**, 213
Mitchell, A.G., see Wong, M.W.Y. **88**, 261
Mitchell, J.C., see Willson, R.J. **132**, 45
Mitchell, J.R., see Melia, C.D. **100**, 263
Mitchell, K., see Ford, J.L. **71**, 95; 213
Mitchell, K., J.L. Ford, D.J. Armstrong, P.N.C. Elliott, J.E. Hogan, C. Rostron, The influence of drugs on the properties of gels and swelling characteristics of matrices containing methylcellulose or hydroxypropylmethylcellulose **100**, 165
The influence of the particle size of hydroxypropylmethylcellulose K15M on its hydration and performance in matrix tablets **100**, 175
Mitchell, K., J.L. Ford, D.J. Armstrong, P.N.C. Elliott, C. Rostron, J.E. Hogan, The influence of additives on the cloud point, disintegration and dissolution of hydroxypropylmethylcellulose gels and matrix tables **66**, 233
The influence of concentration on the release of drugs from gels and matrices containing Methocel^{R} **100**, 155
Mitchell, K., J.L. Ford, D.J.P.N.C. Armstrong Elliott, J.E. Hogan, C. Rostron, The influence of substition type on the performance of methylcellulose and hydroxypropylmethylcellulose in gels and matrices **100**, 143
Mitra, A.K., A.T. Mailloux, Mitomycin C pharmacokinetics in rats—Effect of dose size **32**, 133
Mitra, A.K., M.L. Matthews, Effects of pH and phosphate on the oxidation of iron in aqueous solution **23**, 185
Mitra, A.K., T. Mikkelson, Ophthalmic solution buffer systems. I. The effect of buffer concentration on the ocular absorption of pilocarpine **10**, 219
Ophthalmic solution buffer systems. II. Effects of buffer type and concentration on the ocular absorption of pilocarpine and a method of ocular bioavailability prediction from physicochemical data **37**, 19
Mitra, A.K., M.M. Narurkar, Effect of mercaptan nucleophiles on the degradation of azathioprine in aqueous solution **28**, 119
Kinetics of azathioprine degradation in aqueous solution **35**, 165
Mitra, A.K., see Basu, K. **47**, 195; **65**, 109
Mitra, A.K., see Fort, J.J. **36**, 7; **59**, 271; **100**, 233; **102**, 241; **107**, 9
Mitra, A.K., see Katikaneni, P.R. **123**, 119
Mitra, A.K., see Pujara, C.P. **114**, 197
Mitra, A.K., see Schilling, R.J. **62**, 53
Mitra, A.K., see Shao, Z. **112**, 181
Mitra, A.K., see Spancake, C.W. **75**, 231
Mitra, R., see Banga, A.K. **116**, 211
Mitrevej, A., D. Faroongsarng, N. Sinchaipanid, Compression behavior of spray dried rice starch **140**, 61
Mitrevej, A., R.G. Hollenbeck, Influence of hydrophilic excipients on the interaction of aspirin and water **14**, 243
Mitsuzono, T., see Quan, D. **68**, 239

Mittelman, S.D., see Niven, R.W. **109**, 17
Miyagawa, Y., T. Okabe, Y. Yamaguchi, M. Miyajima, H. Sato, H. Sunada, Controlled-release of diclofenac sodium from wax matrix granule **138**, 215
Miyajima, K., see Aoki, H. **115**, 183; **149**, 15
Miyajima, K., see Kakee, A. **111**, 15
Miyajima, M., M. Yoshida, H. Sato, H. Omichi, R. Katakai, W.I. Higuchi, Controlled release of 9-β-D-arabinofuranosyladenine from thermo-responsive devices based on acryloyl-L-proline methyl ester **95**, 153
Miyajima, M., see Miyagawa, Y. **138**, 215
Miyajima, M., see Miyamoto, Y. **149**, 25
Miyake, M., see Nakada, Y. **89**, 169
Miyake, M., see Nashihata, T. **22**, 147; **33**, 89
Miyamae, A., see Kitamura, S. **56**, 125; **59**, 217
Miyamoto, E., see Murata, Y. **96**, 139
Miyamoto, K., see Hasegawa, T. **56**, 235; **58**, 129
Miyamoto, M., see Yoshimura, Y. **103**, 1
Miyamoto, Y., S. Ogawa, M. Miyajima, M. Matsui, H. Sato, K. Takayama, T. Nagai, An application of the computer optimization technique to wet granulation process involving explosive growth of particles **149**, 25
Miyamoto, Y., see Miyamoto, Y. **149**, 25
Miyata, N., see Kohri, N. **81**, 49
Miyata, T., see Gilbert, D.L. **47**, 79
Miyazaki, K., see Kohri, N. **49**, 213; **68**, 255
Miyazaki, K., see Takada, S. **146**, 147
Miyazaki, M., see Morimoto, K. **81**, 119; **113**, 1
Miyazaki, S., A. Nakayama, M. Oda, M. Takada, D. Attwood, Drug release from oral mucosal adhesive tablets of chitosan and sodium alginate **118**, 257
Miyazaki, S., M. Oshiba, T. Nadai, Unusual solubility and dissolution behavior of pharmaceutical hydrochloride salts in chloride-containing media **6**, 77
Miyazaki, S., T. Yamahira, Y. Morimoto, T. Nadai, Micellar interaction of indomethacin and phenylbutazone with bile salts **8**, 303
Mizukami, Y., see Yamana, T. **11**, 71
Mizumoto, T., see Sako, K. **137**, 225
Mizuno, M., see Naito, S. **17**, 109; **18**, 117
Mizuno, N., see Motozono, H. **108**, 39
Mizuno, N., see Shinkuma, D. **21**, 187
Mlinaric-Majerski, K., see Jaksic, P. **135**, 177
Moczar, E., see Toth, I. **125**, 117
Modamio, P., C.F. Lastra, E.L. Marino, Determination of celiprolol by high-performance liquid chromatography and stability in solution **107**, 243
Modamio, P., C.F. Lastra, O. Montejo, E.L. Marino, Development and validation of liquid chromatography methods for the quantitation of propranolol, metoprolol, atenolol and bisoprolol: Application in solution stability studies **130**, 137
Modamio, P., O. Montejo, C.F. Lastra, E.L. Marino, A valid high-performance liquid chromatography method for oxprenolol stability studies **112**, 93
Modderkolk, J., see Crommelin, D.J.A. **3**, 299
Modena, T., see Pavanetto, F. **48**, 255; **54**, 143; **74**, 175
Moeckly, D.M., L.E. Matheson, The development of a predictive method for the estimation of flux through polydimethylsiloxane membranes. I. Identification of critical variables for a series of substituted benzenes **77**, 151
Moeckly, D.M., see Matheson, L.E. **77**, 163
Moelands, D., N.A. Karnik, R.J. Prankerd, K.B. Sloan, H.W. Stone, J.H. Perrin, Microcalorimetric study of the interactions of aspartame with β-cyclodextrin and hydroxypropyl-β-cyclodextrin: The anomalous heat of dilution of the latter **86**, 263
Moeller, H., S.I. Ali, D. Steinbach, Pharmaceutical and biological availability of sustained release preparations of potassium chloride **7**, 157
Moeller, H., see Steinbach, D. **1**, 197
Moes, A.J., see Beten, D.B. **88**, 31; **103**, 243; **123**, 145
Moes, A.J., see Ingani, H.M. **35**, 157
Moes, A.J., see Oth, M.P. **24**, 275; **55**, 157
Moes, A.J., see Sansdrap, P. **98**, 157
Moes, A.J., see Timmermans, J. **62**, 207
Moghimi, H.R., A.C. Williams, B.W. Barry, A lamellas matrix model for stratum corneum intercellular lipids. I. Characterisation and comparison with stratum corneum intercellular structure **131**, 103
A lamellar matrix model for stratum corneum intercellular lipids. II. Effect of geometry of the stratum corneum on permeation of model drugs 5-flurouracil and oestradiol **131**, 117
A lamellar matrix model for stratum corneum intercellular lipids. III. Effects of terpene penetration enhancers on the release of 5-fluorouracil and oestradiol from the matrix **145**, 37
A lamellar matrix model for stratum corneum intercellular lipids. IV. Effects of terpene penetration enhancers on the permeation of 5-fluorouracil and oestradiol through the matrix **145**, 49
A lamellar matrix model for stratum corneum intercellular lipids. V. Effects of terpene penetration enhancers on the structure and thermal behaviour of the matrix **146**, 41
Moghimi, S.M., C.J.H. Porter, L. Illum, S.S. Davis, The effect of poloxamer-407 on liposome stability and targeting to bone marrow: Comparison with polystyrene microspheres **68**, 121
Moghimi, S.M., see Porter, C.J.H. **83**, 273
Mohamed, A.A., see Kassem, M.A. **29**, 243; **32**, 47
Mohammad, H.A.H., J.T. Fell, Surface free energy characteristics of mixtures **17**, 39
Mohammad, H.A.H., see Fell, J.T. **125**, 327
Mohar, M., see Kerc, J. **68**, 25
Mohr, D., see Dickinson, P.A. **148**, 55
Mohri, S., see Bando, H. **145**, 103
Mohsen Forutan, S., see Ghassabian, S. **130**, 49
Molina, M.M., see Lopez, A.A. **28**, 167
Mollan, M.J., M. Celik, The effects of humidity and storage time on the behavior of maltodextrins for direct compression **114**, 23
Mollan Jr., M.J., M. Celik, The effects of lubrication on the compaction and post-compaction properties of directly compressible maltodextrins **144**, 1

Molle, L., see Verbiese-Genard, N. **2**, 155; **9**, 295
Mollee, H., see Tiemessen, H.L.G.M. **53**, 119
Moller, N., see Hoelgaard, A. **15**, 213
Mollgaard, B., A. Hoelgaard, Permeation of estradiol through the skin: Effect of vehicles **15**, 185
Mollgaard, B., A. Hoelgaard, H. Bundgaard, Pro-drugs as drug delivery systems. XXIII. Improved dermal delivery of 5-fluorouracil through human skin via *N*-acyloxymethyl pro-drug derivatives **12**, 153
Mollgaard, B., see Bundgaard, H. **15**, 285
Mollgaard, B., see Hoelgaard, A. **43**, 233
Mollgaard, B., see Johansen, M. **32**, 199
Mollgaard, B., see Wotton, P.K. **24**, 19
Molokhia, A.M., Effect of storage on crushing strength, disintegration and drug release from mixed tablet bases **22**, 127
Molokhia, A.M., see Al-Gohary, O.M. **55**, 47
Molokhia, A.M., see Gouda, M.W. **5**, 345
Molokhia, A.M., see Niazy, E.M. **56**, 181
Molpeceres, J., M. Guzman, P. Bustamante, Rosario Ab Del, Exothermic-endothermic heat of solution shift of cyclosporin A related to poloxamer 188 behavior in aqueous solutions **130**, 75
Molpeceres, J., see Chacon, M. **141**, 81
Molpeceres, J., see Guzman, M. **80**, 119
Momin, P., see Devleeschouwer, M.J. **32**, 55
Momin, P., see Dony, J. **19**, 239
Momose, Y., see Yamamura, S. **133**, 117
Monedero Perales, M., A. Munoz-Ruiz, Antequera Velasco, M.R. Jimenez-Castellanos, Constant compression-decompression stress rate profiles to obtain rate dependence of maltodextrins for direct compression **132**, 183
Monedero Perales, M., see Munoz-Ruiz, A. **95**, 201; **110**, 37
Monedero Perales, M., see Velasco Antequera, M.V. **103**, 155
Moneghini, M., F. Rubessa, P. Rupena, G. Zingone, Analysis of chlorcyclizine and related compounds by liquid chromatography for stability studies **63**, R1
Moneghini, M., see Mrhar, A. **62**, 15
Mones, J., see Roca, M. **69**, 189
Monger, L., see Hollingsbee, D.A. **77**, 199
Monger, L., see Waring, M.J. **94**, 211
Monkkonen, J., R. Valjakka, M. Hakasalo, A. Urtti, The effects of liposome surface charge and size on the intracellular delivery of clodronate and gallium in vitro **107**, 189
Monnot, E.A., C.G. Kindberg, T.S. Johnson, C.M. Riley, J.F. Stobaugh, M. Slavik, Stability of intravenous admixtures of 5-fluorouracil and spirogermanium, a novel combination of cytotoxic agents **60**, 41
Montanari, C.A., see Montanari, M.L.C. **85**, 199
Montanari, L., see Pavanetto, F. **51**, 55; **54**, 143
Montanari, M.L.C., A.E. Beezer, J.B.P. Sandal, C.A. Montanari, Bioactivity of a series of substituted *N*-(3-phenyl-2-propenylidene)benzeneamines: A microcalorimetric study **85**, 199
Montassier, P., see Amdidouche, D. **111**, 111
Montejo, O., see Modamio, P. **112**, 93; **130**, 137
Montelius, K., see Sjodin, L. **63**, 135
Montenegro, L., J.I. Ademola, F.P. Bonina, H.I. Maibach, Effect of application time of β-methasone-17-valerate 0.1% cream on skin blanching and stratum corneum drug concentration **140**, 51
Montenegro, L., A.M. Panico, F. Bonina, Quantitative determination of hydrophobic compound entrapment in dipalmitoylphosphatidylcholine liposomes by differential scanning calorimetry **138**, 191
Montenegro, L., A.M. Panico, A. Ventimiglia, F.P. Bonina, In vitro retinoic acid release and skin permeation from different liposome formulations **133**, 89
Montenegro, L., see Bonina, F. **145**, 87
Montenegro, L., see Bonina, F.P. **77**, 21; **82**, 171; **98**, 93; **100**, 41, 99; **102**, 19; **106**, 115; **111**, 191; **124**, 45
Montenegro, L., see Palagiano, F. **149**, 171
Montenegro, L., see Saija, A. **124**, 1
Montero, M.T., J. Estelrich, O. Valls, Binding of non-steroidal anti-inflammatory drugs to human serum albumin **62**, 21
Montero, M.T., J. Freixas, J. Hernandez-Borrell, Expression of the partition coefficients of a homologous series of 6-fluoroquinolones **149**, 161
Montero, M.T., A. Marti, J. Hernandez-Borrell, The active trapping of doxorubicin in liposomes by pH gradient: Photon correlation spectroscopy and fluorimetric study **96**, 157
Montero, M.T., D. Saiz, R. Sitges, J.L. Vazquez, J.H. Borrell, Influence of physicochemical properties of fluoroquinolones on encapsulation efficiency in liposomes **138**, 113
Montero, M.T., see Hernandez, J. **57**, 211
Montheard, J.P., see Chafi, N. **52**, 203; **67**, 265
Montheard, J.P., see Kolli, M. **81**, 103
Monti, D., see Saettone, M.F. **72**, 131
Moody, E., see MacLean, D.S. **131**, 243
Moody, R.R., R.B. Taylor, An HPLC assay for the determination of proguanil hydrochloride in tablets **7**, 177
Moody, R.R., see Taylor, R.B. **60**, 185
Moolenaar, F., S. Bakker, J. Visser, T. Huizinga, Biopharmaceutics of rectal administration of drugs in man. IX. Comparative biopharmaceutics of diazepam after single rectal, oral, intramuscular and intravenous administration in man **5**, 127
Moolenaar, F., J.G. Ensing, B.G. Bolhuis, J. Visser, Absorption rate and bioavailability of promethazine from rectal and oral dosage forms **9**, 353
Moolenaar, F., B.G. Kaufmann, J. Visser, D.K.F. Meijer, Rectal absorption of methadone from dissolution-promoting vehicles **33**, 249
Moolenaar, F., B. Koning, T. Huizinga, Biopharmaceutics of rectal administration of drugs in man. Absorption rate and bioavailability of phenobarbital and its sodium salt from rectal dosage forms **4**, 99
Moolenaar, F., P. Meyler, E. Frijlink, T.H. Jauw, J. Visser, H. Proost, Rectal absorption of morphine from controlled release suppositories **114**, 117
Moolenaar, F., J. Pronk, J.G. Hardy, D.K.F. Meijer, Rectal versus oral absorption of diflunisal in man **19**, 161

Moolenaar, F., A.G.G. Stuurman-Bieze, J. Visser, T. Huizinga, Biopharmaceutics of rectal administration of drugs in man. I. Introduction of benzoic acid as a test drug **1**, 323
Moolenaar, F., J. Visser, T. Huizinga, Biopharmaceutics of rectal administration of drugs in man. Absorption rate and bioavailability of glafenine after oral and rectal administration **4**, 195
Moolenaar, F., J. Visser, A. Leuverman, B.J.M. Schoonen, Bioavailability of morphine from suppositories **45**, 161
Moolenaar, F., see Franssen, E.J.F. **80**, R15
Moolenaar, F., see Schoonen, A.J.M. **4**, 141; **7**, 29
Moolenaar, F., see Stuurman-Bieze, A.G.G. **1**, 337
Moolenaar, F., see Vromans, H. **26**, 5
Moore, D.E., Kinetic treatment of photochemical reactions **63**, R5
Moore, D.E., M.P. Fallon, C.D. Burt, Photo-oxidation of tetracycline—A differential pulse polarographic study **14**, 133
Moore, D.E., J.L. Mallesch, Photochemical interaction between triamterene and hydrochlorthiazide **76**, 187
Moore, D.E., see Tcnnesen, H.H. **70**, 95
Moore, D.K., see Zhou, W. **110**, 55
Moore, J.A., S.A. Pletcher, M.J. Ross, Absorption enhancement of growth hormone from the gastrointestinal tract of rats **34**, 35
Moore, R.G., see Ridd, M.J. **11**, 11
Mora, J., see Roca, M. **69**, 189
Mora, M., see Pato, A.J. **104**, 227
Morant, M.J., see Lopez, A. **139**, 205
Morefield, E.M., J.R. Feldkamp, G.E. Peck et al., Preformulation information for suspensions **34**, 263
Moreira, J.N., L.M. Almeida, C.F. Geraldes, V.M.C. Madeira, M.L. Costa, Carboplatin liposomes coated with *O*-palmitoylpullulan: In vitro characterization **147**, 153
Morel, S., M.R. Gasco, R. Cavalli, Incorporation in lipospheres of [D-Trp-6]LHRH **105**, 189
Incorporation in lipospheres of [D-Trp-6]LHRH (Int. J. Pharm. 105 (1994) R1–R3) **119**, 126
Morel, S., E. Ugazio, R. Cavalli, M.R. Gasco, Thymopentin in solid lipid nanoparticles **132**, 259
Morel, S., see Cavalli, R. **117**, 243
Morell, J.L.P., M.D.C. Claramonte, A.P. Vialard, Validation of a release diffusion cell for topical dosage forms **137**, 49
Moren, F., Drug deposition of pressurized inhalation aerosols. I. Influence of actuator tube design **1**, 205
Drug deposition of pressurized inhalation aerosols. II. Influence of vapour pressure and metered volume **1**, 213
Pressurized aerosols for oral inhalation **8**, 1
Moren, F., J. Andersson, Fraction of dose exhaled after administration of pressurized inhalation aerosols **6**, 295
Moren, F., S.E. Jacobsson, In vitro dose sampling from pressurized inhalation aerosols. Investigation of procedures in BPC and NF **3**, 335
Investigation of assay procedures for pressurized inhalation aerosols **5**, 287
Moren, F., see Newman, S.P. **74**, 209
Moreton, R.C., N.A. Armstrong, Design and use of an apparatus for measuring diffusion through glycerogelatin films **122**, 79
Morga, F., see Lamson, M.J. **105**, 259
Morgan, S.M., Al-Shamkhani, D. Callant, E. Schacht, J.F. Woodley, R. Duncan, Alginates as drug carriers: Covalent attachment of alginates to therapeutic agents containing primary amine groups **122**, 121
Morgan, S.M., V. Subr, K. Ulbrich, J.F. Woodley, R. Duncan, Evaluation of *N*-(2-hydroxypropyl)methacrylamide copolymer-peptide conjugates as potential oral vaccines. Studies on their degradation by isolated rat small intestinal peptidases and their uptake by adult rat small intestinal tissue in vitro **128**, 99
Morgan, S.M., see Morgan, S.M. **122**, 121
Mori, A., E. Duda, L. Huanga, Recombinant human tumor necrosis factor-α covalently conjugated to long-circulating liposomes **131**, 57
Mori, H., see Kaneo, Y. **44**, 265
Mori, Y., see Sasaki, H. **44**, 15; **60**, 1, 177
Morigaki, S., see Hatanaka, T. **125**, 195
Morii, M., Zaid Sayed Abu Samir, Size and permeability of liposomes extruded through polycarbonate membranes **17**, 215
Morikawa, K., see Fujita, T. **134**, 47
Morikawa, T., see Katsu, T. **53**, 61
Morimoto, K., E. Kamiya, T. Takeeda et al., Enhancement of rectal absorption of insulin in polyacrylic acid aqueous gel bases containing long chain fatty acid in rats **14**, 149
Morimoto, K., M. Miyazaki, M. Kakemi, Effects of proteolytic enzyme inhibitors on nasal absorption of salmon calcitonin in rats **113**, 1
Morimoto, K., T. Takeeda, Y. Nakamoto, K. Morisaka, Effective vaginal absorption of insulin in diabetic rats and rabbits using polyacrylic acid aqueous gel bases **12**, 107
Morimoto, K., Y. Iwakura, M. Miyazaki, E. Nakatani, Effects of proteolytic enzyme inhibitors of enhancement of transdermal iontophoretic delivery of vasopressin and an analogue i rats **81**, 119
Morimoto, K., see Kamiya, E. **17**, 273; **26**, 197
Morimoto, K., see Kokubo, H. **35**, 181
Morimoto, K., see Takeeda, T. **41**, 21
Morimoto, K.W., see Lee, V.H.L. **13**, 183
Morimoto, Y., H. Natsume, K. Sugibayashi, S. Fujimoto, Effect of chemoembolization of albumin microspheres containing mitomycin C on AH 272 liver metastasis in rats **54**, 27
Morimoto, Y., K. Sugibayashi, K.I. Hosoya, W.I. Higuchi, Penetration enhancing effect of Azone on the transport of 5-fluorouracil across the hairless rat skin **32**, 31
Morimoto, Y., K. Sugibayashi, D. Kobayashi, H. Shoji, J.I. Yamazaki, M. Kimura, A new enhancer-coenhancer system to increase skin permeation of morphine hydrochloride in vitro **91**, 9
Morimoto, Y., see Hatanaka, T. **79**, 21
Morimoto, Y., see Inoue, N. **137**, 75
Morimoto, Y., see Kitamura, S. **56**, 125; **59**, 217

Morimoto, Y., see Kubo, H. **103**, 27
Morimoto, Y., see Manabe, E. **129**, 211
Morimoto, Y., see Miyazaki, S. **8**, 303
Morimoto, Y., see Morimoto, Y. **91**, 9
Morimoto, Y., see Nakamura, H. **141**, 71
Morimoto, Y., see Natsume, H. **58**, 79
Morimoto, Y., see Sato, K. **43**, 31
Morimoto, Y., see Seki, T. **57**, 73
Morimoto, Y., see Sugibayashi, K. **113**, 189
Morimoto, Y., see Tamura, M. **131**, 263
Morimoto, Y., see Ueda, H. **137**, 217; **143**, 37
Morisaka, K., see Kamiya, H. **26**, 197
Morisaka, K., see Morimoto, K. **12**, 107
Morisaka, K., see Takeeda, T. **41**, 21
Morishima, K., Y. Kawashima, H. Takeuchi, T. Niwa, T. Hino, Y. Kawashima, Tabletting properties of bucillamine agglomerates prepared by the spherical crystallization technique **105**, 11
Morishita, I., see Morishita, M. **78**, 1
Morishita, I., M. Morishita, K. Takayama, Y. Machida, T. Nagai, Hypoglycemic effect of novel oral microspheres of insulin with protease inhibitor in normal and diabetic rats **78**, 9
Enteral insulin delivery by microspheres in three different formulations using Eudragit L100 and S100 **91**, 29
Morishita, M., I. Morishita, K. Takayama, Y. Machida, T. Nagai, Novel oral microspheres of insulin with protease inhibitor protecting from enzymatic degradation **78**, 1
Morishita, M., see Morishita, I. **78**, 9; **91**, 29
Morita, E., see Motozono, H. **108**, 39
Morita, T., see Yamamoto, A. **93**, 91
Mork, N., H. Bundgaard, M. Shalmi, S. Christensen, Furosemide prodrugs: Synthesis, enzymatic hydrolysis and solubility of various furosemide esters **60**, 163
Mork, N., see Bundgaard, H. **55**, 91; **63**, 213
Mork, N., see Fredholt, K. **123**, 209
Mork, N., see Hansen, J. **81**, 253
Mork, N., see Thomsen, K.F. **112**, 143
Mork Nielsen, N., T. Kurihara-Bergstrom, Prodrugs as drug delivery system. 68. Chemical and plasma-catalyzed hydrolysis of various esters of benzoic acid: A reference system for designing prodrug esters of carboxylic acid agents **39**, 75
Morphet, C., see Rocco, W.L. **122**, 17
Morris, K., see Wesdyk, R. **65**, 69
Morris, K.R., M.G. Fakes, A.B. Thakur, A.W. Newman, A.K. Singh, J.J. Venit, C.J. Spagnuolo, A.T.M. Serajuddin, An-integrated approach to the selection of optimal salt form for a new drug candidate **105**, 209
Morris, K.R., A.W. Newman, D.E. Bugay, S.A. Ranadive, A.K. Singh, M. Szyper, S.A. Varia, H.G. Brittain, A.T.M. Serajuddin, Characterization of humidity-dependent changes in crystal properties of a new HMG-CoA reductase inhibitor in support of its dosage form development **108**, 195
Morrison, R.A., S.M. Singhvi, Pharmacokinetics and oral bioavailability of pravastatin in dogs **143**, 265
Morrison, R.A., see Nicklin, P.L. **140**, 175
Mortada, L.M., see Khalil, S.A.H. **18**, 157; **19**, 233
Mortazavi, S.A., An in vitro assessment of mucus–mucoadhesive interactions **124**, 173
Mortazavi, S.A., B.G. Carpenter, J.D. Smart, An investigation of the rheological behaviour of the mucoadhesive–mucosal interface **83**, 221
A comparative study on the role played by mucus glycoproteins in the rheological behaviour of the mucoadhesive–mucosal interface **94**, 195
Mortazavi, S.A., J.D. Smart, An investigation of some factors influencing the in vitro assessment of mucoadhesion **116**, 223
Mortimer, B.C., see Lundberg, B.B. **134**, 119
Morton, A.K., H.E. Edwards, J.C. Allen, Some physicochemical and biological characteristics of heparin fractions prepared by gel chromatography **18**, 39
Morton, A.K., H.E. Edwards, J.C. Allen, G.O. Phillips, An evaluation of the oral administration of commercial and fractionated heparin samples in rats **9**, 321
Morton, D.J., Comparison of in vitro release of prostaglandin E_2 from various pessary bases **22**, 131
EDTA reduces antimicrobial efficacy of thiomersal **23**, 357
Morton, D.J., see Ismail, S. **37**, 11
Morton, D.J., see Solanki, P. **42**, 47
Morvan, C., see Skiba, M. **126**, 275
Mosberg, H., see Oie, S. **48**, 223
Moschel, R.C., see Safadi, M. **90**, 239
Moser, K., see Mader, R.M. **68**, 151
Mosharraf, M., C. Nystrom, The effect of particle size and shape on the surface specific dissolution rate of microsized practically insoluble drugs **122**, 35
Solubility characterization of practically insoluble drugs using the Coulter counter principle **122**, 57
Mosher, G.L., T.J. Mikkelson, Permeability of the *n*-alkyl *p*-aminobenzoate esters across the isolated corneal membrane of the rabbit **2**, 239
Mosher, G.L., T.P. Paronen, E. Falch et al., Ocular bioavailability of pilocarpic acid mono- and diester prodrugs as assessed by miotic activity in the rabbit **39**, 113
Mosquera, M.J., M. Cuna, C. Souto, A. Concheiro, R. Martinez-Pacheco, Amoza J.L. Gomez, Effects of hydroxymethylpropylcellulose (HPMC) moisture content on hydrochlorothiazide release from HPMC-based tablets **135**, 147
Mosquera, R., see Fernandez, B. **79**, 199
Moss, J., H. Bundgaard, Prodrugs of peptides. 5. Protection of the pyroglutamyl residue against pyroglutamyl aminopeptidase by bioreversible derivatization with glyoxylic acid derivatives **52**, 255
Prodrugs of peptides. 7. Transdermal delivery of thyro ropin-releasing hormone (TRH) via prodrugs **66**, 39
Prodrugs of peptides. 12. Bioreversible derivatization of thyrotropin-releasing hormone (TRH) by *N*-phthalidylation of its imidazole moiety **74**, 67
Prodrugs of peptides. 17. Bioreversible derivatization of the C-terminal prolineamide residue in peptides to afford protection against prolyl endopeptidase **82**, 91

Moss, J., A. Buur, H. Bundgaard, Prodrugs of peptides. 8. In vitro study of intestinal metabolism and penetration of thyrotropin-releasing hormone (TRH) and its prodrugs **66**, 183
Moss, J., see Bundgaard, H. **45**, 47
Moss, J., see Kahns, A.H. **78**, 199
Moss, J., see Lundin, S. **76**, R1
Mostafa, E.M., see Hamed, M.Y. **69**, 93
Mostafa, E.M., see Ibrahim, S.A. **61**, 1
Motola, S., see Cotton, M.L. **109**, 237
Motozono, H., N. Mizuno, E. Morita, Y. Fujioka, K. Takahashi, Effect of age on gastrointestinal absorption of tobramycin in rats **108**, 39
Mouazen, F., F. Puisieux, M. Seiller, J.T. Carstensen, Dielectric properties of polyoxyethylene-polyoxypropyleneglycols **1**, 275
Moufti, A., see Weingarten, C. **26**, 251
Mougeolle, J.M., see Chevin, J.C. **45**, 101
Moussa, B.A., Colorimetric determination of parasiticides hycanthone and metronidazole **10**, 199
Moussa, I.S., L.H. Cartilier, Evaluation of cross-linked amylose press-coated tablets for sustained drug delivery **149**, 139
Moustafa, M.A., H.I. Al-Shora, M. Gaber, M.W. Gouda, Decreased bioavailability of quinidine sulphate due to interactions with adsorbent antacids and antidiarrhoeal mixtures **34**, 207
Moustafa, M.A., S.A. Babhair, H.I. Kouta, Decreased bioavailability of some antipsychotic phenothiazines due to interactions with adsorbent antacid and antidiarrhoeal mixtures **36**, 185
Moustafa, M.A., M.W. Gouda, M. Tariq, Decreased bioavailability of propranolol due to interactions with adsorbent antacids and antidiarrhoeal mixtures **30**, 225
Moustafa, M.A., see Al-Shora, H.I. **47**, 209
Moustafa, M.A., see Belloto, Jr. R.J. **23**, 195
Moustafa, M.A., see Gouda, M.W. **5**, 345
Moustafa, M.A., see Wafik, Gouda H. **18**, 213
Moyona, J.R., M.J. Arias-Blanco, J.M. Gines, F. Giordano, Solid-state characterization and dissolution characteristics of gliclazide-β-cyclodextrin inclusion complexes **148**, 211
Moyano, J.R., J.M. Gines, M.J. Arias, A.M. Rabasco, Study of the dissolution characteristics of oxazepam via complexation with β-cyclodextrin **114**, 95
Moyano, J.R., see Arias, M.J. **123**, 25
Moyano, J.R., see Arias-Blanco, M.J. **143**, 113
Moyano, J.R., see Gines, J.M. **143**, 247
Mrhar, A., F. Rubessa, R. Karba, M. Moneghini, S. Primosic, Pharmacokinetic evaluation of sustained release formulations of theophylline by analog hybrid simulation **62**, 15
Mrhar, A., see Kmetec, V. **21**, 211
Mrhar, A., see Kristl, A. **57**, 229; **99**, 79; **100**, 33
Mrhar, A., see Srcic, S. **73**, 221
Mroso, P.V., see Li Wan Po, A. **16**, 115; **18**, 287
Mroszczak, E.J., see Kent, J.S. **7**, 245
Mucci, A., see Vandelli, M.A. **118**, 77
Mueller-Goymann, C.C., S.G. Frank, Interaction of lidocaine and lidocaine-HCl with the liquid crystal structure of topical preparations **29**, 147
Mufson, D., see Guo, L.S.S. **75**, 45
Muggetti, L., M. Zurlo, A. Martini, P. Civaroli, R. de Ponti, Preformulation activities of intranasal dosage forms of temazepam **136**, 81
Muggetti, L., see Martini, A. **127**, 239
Muggetti, L., see Torricelli, C. **71**, 19; **75**, 147
Muhammad, N.A., S.C. Dave, J.L. Fox, W.I. Higuchi, The influence of ion activity product on remineralization behavior of carbonate-apatite pellets, hydroxyapatite pellets and hydroxyapatite powder **21**, 295
Muhi-Eldeen, Z., M. Kawahara, A. Dakkuri, A. Hussain, Kinetics and mechanism of the enzymatic hydrolysis of aspirin phenylalanine ethyl ester **26**, 15
Muhuri, G., see Wan, L.S.C. **88**, 159
Muir, N.C., see Hessell, P.G. **52**, 159
Mukai, E., K. Arase, M. Hashida, H. Sezaki, Enhanced delivery of mitomycin C prodrugs through the skin **25**, 95
Mukai, E., see Sasaki, H. **15**, 49, 61
Mukherji, G., N. Aggarwal, Derivative UV-spectroscopic determination of salbutamol sulphate in the presence of gelatin **71**, 187
Quantitative estimation of salbutamol sulphate by derivative UV spectroscopy in the presence of albumin **86**, 153
Mukherji, G., R.S.R. Murthy, B.D. Miglani, Preparation and evaluation of polyglutaraldehyde nanoparticles containing 5-fluorouracil **50**, 15
Preparation and evaluation of cellulose nanospheres containing 5-fluorouracil **65**, 1
Mulder, E.M.A., see Burger, J.J. **23**, 333
Mulinacci, N., F. Melani, G. Mazzi, F.F. Vincieri, Molecular modelling and NMR NOE experiments: Complementary tools for the investigation of complex ibuproxam-β-cyclodextrin topology **90**, 35
Mulinacci, N., F. Melani, F.F. Vincieri, G. Mazzi, A. Romani, ^{1}H NMR NOE and molecular modelling to characterize thymol and carvacrol β-cyclodextrin complexes **128**, 81
Mulinacci, N., see Vincieri, F.F. **48**, 119
Muller, B.W., E. Albers, Complexation of dihydropyridine derivatives with cyclodextrins and 2-hydroxypropyl-β-cyclodextrin in solution **79**, 273
Muller, B.W., U. Brauns, Solubilization of drugs by modified β-cyclodextrins **26**, 77
Muller, B.W., see Backensfeld, T. **74**, 85
Muller, B.W., see Bleich, J. **97**, 111; **106**, 77
Muller, B.W., see Carstensen, H. **67**, 29
Muller, B.W., see Helm, V.J. **70**, 29
Muller, B.W., see Holzner, P.M. **106**, 69; **116**, 11
Muller, B.W., see Lehner, S.J. **93**, 201
Muller, B.W., see Lucks, J.S. **58**, 229; **63**, 183
Muller, B.W., see Lukowski, G. **84**, 23
Muller, B.W., see Reer, O. **104**, 239
Muller, B.W., see Ruchatz, F. **142**, 67
Muller, B.W., see Trenktrog, T. **123**, 199
Muller, B.W., see Van, Dooren A.A. **20**, 217

Muller, B.W., see Vollmer, U. **99**, 51
Muller, B.W., see Waaler, P.J. **87**, 223, 229
Muller, B.W., see Wesemeyer, H. **89**, 33
Muller, C.E., P.T. Daniel, J. Holzschuh, H.J. Roth, Lipophilic disulfide prodrugs. Syntheses and disulfide bond cleavage **57**, 41
Muller, D.G., see Du Plessis, J. **103**, 277, R1; **127**, 273
Muller, D.G., V.J. Stella, A.P. Lotter, Factors influencing the precipitation time of phenytoin in the presence of DDMS, one of its prodrugs **75**, 201
The determination of the amphiphilic properties of a prodrug (DDMS) of phenytoin in aqueous media **86**, 175
Determination of the position of the ester bond in a micellar prodrug of phenytoin **102**, 109
Muller, R.H., S. Heinemann, Fat emulsions for parenteral nutrition. III. Lipofundin MCT/LCT regimens for total parenteral nutrition (TPN) with low electrolyte load **101**, 175
Fat emulsions for parenteral nutrition. IV. Lipofundin MCT/LCT regimens for total parenteral nutrition (TPN) with high electrolyte load **107**, 121
Muller, R.H., C. Lherm, J. Herbort, T. Blunk, P. Couvreur, Alkylcyanoacrylate drug carriers. I. Physicochemical characterization of nanoparticles with different alkyl chain length **84**, 1
Muller, R.H., S. Maassen, H. Weyhers, F. Specht, J.S. Lucks, Cytotoxicity of magnetite-loaded polylactide, polylactide/glycolide particles and solid lipid nanoparticles **138**, 85
Muller, R.H., D. Ruhl, S.A. Runge, Biodegradation of solid lipid nanoparticles as a function of lipase incubation time **144**, 115
Muller, R.H., K.H. Wallis, Surface modification of i.v. injectable biodegradable nanoparticles with poloxamer polymers and poloxamine 908 **89**, 25
Muller, R.H., see Almeida, A.J. **149**, 255
Muller, R.H., see Carstensen, H. **67**, 29
Muller, R.H., see Lherm, C. **84**, 13
Muller, R.H., see Lucks, J.S. **58**, 229; **63**, 183
Muller, R.H., see Lukowski, G. **84**, 23
Muller, R.H., see Rudt, S. **99**, 1
Muller, R.H., see Wesemeyer, H. **89**, 33
Muller, R.H., see Zingel, C. **140**, 13
Muller, U., K. Munz, Distribution of [^{14}C]diazepam associated with two different types of small unilamellar liposomes in mice **47**, 111
Muller, U., see Camber, O. **39**, 229
Muller, U., see Kreuter, J. **55**, 39
Muller, U., see Troster, S.D. **61**, 85
Muller-Goymann, C.C., see Hirsch, R. **120**, 229
Muller-Goymann, C.C., see Kriwet, K. **125**, 231
Muller-Goymann, C.C., see Wilisch, I.L. **96**, 79
Mulski, M.J., see Wood, R.W. **50**, 61
Mumenthaler, M., H. Leuenberger, Atmospheric spray-freeze drying: A suitable alternative in freeze-drying technology **72**, 97
Mummaneni, V., R.C. Vasavada, Solubilization and dissolution of famotidine from solid glass dispersions of xylitol **66**, 71
Mumtaz, A.M., H.S. Ch'ng, Design of a dissolution apparatus suitable for in situ release study of triamcinolone acetonide from bioadhesive buccal tablets **121**, 129
Evaluation of bioadhesive buccal tablets containing triamcinolone acetonide in healthy volunteers **121**, 249
Munday, D.L., A.R. Fassihi, Controlled release delivery: Effect of coating composition on release characteristics of mini-tablets **52**, 109
In vitro–in vivo correlation studies on a novel controlled release theophylline delivery system and on Theo-Dur tablets **118**, 251
Munday, D.L., A.R. Fassihi, C. de Villiers, Bioavailability study of a theophylline oral controlled release capsule containing film coated mini-tablets in beagle dogs **69**, 123
Multiple dose in vivo evaluation of an oral controlled release capsule of theophylline containing film coated mini-tablets in beagle dogs **73**, 89
Munday, D.L., see Sujja-Areevath, J. **139**, 53
Munday, M.R., see Toth, I. **68**, 191
Munguia, O., A. Delgado, J. Farina, C. Evora, M. Llabres, Optimization of DL-PLA molecular weight via the response surface method **86**, 107
Munguia, O., see Dorta, M.J. **100**, 9
Munoz, A., see Castellano, I. **146**, 71
Munoz, A., see Ferrero, M.C. **149**, 233
Munoz, A., see Velasco, M.V. **136**, 107
Munoz, M., J. Bonjoch, J. Prat, M. Pujol, V. Girona, Bolos J. De, Degradation kinetics of ifosfamide in aqueous solution **139**, 249
Munoz, M., see Munoz, M. **139**, 249
Munoz, M., see Pujol, M. **146**, 263
Munoz, N., see Ferrero, C. **147**, 11
Munoz, P., see Arias-Blanco, M.J. **143**, 113
Munoz-Munoz, N., see Munoz-Ruiz, A. **95**, 201; **110**, 37
Munoz-Munoz, N., see Velasco Antequera, M.V. **103**, 155
Munoz-Ruiz, A., M. Monedero Perales, M.V. Velasco Antequera, T. Payan Villar, N. Munoz-Munoz, M.R. Jimenez-Castellanos, Rheology and compression characteristics of lactose based direct compression excipients **95**, 201
Munoz-Ruiz, A., P. Paronen, Particle and powder properties of cyclodextrins **148**, 33
Munoz-Ruiz, A., T. Payan Villar, N. Munoz-Munoz, M. Monedero Perales, M.R. Jimenez-Castellanos, Analysis of the physical characterization and the tabletability of calcium phosphate-based materials **110**, 37
Munoz-Ruiz, A., T.P. Villar, A. Justo, V. Velasco, R. Jimenez-Castellanos, X-ray tablet and raw diffraction as a method to study compression parameters in a direct compression excipient, Compril^{R} **144**, 147
Munoz-Ruiz, A., see Ferrero, C. **147**, 11
Munoz-Ruiz, A., see Monedero Perales, M. **132**, 183
Munoz-Ruiz, A., see Velasco Antequera, M.V. **103**, 155
Munro, S.L.A., see Charman, W.N. **88**, 397
Munz, K., see Kreuter, J. **55**, 39

Munz, K., see Muller, U. **47**, 111
Munz, K., see Waser, P.G. **39**, 213
Mura, P., A. Manderioli, G. Bramanti, S. Furlanetto, S. Pinzauti, Utilization of differential scanning calorimetry as a screening technique to determine the compatibility of ketoprofen with excipients **119**, 71
Mura, P., see Bramanti, G. **49**, 241
Mura, P., see Santoni, G. **50**, 75; **64**, 235; **71**, 1; **93**, 239
Murai, Y., T. Nakagawa, K. Yamaoka, T. Uno, High-performance liquid chromatographic determination and moment analysis of urinary excretion of flucloxacillin and its metabolites in man **15**, 309
Murakami, H., Y. Kawashima, T. Niwa, T. Hino, H. Takeuchi, M. Kobayashi, Influence of the degrees of hydrolyzation and polymerization of poly(vinylalcohol) on the preparation and properties of poly(DL-lactide-*co*-glycolide) nanoparticle **149**, 43
Murakami, M., Y. Kusanoi, K. Takada, S. Muranishi, Assessment of enhancing ability of medium-chain alkyl saccharides as new absorption enhancers in rat rectum **79**, 159
Murakami, M., K. Takada, S. Muranishi, Promoting effect of Azone on intestinal absorption of poorly absorbable drugs in rats **31**, 231
Murakami, M., see Fukui, H. **31**, 239
Murakami, M., see Ohtani, T. **77**, 141
Murakami, T., M. Kishimoto, Y. Higashi, H. Amagase, T. Fuwa, N. Yata, Down-regulation and its effect of epidermal growth factor receptors on the pharmacokinetics of human epidermal growth factor after i.v. administration in rats Prodrugs for dermal delivery **54**, 259
Murakami, T., M. Kishimoto, Y. Higashi, H. Amagase, T. Hayashi, T. Fuwa, N. Yata, Biliary and urinary excretions of immunoreactive human epidermal growth factor after intravenous administration in rats **49**, 173
Murakami, T., see Kadir, S. **33**, 235; **40**, 257
Muramatsu, K., Y. Maitani, Y. Machida, T. Nagai, Effect of soybean-derived sterol and its glucoside mixtures on the stability of dipalmitoylphosphatidylcholine and dipalmitoylphosphatidylcholine/cholesterol liposomes **107**, 1
Effects of soybean-derived sterol and its glucoside mixtures added in dipalmitoylphosphatidylcholine liposomes on the blood circulation and hepatic cellular distribution in mice **124**, 19
Muranishi, S., N. Muranushi, H. Sezaki, Improvement of absolute bioavailability of mormally poor absorbed drugs: Inducement of the intestinal absorption of streptomycin and gentamycin by lipid-bile salt mixed micelles in rat and rabbit **2**, 101
Muranishi, S., see Fujita, T. **134**, 47
Muranishi, S., see Fukui, H. **31**, 239
Muranishi, S., see Hashida, M. **2**, 245
Muranishi, S., see Murakami, M. **31**, 231; **79**, 159
Muranishi, S., see Ohtani, T. **77**, 141
Muranishi, S., see Takada, K. **25**, 155; **44**, 107; **65**, 159
Muranishi, S., see Taniguchi, K. **4**, 219
Muranishi, S., see Yoshikawa, H. **8**, 291; **13**, 321
Muranishi, S., see Yoshioka, T. **8**, 131
Muranushi, N., see Muranishi, S. **2**, 101
Muranushi, N., M. Kinugawa, Y. Nakajima et al., Mechanism for the inducement of the intestinal absorption of poorly absorbed drugs by mixed micelles. I. Effects of various lipid-bile salt mixed micelles on the intestinal absorption of streptomycin in rat **4**, 271
Muranushi, N., Y. Nakajima, M. Kinugawa et al., Mechanism for the inducement of the intestinal absorption of poorly absorbed drugs by mixed micelles. II. Effect of the incorporation of various lipids on the permeability of liposomal membranes **4**, 281
Murata, Y., T. Maeda, E. Miyamoto, S. Kawashima, Preparation of chitosan-reinforced alginate gel beads—Effects of chitosan on gel matrix erosion **96**, 139
Murayama, H., see Tanaka, S. **27**, 29
Muret, P., see Makki, S. **133**, 245
Muro, C., see Shinkuma, D. **9**, 17
Murooka, N., see Kitagawa, S. **126**, 49
Murphy, A., L. Zajic, A.P. Launchbury, Dissolution time, on reconstitution, of a new parenteral formulation of doxorubicin (Doxorubicin Rapid Dissolution) **38**, 257
Murphy, C.A., see Corrigan, O.I. **4**, 67
Murphy, C.A., see Toth, I. **68**, 191
Murphy, T.M., see Gay, C.L. **49**, 39
Murray, G.R., see Wilson, C.G. **40**, 119
Murray, W.J., see Al-Turk, W.A. **41**, 227
Murray, W.J., see Newton, D.W. **11**, 209
Murtha, J.L., see Radebaugh, G.W. **45**, 39
Murthy, R.S.R., see Bhaskar, R. **28**, 59
Murthy, R.S.R., see Mukherji, G. **50**, 15; **65**, 1
Murtoniemi, E., P. Merkku, P. Kinnunen, K. Leiviska, J. Yliruusi, Effect of neural network topology and training end point in modelling the fluidized bed granulation process **110**, 101
Murtoniemi, E., J. Yliruusi, P. Kinnunen, P. Merkku, K. Leiviska, The advantages by the use of neural networks in modelling the fluidized bed granulation process **108**, 155
Murugesan, K., see Jha, P. **63**, 143
Musa, R., see Steed, K.P. **112**, 199
Musatti, L., see Efthymiopoulos, C. **95**, 127
Muscettola, M., see Bocci, V. **32**, 103
Musson, D.G., D.P. Evitts, A.M. Bidgood, O. Olejnik, Stabilization of aqueous thymoxamine using dimethyl-β-cyclodextrin **99**, 85
Mustafa, R., see Hussain, A. **21**, 289
Mustafa, R., see Kagayama, A. **18**, 247
Mustafa, R.M., see Shihab, F.A. **4**, 13
Muti, H., see Othman, S. **41**, 197
Muti, H.Y., see Suleiman, M.S. **50**, 71
Muttonen, E., see Eerikainen, S. **80**, 259
Muzzio, F.J., see Swanson, P.D. **142**, 33
Myers, M.J., see Kearney, P. **38**, 211
Myers, R.A., V.J. Stella, Systemic bioavailability of penclomedine (NSC-338720) from oil-in-water emulsions administered intraduodenally to rats **78**, 217
Factors affecting the lymphatic transport of penclomedine (NSC-338720), a lipophilic cytotoxic drug: Comparison to DDT and hexachlorobenzene **80**, 51

Myers, R.A., see O'Driscoll, C.M. **73**, 177
Myrdal, P.B., see Kipp, J.E. **74**, 215

N

Nabulsi, L., see Sheikh, Salem M. **141**, 257
Nacher, A., see Cejudo-Ferragud, E. **132**, 63
Nachtmann, F., K. Gstrein, Comparison of HPLC and some official test methods for different penicillins **7**, 55
Nachtmann, F., Determination of the L-isomer in D-penicillamine by derivatization liquid chromatography **4**, 337
Nada, A.H., Influence of caffeine on the renal effects and solubility of ketoprofen **70**, 191
Nada, A.H., M.A. Etman, A.R. Ebian, In vitro adsorption of mepyramine maleate onto some adsorbents and antacids **53**, 175
Nada, A.H., see Etman, M.A. **88**, 433
Nada, A.H., see Hammounda, Y.E. **89**, 111
Nada, A.H., see Hassan, E.E. **121**, 149
Nadai, M., see Hasegawa, T. **56**, 235; **58**, 129
Nadai, T., see Araki, M. **81**, 39
Nadai, T., see Miyazaki, S. **6**, 77; **8**, 303
Nadai, T., see Yamashita, S. **108**, 241
Naevestad, A., see Sontum, P.C. **128**, 269
Nagae, I., see Unezaki, S. **144**, 11
Nagahama, E., see Takino, T. **114**, 43
Nagahara, N., see Kurosaki, Y. **67**, 1
Nagai, T., see Akimoto, J. **128**, 251
Nagai, T., see Akimoto, J. **140**, 257
Nagai, T., see Coutel-Egros, A. **84**, 117
Nagai, T., see Fujimori, J. **119**, 47
Nagai, T., see Hamamoto, K. **113**, 9
Nagai, T., see Imasaka, K. **68**, 87; **81**, 31
Nagai, T., see Ito, R. **61**, 109
Nagai, T., see Maitani, Y. **49**, 23; **64**, 139; **69**, 21; **74**, 105; **113**, 165; **117**, 129; **126**, 89; **146**, 11
Nagai, T., see Miyamoto, Y. **149**, 25
Nagai, T., see Morishita, I. **78**, 9; **91**, 29
Nagai, T., see Morishita, M. **78**, 1
Nagai, T., see Muramatsu, K. **107**, 1; **124**, 19
Nagai, T., see Nagano, H. **147**, 119
Nagai, T., see Nakamura, F. **134**, 173; **145**, 29
Nagai, T., see Obata, Y. **89**, 191
Nagai, T., see Ohara, N. **105**, 31; **119**, 125; **123**, 281
Nagai, T., see Onishi, H. **30**, 133
Nagai, T., see Oribe, T. **124**, 27
Nagai, T., see Qi, X. **114**, 33; **146**, 31
Nagai, T., see Quan, D. **68**, 239
Nagai, T., see Song, Y. **98**, 121
Nagai, T., see Takahashi, T. **61**, 35
Nagai, T., see Takayama, K. **61**, 225; **74**, 115
Nagai, T., see Uchida, N. **74**, 95
Nagai, T., see Zhang, Z. **139**, 1
Nagano, H., Y. Machida, M. Iwata, T. Imada, Y. Noguchi, A. Matsumoto, T. Nagai, Preparation of magnetic granules containing bleomycin and its evaluation using model esophageal cancer **147**, 119
Naganuma, H., see Seta, Y. **41**, 263
Nagappa, A.N., see Srivastava, R.C. **38**, 239
Nagappa, A.N., see Tandon, A. **32**, 39
Nagasako, S., M. Hayashibara, Y. Katagiri, K. Iwamoto, Effect of dose on salivary excretion of mexiletine after bolus intravenous administration in rats **81**, 127
Nagase, S., see Hirate, J. **65**, 149
Nagata, M., see Narisawa, S. **148**, 85
Nagayasu, A., see Uchiyama, K. **121**, 195
Nagels, K., see Dickinson, P.A. **148**, 55
Naggar, V.F.B., N.A. Boraie, M.A. Shams-Eldeen, In vitro availability of promethazine-HCl in the presence of some commercial antacids **28**, 239
Naggar, V.F., see Eshra, A.G. **44**, 9
Naggar, V.F., see Etman, M.A. **58**, 177
Naggar, V.F., see Khalil, S.A. **19**, 307
Naggar, V.F., see Samaha, M.W. **42**, 1
Naggi, A., see Ahmed, S.M. **109**, 99
Naggi, A., see Ahmed, S.M. **77**, 247
Naidong, W., S. Hua, E. Roets, R. Busson, J. Hoogmartens, Investigation of keto-enol tautomerism and ionization of doxycycline in aqueous solutions **96**, 13
Naidoo, N.T., Encapsulation and in vitro release of indomethacin from semi-solid matrix capsules. Decomposition kinetics of erythromycin A in acidic aqueous solutions **55**, 53
Naik, A., W.J. Irwin, R.J. Griffin, The reduction of azidoprofen: A model for a soft anti-inflammatory drug for topical application **89**, 51
The hydrolysis of azidoprofen esters: A model for a soft anti-inflammatory drug for topical application **89**, 65
Percutaneous absorption of azidoprofen, a model for a soft anti-inflammatory drug for topical application **90**, 129
Naik, A., see Watkinson, A.C. **87**, 175
Nair, M., Y.W. Chien, Buccal delivery of progestational steroids. I. Characterization of barrier properties and effect of penetrant hydrophilicity **89**, 41
Naisbett, B., J. Woodley, The potential use of tomato lectin for oral drug delivery. 1. Lectin binding to rat small intestine in vitro **107**, 223
The potential use of tomato lectin for oral drug delivery. 2. Mechanism of uptake in vitro **110**, 127
The potential use of tomato lectin for oral drug delivery. 3. Bioadhesion in vivo **114**, 227
The potential use of tomato lectin for oral drug delivery. 4. Immunological consequences **120**, 247
Naisbett, B., see Pato, A.J. **104**, 227
Naito, S., M. Sekino, H. Tanaka, R. Tamai, Aprotinin concentrations in body fluids in rabbits **8**, 253
Naito, S., S. Tanaka, M. Mizuno, Concentrations of human interferons α and β in rabbit body fluids **18**, 117
Naito, S., H. Tominaga, Percutaneous absorption of diclofenac sodium ointment **24**, 115
Naito, S., Tsai Yi-Hung, Percutaneous absorption of indomethacin from ointment bases in rabbits **8**, 263
Naito, S., see Tsai, Y.H. **24**, 101
Naito, S.I., S. Nakamori, M. Awataguchi et al., Observations on and pharmacokinetic discussion of percutaneous absorption of mefenamic acid **24**, 127

Naito, S.I., S. Tanaka, M. Mizuno, H. Kawashima, Determination of human interferons α, β and γ in plasma and/or pharmaceuticals **17**, 109
Naito, S.I., see Tsai, Y.H. **8**, 203; **10**, 125; **23**, 315; **24**, 61; **28**, 47
Najib, N., see Jalal, I. **52**, 63
Najib, N., I. Jalal, Correlation between dissolution and disintegration rate constants for acetaminophen tablets **44**, 43
Najib, N.M., M.A. El-Hinnawi, M.S. Suleiman, Physicochemical characterization of ibuprofen-polyvinylpyrrolidone dispersions **45**, 139
Najib, N.M., M.S. Suleiman, The effect of hydrophilic polymers and surface active agents on the solubility of indomethacin **24**, 165
Characterization of a diflunisal polyethylene glycol solid dispersion system **51**, 225
The kinetics of dissolution of diflunisal and diflunisal-polyethylene glycol solid dispersion **57**, 197
Determination of some parameters influencing the dissolution rate of famotidine **61**, 173
The kinetics of dissolution of diflusinal and diflusinal-polyethylene glycol solid dispersion (Int. J. Pharm. 57 (1989) 197–203) **59**, 83
Najib, N.M., M. Suleiman, A. Malakh, Characteristics of the in vitro release of ibuprofen from polyvinyl pyrrolidone solid dispersions **32**, 229
Najib, N.M., see Al-Nimry, S.S. **149**, 115
Najib, N.M., see El-Hinnawi, M.A. **37**, 175
Najib, N.M., see Hassan, M.A. **58**, 19; **67**, 131; **149**, 227
Najib, N.M., see Kellaway, I.W. **6**, 285; **7**, 285; **9**, 59
Najib, N.M., see Suleiman, M.S. **50**, 71, 103; **54**, 65
Najib, N.M., see Tabba, H.D. **54**, 57
Najjar, T.A., M.I. Al-Hassan, R.M.A. Khan, Theophylline inhibits the elimination of flumazenil in rabbits **98**, 51
Najjar, T.A.O., Effect of cefoperazone on the pharmacokinetics of methotrexate in the rabbit **131**, 67
Naka, T., see Yoshimura, Y. **103**, 1
Nakabayashi, S., see Yamauchi, H. **113**, 141
Nakada, Y., M. Miyake, N. Awata, Some factors affecting the vaginal absorption of human calcitonin in rats **89**, 169
Nakagaki, E., see Sugibayashi, K. **113**, 189
Nakagaki, M., see Maitani, Y. **64**, 89; **69**, 21; **74**, 105
Nakagaki, M., see Uchida, N. **74**, 95
Nakagawa, S., see Hiura, M. **20**, 73
Nakagawa, T., see Murai, Y. **15**, 309
Nakagawa, T., see Tashima, T. **42**, 61
Nakagawa, T., see Yamaoka, K. **10**, 291
Nakahigashi, Y., see Takada, K. **101**, 89
Nakahigashi, Y., see Ushirogawa, Y. **81**, 133
Nakai, Y., see El-Gendy, G.A. **31**, 25
Nakai, Y., see Matsumoto, K. **108**, 167
Nakai, Y., see Oguchi, T. **61**, 27; **113**, 97
Nakajima, Y., see Muranushi, N. **4**, 271, 281
Nakamori, S., see Naito, S.I. **24**, 127
Nakamoto, Y., see Morimoto, K. **12**, 107
Nakamoto, Y., see Takeeda, T. **41**, 21
Nakamura, C., see Matsuo, M. **138**, 225
Nakamura, F., R. Ohta, Y. Machida, T. Nagai, In vitro and in vivo nasal mucoadhesion of some water-soluble polymers **134**, 173
Nakamura, H., Y. Pongpaibul, T. Hayashi, K. Sugibayashi, Y. Morimoto, Effect of lipophilic multicomponent system on the skin permeation of ketotifen fumarate **141**, 71
Nakamura, J., see Sasaki, H. **44**, 15; **60**, 1, 177; **93**, 49
Nakamura, J., M. Kido, K. Nishida, H. Sasaki, Hydrolysis of salicylic acid-tyrosine and salicylic acid-methionine prodrugs in the rabbit **87**, 59
Nakamura, J., M. Kido, C. Tagami, K. Nishida, H. Sasaki, Systemic and presystemic de-conjugation of salicylic acid-arginine conjugate as a prodrug of salicylic acid in rabbits **99**, 189
Nakamura, K., see Yamaguchi, H. **58**, 135
Nakamura, M., see Akimoto, M. **117**, 31
Nakamura, Y., K. Takayama, K. Higashiyama, T. Suzuki, T. Nagai, Promoting effect of *O*-ethylmenthol on the percutaneous absorption of ketoprofen **145**, 29
Nakane, S., see Nishida, K. **65**, 7
Nakano, M., N. Inotsume, N. Kohri, T. Arita, Reversible ring-opening reaction of diazepam in acid media around body temperature **3**, 195
Nakano, M., M. Itoh, K. Juni et al., Sustained urinary excretion of sulfamethizole following oral administration of enteric coated microcapsules in humans **4**, 291
Nakano, M., see Higaki, K. **132**, 165
Nakano, M., see Inotsume, N. **6**, 147; **8**, 111; **17**, 357
Nakano, M., see Matsuo, M. **138**, 225
Nakano, M., see Nakashima, M. **72**, R5
Nakano, N.I., S. Yamaguchi, Y. Shimamori, Y. Fujimoto, Fractionation of a human serum albumin preparation and the salicylate binding characteristics **7**, 335
Nakar, D., see Rubinstein, A. **84**, 141
Nakashima, M., M. Nakano, K. Matsuyama, M. Ichikawa, An application of the microdialysis system to the pharmacokinetic study on striatal distribution of L-DOPA with or without carbidopa in rats **72**, R5
Nakashima, M., see Oguma, T. **63**, 101
Nakatani, E., see Morimoto, K. **81**, 119
Nakayama, A., see Miyazaki, S. **118**, 257
Nakayama, T., T. Sawamoto, Y. kurosaki, T. Kimura, Biopharmaceutical studies on drug/conjugated-metabolite interactions. I. Fate of acetaminophen sulfate, a major conjugated metabolite of acetaminophen, in rats **121**, 55
Nakayama, T., see Kurosaki, Y. **47**, 13; **49**, 47; **51**, 47; **67**, 1; **82**, 165; **146**, 181
Naldini, A., see Bocci, V. **24**, 109; **32**, 103; **34**, 111
Nam-Tran, H., see Gander, B. **129**, 51
Nam-Tran, H., see Huu-Phuoc, N. **34**, 217
Nanayama, T., see Kakee, A. **111**, 15
Nanba, M., see Gomita, Y. **54**, 89
Nangia, A., E. Bloom, B. Berner, H. Maibach, Human keratinocyte cell culture for studying skin irritation in man? **99**, 67
Nangia, A., see Melendres, J.L. **92**, 243

Nankervis, R., S.S. Davis, N.H. Day, P.N. Shaw, Studies on the intravenous pharmacokinetics of three retinoids in the rat **101**, 249
Effect of lipid vehicle on the intestinal lymphatic transport of isotretinoin in the rat **119**, 173
Intestinal lymphatic transport of three retinoids in the rat after oral administration: Effect of lipophilicity and lipid vehicle **130**, 57
Nannipieri, E., see Bilia, A. **130**, 83
Nannipieri, E., see Bonina, F.P. **100**, 41
Nannipieri, E., see Buonaguidi, M. **147**, 1
Nannipieri, E., see Carelli, V. **30**, 9; **35**, 21; **50**, 181; **55**, 199; **88**, 89; **89**, 81; **94**, 103
Nannipieri, E., see Di Colo, G. **30**, 1; **50**, 27
Naon, R., see Privitera, N. **104**, 41; **120**, 73
Narahari, U., see Foster, T.M. **134**, 193
Narayana Reddy, M., see Sastry, C.S.P. **39**, 137
Narayani, R., K.P. Rao, Preparation, characterisation and in vitro stability of hydrophilic gelatin microspheres using a gelatin-methotrexate conjugate **95**, 85
Biodegradable microspheres using two different gelatin drug conjugates for the controlled delivery of methotrexate **128**, 261
Collagen-poly (HEMA) hydrogels for the controlled delivery of methotrexate and cisplatin **138**, 121
Solid tumor chemotherapy using injectable gelatin microspheres containing free methotrexate and conjugated methotrexate **142**, 25
Gelatin microsphere cocktails of different sizes for the controlled release of anticancer drugs **143**, 255
Narisawa, S., M. Nagata, Y. Hirakawa, M. Kobayashi, H. Yoshino, An organic acid-induced sigmoidal release system for oral controlled-release preparations. III. Elucidation of the anomalous drug release behavior through osmotic pumping mechanism **148**, 85
Narisawa, S., H. Yoshino, Y. Hirakawa, K. Noda, Porosity-controlled ethylcellulose film coating. II. Spontaneous porous film formation in the spraying process and its solute permeability **104**, 95
Narisawa, S., see Uekama, K. **13**, 253; **16**, 327
Narurkar, M.M., see Dalal, P.S. **73**, 157
Narurkar, M.M., see Mitra, A.K. **28**, 119; **35**, 165
Nash, S., see Barnes, A.R. **94**, 231; **139**, 255
Nashihata, T., M. Miyake, H. Takahata, A. Kamada, The effect of adjuvants on the colonic absorption of cefmetazole and [Asu^{17}]-eel calcitonin in rats: Concentration dependent absorption pathways **33**, 89
Nastruzzi, C., E. Esposito, C. Pastesini, R. Gambari, E. Menegatti, Comparative study on the release kinetics of methyl-nicotinate from topic formulations **90**, 43
Nastruzzi, C., see Cortesi, R. **105**, 181; **139**, 69
Nastruzzi, C., see Esposito, E. **129**, 263; **142**, 9
Nastruzzi, C., see Gambari, R. **72**, 251
Nation, R.L., see Ridd, M.J. **11**, 11
Natsume, H., K. Sugibayashi, K. Juni, Y. Morimoto, T. Shibata, S. Fujimoto, Preparation and evaluation of biodegradable albumin microspheres containing mitomycin C **58**, 79
Natsume, H., see Kubo, H. **103**, 27
Natsume, H., see Morimoto, Y. **54**, 27
Natsume, H., see Natsume, H. **58**, 79
Navarro, A.S., see Sanchez, B.M. **106**, 229
Navarro, A.S., A. Castano, J.M. Lanao, A. Dominguez-Gil, In vitro interaction between HAPA-gentamicin B and penicillins **37**, 113
Navas, M.J., see Asuero, A.G. **34**, 81; **40**, 43
Naveau, C., see Jacolot, A. **128**, 283
Naviasky, H., see Shah, V.P. **125**, 99
Na'was, T.E., see Alkaysi, H.N. **87**, 73
Nazir, T., see Leigh, T.R. **67**, 275
N'Dongo, M., see Chemtob, C. **29**, 1, 83
Neau, S.H., M.Y. Chow, L. Chafetz, The influence of reaction conditions on the derivatization of carbidopa by β-dicarbonyl compounds for enhanced optical rotation **108**, 95
Neau, S.H., M.Y. Chow, M.J. Durrani, Fabrication and characterization of extruded and spheronized beads containing Carbopol^{R} 974P, NF resin **131**, 47
Neau, S.H., G.L. Flynn, S.H. Yalkowsky, The influence of heat capacity assumptions on the estimation of solubility parameters from solubility data **49**, 223
Neau, S.H., D.R. Howard, Partial molal volumes of *n*-alkyl *p*-aminobenzoates in nonpolar solvents **77**, 239
Neau, S.H., M.K. Shinwari, E.W. Hellmuth, Melting point phase diagrams of free base and hydrochloride salts of bevantolol, pindolol and propranolol **99**, 303
Neau, S.H., see Claudius, J.S. **144**, 71
Neau, S.H., see Katikaneni, P.R. **117**, 13; **123**, 119
Neau, S.H., see Upadrashta, S.M. **112**, 173
Nederman, T., see Sjodin, L. **63**, 135
Needham, T., see Dua, R. **147**, 233
Needham, T.E., see Dondeti, P. **122**, 91; **127**, 115
Neervannan, S., J.D. Reinert, V.J. Stella, M.Z. Southard, A numerical convective-diffusion model for dissolution of neutral compounds under laminar flow conditions **96**, 167
Neil, J.M., A.F. Fell, G. Smith, Evaluation of the stability of frusemide in intravenous infusions by reversed-phase high-performance liquid chromatography **22**, 105
Nejmeh, M., see Pilpel, N. **5**, 273
Nelis, H., see Voorspoels, J. **133**, 9
Nelson, K.F., see Choudhury, S. **85**, 175
Nemati, F., C. Dubernet, A.C. de Verdiere, M.F. Poupon, L. Treupel-Acar, F. Puisieux, P. Couvreur, Some parameters influencing cytotoxicity of free doxorubicin and doxorubicin-loaded nanoparticles in sensitive and multidrug resistant leucemic murine cells: Incubation time, number of nanoparticles per cell **102**, 55
Nemati, F., C. Dubernet, H. Fessi, A.C. de Verdiere, M.F. Poupon, F. Puisieux, P. Couvreur, Reversion of multidrug resistance using nanoparticles in vitro: Influence of the nature of the polymer **138**, 237
Nemati, F., see Skiba, M. **145**, 241
Nequist, G., see Patel, J.P. **65**, 195
Nesbitt, J.D., see Wagner, D.L. **57**, 177
Nesbitt, R.U., F.W. Goodhart, R.H. Gordon, Evaluation of polyvinyl acetate phthalate as an enteric coating material **26**, 215

Nesbitt, R.U., see Ghebre-Sellassie, I. **37**, 211
Neubert, R., C. Bendas, W. Wohlrab, B. Gienau, W. Furst, A multilayer membrane system for modelling drug penetration into skin **75**, 89
Neubert, R., see Bendas, B. **116**, 19
Neubert, R., see Dongowski, G. **144**, 233
Neubert, R., see Furst, W. **61**, 43
Neubert, R., see Schneider, I.M. **145**, 187
Neubert, R., see Wegener, M. **128**, 203
Newman, A., see Wesdyk, R. **65**, 69
Newman, A.W., see Desai, D.S. **91**, 217
Newman, A.W., see Morris, K.R. **105**, 209; **108**, 195
Newman, A.W., see Wesdyk, R. **93**, 101
Newman, M.S., see Woodle, M.C. **88**, 327
Newman, S., see Borgstrom, L. **97**, 47
Newman, S., see Pitcairn, G. **102**, 11
Newman, S.P., A.R. Clark, N. Talaee, S.W. Clarke, Lung deposition of 5 mg Intal from a pressurised metered dose inhaler assessed by radiotracer technique **74**, 203
Newman, S.P., A. Hollingworth, A.R. Clark, Effect of different modes of inhalation on drug delivery from a dry powder inhaler **102**, 127
Newman, S.P., M. Killip, D. Pavia et al., The effect of changes in particle size on the deposition of pressurized inhalation aerosols **19**, 333
Newman, S.P., F. Moren, E. Trofast, N. Talaee, S.W. Clarke, Terbutaline sulphate Turbuhaler: Effect of inhaled flow rate on drug deposition and efficacy **74**, 209
Newman, S.P., P.G.D. Pellow, S.W. Clarke, Efficient nebulisation of powdered antibiotics **36**, 55
Newman, S.P., see Dewsbury, N.J. **137**, 261
Newman, S.P., see Steed, K.P. **123**, 291
Newton, D.W., S. Ratanamaneichatara, W.J. Murray, Dissociation, solubility and lipophilicity of azathioprine **11**, 209
Newton, D.W., see Al-Turk, W.A. **41**, 227
Newton, J.M., G. Alderborn, C. Nystrom, R.J. Prankerd, The compressive to tensile strength ratio of pharmaceutical compacts **93**, 249
Newton, J.M., S.R. Chapman, R.C. Rowe, The assessment of the scale-up performance of the extrusion/spheronisation process **120**, 95
The influence of process variables on the preparation and properties of spherical granules by the process of extrusion and spheronisation **120**, 101
Newton, J.M., see Baert, L. **86**, 187
Newton, J.M., see Bains, D. **69**, 233
Newton, J.M., see Blanque, D. **119**, 203
Newton, J.M., see Chawla, A. **108**, 233
Newton, J.M., see Chohan, R.K. **131**, 201
Newton, J.M., see Clarke, G.M. **100**, 81; **114**, 1
Newton, J.M., see Craig, D.Q.M. **74**, 33, 43; **76**, 17; **78**, 175
Newton, J.M., see Eriksson, M. **148**, 149
Newton, J.M., see Fielden, K.E. **79**, 47; **81**, 205, 225; **97**, 79
Newton, J.M., see Harrison, P.J. **35**, 235
Newton, J.M., see Jerwanska, E. **149**, 131
Newton, J.M., see Jerwanska, E. **121**, 65
Newton, J.M., see Lundqvist, A.E.K. **147**, 95
Newton, J.M., see Mackellar, A.J. **112**, 65, 79
Newton, J.M., see Odidi, I.O. **72**, 43; **90**, 203
Newton, J.M., see Pinto, J.F. **83**, 187; **118**, 95; **147**, 79
Newton, J.M., see Podczeck, F. **124**, 253; **145**, 221; **149**, 151
Newton, J.M., see Sousa, J.J. **144**, 159
Newton, J.M., see Sutananta, W. **110**, 75; **111**, 51; **125**, 123; **132**, 1
Newton, J.M., see Tan, S.B. **61**, 145; **63**, 275; **64**, 227; **66**, 207, 273, 283
Newton, J.M., see Tapia, C. **92**, 211
Newton, J.M., see Yuen, K.H. **97**, 61
Nghiem, B., see Wong, O. **52**, 191
Nghiem, B.T., T. Higuchi, Esterase activity in snake skin **44**, 125
Nghiem, B.T., see Akazawa, M. **50**, 53
Ngombo, Mukendi J., see Vandamme, Th.F. **132**, 153; **145**, 77
Nguyen, F., see Peyrot, M. **102**, 25
Nguyen, N., see Laurin, P. **28**, 177
Nguyen, N.T., see Kaul, D. **88**, 345
Nguyen, P.A., see Maa, Y.F. **144**, 47
Nguyen, T., see Small, D. **149**, 203
Nguyen, T.H., K.J. Himmelstein, T. Higuchi, Some equilibrium and kinetic aspects of water sorption in poly(*ortho*-ester)s **25**, 1
Nguyen, T.H., see Lam, X.M. **142**, 85
Nguyen, T.H., see Wilke, T.L. **103**, 187
Ni, X.R., see Jansen, A.C.A. **75**, 193
Nia, B., E.M. Ouriemchi, J.M. Vergnaud, Calculation of the blood level of a drug taken orally with a diffusion controlled dosage form **119**, 165
Niazy, E.M., Influence of oleic acid and other permeation promoters on transdermal delivery of dihydroergotamine through rabbit skin **67**, 97
Differences in penetration-enhancing effect of Azone through excised rabbit, rat, hairless mouse, guinea pig and human skins **130**, 225
Bioavailability of meclofenamate from experimental sustained release microcapsules in beagle dogs **134**, 129
Niazy, E.M., A.M. Molokhia, A.S. El-Gorashi, Effect of Azone and other penetration enhancers on the percutaneous absorption of dihydroergotamine **56**, 181
Niazy, E.M., see Al-Shora, H.I. **47**, 209
Niazy, E.M., see El-Sayed, Y.M. **123**, 113
Niazy, E.M., see Hosny, E.A. **117**, 147; **136**, 37
Nicholls, D., see Walker, I. **104**, 157; **108**, 225
Nicholls, P.J., see Hoshino, T. **38**, 265
Nicholls, T.J., K.L. Green, D.J. Rogers, J.D. Cook, S. Wolowacz, J.D. Smart, Lectins in ocular drug delivery: An investigation of lectin binding sites on the corneal and conjunctival surfaces **138**, 175
Nicklasson, M., A. Brodin, The relationship between intrinsic dissolution rates and solubilities in the water–ethanol binary solvent system **18**, 149
Nicklasson, M., A. Brodin, L.O. Sundelof, On the determination of true dissolution rate parameters from rotating disc experiments **15**, 87

Studies of some characteristics of molecular dissolution kinetics from rotating discs **23**, 97
Nicklasson, M., C. Graffner, M. Nilsson, Assessment of in vivo drug dissolution by means of numerical deconvolution considering gastrointestinal availability **40**, 165
Nicklasson, M., A. Orbe, J. Lindberg, B. Borga, A.B. Magnusson, G. Nilsson, R. Ahlgren, L. Jacobsen, A collaborative study of the in vitro dissolution of phenacetin crystals comparing the flow through method with the USP Paddle method **69**, 255
Nicklasson, M., B. Wennergren, J. Lindberg et al., A collaborative in vitro dissolution study using the flow-through method **37**, 195
Nicklasson, M., see Arwidsson, H. **58**, 73
Nicklasson, M., see Fyhr, P. **40**, 193
Nicklasson, M., see Wennergren, B. **53**, 35
Nicklin, P., B. Irwin, I. Hassan, I. Williamson, M. Mackay, Permeable support type influences the transport of compounds across Caco-2 cells **83**, 197
Nicklin, P., A.C. Keates, T. Page, C.J. Bailey, Transfer of metformin across monolayers of human intestinal Caco-2 cells and across rat intestine **128**, 155
Nicklin, P.L., W.J. Irwin, I.F. Hassan, M. Mackay, Development of a minimum-calcium Caco-2 monolayer model: Calcium and magnesium ions retard the transport of pamidronate **123**, 187
Nicklin, P.L., W.J. Irwin, P. Timmins, R.A. Morrison, Uptake and transport of the ACE-inhibitor ceronapril (SQ 29852) by monolayers of human intestinal absorptive (Caco-2) cells in vitro **140**, 175
Nicol, A.C., see McElnay, J.C. **19**, 89
Nicolas, A., see Leroy, P. **82**, 157
Nicolay, K., see Talsma, H. **37**, 171
Niederer, R.R., see May, H.A. **19**, 169
Nielsen, G.H., see Bundgaard, H. **27**, 71
Nieisen, H.M., see Brondsted, H. **114**, 151
Nielsen, H.M., see Hovgaard, L. **114**, 141
Nielsen, L.S., Improved peroral bioavailability of mebendazole in rabbits by administration of various *N*-alkoxycarbonyl derivatives of mebendazole **104**, 175
Nielsen, L.S., F. Slok, H. Bundgaard, *N*-alkoxycarbonyl prodrugs of mebendazole with increased water solubility **102**, 231
Nielsen, N.M., H. Bundgaard, Prodrugs as drug delivery systems. Part 42. 2-Hydroxymethylbenzamides and 2-acyloxymethylbenzamides as potential prodrug forms for amines **29**, 9
Nielsen, N.M., see Bundgaard, H. **42**, 217; **43**, 101; **44**, 151
Nielsen, O., see Hansen, J. **16**, 141
Nielsen, V.F., see Pedersen, M. **90**, 247
Niemi, L., E. Laine, Effect of water content on the microstructure of an o/w cream **68**, 205
Nightingale, A.S., see Liu, P. **90**, 171
Nightingale, J., see Goldberg-Cettina, M. **114**, 237
Nightingale, J., see Sclafani, J. **124**, 213
Nightingale, J., see Wilding, I.R. **132**, 81
Niibuchi, J.J., Y. Aramaki, S. Tsuchiya, Binding of antibiotics to rat intestinal mucin **30**, 181
Nikore, R.L., see Pathak, Y.V. **24**, 351
Nikupaavo, P., see Heinamaki, J.T. **112**, 191
Nilsson, G., see Wennergren, B. **53**, 35
Nilsson, M., see Nicklasson, M. **40**, 165
Nilsson, P., M. Westerberg, C. Nystrom, Physicochemical aspects of drug release. V. The importance of surface coverage and compaction on drug dissolution from ordered mixtures **45**, 111
Nimier, K., D. Robic, R. Bourbouze, Macromolecular aggregation of F1 and P1 fractions of RU 41740 an immunomodulating compound isolated from *Klebsiella pneumoniae*, as revealed by size exclusion high-performance liquid chromatography and light scattering detection **105**, 235
Nimmerfall, F., J. Rosenthaler, Modified release of drug: A way to its quantification **32**, 1
Ning, A., see Palin, K.J. **33**, 99
Niphadkar, P., see D'Souza, M. **31**, 165
Nisato, D., see Delie, F. **115**, 45
Nishida, K., C. Tonegawa, S. Nakane, Y. Takakura, M. Hashida, H. Sezaki, Effect of electric charge on the hepatic uptake of macromolecules in the rat liver **65**, 7
Nishida, K., T. Takino, Y. Eguchi, F. Yamashita, M. Hashida, H. Sezaki, Pharmacokinetic analysis of uptake process of lactosaminated albumin in rat liver constant infusion experiments **80**, 101
Nishida, K., see Nakamura, J. **87**, 59; **99**, 189
Nishida, K., see Sasaki, H. **93**, 49
Nishida, T., see Uchiyama, K. **121**, 195
Nishigaki, R., see Shim, C.K. **8**, 143
Nishihata, T., Simple formulation of sustained-release tablets of sodium diclofenac and examination in humans **40**, 125
Nishihata, T., G.M. Ghaly, H. Inagaki, A. Kamada, T. Yagi, R. Kawamori, M. Shichiri, An effective formulation for an insulin suppository; examintion in normal dogs **38**, 83
Nishihata, T., Y. Hirotani, A. Yamazi, K. Takahashi, H. Yoshitomi, Preliminary study on sustained-release particles prepared with hydrogenated soya phospholipid and cholesterol **42**, 257
Nishihata, T., S. Kawabe, M. Miyake et al., Effect of sodium ion on the adjuvant action of sodium salicylate in enhancing rectal ampicillin absorption in rats **22**, 147
Nishihata, T., M. Keigami, A. Kamada, T. Fujimoto, S. Kamide, N. Tatsumi, Clinical investigation of sodium diclofenac sustained-release suppositories **42**, 251
Nishihata, T., S. Kim, G.G. Liversidge, T. Higuchi, Rectal absorption and lymphatic uptake of Ara-C in rats **31**, 185
Nishihata, T., Y. Okamura, H. Inagaki et al., Trials of rectal insulin suppositories in healthy humans **34**, 157
Nishihata, T., J.H. Rytting, T. Higuchi et al., Enhancement of rectal absorption of water-soluble antibiotics in dogs **21**, 239
Nishihata, T., M. Sudho, A. Kamada et al., Investigation of sustained-release suppository of sodium diclofenac in humans **33**, 181

Nishihata, T., H. Wada, A. Kamada, Sustained-release of sodium diclofenac from suppository **27**, 245
Nishihata, T., see Bhattachar, S.N. **79**, 263
Nishihata, T., see Kim, S. **21**, 179
Nishihata, T., see Liversidge, G.G. **23**, 87; **30**, 247
Nishihata, T., see Tahara, K. **99**, 311; **133**, 17
Nishihata, T., see Takahashi, K. **49**, 179
Nishihata, T., see Yamamoto, K. **102**, 133
Nishikawa, M., Y. Ohtsubo, J. Ohno, T. Fujita, Y. Koyama, F. Yamashita, M. Hashida, H. Sezaki, Pharmacokinetics of receptor-mediated hepatic uptake of glycosylated albumin in mice **85**, 75
Nishikawa, M., see Takakura, Y. **105**, 19
Nishimura, H., see Kurosaki, Y. **67**, 1; **82**, 165
Nishimura, K., see Seta, Y. **41**, 245, 255
Nishimura, K., see Wakiyama, N. **78**, 95
Nishimura, M., see Yamaguchi, T. **85**, 87
Nishio, M., see Adachi, T. **124**, 129
Nitta, C., see Kadir, S. **40**, 257
Nitta, K., see Ohwaki, T. **85**, 19
Niu, S., see Zhang, J. **101**, 15
Niven, R.W., J.D. Brain, Some functional aspects of air-jet nebulizers **104**, 73
Niven, R.W., A.Y. Ip, S.D. Mittelman, C. Farrar, T. Arakawa, S.J. Prestrelski, Protein nebulization. I. Stability of lactate dehydrogenase and recombinant granulocyte-colony stimulating factor to air-jet nebulization **109**, 17
Niven, R.W., F.D. Lott, A.Y. Ip, K.D. Somaratne, M. Kearney, Development and use of an in vitro system to evaluate inhaler devices **101**, 81
Niven, R.W., S.J. Prestrelski, M.J. Treuheit, A.Y. Ip, T. Arakawa, Protein nebulization. II. Stabilization of G-CSF to air-jet nebulization and the role of protectants **127**, 191
Niwa, T., H. Takeuchi, T. Hino, M. Nohara, Y. Kawashima, Biodegradable submicron carriers for peptide drugs: Preparation of DL-lactide/glycolide copolymer (PLGA) nanospheres with nafarelin acetate by a novel emulsion-phase separation method in an oil system **121**, 45
Niwa, K., see Takaya, T. **110**, 47
Niwa, T., see Kawashima, Y. **72**, 65; **75**, 25; **89**, 9; **99**, 229; **119**, 139
Niwa, T., see Morishima, K. **105**, 11
Niwa, T., see Murakami, H. **149**, 43
Niwa, T., see Ohwaki, T. **85**, 19; **93**, 61
Niwa, T., see Schade, A. **117**, 209
Niwa, T., see Takeuchi, H. **86**, 25
Nixon, J.R., G.A. Agyilirah, The influence of colloidal proportions on the release of phenobarbitone from microcapsules **6**, 277
Nixon, J.R., K.T. Wong, Evaluation of permeation through polymeric membranes as a model for the release of drugs from gelatin-acacia walled microcapsules **50**, 205
Evaluation of drug permeation through polymeric membranes as a model for release (II) ethylcellulose-walled microcapsules **58**, 31
Nixon, J.R., see Avgoustakis, K. **70**, 77; **99**, 239, 247
Nixon, P.R., see Sinko, C.M. **81**, 243
Nixon, P.R., see Sung, K.C. **142**, 53
Noach, A.B.J., Y. Kurosaki, M.C.M. Blom-Roosemalen, Boer A.G. De, D.D. Breimer, Cell-polarity dependent effect of chelation on the paracellular permeability of confluent Caco-2 cell monolayers **90**, 229
Nobile, F., see Lovrecich, M. **131**, 247
Nobile, L., V. Cavrini, M.A. Raggi, Analysis of thiamphenicol glycinate salts in pharmaceutical formulations by derivative UV spectroscopy **40**, 85
Nobile, L., see Raggi, M.A. **61**, 199
Noda, K., see Narisawa, S. **104**, 95
Noe, C., see Pirker, S. **128**, 189
Noguchi, A., see Ogiso, T. **102**, 141
Noguchi, M., see Sugito, K. **47**, 157; **60**, 89
Noguchi, T., W.N.A. Charman, V.J. Stella, The effect of drug lipophilicity and lipid vehicles on the lymphatic absorption of various testosterone esters **24**, 173
Lymphatic appearance of DDT in thoracic or mesenteric lymph duct cannulated rats **24**, 185
Noguchi, T., see Charman, W.N.A. **33**, 155, 173
Noguchi, T., see Yamahira, Y. **3**, 23
Noguchi, Y., see Nagano, H. **147**, 119
Nohara, M., see Niwa, T. **121**, 45
Nokhodchi, A., J.L. Ford, P.H. Rowe, M.H. Rubinstein, The effects of compression rate and force on the compaction properties of different viscosity grades of hydroxypropylmethylcellulose 2208 **129**, 21
Nokhodchi, A., M.H. Rubinstein, J.L. Ford, The effect of particle size and viscosity grade on the compaction properties of hydroxypropylmethylcellulose 2208 **126**, 189
Nokhodchi, A., M.H. Rubinstein, H. Larhrib, J.C. Guyot, The effect of moisture on the properties of ibuprofen tablets **118**, 191
The effect of moisture content on the energies involved in the compaction of ibuprofen **120**, 13
Nolan, J.C., see Cefali, E.A. **127**, 147
Nolen III, H.W., P. Catz, D.R. Friend, Percutaneous penetration of methyl phosphonate antisense oligonucleotides **107**, 169
Nomura, A., see Kohri, N. **81**, 49
Nomura, H., see Watanabe, Y. **110**, 93
Nook, T., E. Doelker, P. Buri, Intestinal absorption kinetics of various model drugs in relation to partition coefficients **43**, 119
Noonan, P.K., L.Z. Benet, Formation of mono- and di-nitrate metabolites of nitroglycerin following incubation with human blood **12**, 331
Noonan, P.K., see Wester, R.C. **7**, 99
Noormohammadi, A., see Avgerinos, A. **68**, 97
Noormohammadi, A., see Hadzija, B.W. **67**, 185
Noory, A., see Shah, V.P. **125**, 99
Noory, C., see Shah, V.P. **125**, 99
Nord, K., see Bjerga Bjaen, A.K **92**, 183
Nord, K., J. Karlsen, H.H. Tonnesen, Photochemical stability of biologically active compounds. IV. Photochemical degradation of chloroquine **72**, 11
Nordstrom, A.J., see Heinamaki, J.T. **109**, 9

Norgaard, T., see Bundgaard, H. **42**, 217
Norman, S., see Washington, N. **57**, 17
Norstedt, G., see Prisell, P.T. **85**, 51
Noszal, B., see Takacs-Novak, K. **89**, 261
Notari, R.E., M.Y. Huang, P.R. Byron, Calculations of optimum pharmacokinetic drug supply rates for maximum duration during multiple dose therapy by prodrug administration **1**, 233
Notari, R.E., see Byron, P.R. **1**, 219
Notari, R.E., see D'Silva, J.B. **8**, 45
Nour, I.M., see Ibrahim, K.E. **98**, 231
Nouravarsani, F., J. Cobby, Measurement of protein binding for drugs that are unstable in aqueous solution **11**, 45
Novak, C., see Arias-Blanco, M.J. **143**, 113
Novikov, D.K., see Kosykh, V.A. **57**, 223
Ntawukulilyayo, J.D., S. Bouckaert, J.P. Remon, Low substituted *N*-octenylsuccinate starch as a dissolution rate enhancer in solid dosage forms **91**, 23
Enhancement of dissolution rate of nifedipine using sucrose ester coprecipitates **93**, 209
Ntawukulilyayo, J.D., C. Demuynck, J.P. Remon, Microcrystalline cellulose-sucrose esters as tablet matrix forming agents **121**, 205
Ntawukulilyayo, J.D., S.C. De Smedt, J. Demeester, J.P. Remon, Stabilisation of suspensions using sucrose esters and low substituted *n*-octenylsuccinate starch-xanthan gum associations **128**, 73
Ntawukulilyayo, J.D., C. Vervaet, J.P. Remon, J.P. Gortz, J.A. Berlo, In vitro and in vivo evaluation of a xanthan gum-*n*-octenylsuccinate starch matrix tablet containing ibuprofen as a model drug **139**, 79
Numajiri, S., see Yamaguchi, H. **58**, 135
Nunez, A., see Remunan, C. **80**, 151
Nurnberg, E., see Jachowicz, R. **99**, 321
Nuutinen, J., see Vidgren, M. **42**, 211; **64**, 1; **73**, 131
Nyberg, G., see Wennergren, B. **53**, 35
Nykanen, S., see Marvola, M. **53**, 145
Nyqvist, H., see Davidson, R. **100**, 49
Nyqvist, H., see Ek, R. **124**, 9
Nyska, A., see Hoffman, A. **111**, 197
Nystrom, C., J. Mazur, J. Sjogren, Studies on direct compression of tablets. II. The influence of the particle size of a dry binder on the mechanical strength of tablets **10**, 209
Nystrom, C., M. Glazer, Studies on direct compression of tablets. XIII. The effect of some dry binders on the tablet strength of compounds with different fragmentation propensity **23**, 255
Nystrom, C., see Adolfsson, A. **132**, 95
Nystrom, C., see Alderborn, G. **23**, 79
Nystrom, C., see Anderberg, E.K. **47**, 67; **62**, 143
Nystrom, C., see Angberg, M. **61**, 67; **73**, 209; **77**, 269; **81**, 153; **83**, 11; **90**, 19
Nystrom, C., see Bisrat, M. **47**, 223; **63**, 49; **80**, 191
Nystrom, C., see Ek, R. **111**, 43; **122**, 49; **124**, 9; **125**, 257
Nystrom, C., see Elamin, A.A. **111**, 159
Nystrom, C., see Eriksson, M. **63**, 189; **99**, 197; **148**, 149
Nystrom, C., see Gren, T. **74**, 49
Nystrom, C., see Jerwanska, E. **121**, 65; **149**, 131
Nystrom, C., see Karehill, P.G. **61**, 251; **64**, 27, 35
Nystrom, C., see Mosharraf, M. **122**, 35, 57
Nystrom, C., see Newton, J.M. **93**, 249
Nystrom, C., see Nilsson, P. **45**, 111
Nystrom, C., see Olsson, H. **143**, 233
Nystrom, C., see Sjokvist, E. **47**, 51; **54**, 161; **67**, 139; **69**, 53; **79**, 123
Nystrom, C., see Sjokvist, Saers E. **90**, 105
Nystrom, C., see Westerberg, M. **28**, 23; **69**, 129; **90**, 1

O

Obach, R., see Calpena, A.C. **103**, 179
Obara, S., J.W. McGinity, Influence of processing variables on the properties of free films prepared from aqueous polymeric dispersions by a spray technique **126**, 1
Obata, Y., K. Takayama, Y. Maitani, Y. Machida, T. Nagai, Effect of ethanol on skin permeation of nonionized and ionized diclofenac **89**, 191
Obata, Y., see Takayama, K. **61**, 225
Obeng, E.K., see Hadzija, B.W. **67**, 185
Obi, C.E., see Okor, R.S. **58**, 89; **59**, 83
Obiorah, B.A., Possible prediction of compression characteristics from pressure cycle plots **1**, 249
O'Callaghan, R.J., see Callegan, M.C. **123**, 173
Ochekpe, N.A., see Taylor, R.B. **60**, 185
Ochoa, Machiste E., G. Buckton, Dynamic surface tension studies of hydroxypropylmethylcellulose film-coating solutions **145**, 197
Ochoa, Machiste E., P. Giunchedi, M. Setti, U. Conte, Characterization of carbamazepine in systems containing a dissolution rate enhancer **126**, 65
O'Connor, C.A., see Attwood, D. **65**, 201; **70**, 147
O'Connor, R.E., see Clas, S.D. **121**, 73
Oda, M., see Miyazaki, S. **118**, 257
Odidi, I.O., J.M. Newton, Cylindrical tube and surface tension viscous flow models in the assessment of capillary flow and liquid contact angles in pharmaceutical powders **90**, 203
Odidi, I.O., J.M. Newton, G. Buckton, The effect of surface treatment on the values of contact angles measured on a compressed powder surface **72**, 43
Odoardi, P., see Di Giulio, A. **74**, 183
O'Driscoll, C.M., R.A. Myers, V.J. Stella, Blood and lymph transport of DDT after oral and parenteral administration to anaesthetised rats **73**, 177
O'Driscoll, C.M., see O'Reilly, J.R. **105**, 137; **109**, 147
Oechslein, C.R., G. Fricker, T. Kissel, Nasal delivery of octreotide: Absorption enhancement by particulate carrier systems **139**, 25
Offringa, J.C.A., see De, Smidt J.H. **77**, 255
Ogata, H., N. Aoyagi, N. Kaniwa et al., Bioavailability of metronidazole from sugar-coated tablets in humans. I. Effect of gastric acidity and correlation with in vitro dissolution rate **23**, 277
Bioavailability of metronidazole from sugar-coated tablets in humans. II. Evaluation of beagle dogs as an animal model **23**, 289

Gastric acidity dependent bioavailability of cinnarizine from two commercial capsules in healthy volunteers **29**, 113
Evaluation of beagle dogs as an animal model for bioavailability testing of cinnarizine capsules **29**, 121
Ogata, H., H.L. Fung, Effect of sesame oil emulsion on nitroglycerin pharmacokinetics **5**, 335
Ogata, H., see Akimoto, M. **117**, 31
Ogata, H., see Sugito, K. **47**, 157; **60**, 89
Ogawa, K., see Kaneo, Y. **57**, 263
Ogawa, N., see Jay, M. **19**, 97
Ogawa, S., see Miyamoto, Y. **149**, 25
Ogawa, Y., see Heya, T. **69**, 69; **72**, 199
Ogawa, Y., see Iga, K. **57**, 241
Ogawa, Y., see Okada, H. **54**, 231
Ogawa, Y., see Takada, S. **146**, 147
Ogawa, Y., see Yanai, S. **123**, 237
Ogihara, M., see Inoue, N. **137**, 75
Ogihara, M., see Ueda, H. **137**, 217; **143**, 37
Ogiso, T., M. Iwaki, Z. Jing-ping, T. Tanino, A. Noguchi, Percutaneous absorption of betahistine through rat skin and pharmacokinetic analysis of the plasma concentration **102**, 141
Ogiso, T., M. Iwaki, T. Tanino, M. Yatomi, C. Tsujimoto, Percutaneous absorption of terodiline and the membrane-controlled transdermal therapeutic system **98**, 113
O'Grady, J., see Carter, K.C. **53**, 129
Oguchi, T., K. Matsumoto, E. Yonemochi, Y. Nakai, K. Yamamoto, Dissolution studies in organic solvents for evaluating hydrogen-bond matrix of cellulose in the ground mixture **113**, 97
Oguchi, T., M. Okada, E. Yonemochi, K. Yamamoto, Y. Nakai, Freeze-drying of drug-additive binary systems. III. Crystallization of α-cyclodextrin inclusion complex in freezing process **61**, 27
Oguchi, T., see Matsumoto, K. **108**, 167
Oguma, T., K. Shimamura, Y. Ushio, H. Yamada, K. Takahashi, N. Muranushi, T. Yoshikawa, N. Nakashima, T. Uematsu, Pharmacokinetic analysis of discontinuous absorption process of ceftibuten in humans **63**, 101
Ogunbona, F.A., O.O. Oluwatudimu, Effect of a Non-European (Nigerian) meal on the bioavailability of nitrofurantoin in man **29**, 191
Oh, D.M., P.J. Sinko, G.L. Amidon, Characterization of the oral absorption of several aminopenicillins: Determination of intrinsic membrane absorption parameters in the rat intestine in situ **85**, 181
Oh, I., S.C. Chi, B.R. Vishnuvajjala, B.D. Anderson, Stability and solubilization of oxathiin carboxanilide, a novel anti-HIV agent **73**, 23
Oh, I.J., H.M. Song, K.C. Lee, Effect of 2-hydroxypropyl-β-cyclodextrin on the stability of prostaglandin E_2 in solution **106**, 135
Oh, I.J., see Kim, C.K. **124**, 61
Oh, I.J., see Lee, K.C. **106**, 167
Oh, K., see Uekama, K. **25**, 339
Oh, K.H., see Hwang, S.J. **116**, 125
Oh, K.H., see Kim, C.K. **106**, 213; **124**, 61; **146**, 63
Oh, K.H., see Lim, C.R. **132**, 175
Oh, K.H., see Bodmeier, R. **51**, 1
Oh, S.Y., R.H. Guy, Effects of iontophoresis on the electrical properties of human skin in vivo **124**, 137
Oh, U., see Kim, C.K. **146**, 63
Oh, Y.K., see Kim, C.K. **47**, 163; **101**, 191; **128**, 65
O'Hagan, D.T., H. Jeffery, S.S. Davis, The preparation and characterization of poly(lactide-*co*-glycolide) microparticles. III. Microparticle/polymer degradation rates and the in vitro release of a model protein **103**, 37
O'Hagan, D.T., J.P. McGee, M. Lindblad, J. Holmgren, Cholera toxin B subunit (CTB) entrapped in microparticles shows comparable immunogenicity to CTB mixed with whole cholera toxin following oral immunization **119**, 251
O'Hagan, D.T., D. Rahman, H. Jeffery, S. Sharif, S.J. Challacombe, Controlled release microparticles for oral immunization **108**, 133
O'Hagan, D.T., see Fisher, A.N. **74**, 147
O'Hagan, D.T., see Illum, L. **63**, 207
O'Hagan, D.T., see Jeffery, H. **77**, 169
O'Hagan, D.T., see Sharif, S. **115**, 259; **119**, 239
O'Hanrahan, M.T., see Harte, V.J. **1**, 165
Ohara, N., K. Takayama, Y. Machida, T. Nagai, Combined effect of D-limonene and temperature on the skin permeation of ketoprofen **105**, 31
Combined effect of D-limonene and temperature on the skin permeation of ketoprofen (Int. J. Pharm. 105 (1994) 31–38) **119**, 125
Ohara, N., K. Takayama, T. Nagai, Influence of temperature on the percutaneous absorption for lipophilic and hydrophilic drugs across the rat skin pretreated with oleic acid **123**, 281
Ohm, A., see Lippold, B.C. **28**, 67
Ohmura, T., see Sako, K. **137**, 225
Ohno, J., see Nishikawa, M. **85**, 75
Ohno, Y., see Katayama, N. **115**, 87
Ohta, N., D. Chen, S. Ito, T. Futo, T. Yotsuyanagi, K. Ikeda, Effect of *trans*-diamminedichloroplatinum(II) on human serum albumin: Conformational changes through partial disulfide bond cleavage **118**, 85
Ohta, N., D. Chen, M. Ukai, T. Matsuo, M. Masuda, T. Yotsuyanagi, Aggregation of γ-globulin by *cis*-diamminedichloroplatinum(II): Alteration of Fc region and restoration by diethyldithiocarbamate **124**, 165
Ohta, N., T. Yotsuyanagi, D. Chen, R. Ono, S. Ito, K. Ikeda, Disulfide bond cleavage of human serum albumin and alterations of its secondary structure by *cis*-diamminedichloroplatinum(II) **85**, 39
Ohta, N., T. Yotsuyanagi, K. Ikeda, Degradation of gabexate mesilate catalyzed by human serum albumin **29**, 137
Ohta, N., see Chen, D. **106**, 249
Ohta, R., see Nakamura, F. **134**, 173
Ohtani, T., M. Murakami, A. Yamamoto, K. Takada, S. Muranishi, Effect of absorption enhancers on pulmonary absorption of fluorescein isothiocyanate dextrans with various molecular weights **77**, 141
Ohtsubo, Y., see Nishikawa, M. **85**, 75

Ohwaki, T., K. Nitta, H. Ozawa, Y. Kawashima, T. Hino, H. Takeuchi, T. Niwa, Improvement of the formation percentage of water-in-oil-in-water multiple emulsion by the addition of surfactants in the internal aqeous phase of the emulsion **85**, 19
Ohwaki, T., R. Machida, H. Ozawa, Y. Kawashima, T. Hino, H. Takeuchi, T. Niwa, Improvement of the stability of water-in-oil-in-water multiple emulsions by the addition of surfactants in the internal aqueous phase of the emulsions **93**, 61
Ohwaki, T., see Ando, H. **34**, 153
Ohwaki, T., see Aoki, S. **85**, 29
Ohwaki, T., see Ohwaki, T. **85**, 19; **93**, 61
Ohya, M., see Kawashima, Y. **99**, 229
Oie, S., D. Trenk, T.W. Guentert, H. Mosberg, E. Jahnchen, Disposition of vitamin K_1 after intravenous and oral administration to subjects on phenprocoumon therapy **48**, 223
Oie, S., Effect of 4-*N*-acetyl-sulfisoxazole on the disposition of sulfisoxazole in the rat **3**, 311
Oie, S., Pharmacokinetics of sulfisoxazole in rabbits with experimental renal failure after single and multiple dosing **23**, 345
Oie, S., see Jung, D. **7**, 179
Ojile, J.E., C.B. Macfarlane, A.B. Selkirk, Drug distribution during massing and its effect on dose uniformity in granules **10**, 99
Okabe, H., see Takayama, K. **61**, 225
Okabe, T., see Miyagawa, Y. **138**, 215
Okada, H., T. Heya, Y. Igari, Y. Ogawa, H. Toguchi, T. Shimamoto, One-month release injectable microspheres of leuprolide acetate inhibit steroidogenesis and genital organ growth in rats **54**, 231
Okada, H., see Heya, T. **69**, 69; **72**, 199
Okada, H., see Okada, H. **54**, 231
Okada, H., see Yanai, S. **123**, 237
Okada, M., see Oguchi, T. **61**, 27
Okada, R., see Seta, Y. **41**, 245, 263
Okamoto, H., H. Komatsu, M. Hashida, H. Sezaki, Effects of β-cyclodextrin and di-*O*-methyl-β-cyclodextrin on the percutaneous absorption of butylparaben, indomethacin and sulfanilic acid **30**, 35
Okamoto, R., see Yamaguchi, T. **85**, 87
Okamura, Y., see Nishihata, T. **34**, 157
Okano, T., see Gilbert, D.L. **47**, 79
Okano, T., see Yuk, S.H. **77**, 221, 231
Okhamafe, A.O., J.O. Akerele, Some aspects of the bioavailability of orally administered ofloxacin in healthy human volunteers **50**, 83
Okhamafe, A.O., J.O. Akerele, C.S. Chukuka, Pharmacokinetic interactions of norfloxacin with some metallic medicinal agents **68**, 11
Okhamafe, A.O., R. Bjankova, Interaction phenomena in pharmaceutical film coatings and testing methods **39**, 1
Okhamafe, A.O., P. York, Effects of solids–polymer interactions on the properties of some aqueous-based tablet film coating formulations. I. Moisture permeability **22**, 265
Effects of solids–polymer interactions on the properties of some aqueous-based tablet film coating formulations. II. Mechanical characteristics **22**, 273
Okor, R.S., Enhanced dissolution rate of aspirin from aspirin-methacrylic acid, methyl methacrylate coacervates in a simulated intestinal fluid **47**, 263
Retard release from a flocculated emulsion system **65**, 133
Okor, R.S., C.E. Obi, Drug release through aqueous-based film coatings of acrylate-methacrylate, a water-insoluble copolymer **58**, 89
Drug release through aqueous-based film coatings of acrylate-methacrylate, a water-insouble copolymer (Int. J. Pharm. 58 (1990) 89–91) **59**, 83
Oksanen, M., see Ketolainen, J. **125**, 45
Okuizumi, S., see Maruyama, K. **111**, 103
Okumura, K., S. Iwakawa, T. Yoshida, T. Seki, F. Komada, Intratracheal delivery of insulin. Absorption from solution and aerosol by rat lung **88**, 63
Okutgen, E., J.E. Hogan, M.E. Aulton, Quantitative estimation of internal stress development in aqueous HPMC tablet film coats **119**, 193
Olafsdottir, B.J., see Loftsson, T. **66**, 289; **67**, R5; **79**, 107
Olafsson, J.H., see Loftsson, T. **115**, 255
Olajos, S., see Cserhati, T. **61**, 189
Oldershaw, G.A., see Gould, P.L. **51**, 195
Olejnik, O., S.S. Davis, The stoichiometric interaction and influence of ionic strength on the organic-aqueous distribution behaviour of a tetrazole ion pair **30**, 101
Olejnik, O., see Musson, D.G. **99**, 85
Olesen, H.P., see Harboe, E. **53**, 157
Oliva, A., J.B. Farina, M. Llabres, Influence of temperature and shaking on stability of insulin preparations: Degradation kinetics **143**, 163
Oliva, M., see Coderch, L. **111**, 7; **139**, 197
Oliyai, R., S. Lindenbaum, Stability testing of pharmaceuticals by isothermal heat conduction calorimetry: Ampicillin in aqueous solution **73**, 33
Olliff, C.J., see Lloyd, A.W. **131**, 257
Olsen, C.E., see Jensen, J. **88**, 177
Olsen, G.B., see Seminoff, L.A. **54**, 241, 251
Olsson, B., Z.G. Wagner, P. Mansson, G. Ragnarsson, A γ scintigraphic study of the absorption of morphine from controlled-release tablets **119**, 223
Olsson, H., A. Adolfsson, C. Nystrom, Compaction and measurement of tablets in liquids with different dielectric constants for determination of bonding mechanisms—Evaluation of the concept **143**, 233
Olsson, L.I., see Camber, O. **29**, 259
Oluwatudimu, O.O., see Ogunbona, F.A. **29**, 191
Omar, S.M., see Kassem, M.A. **12**, 1
Omar, S.M., see Mattha, A.G. **11**, 27
Omer, A.I.H., see Salama, R.B. **17**, 325
Omichi, H., see Miyajima, M. **95**, 153
Omichi, H., see Yoshida, M. **115**, 61
Omotosho, J.A., The effect of acacia, gelatin and polyvinylpyrrolidone on chloroquine transport from multiple w/o/w emulsions **62**, 81

Omotosho, J.A., A.T. Florence, T.L. Whateley, Absorption and lymphatic uptake of 5-fluorouracil in the rat following oral administration of w/o/w multiple emulsions **61**, 51
O'Mullane, J.E., K. Petrak, L.E.F. Hutchinson, E. Tomlinson, The effect of adsorbed coats of Poloxamers 237 and 338 on the in vitro aggregation and in vivo distribution of polystyrene latex (PSL) particles **63**, 177
Onaga, I.C., A.J. Baillie, The toxicity of liposomal chloramphenicol for *Tetrahymena pyriformis* **7**, 89
The uptake of liposomal and free [^{14}]chloramphenicol by *Tetrahymena pyriformis* **12**, 323
O'Neal, J.S., S.G. Schulman, der Giesen Van, A.C.J. Roomer, Alkaline hydrolytic lability of some hydroxy- and methoxycoumarins and related anticoagulants **12**, 355
O'Neill, C.T., P.B. Deasy, Development and evaluation using hairless mouse skin of a transdermal timolol product **48**, 247
Ong, H., see Juhasz, J. **77**, 309
Ong, J.T.H., Z.T. Chowhan, G.J. Samuels, Drug–excipient interactions resulting from powder mixing. VI. Role of various surfactants **96**, 231
Onishi, H., T. Nagai, Characterization and evaluation of dialdehyde starch as an erodible medical polymer and a drug carrier **30**, 133
Onishi, H., see Song, Y. **98**, 121
Ono, R., see Ohta, N. **85**, 39
Onur, F., N. Acar, Simultaneous determination of methylene blue, hexamethylene tetramine and resorcinol in pharmaceutical formulations by first-derivative UV spectrophotometry **78**, 89
Onur, M.A., see Onur, M.A. **88**, 313
Onur, M.A., C.J. Kirby, A. Isimer, S. Beksac, N. Basci, R. Pamir, T. Coskun, A. Tumer, Effect of liposomal encapsulation of chloramphenicol on its transfer across the human placenta in a dual in vitro perfusion system **88**, 313
Onyeji, C.O., C.P. Babalola, The effect of magnesium trisilicate on proguanil absorption **100**, 249
Onyekweli, A.O., see Pilpel, N. **70**, 241
Opdebeeck, J.P., see Khan, M.Z.I. **76**, 161; **90**, 255
Oppenheim, R.C., Solid colloidal drug delivery systems: Nanoparticles **8**, 217
Oppenheim, R.C., see D'Onofrio, G.P. **2**, 91
Oppermann, J.A., see Cook, C.S. **60**, 125
Orbe, A., L.O. Sundelof, A new in vitro model for quantitative study of cream permeability **41**, 49
Orbe, A., see Nicklasson, M. **69**, 255
Orecchioni, A.M., see Ezpeleta, I. **131**, 191
Orecchioni, A.M., see Irache, J.M. **126**, 103
O'Reilly, J.R., O.I. Corrigan, C.M. O'Driscoll, The effect of simple micellar systems on the solubility and intestinal absorption of clofazimine (B663) in the anaesthetised rat **105**, 137
The effect of mixed micellar systems, bile salt/fatty acids, on the solubility and intestinal absorption of clofazimine (B663) in the anaesthetised rat **109**, 147
O'Reilly, S., C.G. Wilson, J.G. Hardy, The influence of food on the gastric emptying of multiparticulate dosage forms **34**, 213
Oren, I., see Sener, E. **110**, 109
Oribe, T., M. Yamada, K. Takeuchi, S. Tsunemi, K. Imasaka, O. Shirakura, S. Ishimaru, K. Takayama, T. Nagai, Formulation and in vivo–in vitro correlation of the dissolution property of lemildipine solid dispersions-incorporated suppositories **124**, 27
Oribe, T., see Oribe, T. **124**, 27
Orner, D., see Smith, P.L. **106**, 95
Orr, C.A., see Mackellar, A.J. **112**, 65, 79
Orr, N.A., see Jones, S.P. **53**, 43
Ortega-Alvaro, A., see Alvarez-Fuentes, J. **139**, 237
Ortigosa, C., see Sonanglio, D. **115**, 53
Orzalesi, G., S. Selleri, P. Gratteri, Chromatographic analysis of picotamide and its impurities **52**, 225
Oschmann, R., see Braun, M.A. **135**, 53
Oshiba, M., see Miyazaki, S. **6**, 77
Ossembe, N., see Van, Schepdael, A. **73**, 105
Ostberg, T., E.M. Lund, C. Graffner, Calcium alginate matrices for oral multiple unit administration. IV. Release characteristics in different media **112**, 241
Ostberg, T., L. Vesterhus, C. Graffner, Calcium alginate matrices for oral multiple unit administration. II. Effect of process and formulation factors on matrix properties **97**, 183
Ostenberg, T., C. Graffner, Calcium alginate matrices for oral multiple unit administration. III. Influence of calcium concentration, amount of drug added and alginate characteristics on drug release **111**, 271
Ostlius, J., see Ek, R. **125**, 257
Ostrenga, J.A., see Matheson, Jr. L.E. **4**, 309
Otagiri, M., Y. Otagiri, J.H. Perrin, Some fluorescent investigations of the interaction between the enantiomers of warfarin and human serum albumin **2**, 283
Otagari, M., see Suenaga, A. **59**, 121
Otagiri, M., see Bekers, O. **52**, 239; **72**, 123
Otagiri, M., see Imai, T. **53**, 7; **67**, 11
Otagiri, M., see Maruyama, T. **59**, 137
Otagiri, M., see Suenaga, A. **82**, 29
Otagiri, M., see Uekama, K. **16**, 327; **23**, 35
Otagiri, Y., see Otagiri, M. **2**, 283
Otero-Espinar F., S. Anguiano-Igea, J. Blanco-Medez, J.L. Vila-Jato, Reduction in the ulcerogenicity of naproxen by complexation with β-cyclodextrin **70**, 35
Otero-Espinar, F.J., S. Anguiano-Igea, N. Garcia-Gonzalez, J.L. Vila-Jato, J. Blanco-Mendez, Oral bioavailability of naproxen-β-cyclodextrin inclusion compound **75**, 37
Interaction of naproxen with β-cyclodextrin in solution and in the solid state **79**, 149
Otero-Espinar, F.J., see Anguiano-Igea, S. **135**, 161
Otero-Espinar, F.J., see Blanco-Fuente, H. **138**, 103; **142**, 169
Otero-Espinar, F.J., see Ganza-Gonzalez, A. **106**, 179
Oth, M.P., A.J. Moes, Enhanced in vitro release of indomethacin from non-aqueous suspensions using indomethacin-polyvinylpyrrolidone coprecipitate **24**, 275
Sustained release solid dispersions of indomethacin with Eudragit RS and RL **55**, 157

Othman, S., Multicomponent derivative spectroscopic analysis of sulfamethoxazole and trimethoprim **63**, 173
Othman, S., H. Muti, O. Shaheen, A. Awidi, W.A. Al-Turk, Studies on the adsorption and solubility of nalidixic acid **41**, 197
Othman, S., see Al-Turk, W.A. **41**, 227
Othman, S., see Shaheen, O. **38**, 123
Otsuka, A., see Takeeda, T. **41**, 21
Otsuka, M., N. Kaneniwa, Effect of grinding on the crystallinity and chemical stability in the solid state of cephalothin sodium **62**, 65
Otsuka, T., see Seta, Y. **41**, 255, 263
Otten, M., see Boussard, P. **32**, 91
Ouriemehi, E.M., J. Bouzon, J.M. Vergnaud, Oral dosage forms with a core and shell with the same polymer containing different drug concentrations **102**, 47
Modelling the process of controlled release of drug in in vitro and in vivo tests **113**, 231
Ouriemchi, E.M., J.M. Vergnaud, Calculation of the plasma drug level with oral controlled release dosage forms. Effect of the dose frequency **127**, 177
Ouriemchi, E.M., see Bidah, D. **80**, 145
Ouriemchi, E.M., see Nia, B. **119**, 165
Oussoren, C., see Zuidema, J. **105**, 189
Overcashier, D.E., see Lam, X.M. **142**, 85
Owada, E., see Kurosawa, N. **27**, 81
Owais, L., see Badwan, A.A. **28**, 41
Owoyale, J.A., Decomposition profile of ultraviolet-irradiated chloroquine **52**, 179
Amodiaquine less sensitive than chloroquine to photochemical reactions **56**, 213
Owoyale, J.A., Z. Elmarakby, On the ultraviolet 'spectral shift' phenomenon of chloroquine **12**, 265
Effect of sunlight, ultraviolet irradiation and heat on proguanil **50**, 219
Owusu-Ababio, G., J.A. Rogers, H. Anwar, Method of evaluation of sustained release microsphere formulations using the open chemostat system **122**, 9
Ozawa, H., see Aoki, S. **85**, 29, 65; **95**, 67
Ozawa, H., see Ohwaki, T. **85**, 19; **93**, 61
Ozawa, H., see Takeuchi, H. **14**, 279; **86**, 25
Ozer, A.Y., H. Talsma, Preparation and stability of liposomes containing 5-fluorouracil **55**, 185
Ozyazici, M., F. Sevgi, G. Ertan, Micromeritic studies on nicardipine hydrchloride microcapsules **138**, 25

P

Paalman, A.C.A., see Beijnen, J.H. **41**, 169
Pablos, F., see Gonzalez, A.G. **72**, 193
Pabmanabhan, R., see Wong, O. **76**, 171
Paborji, M., C.M. Riley, V.J. Stella, A novel use of Intralipid for the parenteral delivery of perilla ketone (NSC-348407), an investigational cytotoxic drug with a high affinity for plastic **42**, 243
Paddock, S.W., see Rojanasakul, Y. **61**, 163
Padfield, J.M., see Liversidge, G.G. **7**, 211
Padmadisastra, Y., R.A. Kennedy, P.J. Stewart, Influence of carrier moisture adsorption capacity on the degree of adhesion of interactive mixtures **104**, R1
Solid bridge formation in sulphonamide–Emdex interactive systems **112**, 55
Padmalatha Devi, K., see Baveja, S.K. **24**, 355; **27**, 157
Padmalatha Devi, K., see Ranga Rao, K.V. **48**, 1
Paeratakul, O., see Bodmeier, R. **59**, 197; **70**, 59; **96**, 129; **103**, 47
Paesen, J., K. Khan, E. Roets, J. Hoogmartens, Study of the stability of erythromycin in neutral and alkaline solutions by liquid chromatography on poly(styrene-divinylbenzene) **113**, 215
Pagano, A., see Saettone, M.F. **86**, 159
Page, T., see Nicklin, P. **128**, 155
Pai, S.H., see Law, S.L. **32**, 237; **43**, 257
Paik, W.H., see Lee, H.J. **121**, 1; **131**, 91
Pajean, M., D. Herbage, Effect of collagen on liposome permeability **91**, 209
Pajean, M., A. Huc, D. Herbage, Stabilization of liposomes with collagen **77**, 31
Palagiano, F., L. Arenare, F. Barbato, Rotonda M. La, F. Quaglia, F.P. Bonina, L. Montenegro, P. De Caprariis, In vitro and in vivo evaluation of terpenoid esters of indomethacin as dermal prodrugs **149**, 171
Palazzo, A.S., see Giammona, G. **105**, 57
Palazzo, S., see Cavallaro, G. **90**, 195
Palepu, N.R., see Bogardus, J.B. **4**, 159
Palepu, N.R., see Kearney, A.S. **127**, 229
Palermiti, L.G., see Memoli, A. **117**, 159
Palicharla, P., see Bonina, F.P. **98**, 93
Palin, K.J., A.J. Phillips, A. Ning, The oral absorption of cefoxitin from oil and emulsion vehicles in rats **33**, 99
Palin, K.J., D.R. Whalley, C.G. Wilson et al., Determination of gastric-emptying profiles in the rat: Influence of oil structure and volume **12**, 315
Palin, K.J., see Reid, A.S. **40**, 181
Palin, K.J., see Thomas, N.W. **20**, 163; **44**, 261
Palin, K.J., see Whiston, R.J. **23**, 247
Palin, K.J., see Young, C. **40**, 187
Palm, B., see Wennergren, B. **53**, 35
Palm, K., see Lennernas, H. **127**, 103
Palmer, A.J., see Sewell, G.J. **70**, 119; **72**, 57
Palsdottir, G., see Loftsson, T. **57**, 63
Pande, G.S., R.F. Shangraw, Characterization of β-cyclodextrin for direct compression tableting **101**, 71
Characterization of β-cyclodextrin for direct compression tableting. II. The role of moisture in the compactibility of β-cyclodextrin **124**, 231
Panderi, I., M. Parissi-Poulou, Determination of captopril and captopril-hydrochlorothiazide combination in tablets by derivative UV spectrophotometry **86**, 99
Determination of clopamide-pindolol combination in tablets by fourth-order derivative UV spectrophotometry **99**, 327
Pandit, J.K., S.K. Gupta, K.D. Gode, B. Mishra, Effect of crystal form on the oral absorption of phenylbutazone **21**, 129

Pandit, N.K., J. Kanjia, Phase behavior of nonionic surfactant solutions in the presence of polyvinylpyrrolidone **141**, 197
Pandit, N.K., J. Kanjia, K. Patel, D.G. Pontikes, Phase behavior of aqueous solutions containing nonionic surfactant-polyethylene glycol mixtures **122**, 27
Pandit, N.K., J. Kisaka, Loss of gelation ability of Pluronic^{R} F127 in the presence of some salts **145**, 129
Pandit, N.K., J.M. Strykowski, L. Shtohryn, The effect of salt on the distribution and solubility of an acidic drug **50**, 7
Panduranga Rao, K., see Jeyanthi, R. **35**, 177; **55**, 31
Panduranga Rao, K., see Madhan Kumar, A.B. **149**, 107
Panduranga Rao, K., see Narayani, R. **95**, 85; **138**, 121; **142**, 25; **143**, 255
Panduranga Rao, K., see Thacharodi, D. **96**, 33; **134**, 239
Panetta, J.A., see Pop, E. **140**, 33
Panichi, L., see Burgalassi, S. **133**, 1
Panico, A.M., R. Pignatello, S. Mazzone, V. Cardile, M. Bindoni, F. Cordopatri, A. Villari, N. Micali, G. Mazzone, Lipid vesicles loaded with thymopentin: Characterization and in vitro activity on tumoral cells **98**, 19
Panico, A.M., R. Pignatello, G. Puglisi, G. Mazzone, G. Petrone, Use of liposomes as carriers for immunomodulatory polypeptides: Studies on thymostimulin encapsulation and retention **80**, 93
Panico, A.M., see Montenegro, L. **133**, 89; **138**, 191
Panini, R., see Vandelli, M.A. **118**, 77
Pannatier, A., B. Testa, J.C. Etter, Enzymatic hydrolysis by mouse skin homogenates: Structure–metabolism relationships of *para*-nitrobenzoate esters **8**, 167
Panomsuk, S.P., T. Hatanaka, T. Aiba, K. Katayama, T. Koizumi, A study of the hydrophilic cellulose matrix: Effect of indomethacin and a water-soluble additive on swelling properties **126**, 147
Panomsuk, S.P., see Koizumi, T. **116**, 45
Pantaleoni, G., see Di Giulio, A. **144**, 27
Pantzar, N., see Lundin, S. **59**, 263
Panza, L., see Ventura, C.A. **149**, 1
Panzova, B., B. Bogdanov, Physicochemical basis for the microbicidal action of disinfection solutions. I. Polyvinylpyrrolidinone-iodine **65**, 35
Panzova, B., M. Ilievska, G. Trendovska, B. Bogdanov, Simultaneous determination of polythiazide and prazosin in tablets by second-order derivative UV spectroscopy **70**, 187
Papadimitriou, E., G. Buckton, M. Efentakis, Probing the mechanism of swelling of hydroxypropylmethylcellulose matrices **98**, 57
Papadimitriou, E., M. Efentakis, N.H. Choulis, Evaluation of maltodextrins as excipients for direct compression tablets and their influence on the rate of dissolution **86**, 131
Papaioannou, G., see Archondikis, A. **55**, 217
Papaioannou, G., see Velissaratou, A.S. **52**, 83
Pape, D., see Cognez, P. **123**, 137
Papisov, M.I., V.Y. Savelyev, V.B. Sergienko, V.P. Torchilin, Magnetic drug targeting. I. In vivo kinetics of radiolabelled magnetic drug carriers **40**, 201
Papisov, M.I., V.P. Torchilin, Magnetic drug targeting. II. Targeted drug transport by magnetic microparticles: Factors influencing therapeutic effect **40**, 207
Papp, K.J., see Cannon, J.B. **114**, 65
Paragamian, V., see Walkling, W.D. **4**, 39
Parera, A., see Gallardo, V. **71**, 223
Parera Vialard, A., see Contreras Claramonte, M.D. **98**, 37
Parfitt, R.T., see Kraisintu, K. **10**, 67
Parfitt, R.T., see Lacy, J.E. **43**, 111
Parimoo, P., C.V.N. Prasad, A. Gautam, Simultaneous quantitative determination of metronidazole and diloxanide furoate in a tablet preparation by difference spectroscopy **134**, 231
Parimoo, P., P. Umapathi, K. Ilango, Simultaneous quantitative determination of salbutamol sulfate and bromhexine hydrochloride in drug preparations by difference spectrophotometry **100**, 227
Paris, J., see Vaution, C. **1**, 349
Paris, J., see Walchshofer, N. **77**, 265; **100**, 271
Parissi-Poulou, M., V. Reizopoulou, M. Koupparis, P. Macheras, Second derivative UV spectrophotometric determination of hydrochlorothiazide and hydrochlorothiazide-amiloride combination in tablets **51**, 169
Parissi-Poulou, M., see Panderi, I. **86**, 99; **99**, 327
Park, E.S., see Chi, S.C. **126**, 267
Park, J.Y., M. Langworthy, C.R. Behl et al., Vaginal drug absorption in the rhesus monkey: Bioavailability method and assessment of estrogens **2**, 215
Park, J.Y., see Komiya, I. **4**, 249
Park, K., J.R. Robinson, Bioadhesive polymers as platforms for oral-controlled drug delivery: Method to study bioadhesion **19**, 107
Park, K.M., see Kim, C.K. **128**, 65; **146**, 63; **147**, 131
Park, M.K., see Chang, H.W. **79**, 71
Park, M.K., see Yoon, E.J. **67**, 177
Park, Y.H., see Han, S.K. **126**, 35
Parkash, A.K., J.K. Sugden, Z. Yucan, W. Mingxin, A rapid method for screening non steroidal anti-inflammatory agents for hydroxyl radical scavenging **92**, 151
Parker, M.D., P. York, R.C. Rowe, Binder-substrate interactions in wet granulation. 1. The effect of binder characteristics **64**, 207
Binder–substrate interactions in wet granulation. 2. The effect of binder molecular weight **72**, 243
Binder–substrate interactions in wet granulation. 3. The effect of excipient source variation **80**, 179
Parker, M.D., see Hancock, B.C. **76**, 239
Parker, M.D., see Rowe, R.C. **91**, 247
Parker, M.S., see Fassihi, A.R. **31**, 271
Parkins, D.A., see Wakerly, Z. **129**, 73
Parkinson, H., J.B. Stanford, J.K. Sugden, The influence of sonic energy on the hydrolysis of procaine hydrochloride in aqueous buffer at pH 9.15 **51**, 189
Parks, E.H., see Stewart, J.T. **17**, 161
Parnes, J.H., see Prankerd, R.J. **78**, 59
Paronen, P., see Hirvonen, J. **99**, 253
Paronen, P., see Ilkka, J. **94**, 181

Paronen, P., see Ketolainen, J. **92**, 157
Paronen, P., see Marjukka Turunen, T. **92**, 89
Paronen, P., see Munoz-Ruiz, A. **148**, 33
Paronen, P., see Pesonen, T. **57**, 139
Paronen, P., see Pirttimaki, J. **95**, 93
Paronen, P., see Silvennoinen, R. **131**, 209
Paronen, P., see Sutinen, R. **57**, 149, 155; **62**, 113; **92**, 177
Paronen, P., see Tirkkonen, S. **88**, 39; **92**, 55; **124**, 219
Paronen, P., see Vidgren, M. **42**, 211; **64**, 1; **73**, 131; **96**, 183
Paronen, T.P., see Mosher, G.L. **39**, 113
Paronen, T.P., see Vidgren, M.T. **35**, 139; **37**, 239; **39**, 101
Parr, A., R.M. Beihn, M. Jay, Radiolabeling of an enteric coated tablet by (n, γ) radioactivation of erbium-170 for scintigraphic imaging **32**, 251
Parr, G.D., see Davis, S.S. **49**, 183
Parr, G.D., see Farraj, N.F. **38**, 39; **43**, 83, 93; **52**, 19
Parr, G.D., see Hessell, P.G. **52**, 159
Parr, G.D., see Jones, S.P. **33**, 105; **36**, 223
Parr, G.D., see Wilson, C.G. **18**, 1
Parra, J.L., see Coderch, L. **111**, 7
Parrott, K.A., see Lee, B.J. **124**, 119
Parry-Jones, D.R., see Farr, S.J. **26**, 303
Parsons, D.L., see Betageri, G.V. **81**, 235
Parsons, G.E., G. Buckton, S.M. Chatham, The extent of the errors associated with contact angles obtained using liquid penetration experiments **82**, 145
The use of surface energy and polarity determinations to predict physical stability of non-polar, non-aqueous suspensions **83**, 163
Partridge, W.M., see Terasaki, T. **81**, 143
Pasanen, K., see Alderborn, G. **23**, 79
Pascual-Costa, R.M., see Martin-Algarra, V. **122**, 1
Passmore, P., see McElnay, J.C. **33**, 195
Pastesini, C., see Esposito, E. **117**, 151
Pastesini, C., see Nastruzzi, C. **90**, 43
Patat, F., see Matchet, L. **133**, 39
Patat, F., see Pelucio-Lopes, C. **96**, 249
Pate, D.W., see Jarho, P. **137**, 209
Patel, A.N., see Armstrong, N.A. **48**, 173
Patel, C.G., see Grant, D.J.W. **5**, 109
Patel, C.I., J. Zuidema, M. Lindley, Fatigue failure behaviour of direct compression excipients **39**, 93
Patel, C.I., see Shah, N.H. **106**, 15
Patel, H.K., R.C. Rowe, J. McMahon, R.F. Stewart, A systematic microscopical examination of gels and emulsions containing cetrimide and cetostearyl alcohol **25**, 13
Properties of cetrimide/cetostearyl alcohol ternary gels; preparation effects **25**, 237
Patel, H.K., see Attwood, D. **49**, 129
Patel, H.K., see Louden, J.D. **25**, 179
Patel, J.M., D.E. Wurster, Effect of hydrocarbon chain length on the hydrolysis of several naphthyl esters in the presence of *o*-iodosobenzoic acid and CTAB micelles **86**, 43
Patel, J.M., see Wurster, D.E. **78**, 157
Patel, J.P., K. Marsh, L. Carr, G. Nequist, Factorial designs in ophthalmic formulation development of Enalkiren **65**, 195
Patel, J.P., A.J. Repta, Enol esters as potential prodrugs. I. Stability and enzyme-mediated hydrolysis of α-acetoxystyrene **5**, 329
Enol esters as potential pro-drugs. II. In vitro aqueous stability and enzyme-mediated hydrolysis of several enol esters of acetophenone **9**, 29
Patel, J.P., see Repta, A.J. **10**, 29, 239
Patel, K., see Kearney, A.S. **127**, 229
Patel, K., see Pandit, N.K. **122**, 27
Patel, K.K., see Buckton, G. **56**, 151
Patel, M.J., see McCallion, O.N.M. **130**, 245
Patel, M.V., see Sung, K.C. **142**, 53
Patel, N., see Aboofazeli, R. **125**, 107
Patel, N.K., A.H. Upadhyay, J.S. Bergum, G.E. Reier, An evaluation of microcrystalline cellulose and lactose excipients using an instrumented single station tablet press **110**, 203
Patel, N.R., see Sinko, P.J. **109**, 125
Patel, R., F. Podczeck, Investigation of the effect of type and source of microcrystalline cellulose on capsule filling **128**, 123
Patel, V.S., W.G. Kramer, Allopurinol rectal absorption in the rabbit **30**, 221
Patel, V.S., see Wagner, D.L. **114**, 171
Pathak, K., see Singla, A.K. **60**, 27
Pathak, Y.V., R.L. Nikore, A.K. Dorle, Study of Rosin and Rosin esters as coating materials **24**, 351
Pathak, Y.V., A. Rubinstein, S. Benita, Enhanced stability of physostigmine salicylate in submicron o/w emulsion **65**, 169
Patil, S.M., P. Singh, H.J. Maibach, Cumulative irritancy in man to sodium lauryl sulfate: The overlap phenomenon **110**, 147
Pato, A.J., M. Mora, B. Naisbett, J.F. Woodley, R. Duncan, Uptake and transport of poly(*N*-vinylpyrrolidone-*co*-maleic acid) by the adult rat small intestine cultured in vitro: Effect of chemical structure **104**, 227
Patriarche, G.J., see Vincke, B.J. **21**, 265; **32**, 61
Pattarino, F., E. Marengo, M.R. Gasco, R. Carpignano, Experimental design and partial least squares in the study of complex mixtures: Microemulsions as drug carriers **91**, 157
Pattarino, F., see Gasco, M.R. **62**, 119; **69**, 193
Patterson, L.H., see Lilley, J.C. **107**, 149
Patton, T.F., A.J. Repta, L.A. Sternson, R.J. Belt, Pharmacokinetics of intact cisplatin in plasma. Infusion versus bolus dosing **10**, 77
Patton, T.F., see Ahmed, I. **19**, 215; **34**, 163
Patton, T.F., see Francoeur, M. **2**, 337; **16**, 203
Patton, T.F., see Francoeur, M.L. **25**, 275
Patton, T.F., see Magron, P. **38**, 91
Patton, T.F., see Reynolds, D.L. **9**, 67
Paul, C., see Vincentelli, J. **134**, 147
Paule, B., see Bonhomme-Faivre, L. **134**, 99
Paulet, J., see Laghoueg, N. **50**, 133
Paulo, M., see Lewis, D. **148**, 177
Pavanetto, F., B. Conti, I. Genta, P. Giunchedi, Solvent evaporation, solvent extraction and spray drying for polyactide microsphere preparation **84**, 151

Pavanetto, F., B. Conti, I. Genta, T. Modena, Particulate contamination from siliconized rubber stoppers **74**, 175
Pavanetto, F., B. Conti, I. Genta, R. Ponci, L. Montanari, S. Vianello, Particulate matter contamination of small volume parenterals **51**, 55
Pavanetto, F., B. Conti, T. Modena, I. Genta, R. Ponci, Particulate contamination in parenteral type medical devices **48**, 255
Pavanetto, F., I. Genta, B. Conti, T. Modena, L. Montanari, Aluminium, cadmium and lead in large volume parenterals: Contamination levels and sources **54**, 143
Pavia, D., see Newman, S.P. **19**, 333
Pavliv, L., K. Freebern, T. Wilke, C.C. Chiang, B. Shetty, P. Tyle, Topical formulation development of a novel thymidylate synthase inhibitor for the treatment of psoriasis **105**, 227
Pavlov, M.Y., see Sviridov, D.D. **96**, 1
Pawlik, B., see Knapczyk, J. **80**, 33
Payan Villar, T., see Munoz-Ruiz, A. **95**, 201; **110**, 37
Payling, D.W., see Lee, G. **23**, 43
Payne, R.S., R.J. Roberts, R.C. Rowe, M. McPartlin, A. Bashal, The mechanical properties of two forms of primidone predicted from their crystal structures **145**, 165
Payton, R., B. Rabinow, N. Raghavan, Application of radiotracer method for precise determination of water vapor transmission characteristics of unit dose packaging systems **33**, 257
Pazzi, P., see Scalia, S. **115**, 249
Peach, J., see Wilson, C.G. **40**, 119
Peach, J.M., see Washington, N. **57**, 17
Pearce, K.J., see Jones, D.S. **118**, 199; **131**, 25
Pearlman, R., B.S. Rutherford, K.M. Pozsgai, C.A. Hirsch, The isolation and identification of some degradation products of flurandrenolide in Cordran cream **18**, 53
Pech, B., O. Duval, P. Richomme, J.P. Benoit, A timolol prodrug for improved ocular delivery: Synthesis, conformational study and hydrolysis of palmitoyl timolol malonate **128**, 179
Peck, C.C., W.H. Barr, L.Z. Benet, J. Collins, R.E. Desjardins, D.E. Furst, J.G. Harter, G. Levy, T. Ludden, J.H. Rodman, L. Sanathanan, J.J. Schentag, V.P. Shah, L.B. Sheiner, J.P. Skelly, D.R. Stanski, R.J. Temple, C.T. Viswanathan, J. Weissinger et al., Opportunities for integration of pharmacokinetics, pharmacodynamics and toxicokinetics in rational drug development **82**, 9
Peck, G.E., see Morefield, E.M. **34**, 263
Peck, J., see Hadgraft, J. **141**, 17
Peck, K.D., A.H. Ghanem, W.I. Higuchi, V. Srinivasan, Improved stability of the human epidermal membrane during successive permeability expriments **98**, 141
Pedersen, M., M. Edelstein, V.F. Nielsen, A. Scarpellini, S. Skytte, C. Slot, Formation and antimycotic effect of cyclodextrin inclusion complexes of econazole and miconazole **90**, 247
Pedersen, M., see Jacobsen, J. **125**, 165; **141**, 217
Pedersen, S.B., see Bundgaard, H. **27**, 71; **64**, 75
Pedraz, J.L., E.L. Marino, F.J. Burguillo, A. Dominguez-Gil, In vitro binding of ketamine to human serum albumin **25**, 147
Pedraz, J.L., see Calvo, M.B. **67**, 259
Pedraz, J.L., see Hernandez, R.M. **139**, 45
Peeters, R., R. Kinget, Film-forming polymers for colonic drug delivery. I. Synthesis and physical and chemical properties of methyl derivatives of Eudragit S **94**, 125
Peh, K.K., K.H. Yuen, Indirect gastrointestinal transit monitoring and absorption of theophylline **139**, 95
Pellett, M.A., A.F. Davis, J. Hadgraft, Effect of supersaturation on membrane transport. 2. Piroxicam **111**, 1
Pellion, B., see Zuber, M. **47**, 31
Pellow, P.G.D., see Newman, S.P. **36**, 55
Pelplinska, T., see Regosz, A. **88**, 437
Pelucio-Lopes, C., L. Machet, L. Vaillant, F. Patat, M. Lethiecq, Y. Furet, L. Pourcelot, G. Lorette, Phonophoresis of azidothymidine (AZT) **96**, 249
Pena, L.E., P.L. Possert, J.F. Stearns, B.L. Lee, M.J. Hageman, Rheological characterization of rbSt oil suspensions **113**, 89
Pena, Romero A., C. Caramella, M. Ronchi, F. Ferrari, D. Chulia, Water uptake and force development in an optimized prolonged release formulation **73**, 239
Penaud, J.F., M.O. Decroix, P. Arnaud, D. Magne, J.G. Gobert, J.C. Chaumeil, In vitro study of cyclosporin absorption: Vehicles and intestinal immaturity **142**, 1
Pennington, A.K., J.H. Ratcliffe, C.G. Wilson, J.G. Hardy, The influence of solution viscosity on nasal spray deposition and clearance **43**, 221
Pens, L., see Coderch, L. **111**, 7
Pense, A.M., C. Vauthier, F. Puisieux, J.P. Benoit, Microencapsulation of benzalkonium chloride **81**, 111
Pentari, J.G., C.E. Efstathiou, M.A. Koupparis, Kinetic determination of carbocysteine in syrup based on its reaction with 1-fluoro-2,4-dinitrobenzene monitored with a fluoride-selective electrode **77**, 41
Peppas, N.A., P. Colombo, U. Conte, A. Gazzaniga, C. Caramella, F. Ferrari, Energetics of tablet disintegration **51**, 77
Peppas, N.A., P.J. Hansen, P.A. Buri, A theory of molecular diffusion in the intestinal mucus **20**, 107
Peppas, N.A., J.J. Sahlin, A simple equation for the description of solute release. III. Coupling of diffusion and relaxation **57**, 169
Peppas, N.A., see am Ende, M.T. **120**, 33
Peppas, N.A., see Bell, C.L. **134**, 167
Peppas, N.A., see Brannon-Peppas, L. **70**, 53
Peppas, N.A., see Caramella, C. **44**, 177
Peppas, N.A., see Colombo, P. **88**, 99
Peppas, N.A., see Hariharan, D. **112**, 47
Peppas, N.A., see Maitani, Y. **126**, 89
Peppas, N.A., see Mikos, A.G. **53**, 1
Peppas, N.A., see Ponchel, G. **38**, 65
Perard, D., see Luessen, H.L. **141**, 39
Perboni, I., see Raggi, M.A. **61**, 199
Pereira de Almeida, L., see Simoes, S. **139**, 169

Pereswetoff-Morath, L., S. Bjurstrom, R. Khan, M. Dahlin, P. Edman, Toxicological aspects of the use of dextran microspheres and thermogelling ethyl(hydroxyethyl) cellulose (EHEC) as nasal drug delivery systems **128**, 9
Pereswetoff-Morath, L., P. Edman, Dextran microspheres as a potential nasal drug delivery system for insulin—in vitro and in vivo properties **124**, 37
Influence of osmolarity on nasal absorption of insulin from the thermogelling polymer ethyl(hydroxyethyl) cellulose **125**, 205
Immunological consequences of nasal drug delivery in dextran microspheres and ethyl(hydroxyethyl)cellulose in rats **128**, 23
Perez-Buendia, M., see Pla-Delfina, J.M. **37**, 49
Perez-Buendia, M.D., T.M. Garrigues, J.E. Peris-Ribera, V.G. Casabo, J.M. Pla-Delfina, Correlation between in situ absorption and in vitro dialysis data found in the presence of surfactants **55**, 165
Perez-Buendia, M.D., see Garrigues, T.M. **57**, 189
Perez-Marcos, B., J.L. Ford, D.J. Amstrong, P.N.C. Elliott, C. Rostron, J.E. Hogan, Release of propranolol hydrochloride from matrix tablets containing hydroxypropylmethylcellulose K4M and carbopol 974 **111**, 251
Perez-Marcos, B., C. Gutierrez, J.L. Gomez-Amoza, R. Martinez-Pacheco, C. Souto, A. Concheiro, Usefulness of certain varieties of Carbomer in the formulation of hydrophilic furosemide matrices **67**, 113
Perez-Marcos, B., R. Martinez-Pacheco, J.L. Gomez-Amoza, C. Souto, A. Concheiro, R.C. Rowe, Interlot variability of carbomer 934 **100**, 207
Perez-Marcos, B., see Landin, M. **107**, 247
Perez-Marcos, M.B., see Esclusa-Diaz, M.T. **142**, 183; **143**, 203
Perez-Martinez, J.I., see Arias, M.J. **123**, 25
Perez-Sayas, E., see Diez-Sales, O. **100**, 1
Perez-Soler, R., see Cruz, M.E.M. **96**, 67
Perez-Soler, R., see Vadiei, K. **57**, 125, 133
Perez-Varona, A.T., see Bermejo, M.V. **69**, 221
Perez-Varona, A.T., see Garrigues Pelufo, T.M **79**, 135
Perez-Varona, A.T., see Garrigues, T.M. **64**, 127
Perini, G., see Saettone, M.F. **126**, 83
Peris-Ribera, J.E., see Collado, E.F. **44**, 187
Peris-Ribera, J.E., see Perez-Buendia, M.D. **55**, 165
Periviita, L., see Salminen, L. **18**, 17
Perkins, A.C., see Hardy, J.G. **142**, 129
Perkins, A.C., see Harnor, K.J. **95**, 111
Perlia, X., see Werner, I.A. **63**, 155
Permentier, D., see Vermeersch, H. **60**, 253
Perrier, D.G., see Gallo, J.M. **22**, 63
Perrier, D.G., see Gupta, P.K. **33**, 137, 147; **43**, 167
Perrier, D.G., see Hung, C.T. **44**, 117
Perrin, J.H., see Fleitman, J. **11**, 215, 227
Perrin, J.H., see Imamura, Y. **5**, 25
Perrin, J.H., see Otagiri, M. **2**, 283
Perrin, J.H., see Prankerd, R.J. **88**, 189
Perry, D.F., see Blanchard, J. **18**, 259
Pershing, L.K., L.D. Lambert, V.P. Shah, S.Y. Lam, Variability and correlation of chromameter and tape-stripping methods with the visual skin blanching assay in the quantitative assessment of topical 0.05% betamethasone dipropionate bioavailability in humans **86**, 201
Pershing, L.K., see Law, M.Y.L. **88**, 251
Persicaner, P.H.R., see Fassihi, A.R. **37**, 167
Persson, C., see Wennergren, B. **53**, 35
Pes, M.A., see Garcia-Celma, M.J. **105**, 77
Peschier, L.J.C., see Bouwstra, J.A. **52**, 47
Peschier, L.J.C., see Verhoeven, J. **45**, 65
Pesonen, T., P. Paronen, J. Ketolainen, Disintegrant properties of an agglomerated cellulose powder **57**, 139
Peter, J.D., see Bourget, P. **128**, 223
Peters, K.R., see Lamson, M.J. **105**, 259
Peters, T., see Vollmer, U. **99**, 51
Petersson, T., S. Engstrom, Surface and interfacial properties of clomethiazole **132**, 231
Petiprez, S., see Fournier, C. **106**, 41
Petit, I., see Martel, P. **149**, 37
Petrak, K., see O'Mullane, J.E. **63**, 177
Petrone, G., see Panico, A.M. **80**, 93
Petrov, L., see Kalinkova, G.N. **115**, 245
Petrovick, P.D., M. Jacob, D. Gaudy, V.L. Bassini, S.S. Guterres, Influence of adjuvants on the in vitro dissolution of hydrochlorothiazide from hard gelatin capsules **76**, 49
Petszulat, M., see Karau, C. **128**, 89
Pettersson, A., see Gottfries, J. **106**, 141
Peura, P., see Jarvinen, T. **75**, 259; **79**, 243
Peura, P., see Suhonen, P. **74**, 221
Peveri, T., see Redenti, E. **129**, 289
Peyrot, M., A.M. Sautereau, J.M. Rabanel, F. Nguyen, J.F. Tocanne, D. Samain, Supramolecular biovectors (SMBV): A new family of nanoparticulate drug delivery systems. Synthesis and structural characterization **102**, 25
Pfaff, G., P. Briegel, I. Lamprecht, Inter-individual variation in the metabolism of dextromethorphan **14**, 173
Pfeiffer, R.R., see Stephenson, G.A. **146**, 93
Pfluger, B., see Waser, P.G. **39**, 213
Phang, S., see Aungst, B.J. **117**, 95
Phillips, A.J., see Palin, K.J. **33**, 99
Phillips, C.A., B.B. Michniak, Topical application of Azone analogs to hairless mouse skin: A histopathological study **125**, 63
Phillips, C.A., see Michniak, B.B. **116**, 201
Phillips, E.M.., An approach to estimate the amorphous content of pharmaceutical powders using calorimetry with no calibration standards **149**, 267
Phillips, E.M., P.R. Byron, Surfactant promoted crystal growth of micronized methylprednisolone in trichloromonofluoromethane **110**, 9
Phillips, E.M., V.J. Stella, Rapid expansion from supercritical solutions: Application to pharmaceutical processes **94**, 1
Phillips, G.O., see Morton, A.K. **9**, 321
Phillips, N.C., see Heiati, H. **146**, 123
Pickup, M.E., M.S. Rogers, A.P. Launchbury, Temazepam elixir: Comparative bioavailability with a capsule formulation **22**, 311

Piepma, K.A., H. Vromans, C.F. Lerk, A coherent matrix model for the consolidation and compaction of an excipient with magnesium stearate **97**, 195
Pieters, F.A.J.M., J. Zuidema, Some comments on a method to quantify modified drug release **39**, 267
Pieters, F.A.J.M., see Zuidema, J. **47**, 1
Pignatello, R., see Castelli, F. **52**, 115
Pignatello, R., see Giammona, G. **57**, 55
Pignatello, R., see Panico, A.M. **80**, 93; **98**, 19
Pijpers, A., see Kadir, F. **64**, 171
Pikal, M.J., K.M. Dellerman, Stability testing of pharmaceuticals by high-sensitivity isothermal calorimetry at 25°C: Cephalosporins in the solid and aqueous solution states **50**, 233
Pikal, M.J., J.E. Lang, S. Shah, Desolvation kinetics of cefamandole sodium methanolate: The effect of water vapor **17**, 237
Pikal, M.J., S. Shah, The collapse temperature in freeze drying: Dependence on measurement methodology and rate of water removal from the glassy phase **62**, 165
Pikal, M.J., S. Shah, M.L. Roy, R. Putman, The secondary drying stage of freeze drying: Drying kinetics as a function of temperature and chamber pressure **60**, 203
Pikkarainen, P., see Vidgren, M. **96**, 183
Pillai, G.K., see Chandraprakash, K.S. **61**, R1
Pillai, G.K., see Sheikh, Salem M. **141**, 257
Pilpel, N., J.R. Britten, A.O. Onyekweli, S. Esezobo, Compression and tableting of pharmaceutical powders at elevated temperatures **70**, 241
Pilpel, N., C.I. Igwilo, S. Malamataris, Effects of molecular coatings on the compression and tableting of some pharmaceutical powders **68**, 157
Pilpel, N., M. Nejmeh, W.H.K. Sanniez, In vitro studies on photosensitization by phenothiazines, sulphonamides and tetracyclines **5**, 273
Pilpel, N., see Adeyemi, M.I. **15**, 293
Pilpel, N., see Adeyemi, M.O. **20**, 171
Pilpel, N., see Igwilo, C.I. **15**, 73
Pilpel, N., see Itiola, O.A. **31**, 99
Pilpel, N., see Sakr, F.M. **10**, 43, 57
Pilpel, N., see Wong, L.W. **59**, 145
Pimienta, C., see Juhasz, J. **77**, 309
Pimienta, C., F. Chouinard, A. Labib, V. Lenaerts, Effect of various poloxamer coatings on in vitro adhesion of isohexylcyanoacrylate nanospheres to rat ileal segments under liquid flow **80**, 1
Pimm, M.V., J.A. Clegg, F. Hudecz, R.W. Baldwin, ^{111}In labelling of a branched polypeptide drug carrier with a poly(L-lysine) backbone **79**, 77
Pina, M.E., A.T. Sousa, A.P. Brojo, Enteric coating of hard gelatin capsules. Part 1. Application of hydroalcoholic solutions of formaldehyde in preparation of gastro-resistant capsules **133**, 139
Enteric coating of hard gelatin capsules. Part 2. Lioavailability of formaldehyde treated capsules **148**, 73
Pinelli, M., see Iannuccelli, V. **143**, 187
Ping, Q., T.K. Law, T.L. Whateley, A.T. Florence, Aggregation and gelation of a polymerisable diacryloyl derivative of poloxamer 407 **61**, 79
Pinguet, F., see Martel, P. **149**, 37
Pinio, F., see Giammona, G. **64**, 239
Pinnamaraju, P., see Seth, S.C. **106**, 7
Pinto, J.F., G. Buckton, J.M. Newton, The influence of four selected processing and formulation factors on the production of spheres by extrusion and spheronisation **83**, 187
A relationship between surface free energy and polarity data and some physical properties of spheroids **118**, 95
Pinto, J.F., F. Podczeck, J.M. Newton, Investigations of tablets prepared from pellets produced by extrusion and spheronisation. Part I. The application of canonical analysis to correlate the properties of the tablets to the factors studied in combination with principal component analysis **147**, 79
Pinton, J., see Machet, L. **72**, R1
Pinton, J., see Matchet, L. **133**, 39
Pinzauti, S., see Chafi, N. **52**, 203
Pinzauti, S., see Corti, P. **111**, 83
Pinzauti, S., see Mura, P. **119**, 71
Pinzauti, S., see Santoni, G. **50**, 75; **64**, 235; **80**, 263
Piotrovskij, V.K., M. Gajdos, B. Illek, R. Dzurik, T. Trnovec, Application of a population pharmacokinetic modeling to bioavailability/bioequivalence study of cefadroxil preparations **119**, 213
Pipkin, J.D., see Urtti, A. **61**, 235, 241
Pipkin, J.D., V.J. Stella, Phenindione solubility in mixed organic solvents: Analysis of the role of specific hydrogen and non-hydrogen bonding interactions **14**, 263
Pirjola, J., see Marvola, M. **3**, 13
Pirker, S., J. Kruse, C. Noe, K. Langer, A. Zimmer, J. Kreuter, Characterization of polybutyleyanoacrylate nanoparticles. Part II. Determination of polymer content by NMR-analysis **128**, 189
Pirttimaki, J., E. Laine, J. Ketolainen, P. Paronen, Effects of grinding and compression on crystal structure of anhydrous caffeine **95**, 93
Pitarresi, G., see Cavallaro, G. **90**, 195
Pitarresi, G., see Giammona, G. **105**, 57; **127**, 165
Pitarresi, G., see Turco Liveri, V. **99**, 285
Pitcairn, G., G. Lunghetti, P. Ventura, S. Newman, A comparison of the lung deposition of salbutamol inhaled from a new dry powder inhaler, at two inhaled flow rates **102**, 11
Pitha, J., T. Hoshino, Effects of ethanol on formation of inclusion complexes of hydroxypropylcyclodextrins with testosterone or with methyl orange **80**, 243
Pitha, J., T. Hoshino, J. Torres-Labandeira, T. Irie, Preparation of drug: Hydroxypropylcyclodextrin complexes by a method using ethanol or aqueous ammonium hydroxide as co-solubilizers **80**, 253
Pitha, J., J. Milecki, H. Fales et al., Hydroxypropyl-β-cyclodextrin: Preparation and characterization; effects on solubility of drugs **29**, 73
Pitha, J., see Hoshino, T. **98**, 239
Pitha, J., see Irie, T. **43**, 41

Pitman, I.H., see Williams, D.B. **14**, 113
Pitman, I.H., see Young-Harvey, J.A. **30**, 151
Pitt, C.G., The controlled parenteral delivery of polypeptides and proteins **59**, 173
Pitt, C.G., Y.T. Bao, A.L. Andrady, P.N.K. Samuel, The correlation of polymer-water and octanol-water partition coefficients: Estimation of drug solubilis in polymers **45**, 1
Pitt, K.G., see Craig, D.Q.M. **96**, 147
Pittman, K.A., see Shah, V.P. **82**, 1
Piusieux, F., see Fessi, H. **55**, R1
Pla-Delfina, J.M., Buendia M. Perez, V.G. Casabo et al., Absorption–partition relationships for true homologous series of xenobiotics as a possible approach to study mechanisms of surfactants in absorption. I. Aromatic amines in rat colon **37**, 49
Pla-Delfina, J.M., see Bermejo, M.V. **69**, 221
Pla-Delfina, J.M., see Fabra-Campos, S. **109**, 197
Pla-Delfina, J.M., see Garrigues, T.M. **101**, 209
Pla-Delfina, J.M., see Perez-Buendia, M.D. **55**, 165
Pla-Delfina, J.M., see Ramos, A.J. **128**, 129
Plaizier-Vercammen, J.A., Investigation of the bioavailability of codeine from a cation ion-exchange sulfonic acid. 1. Effect of parameters **85**, 45
Investigation of the bioavailability of codeine from a cation-exchange sulphonic acid. 2. Evaluation of release kinetics of codeine from the resinate and uptake of Na^+ from the solution **87**, 31
Plakogiannis, F., see Mesiha, M. **111**, 213
Plane, J.H., see Allwood, M.C. **19**, 207; **31**, 1
Plate, N.A., see Feldstein, M.M. **131**, 229
Player, M.R., see Michniak, B.B. **91**, 85; **94**, 203; **110**, 231; **116**, 201
Plazonnet, B., see Grove, J. **66**, 23
Plazonnet, B., see Meseguer, G. **95**, 229
Plazonnet, B., see Rozier, A. **57**, 163
Plazonnet, B., see Thermes, F. **81**, 59
Pletcher, S.A., see Moore, J.A. **34**, 35
Plumpton, E.J., P. Gilbert, J.T. Fell, Effect of spatial distribution of contaminant microorganisms within tablet formulations on subsequent inactivation through compaction **30**, 237
The survival of microorganisms during tabletting **30**, 241
Po, A.L.W., see Chan, S.Y. **55**, 1
Po, A.L.W., see Hashim, H. **35**, 201
Po, A.L.W., see Jian, L. **107**, 77
Po, A.L.W., see McCallion, R. **105**, 103; **113**, 247
Pochopin, N.L., W.N. Charman, V.J. Stella, Kinetics of rearrangement and hydrolysis of amino acid derivatives of prazosin **105**, 169
Amino acid derivatives of dapsone as water-soluble prodrugs **121**, 157
Pochopin, N.L., see Charman, W.N. **88**, 397
Podczeck, F., Comparison of in vitro dissolution profiles by calculating mean dissolution time (MDT) or mean residence time (MRT) **97**, 93
The calculation of drug absorption rates of morphine sulphate from a sublingual aerosol preparation using quantified maximum entropy **138**, 129
Assessment of the mode of adherence and the deformation characteristics of micronized particles adhering to various surfaces **145**, 65
Optimization of the operation conditions of an Andersen–Cascade impactor and the relationship to centrifugal adhesion measurements to aid the development of dry powder inhalations **149**, 51
Podczeck, F., S. Dhillon, A.P.R. Wilson, The assessment of pharmacokinetic parameters of teicoplanin in burns comparing the methods of nonlinear curve fitting and quantified maximum entropy **142**, 235
Podczeck, F., G. Lee-Amies, The bulk volume changes of powders by granulation and compression with respect to capsule filling **142**, 97
Podczeck, F., G. Merkel, P. Revesz, The application of canonical analysis to the evaluation of the influence of changes in components of standard direct compression tablet formulations **97**, 15
Podczeck, F., Y. Miah, The influence of particle size and shape on the angle of internal friction and the flow factor of unlubricated and lubricated powders **144**, 187
Podczeck, F., J.M. Newton, The evaluation of a three-dimensional shape factor for the quantitative assessment of the sphericity and surface roughness of pellets **124**, 253
Podczeck, F., J.M. Newton, M.B. James, The influence of constant and changing relative humidity of the air on the autoadhesion force between pharmaceutical powder particles **145**, 221
Variations in the adhesion force between a drug and carrier particles as a result of changes in the relative humidity of the air **149**, 151
Podczeck, F., P. Revesz, Evaluation of the properties of microcrystalline and microfine cellulose powders **91**, 183
Podczeck, F., M. Sharma, The influence of particle size and shape of components of binary powder mixtures on the maximum volume reduction due to packing **137**, 41
Podczeck, F., see Blanque, D. **119**, 203
Podczeck, F., see Lundqvist, A.E.K. **147**, 95
Podczeck, F., see Patel, R. **128**, 123
Podczeck, F., see Pinto, J.F. **147**, 79
Podczeck, F., see Sousa, J.J. **144**, 159
Podres, E.A., see Kosykh, V.A. **57**, 223
Podzceck, F., see Eriksson, M. **148**, 149
Poelma, F.G.J., R. Breas, J.J. Tukker, Intestinal absorption of drugs. IV. The influence of taurocholate and L-cysteine on the barrier function of mucus **64**, 161
Poelma, F.G.J., see Lehr, C.M. **70**, 235
Poelman, M.C., see Amdidouche, D. **54**, 175; **111**, 111
Poelman, M.C., see Lin, S.Z. **69**, 211; **106**, 63
Poelman, M.C., see Thevenin, M.A. **137**, 177
Poindron, P., see Goldbach, P. **123**, 33
Polache, A., see Cejudo-Ferragud, E. **132**, 63
Polack, A.E., see Donaldson, J.D. **88**, 351
Polack, A.E., see Prayurnprohm, P. **100**, 189
Polack, A.E., see Roberts, M.S. **2**, 295; **17**, 145
Polack, A.E., see Siddiqui, O. **27**, 193

Polard, E., P. Le Corre, F. Chevanne, R. Le Verge, In vitro and in vivo evaluation of polylactide and polylactide-*co*-glycolide microspheres of morphine for site-specific delivery **134**, 37
Polderman, J., see Rutten-Kingma, J.J. **3**, 39, 179, 187
Poliszczuk, A., see Maier, G.A. **4**, 75
Politov, A.A., see Shakhtshneider, T.P. **130**, 25
Pollack, G.M., B. Baswell, S.J. Szefler, D.D. Shen, Comparison of inter- and intra-subject variation in oral absorption of theophylline from sustained-release products **21**, 3
Polli, G.P., see Williams, N.A. **44**, 205
Pomeroy, A.R., see Graham, D.T. **1**, 315
Ponchel, G., A.D. Woolfson, D. Wouessidjewe, D. Duchene, N.A. Peppas, Bioadhesive analysis of controlled-release systems. III. Bioadhesive and release behavior of metronidazole-containing poly(acrylic acid)-hydroxypropyl methylcellulose systems **38**, 65
Ponchel, G., see Irache, J.M. **90**, R9
Ponci, R., see Pavanetto, F. **48**, 255; **51**, 55
Ponec, M., see de Vries, M. **76**, 25
Pongpaibul, Y., C.W. Whitworth, Preparation and in vitro dissolution characteristics of propranolol microcapsules **33**, 243
Pongpaibul, Y., see Nakamura, H. **141**, 71
Pons, M., M. Foradada, J. Estelrich, Liposomes obtained by the ethanol injection method **95**, 51
Pons, M., see Coderch, L. **139**, 197
Pons, S., see Masada, T. **49**, 57
Pontikes, D.G., see Pandit, N.K. **122**, 27
Pontikos, N.M., see Acree, Jr. W.E. **31**, 225
Poochikian, G.K., J.C. Cradock, J.P. Davignon, Heroin: Stability and formulation approaches **13**, 219
Poole, J.W., see Tsau, J. **12**, 185
Poole, S.W., see Lashmar, U.T. **103**, 119
Pop, E., F. Soti, W.R. Anderson, J.A. Panetta, K.S. Estes, N.S. Bodor, M.E. Brewster, Redox targeting of LY231617, an antioxidant with potential use in the treatment of brain damage **140**, 33
Pop, E., W. Anderson, J. Vlasak, M.E. Brewster, N. Bodor, Enhanced brain delivery of 2′-fluoro-5-methylarabinosyluracil **84**, 39
Pop, E., see Amselem, S. **125**, 31
Pop, E., see Brewster, M.E. **125**, 17
Popat, S.C., see Ho, A.H.L. **104**, 247
Pope, D.G., J.D. Baggot, Special considerations in veterinary formulation design **14**, 123
Pope, D.G., see Krycer, I. **12**, 113
Porter, C.J.H., S.A. Charman, R.D. Williams, M.V. Bakalova, W.N. Charman, Evaluation of emulsifiable glasses for the oral administration of cyclosporin in beagle dogs **141**, 227
Porter, C.J.H., S.M. Moghimi, M.C. Davies, S.S. Davis, L. Illum, Differences in the molecular weight profile of poloxamer 407 affect its ability to redirect intravenously administered colloids to the bone marrow **83**, 273
Porter, C.J.H., see Bibby, D.C. **144**, 61
Porter, C.J.H., see Moghimi, S.M. **68**, 121
Porter, J., see Hudson, A.J. **133**, 257
Porter, W.R., see Foster, T.M. **134**, 193
Porutiu, D., see Leucuta, S.E. **41**, 213
Possert, P.L., see Pena, L.E. **113**, 89
Postaire, E., M.D. Le Hoang, P. Anglade, D. Martinez, F. Brion, J. Navarro, P. Prognon, D. Pradeau, Stability and behaviour of selenium in total parenteral nutrition solutions **55**, 99
Postaire, E., see Gimenez, F. **43**, 23
Postaire, E., see Regnault, C. **132**, 263
Postaire, E., see Stella, V. **121**, 117
Posti, J., M. Salonen, Effect of formulation factors and food intake on the bioavailability of erythromycin stearate tablets **17**, 225
Potman, R.P., see Beijnen, J.H. **34**, 247
Poujol, S., see Martel, P. **149**, 37
Poulou, M., P. Macheras, Determination of nitrofurantoin in urine by derivative spectroscopy **34**, 29
Poupaert, J.H., see Mbela, T.K.M. **79**, 29; **92**, 71
Poupon, M.F., see Nemati, F. **102**, 55; **138**, 237
Pourcelly, G., see Raffin, F. **120**, 205
Pourcelot, L., see Matchet, L. **133**, 39
Pourcelot, L., see Pelucio-Lopes, C. **96**, 249
Pourcelot, Y., see Andres, C. **144**, 141
Pourkavoos, N., Antimicrobial activity of cephaloridine in the presence of crude and purified preparations of bacterial β-lactamases before and after liposome encapsulation **82**, 53
Pouton, C.W., Self-emulsifying drug delivery systems: Assessment of the efficiency of emulsification **27**, 335
Powell, M.F., A. Magill, A.R. Becker, Non-steroidal anti-psoriatic prodrugs. Hydrolysis and aminolysis of naphthyl ester in aqueous solution **35**, 61
Powell, M.F., A. Magill, A.R. Becker, R.A. Kenley, S. Chen, G.C. Visor, Non-steroidal anti-psoriatic prodrugs. II. Citric acid stabilization of lonapalene in various alcohol media and an ointment formulation **44**, 225
Powers, M.E., A. Adjei, Lu M.F. Fu, M.C. Manning, Solution behavior of leuprolide acetate, an LHRH agonist, as determined by circular dichroism spectroscopy **108**, 49
Pozsgai, K.M., see Pearlman, R. **18**, 53
Pozzi, F., see Wilding, I.R. **111**, 99
Pradat, S., see Aiache, J.M. **16**, 241
Pradeau, D., see Aignasse, M.F. **113**, 241
Pradeau, D., see Postaire, E. **55**, 99
Pradhan, S., A.T. Wu, L.J. Lesko, M.L. Chen, R.L. Williams, Bioavailability measurements of methazolamide in plasma, red blood cells and whole blood: Implications for bioequivalence studies **138**, 207
Prahl, M., see Sjodin, L. **63**, 135
Prakongpan, S., see Swarbrick, J. **32**, 21
Prankerd, R., see Beall, H. **111**, 223
Prankerd, R.J., Solid-state properties of drugs. I. Estimation of heat capacities for fusion and thermodynamic functions for solution from aqueous solubility-temperature dependence measurements **84**, 233
Prankerd, R.J., R.H. McKeown, Physico-chemical properties of barbituric acid derivatives. Part I. Solubility-temperature

dependence for 5,5-disubstituted barbituric acids in aqueous solutions **62**, 37
Physico-chemical properties of barbituric acid derivatives. II. Partition coefficients of 5,5-disubstituted barbituric acids at 25°C **83**, 25
Physico-chemical properties of barbituric acid derivatives. III. Partition coefficients of cycloalkane-1′,5-spirobarbituric acids at 25°C **83**, 39
Physico-chemical properties of barbituric acid derivatives. IV. Solubilities of 5,5-disubstituted barbituric acids in water **112**, 1
Prankerd, R.J., V.J. Stella, Equilibria and kinetics of hydrolysis of ebifuramin (NSC-201047), an azomethine-containing structure exhibiting a reversible degradation step in acidic solutions **52**, 71
Prankerd, R.J., H.W. Stone, K.B. Sloan, J.H. Perrin, Degradation of aspartame in acidic aqueous media and its stabilization by complexation with cyclodextrins or modified cyclodextrins **88**, 189
Prankerd, R.J., J.M. Walters, J.H. Parnes, Kinetics for degradation of rifampicin, an azomethine-containing drug which exhibits reversible hydrolysis in acidic solutions **78**, 59
Prankerd, R.J., see Elsabee, M. **86**, 211, 221
Prankerd, R.J., see Moelands, D. **86**, 263
Prankerd, R.J., see Newton, J.M. **93**, 249
Prapaitrakul, W., see Sprockel, O.L. **48**, 217; **58**, 123
Prasad, C.V.N., see Parimoo, P. **134**, 231
Prasad, K.P.P., see Wan, L.S.C. **41**, 159; **50**, 147; **55**, 115
Prasad, V.K., R.S. Rapaka, P.W. Knight, B.E. Cabana, Dissolution medium—a critical parameter to identify bioavailability problems of furosemide tablets **11**, 81
Prasad, V.K., V.P. Shah, P. Knight et al., Importance of media selection in establishment of in vitro–in vivo relationships for quinidine gluconate **13**, 1
Prasad, V.K., see Goehl, T.J. **11**, 181; **15**, 115
Prasad, V.K., see Rapaka, R.S. **11**, 237; **12**, 285
Prat, J., see Munoz, M. **139**, 249
Prat, J., see Pujol, M. **58**, 103; **146**, 263
Prat, J., see Torres, F. **129**, 275
Pratt, M., see Adjei, A. **61**, 135
Prayurnprohm, P., A.E. Polack, The effect of each of three physicochemical variables on solute uptake by polyvinylchloride infusion bags **100**, 189
Preat, V., S. Thysman, Transdermal iontophoretic delivery of sufentanil **96**, 189
Preat, V., see Gaspar, R. **68**, 111
Preat, V., see Jadoul, A. **120**, 221
Preat, V., see Tasset, C. **58**, 41; **81**, R5
Preat, V., see Thysman, S. **101**, 105
Predella, P., see Catellani, P.L. **51**, 63
Preiss, R., see Antonin, K.H. **130**, 33
Preston, M.S., see Hendriksen, B.A. **118**, 1
Prestrelski, S.J., see Niven, R.W. **127**, 191
Pretzer, D., A.J. Repta, Stability of the experimental anticancer agent [[(4-methoxyphenyl)sulfonyl]hydrazono]acetic acid (NSC-267213). I. Kinetics and mechanism of degradation **38**, 147
Stability of the experimental anticancer agent [[(4-methoxyphenyl)sulfonyl]hydrazono]acetic acid (NSC-267213). II. Effect of structural modification on stability and mechanism of degradation **38**, 161
Prevost, R., see Herremans, J.L.M. **129**, 191
Price, J.H., see Woolfson, A.D. **84**, 69
Price, J.M.C., S.S. Davis, I.R. Wilding, The effect of fibre on gastrointestinal transit times in vegetarians and omnivores **76**, 123
Pridal, L., H. Agerbaek, L.N. Christensen, K. Thomsen, O. Kirk, Absorption of glucagon-like peptide-1 can be protracted by zinc or protamine **136**, 53
Prieto, J.G., see Del Estal, J.L. **91**, 105; **102**, 257
Primorac, M., see Vulovic, N. **55**, 123
Primosic, S., see Mrhar, A. **62**, 15
Prince, A.P., see Bowtle, W.J. **4**, 263
Pringle, J., see Meisner, D. **55**, 105
Prisco, M.R., see Kuu, W.Y. **55**, 77
Prisell, P.T., O. Camber, J. Hiselius, G. Norstedt, Evaluation of hyaluronan as a vehicle for peptide growth factors **85**, 51
Pritchard, K., A.B. Lansley, G.P. Martin, M. Helliwell, C. Marriott, L.M. Benedetti, Evaluation of the bioadhesive properties of hyaluronan derivatives: Detachment weight and mucociliary transport rate studies **129**, 137
Pritchard, R.A., see Johnson, D.M. **31**, 125
Privitera, N., R. Naon, J.G. Riess, Phagocytosis by cultured moused peritoneal macrophages of microspheres coated with perfluoroalkylated telomeric surfactants derived from tris(hydroxymethyl)aminomethane **104**, 41
Privitera, N., R. Naon, P. Vierling, J.G. Riess, Phagocytic uptake by mouse peritoneal macrophages of microspheres coated with phosphocholine or polyethylene glycol phosphate-derived perfluoroalkylated surfactants **120**, 73
Prognon, P., see Aignasse, M.F. **113**, 241
Prognon, P., see Gimenez, F. **43**, 23
Prognon, P., see Postaire, E. **55**, 99
Prognon, P., see Tokgoz, N.S. **141**, 27
Pronk, J., see Moolenaar, F. **19**, 161
Proost, J.H., G.K. Bolhuis, C.F. Lerk, The effect of the swelling capacity of disintegrants on the in vitro and in vivo availability of diazepam tablets, containing magnesium stearate as a lubricant **13**, 287
Proost, J.H., see Bootsma, H.P.R. **51**, 213
Prot, O.M., see Benoit, E.F. **118**, 121
Provasi, D., see Lashmar, U.T. **103**, 119
Provost, C., H. Herbots, R. Kinget, Transparent oil-water gels. Part II. A study of the gel structure in transparent oil-water gels by differential scanning calorimetry **62**, 217
Prudat-Christiaens, C., P. Arnaud, P. Allain, J.C. Chaumeil, Aminophylline bioadhesive tablets attempted by wet granulation **141**, 109
Pryce-Jones, R.H., G.M. Eccleston, B.B. Abu-Bakar, Aminophylline suppository decomposition: An investigation using differential scanning calorimetry **86**, 231
Puech, A., see Delalonde, M. **130**, 147
Puech, A., see El Moussaoui, A. **106**, 109

Pugh, W.J., J. Hadgraft, Ab initio prediction of human skin permeability coefficients **103**, 163
Pugh, W.J., M.S. Roberts, J. Hadgraft, Epidermal permeability–penetrant structure relationships. 3. The effect of hydrogen bonding interactions and molecular size on diffusion across the stratum corneum **138**, 149
Pugh, W.J., see Cabral Marques, H. **63**, 267
Pugh, W.J., see Hadgraft, J. **123**, 285; **141**, 17
Pugh, W.J., see Roberts, M.S. **126**, 219; **132**, 23
Pugh, W.J., see Russell, A.D. **27**, 163; **35**, 227
Puglisi, G., M. Fresta, G. Giammona, C.A. Ventura, Influence of the preparation conditions on poly(ethylcyanoacrylate) nanocapsule formation **125**, 283
Puglisi, G., M. Fresta, G. Mazzone, P.M. Furneri, Tempera, Formulation parameters of fluoroquinolone-loaded liposomes and in vitro antimicrobial activity **118**, 65
Puglisi, C., see Bonina, F. **145**, 87
Puglisi, G., see Castelli, F. **52**, 115; **59**, 19; **70**, 43; **88**, 1; **103**, 217
Puglisi, G., see Cavallaro, G. **111**, 31
Puglisi, G., see Fresta, M. **99**, 145; **134**, 89
Puglisi, G., see Giammona, G. **57**, 55
Puglisi, G., see Panico, A.M. **80**, 93
Puglisi, G., see Santagati, N.A. **65**, 137
Puglisi, G., see Ventura, C.A. **149**, 1
Puglisi, G., see Villari, A. **127**, 185
Puiol, M., see Torres, F. **129**, 275
Puisieux, F., see Bamba, M. **2**, 307
Puisieux, F., see Belbella, A. **129**, 95
Puisieux, F., see Benkerrour, L. **19**, 27
Puisieux, F., see Blanchon, S. **72**, 1
Puisieux, F., see Blanco-Prieto, M.J. **111**, 137
Puisieux, F., see Carstensen, J.T. **1**, 65
Puisieux, F., see Clemessy, M. **101**, 219
Puisieux, F., see Couarraze, G. **56**, 197
Puisieux, F., see Delie, F. **115**, 45
Puisieux, F., see Fessi, H. **1**, 265; **30**, 209
Puisieux, F., see Guterres, S.S. **113**, 57
Puisieux, F., see Husson, I. **86**, 113
Puisieux, F., see Jansen, J. **103**, 19
Puisieux, F., see Leroy-Lechat, F. **101**, 97
Puisieux, F., see Lherm, C. **84**, 13
Puisieux, F., see Mayorga, P. **132**, 71
Puisieux, F., see Mestiri, M. **89**, 229
Puisieux, F., see Mouazen, F. **1**, 275
Puisieux, F., see Pense, A.M. **81**, 111
Puisieux, F., see Rabaron, A. **99**, 29
Puisieux, F., see Ramaldes, A. **143**, 1
Puisieux, F., see Ribardiere, A. **144**, 195
Puisieux, F., see Roblot-Treupel, L. **31**, 131
Puisieux, F., see Rodrigues, Jr. J.M. **126**, 253
Puisieux, F., see Skiba, M. **120**, 1; **126**, 275; **129**, 113; **142**, 121; **145**, 241
Puisieux, F., see Thioune, O. **146**, 233
Puisieux, F., see Vaution, C. **1**, 349
Puisieux, F., see Wesoluch, F. **2**, 343
Pujara, C.P., Z. Shao, M.R. Duncan, A.K. Mitra, Effects of formulation variables on nasal epithelial cell integrity: Biochemical evaluations thermal, mechanical and functional properties of cellulose acetate phthalate (CAP) coatings obtained from neutralized aqueous solutions **114**, 197
Pujol, M., V. Girona, J. Prat, J. Cots, HPLC determination of Azlocillin sodium for stability studies **58**, 103
Pujol, M., V. Girona, J. Prat, M. Munoz, J. De Bolos, Degradation pathway of carboplatin in aqueous solution **146**, 263
Pujol, M., see Girona, V. **41**, 241
Pujol, M., see Munoz, M. **139**, 249
Puozzo, C., see Galleyrand, J.C. **42**, 225
Puranik, P.K., see Ramani, C.C. **137**, 11
Purewal, T.S., see Bower, C. **118**, 229
Puri, A., see Ho, A.H.L. **107**, 199
Purmann, T., see Michel, C. **84**, 93
Purra, K., see Malkki-Laine, L. **117**, 189
Putman, R., see Pikal, M.J. **60**, 203
Pyne, M.T., see Hansen, L.D. **137**, 1
Pyter, R.A., see Garren, K.W. **63**, 167

Q

Qi, X., Y. Maitani, T. Nagai, Effect of soybean-derived sterols on the in vitro stability and the blood circulation of liposomes in mice **114**, 33
Qi, X.R., Y. Maitani, T. Nagai, S.L. Wei, Comparative pharmacokinetics and antitumor efficacy of doxorubicin encapsulated in soybean-derived sterols and poly(ethylene glycol) liposomes in mice **146**, 31
Qiu, Y., P. Gupta, J. Briskin, H. Cheskin, S. Semla, Sustained-release multiparticulate formulations of Zileuton. I. In vitro and in vivo evaluation **143**, 179
Qiu, Y., P. Gupta, H. Cheskin, A. Adjei, Pulmonary bioavailability and absorption characteristics of the 5-lipoxygenase inhibitor, Abbott-79175, in beagle dogs **115**, 95
Qiu, Y., see Gupta, P. **147**, 207
Quaglia, F., see Palagiano, F. **149**, 171
Quaglia, M.G., see Carlucci, G. **49**, 79; **53**, 257
Quan, D., H.I. Maibach, An electron spin resonance study. I. Effect of Azone^{R} on 5-doxyl stearic acid-labeled human stratum corneum **104**, 61
Quan, D., K. Takayama, T. Mitsuzono, K. Isowa, T. Nagai, Influence of novel percutaneous absorption enhancers, cyclohexanone and piperidone derivatives, on histopathology of rat skin **68**, 239
Quarton, M., see Lenaerts, V. **39**, 121
Queen, D., G.P. Martin, C. Marriott, J.E. Fairbrother, Assessment of the potential of a new hydrocolloid dermatological patch (Actiderm) in the treatment of steroid-responsive dermatoses **44**, 25
Quiding, H., see Gjellan, K. **102**, 71
Quigley, J.M., J.M. Blake, F.J. Bonner, The effect of ionization on the partitioning of clofazimine in the 2,2,4-trimethylpentane-water system **54**, 155

Quigley, J.M., K.M.S. Fahelelbom, R.F. Timoney, O.I. Corrigan, Temperature dependence and thermodynamics of partitioning of clofazimine analogues in the *n*-octanol/water system **58**, 107
Quigley, J.M., C.G.M. Jordan, R.F. Timoney, The synthesis, hydrolysis kinetics and lipophilicity of *O*-acyl esters of propranolol **101**, 145
Quigley, J.M., see Irwin, V.P. **34**, 241; **43**, 187
Quigley, J.M., see Jordan, C.G.M. **84**, 175
Quigley, K.J., see Deasy, P.B. **73**, 117
Quincey, S.M., see Staniforth, J.N. **32**, 177
Quinquis-Desmaris, V., see Bourget, P. **128**, 223
Quint, M.P., see Grove, J. **66**, 23
Quintanar-Guerrero, D., H. Fessi, E. Allemann, E. Doelker, Influence of stabilizing agents and preparative variables on the formation of poly(D,L-lactic acid) nanoparticles by an emulsification-diffusion technique **143**, 133
Quraishi, O., see Bechard, S.R. **87**, 133

R

Raatikainen, P., see Silvennoinen, R. **131**, 209
Raatikainen, P., see Sutinen, R. **92**, 177
Rabanel, J.M., see Peyrot, M. **102**, 25
Rabaron, A., G. Cave, F. Puisieux, M. Seiller, Physical methods for measurement of the HLB of ether and ester nonionic surface-active agents: H-NMR and dielectric constant **99**, 29
Rabasco, A.M., J.M. Gines, M. Fernandez-Arevalo, M.A. Holgado, Dissolution rate of diazepam from polyethylene glycol 6000 solid dispersions **67**, 201
Rabasco, A.M., see Alvarez-Fuentes, J. **139**, 237
Rabasco, A.M., see Arias, M.J. **123**, 25
Rabasco, A.M., see Fernandez-Hervas, M.J. **108**, 187
Rabasco, A.M., see Caraballo, I. **89**, 213; **96**, 175; **109**, 229; **139**, 177
Rabasco, A.M., see Fernandez-Hervas, M.J. **113**, 39; **130**, 115; **136**, 101
Rabasco, A.M., see Fini, A. **121**, 19
Rabasco, A.M., see Gines, J.M. **126**, 287
Rabasco, A.M., see Gonzalez-Rodriguez, M.L. **146**, 175
Rabasco, A.M., see Holgado, M.A. **120**, 157; **121**, 187; **142**, 143
Rabasco, A.M., see Lucero, M.J. **104**, 49
Rabasco, A.M., see Moyano, J.R. **114**, 95
Rabasco-Alvarez, A.M., see Fernandez-Arevalo, M. **95**, 117
Rabinow, B., see Payton, R. **33**, 257
Rabinow, B.E., see Jenke, D.R. **78**, 115
Raby, C., see Jambut-Absil, A.C. **35**, 129
Rachmilewitz, E.A., see Grinberg, L.N. **132**, 251
Racz, I., see Dredan, J. **145**, 61
Radebaugh, G.W., S.R. Babu, J.N. Bondi, Characterization of the viscoelastic properties of compacted pharmaceutical powders by a novel nondestructive technique **57**, 95
Radebaugh, G.W., J.L. Murtha, T.N. Julian, J.N. Bondi, Methods for evaluating the puncture and shear properties of pharmaceutical polymeric films **45**, 39
Radebaugh, G.W., see Kearney, A.S. **78**, 25; **92**, 63; **102**, 63; **104**, 169
Radebaugh, G.W., see Mehta, S.C. **108**, 85
Radebaugh, G.W., see Senderoff, R.I. **83**, 65
Radebaugh, G.W., see Zingerman, J.P. **88**, 303
Radu, A., see Atanasiu, R. **90**, 119
Radwan, M.A., see Zaghloul, I.Y. **131**, 131
Rae, I.D., see Young-Harvey, J.A. **30**, 151
Raffin, F., C. Duru, M. Jacob, Permeability to hydrogen ions of an enteric coating polymer and interaction of film formulation factors **145**, 247
Raffin, F., C. Duru, M. Jacob, P. Sistat, J. Sandeaux, R. Sandeaux, G. Pourcelly, C. Gavach, Physico-chemical characterization of the ionic permeability of an enteric coating polymer **120**, 205
Rafiee, Therani M., see Jamali, F. **9**, 221
Ragazzi, E., see Realdon, N. **148**, 155
Raggi, M.A., L. Nobile, I. Perboni, M. Ramini, Extractive colorimetric determination of iron(III) in pharmaceutical formulations by means of trifluoroacetylacetone **61**, 199
Raggi, M.A., see Cavrini, V. **10**, 119; **13**, 333
Raggi, M.A., see Di Pietra, A.M. **35**, 13
Raggi, M.A., see Nobile, L. **40**, 85
Raghavan, N., see Payton, R. **33**, 257
Raghaven, N., see Jacobs, G.P. **32**, 151
Ragnarsson, G., A.W. Holzer, J. Sjogren, The influence of mixing time and colloidal silica on the lubricating properties of magnesium stearate **3**, 127
Ragnarsson, G., A. Sandberg, M.O. Johansson, B. Lindstedt, J. Sjogren, In vitro release characteristics of a membrane-coated pellet formulation—Influence of drug solubility and particle size **79**, 223
Ragnarsson, G., J. Sjogren, Influence of the granulating method on bulk properties and tablettability of a high dosage drug **12**, 163
Ragnarsson, G., J. Sjogren, Tablet compaction to a constant load **16**, 349
Ragnarsson, G., see Davidson, R. **100**, 49; **112**, 153
Ragnarsson, G., see Ek, R. **122**, 49
Ragnarsson, G., see Gren, T. **141**, 53
Ragnarsson, G., see Lindstedt, B. **56**, 261
Ragnarsson, G., see Olsson, B. **119**, 223
Ragno, G., M. Veronico, C. Vetuschi, Gas chromatographic and UV derivative determination of nitrendipine and its photodegradation product **99**, 351
Analysis of nimodipine and its photodegradation product by derivative spectrophotometry and gas chromatography **119**, 115
Ragno, G., see Veronico, M. **119**, 109
Ragno, G., see Vetuschi, C. **65**, 177
Rahman, D., see O'Hagan, D.T. **108**, 133
Raikov, Z., see Simeonova, M. **43**, 267
Raikova, E., see Simeonova, M. **43**, 267
Rainer, H., see Mader, R.M. **68**, 151
Rainova, L., see Bogdanova, S. **38**, 99
Raisanen, J., see Mannermaa, J.P. **103**, 125
Rajabi-Siahboomi, A.R., see Melia, C.D. **100**, 263

Rajagopalan, N., S.C. Chen, W.S. Chow, A study of the inclusion complex of amphotericin-B with γ-cyclodextrin **29**, 161
Rajagopalan, S., see Gopal, N.G.S. **9**, 359
Rajala, R., see Laine, E. **38**, 33
Rajan, Z., see Buckton, G. **74**, 169
Rajan, Z., see Efentakis, M. **70**, 153
Rajasekharan, K.N., see Anto, R.J. **131**, 1
Rajendraprasad, Singh N., see Dowse, R. **39**, 141
Rajewski, L.G., A.A. Stinnett, V.J. Stella, E.M. Topp, Enzymic and non-enzymic hydrolysis of a polymeric prodrug: Hydrocortisone esters of hyaluronic acid **82**, 205
Rak, R., see Antonin, K.H. **130**, 33
Rakic, R., Rao K.V. Ranga, Z. Djuric, Relation between viscosity of carboxymethylcellulose aqueous solutions and charge of its particles in the presence of preservatives **39**, 47
Rakic, R., see Jovanovic, M. **75**, 155
Ralambosoa, C., see Ceolin, R. **74**, 77
Ralph, L., see Habberfield, A. **145**, 1
Ramachandran, C., see Du Plessis, J. **103**, 277; **127**, 273
Ramachandran, C., see Ma, L. **70**, 209; **77**, 127
Ramaldes, A., J.R. Deverre, J.M. Grognet, F. Puisieux, E. Fattal, Use of an enzyme immunoassay for the evaluation of entrapment efficiency and in vitro stability in intestinal fluids of liposomal bovine serum albumin **143**, 1
Ramana, V., S. Uppoor, J.C. Shah, Selective complexing agents designed after adenosine receptor as percutaneous permeation enhancers of drugs **145**, 145
Ramani, C.C., P.K. Puranik, A.K. Dorle, Study of diabietic acid as matrix forming material **137**, 11
Rambaud, J., L. Maury, C. Lefebvre, R. Roques, Comparative physicochemical study of forms I and II of sulphabenzamide **15**, 199
Rambourg, P., see Levy, M.C. **69**, R1
Ramdour, S., see Ford, J.L. **71**, 213
Ramini, M., see Raggi, M.A. **61**, 199
Ramires, P.A., see Richardson, J.L. **115**, 9
Ramos, A.J., E. Hernandez, J.M. Pla-Delfina, M. Merino, Intestinal absorption of zearalenone and in vitro study of non-nutritive sorbent materials **128**, 129
Ramos, T., see Martinez-Pacheco, R. **32**, 99
Ramsey, P.J., see Rowlings, C.E. **116**, 191
Ramzan, I.M., see Chou, R.C. **18**, 217
Ranadive, S.A., see Desai, D.S. **144**, 153
Ranadive, S.A., see Morris, K.R. **108**, 195
Randall, C.S., B.K. DiNenno, R.K. Schultz, L. Dayter, M. Konieczny, S.L. Wunder, Solid-state transformation of a leukotriene antagonist acid◇ **120**, 235
Ranga Rao, K.V., P. Buri, A novel in situ method to test polymers and coated microparticles for bioadhesion **52**, 265
Ranga Rao, K.V., K. Padmalatha Devi, Swelling controlled-release systems: Recent developments and applications **48**, 1
Ranga Rao, K.V., see Baveja, S.K. **19**, 229; **24**, 355; **27**, 157; **31**, 169
Ranga Rao, K.V., see Rakic, R. **39**, 47
Ranson, C., see Allman, J. **115**, 241
Ranson, C., see Billany, M.R. **121**, 107
Rantala, J., see Ketolainen, J. **125**, 45
Rao Jasti, B., J. Du, R.C. Vasavada, Characterization of thermal behavior of etoposide **118**, 161
Rao, K.P., see Narayani, R. **128**, 261
Rao, K.P., see Thacharodi, D. **111**, 235; **120**, 115; **131**, 97
Rao, K.P., see Vasantha, R. **47**, 95
Rao, K.V.R., see Baveja, S.K. **41**, 55; **47**, 133
Rao, L.S., J.G. Hardy, C.G. Wilson, Tissue distribution and fate of free and liposome-encapsulated [^{125}Sb]sodium stibogluconate by γ scintigraphy **17**, 283
Rao, M.N.A., see Kunchandy, E. **57**, 173; **58**, 237; **65**, 261
Rao, M.N.A., see Mathews, S. **76**, 257
Rao, M.N.A., see Sreejayan **100**, 93
Rao, R.C., see Gehenot, A. **45**, 13
Rao, S., see Adjei, A. **90**, 141
Rao, S.S., W.A. Ritschel, Development and in vitro/in vivo evaluation of a colonic release capsule of vasopressin **86**, 35
Rao, V.M., see Hudson, A.J. **136**, 23
Rao, Y.K., see Miller, II K.J. **98**, 101
Rapaka, R.S., J. Roth, G.A. Brine, V.K. Prasad, A simple HPLC method for the dissolution studies on levothyroxine sodium tablets **12**, 285
Rapaka, R.S., J. Roth, V.K. Prasad, The analysis of 4-chloro-5-sulfamoylanthranilic acid in the bulk material and pharmaceutical preparations of furosemide by a high-performance liquid chromatographic method **11**, 237
Rapaka, R.S., see Prasad, V.K. **11**, 81
Rasadi, B., see Harper, Bellantone **30**, 63
Raschkov, I., see Dimitrova, E. **93**, 21
Rashidbaigi, Z.A., see Mahjour, M. **95**, 161
Rasmussen, G.J., H. Bundgaard, Prodrugs of peptides. 10. Protection of di- and tri-peptides against aminopeptidase by formation of bioreversible 4-imidazolidinone derivatives **71**, 45
Prodrugs of peptides. 15. 4-Imidazolidinone prodrug derivatives of enkephalins to prevent aminopeptidase-catalyzed metabolism in plasma and absorptive mucosae **76**, 113
Rasmussen, S.N., see Christrup, L.L. **66**, 169
Rasmussen, S.N., see Hansen, L.B. **88**, 243
Rassing, J., D. Attwood, Ultrasonic velocity and light-scattering studies on the polyoxyethylene-polyoxypropylene copolymer Pluronic F127 in aqueous solution **13**, 47
Rassing, J., see Attwood, D. **9**, 285
Rassing, J., see Tolley, J.A. **14**, 223
Rassing, M., see Buur, A. **16**, 41
Rassing, M.R., see Burgalassi, S. **133**, 1
Rastelli, G., see Costantino, L. **86**, 17
Ratanamaneichatara, S., see Newton, D.W. **11**, 209
Ratcliffe, J.H., see Pennington, A.K. **43**, 221
Rathbone, M.J., Human buccal absorption. I. A method for estimating the transfer kinetics of drugs across the human buccal membrane **69**, 103

Human buccal absorption. II. A comparative study of the buccal absorption of some parahydroxybenzoic acid derivatives using the buccal absorption test and a buccal perfusion cell **74**, 189
Rathbone, M.J., J. Hadgraft, Absorption of drugs from the human oral cavity **74**, 9
Rathbone, M.J., see Byron, P.R. **21**, 107; **29**, 103
Rathbone, M.J., see Medlicott, N.J. **143**, 25
Rathi, R., see Singh, M. **76**, R5
Rathi, R.C., see Rihova, B. **87**, 105
Rattie, E.S., see Lindenbaum, S. **26**, 123
Rau, H., see Sher, A. **133**, 237
Raudino, A., see Castelli, F. **70**, 43
Ravecca, S., see Saettone, M.F. **20**, 187
Ravelli, V., see Wilding, I.R. **76**, 133
Rawicki, H.B., see Sitaram, B.R. **118**, 181
Rawlins, D.A., J.B. Kayes, Pharmaceutical suspension studies. I. A comparison of adsorption of polyvinylalcohol at the diloxanide furoate B.P. and polystyrene latex–water interface **13**, 145
Pharmaceutical suspension studies. II. The interaction between particles of diloxanide furoate B.P. coated with an adsorbed polymeric layer **13**, 159
Pharmaceutical suspension studies. III. The redispersibility of suspensions **13**, 171
Rawlins, D.A., see Ahmed, H. **130**, 195
Rawlins, D.A., see Mall, S. **131**, 41
Rawnsley, S., see Summers, R.S. **30**, 83
Raynolds, J.R., see Hardy, J.G. **31**, 151
Raz, I., see Hussein, Z. **37**, 97
Razdan, B., P.K. Verma, Evaluation of dosage forms. III: Studies on commercial acetaminophen tablet dosage forms **79**, 83
Realdon, N., E. Ragazzi, Zotto M. Dal, Fini Dalla, Layered excipient suppositories: The possibility of modulating drug availability **148**, 155
Reardon, P.M., K.L. Audus, Ammonium glycyrrhizinate (AMGZ) effects on membrane integrity **94**, 161
Recamales, A.F., see Asuero, A.G. **34**, 81
Reddy, I.K., see Singh, S.K. **141**, 179
Redell, M.A., D.C. Yang, V.H.L. Lee, The role of esterase activity in the ocular disposition of dipivalyl epinephrine in rabbits **17**, 299
Redell, M.A., see Lee, V.H.L. **16**, 163
Redell, M.A., see Stratford Jr., R.E. **13**, 263
Redenti, E., T. Peveri, M. Zanol, P. Ventura, G. Gnappi, A. Montenero, A study on the differentiation between amorphous piroxicam: β-cyclodextrin complex and a mixture of the two amorphous components **129**, 289
Redgrave, T.G., see Lundberg, B.B. **134**, 119
Reer, O., B.W. Muller, Determination of the degree of substitution of chemically modified cyclodextrins by a microcalorimetric titration technique **104**, 239
Rees, J.A., see Wilson, C.G. **50**, 155
Rees, J.E., K.D. Tsardaka, Compaction stress relaxation interpreted using a hyperbolic relation **92**, 137
Rees, J.E., see Malamataris, S. **92**, 123; **141**, 101
Regan, P., see Smith, A. **59**, 115
Reginault, P., see Andres, C. **144**, 141
Regnault, C., C. Benoist, H. Fessi, M. Roch-Arveiller, E. Postaire, G. Hazebroucq, Preparation of superoxide dismutase entrapped in ceramide-containing liposomes for oral administration **132**, 263
Regosz, A., T. Pelplinska, P. Kowalski, Z. Thiel, Prediction of solubility of sulfonamides in water and organic solvents based on the extended regular solution theory **88**, 437
Reid, A.S., N.W. Thomas, K.J. Palin, P.L. Gould, Formulation of fenbufen suppositories. I. Quantitative histological assessment of the rectal mucosa of rats following treatment with suppository bases **40**, 181
Reid, A.S., see Young, C. **40**, 187
Reid, B.D., see El-Bagory, I. **105**, 255
Reier, G.E., see Patel, N.K. **110**, 203
Reig, F., M.A. Busquets, Anton J.M. Garcia, G. Valencia, M.A. Alsina, Interaction of analgesics with lecithin and ganglioside monolayers **44**, 257
Reig, F., C. Mestres, Anton J.M. Garcia, G. Valencia, M.A. Alsina, Penetration of morphine and naloxone in phosphatidylserine monolayers **44**, 269
Reig, F., see Colome, C. **90**, 59
Reig, F., see Fonseca, M.J. **133**, 265
Reig, F., see Mestres, C. **76**, 145
Reinert, J.D., see Neervannan, S. **96**, 167
Reits, D.J., see Gordon, R.E. **21**, 99
Reizopoulou, V., see Parissi-Poulou, M. **51**, 169
Rekkas, D.M., see Athanassiou, G.C. **72**, 141; **85**, 209; **90**, 51
Rekkas, D.M., see Giannakou, S.A. **125**, 7
Rekkas, D.M., see Vlachou, M.D. **82**, 47
Remon, J.P., see Inghelbrecht, S. **148**, 103
Remon, J.P., see Vervaet, C. **106**, 157; **107**, 29
Remon, J.P., see Baert, L. **86**, 187; **95**, 135; **96**, 225; **99**, 7, 181
Remon, J.P., see Bottenberg, P. **49**, 147; **58**, 203
Remon, J.P., see Bouckaert, S. **130**, 257
Remon, J.P., see de Muynck, C. **85**, 103; **104**, 1, 87
Remon, J.P., see de Vroe, C. **65**, 49
Remon, J.P., see Herman, J. **42**, 15; **55**, 143; **56**, 51, 65; **63**, 201
Remon, J.P., see Kiekens, A. **146**, 239
Remon, J.P., see Ntawukulilyayo, J.D. **91**, 23; **93**, 209; **121**, 205; **128**, 73; **139**, 79
Remon, J.P., see Van Aerde, P. **45**, 145
Remon, J.P., see Vandenbossche, G.M.R. **67**, 195
Remon, J.P., see Vermeersch, H. **60**, 253
Remon, J.P., see Vermeire, A. **146**, 213
Remon, J.P., see Vervaet, C. **108**, 207; **116**, 131; **133**, 29
Remon, J.P., see Visavarungroj, N. **59**, 73; **62**, 125; **65**, 43; **69**, 43
Remon, J.P., see Voorspoels, J. **133**, 9
Remon, J.P., see Zhou, F. **133**, 155; **147**, 23
Remunan, C., M.J. Bretal, A. Nunez, Jato J.L. Vila, Accelerated stability study of sustained-release nifedipine tablets prepared with Gelucire^R **80**, 151

Remunan-Lopez, C., R. Bodmeier, Effect of formulation and process variables on the formation of chitosan-gelatin coacervates **135**, 63
Renaux, C., see Arnaud, P. **80**, 129
Renaux, Ch., see Sarbach, Ch. **140**, 169
Renaux, J., see Bonhomme, L. **80**, 271
Renoux, R., see Aiache, J.M. **39**, 235
Renzi, G., see Santoni, G. **71**, 1; **80**, 263
Repin, V.S., see Sviridov, D.D. **96**, 1
Repond, C., S.S. Davis, H. Van de Waterbeemd, B. Testa, W. Linert, Thermodyanmics and mechanism of partitioning of pyridylalkanamides in *n*-octanol/water and di-*n*-butyl ether/water **38**, 47
Reppas, C., see Koupparis, M. **20**, 325
Reppas, C., see Macheras, P. **37**, 103
Repta, A.J., M.J. Hageman, J.P. Patel, Enol esters as potential prodrugs. IV. Enhanced delivery of the quaternary species coralyne to rat brain using 6′-acetylpapaverin and its enol esters as prodrugs **10**, 239
Repta, A.J., A.A. Hincal, Complexation and solubilization of acronine with alkylgentisates **5**, 149
Repta, A.J., J.P. Patel, Enol esters as potential prodrugs. III. Stability and enzyme-mediated hydrolysis of enol esters of 6′-acetylpapaverin **10**, 29
Repta, A.J., L.A. Sternson, K.A. Mereish, N.M. Meltzer, Complexation in the retention of benzoic acids on a bonded xanthine HPLC stationary phase **18**, 277
Repta, A.J., see El-Sayed A.A. **13**, 303
Repta, A.J., see Garren, K.W. **24**, 91; **48**, 189
Repta, A.J., see Gogate, U.S. **40**, 235, 249
Repta, A.J., see Long, D.F. **6**, 167
Repta, A.J., see Patel, J.P. **5**, 329; **9**, 29
Repta, A.J., see Patton, T.F. **10**, 77
Repta, A.J., see Pretzer, D. **38**, 147, 161
Repta, A.J., see Urtti, A. **61**, 235
Reschova, S., see Hampl, J. **144**, 107
Resetarits, D.E., K.C. Cheng, B.A. Bolton et al., Dissolution behavior of 17β-estradiol (E_2) from povidone coprecipitates. Comparison with microcrystalline and macrocrystalline E_2 **2**, 113
Rettig, W., see Wegener, M. **128**, 203
Reunamaki, T., see Suhonen, P. **127**, 85
Reuning, R.H., see Yoshimura, Y. **38**, 179
Reuvers, K.A., see Schoonen, A.J.M. **7**, 29
Reverdy, M.E., see Lafforgue, C. **147**, 243
Revesz, P., see Podczeck, F. **91**, 183; **97**, 15
Reyderman, L., S. Stavchansky, Electrostatic spraying and its use in drug delivery—cholesterol microspheres **124**, 75
Reynes, M., see Leomy, C. **115**, 235
Reynolds, D.L., K.K. Ulrich, T.F. Patton et al., Plasma levels of 1,4-dihydroxy-5,8-bis(([2-[(2-hydroxylethyl)amino]ethyl]-amino)-9,10-anthrace nedione dihydrochloride (DHAD) in humans **9**, 67
Rhee, G.J., see Hwang, S.J. **116**, 125
Rhie, J.K., see Gordon, M.S. **97**, 119
Rhodes, C.T., see Jimenez-Castellanos, M.R. **89**, 223; **105**, 65
Rhodes, C.T., see Romero, A.J. **99**, 125
Ribardiere, A., P. Tchoreloff, G. Couarraze, F. Puisieux, Modification of ketoprofen bead structure produced by the spherical crystallization technique with a two-solvent system **144**, 195
Ricart, G., see Larhrib, H. **147**, 187
Riccieri, F., see Touitou, E. **103**, 131
Riccieri, F.M., see Touitou, E. **78**, 85
Richards, F.M., see Alpar, H.O. **62**, 133
Richards, J.H., see Barzegar-Jalali, M. **2**, 195; **3**, 133; **4**, 127, 171
Richards, R.M.E., R.B. Baylor, J.Z. Xing, Investigations of the correlation between bacterial uptakes of trimethoprim and sulphadiazine with antibacterial activities against *Enterococcus faecalis* **92**, 237
Richards, R.M.E., D.K.L. Xing, Evaluation of synergistic effects of combinations of antibacterials having relevance to treatment of burn wound infections **75**, 81
Enhancement of antibacterial activity by *p*-aminobenzoic acid and sulphadiazine **82**, 107
Investigation of the antibacterial activity of *p*-aminobenzoic acid against *P. aeruginosa* and *E. cloacae* **87**, 195
The effect of *p*-aminobenzoic acid on the uptake of thymidine and uracil by *Escherichia coli* **116**, 217
Richards, R.M.E., see Taylor, R.B. **102**, 201
Richardson, J.L., N.F. Farraj, L. Illum, Enhanced vaginal absorption of insulin in sheep using lysophosphatidylcholine and a bioadhesive microsphere delivery system **88**, 319
Richardson, J.L., P.S. Minhas, N.W. Thomas, L. Illum, Vaginal administration of gentamicin to rats. Pharmaceutical and morphological studies using absorption enhancers **56**, 29
Richardson, J.L., P.A. Ramires, M.R. Miglietta, M. Rochira, L. Bacelle, L. Callegaro, L. Benedetti, Novel vaginal delivery systems for calcitonin. I. Evaluation of HYAFF/calcitonin microspheres in rats **115**, 9
Richardson, J.L., N.W. Thomas, L. Illum, Recovery of the rat vaginal epithelium from the histological effects of absorption enhancers **77**, 75
Richardson, J.L., see Rochira, M. **144**, 19
Richardson, J.P., see Lashmar, U.T. **125**, 315
Richardson, M., see Fitzgerald, P. **83**, 177
Richardson, P.J., D.F. McCafferty, A.D. Woolfson, Determination of three-component partial solubility parameters for temazepam and the effects of change in partial molal volume on the thermodynamics of drug solubility **78**, 189
Richomme, P., see Pech, B. **128**, 179
Richter, P., Second-derivative spectrophotometric assay of bromazepam in pharmaceutical formulations **72**, 207
Ridd, M.J., K.F. Brown, R.G. Moore, R.L. Nation, Drug plasma binding and non-esterified fatty acids: Methodologic considerations **11**, 11
Ridout, G., see Hadgraft, J. **42**, 97
Ridout, G., see Washington, N. **39**, 163
Rieg-Falson, F., see Blanchon, S. **72**, 1
Riegelman, S., see Collier, P.S. **26**, 99
Riepma, K.A., B.G. Dekker, C.F. Lerk, The effect of moisture sorption on the strength and internal surface area of lactose tablets **87**, 149

Riepma, K.A., B.G. Dekker, R.S. Jager, P.A. Elberse, C.F. Lerk, The effect of storage at ambient humidity on the BET-specific surface area of tablets compacted from different materials **90**, R1
Riepma, K.A., C.F. Lerk, Boer A.H. De, G.K. Bolhuis, K.D. Kussendrager, Consolidation and compaction of powder mixtures. I. Binary mixtures of same particle size fractions of different types of crystalline lactose **66**, 47
Riepma, K.A., J. Veenstra, A.H. De Boer, G.K. Bolhuis, K. Zuurman, C.F. Lerk, H. Vromans, Consolidation and compaction of powder mixtures. II. Binary mixtures of different particle size fractions of α-lactose monohydrate **76**, 9
Riepma, K.A., H. Vromans, K. Zuurman, C.F. Lerk, The effect of dry granulation on the consolidation and compaction of crystalline lactose **97**, 29
Riepma, K.A., K. Zuurman, G.K. Bolhuis, A.H. De Boer, C.F. Lerk, Consolidation and compaction of powder mixtures. III. Binary mixtures of different particle size fractions of different types of crystalline lactose **85**, 121
Riepma, K.A., see Zuurman, K. **102**, 1
Riess, J.G., see Privitera, N. **104**, 41; **120**, 73
Rieutord, A., P. Bourget, G. Troche, J.F. Zazzo, In vitro study of the protein binding of fusidic acid: A contribution to the comprehension of its pharmacokinetic behaviour **119**, 57
Rihova, B., R.C. Rathi, P. Kopeckova, J. Kopecek, In vitro bioadhesion of carbohydrate-containing *N*-(2-hydroxypropyl)methacrylamide copolymers to the GI tract of guinea pigs **87**, 105
Riis, K.J., see Bechgaard, E. **106**, 237
Rijli, P., see Saettone, M.F. **126**, 83
Riley, C.M., see Allsopp, M.A. **69**, 197
Riley, C.M., see Kennedy, P.E. **48**, 179
Riley, C.M., see Monnot, E.A. **60**, 41
Riley, C.M., see Paborji, M. **42**, 243
Riley, C.M., see Ross, D.L. **63**, 237; **83**, 267; **87**, 203; **88**, 379; **90**, 179; **93**, 121, 131
Riley, C.M., see Stout, S.A. **24**, 193; **37**, 257
Rillosi, M., G. Buckton, Modelling mucoadhesion by use of surface energy terms obtained by the Lewis acid–Lewis base approach **117**, 75
Rim, S., K.L. Audus, R.T. Borchardt, Relationship of octanol/buffer and ocatanol/water partition coefficients to transcellular diffusion across brain microvessel endothelial cell monolayers **32**, 79
Rim, S., see Harper, Bellantone **30**, 63
Rimer, A., see Friedman, M. **44**, 243
Rimoy, G.H., see Wilding, I.R. **132**, 81
Ringard-Lefebvre, C., see Santi, P. **68**, 43
Risca, R., see Leucuta, S.E. **41**, 213
Risha, P.A., see Vervaet, C. **107**, 29
Ritchie, K., A.J. Winfield, An investigation of phenobarbitone-sucrose glass systems **18**, 89
Ritschel, W.A., W. Cacini, Biological model for the study of the digoxin–quinidine drug interaction **13**, 353
Ritschel, W.A., see Banerjee, P.S. **49**, 189, 199
Ritschel, W.A., see Lootvoet, G. **85**, 113
Ritschel, W.A., see Rao, S.S. **86**, 35
Ritschel, W.A., see Sathyan, G. **114**, 75
Ritschel, W.A., see Turkoglu, M. **103**, 115
Rius, F., see Canton, E. **92**, 47
Rivera, S.L., S. Ghodbane, In vitro adsorption-desorption of famotidine on microcrystalline cellulose **108**, 31
Riviere, J.E., J.D. Brooks, P.L. Williams, E. McGown, M.L. Francoeur, Cutaneous metabolism of isosorbide dinitrate after transdermal administration in isolated perfused porcine skin **127**, 213
Rizk, S., J.C. Guyot, C. Duru, D. Gaudy, Influence of lubricant properties on compression behaviour and drug dissolution rate of scleroglucan hydrophilic matrix **126**, 57
Rizovski, B., see Mader, R.M. **68**, 151
Robak, T., see Szwarocka, A. **135**, 167
Robb, D.A., see Saleh, A.M. **57**, 205
Roberts, C.J.C., see Ali, H.M. **42**, 155
Roberts, J.M.C., see Kohli, R. **26**, 275
Roberts, L.K., see Law, M.Y.L. **88**, 251
Roberts, M.S., P.A. Cossum, E.A. Kowaluk, A.E. Polack, Factors affecting the availability of organic nitrates from plastic infusion systems: structure of organic nitrate, nature of plastic and effect of temperature **17**, 145
Roberts, M.S., A.E. Polack, G. Martin, H.D. Blackburn, The storage of selected substances in aqueous solution in polyethylene containers: The effect of some physicochemical factors on the disappearance kinetics of the substances **2**, 295
Roberts, M.S., W.J. Pugh, J. Hadgraft, Epidermal permeability: Penetrant structure relationships. 2. The effect of H-bonding groups in penetrants on their diffusion through the stratum corneum **132**, 23
Roberts, M.S., W.J. Pugh, J. Hadgraft, A.C. Watkinson, Epidermal permeability–penetrant structure relationships. 1. An analysis of methods of predicting penetration of monofunctional solutes from aqueous solutions **126**, 219
Roberts, M.S., see Donaldson, J.D. **88**, 351
Roberts, M.S., see Pugh, W.J. **138**, 149
Roberts, M.S., see Siddiqui, O. **27**, 193
Roberts, M.S., see Singh, P. **101**, R5
Roberts, M.S., see Wanwimolruk, S. **75**, 55
Roberts, R.J., R.C. Rowe, Brittle/ductile behaviour in pharmaceutical materials used in tabletting **36**, 205
The Young's modulus of pharmaceutical materials **37**, 15
Determination of the critical stress intensity factor (K(IC)) of microcrystalline cellulose using radially edge-cracked tablets **52**, 213
The solubility parameter and fractional polarity of microcrystalline cellulose as determined by mechanical measurement **99**, 157
The unit cell dimensions of (*R*,*S*)-propranolol hydrochloride—A confirmatory study using data from powder X-ray diffraction **109**, 83
Influence of polymorphism on the Young's modulus and yield stress of carbmazepine, sulfathiazole and sulfanilamide **129**, 79

Roberts, R.J., R.C. Rowe, P. York, The measurement of the critical stress intensity factor (K(IC)) of pharmaceutical powders using three point single edge notched beam (SENB) testing **91**, 173
The Poisson's ratio of microcrystalline cellulose **105**, 177
The relationship between the fracture properties, tensile strength and critical stress intensity factor of organic solids and their molecular structure **125**, 157
Roberts, R.J., see Bassam, F. **64**, 55
Roberts, R.J., see Bateman, S.D. **49**, 209
Roberts, R.J., see Payne, R.S. **145**, 165
Roberts, R.J., see Rowe, R.C. **78**, 49; **86**, 49
Roberts, R.J., see York, P. **66**, 143
Robertson, C., see Davis, S.S. **68**, 199
Robertson, C.S., see Hardy, J.G. **48**, 79
Robertson, J., see Cole, S.K. **88**, 211
Robertson, J.D., see MacLean, D.S. **131**, 243
Robertson, S.S.D., see Fassihi, A.R. **50**, 79; **72**, 175
Robic, D., see Nimier, K. **105**, 235
Robins, R.H., see Snider, B.G. **66**, 63
Robinson, D.H., see Ezzedeen, F.W. **59**, 255
Robinson, D.H., see Sampath, S.S. **78**, 165
Robinson, G., see Bryan, A.J. **7**, 145
Robinson, J., see Gandhi, R. **85**, 129
Robinson, J.R., see Calvo, P. **103**, 283
Robinson, J.R., see Chang, R.K. **80**, 171
Robinson, J.R., see Diepold, R. **54**, 149
Robinson, J.R., see Dowty, M.E. **88**, 293
Robinson, J.R., see Gupta, P.K. **43**, 45; **109**, 91; **125**, 185
Robinson, J.R., see Hui, H.W. **26**, 203
Robinson, J.R., see Lee, V.H.L. **11**, 155
Robinson, J.R., see Li, V.H.K. **53**, 219
Robinson, J.R., see Liaw, J. **59**, 105; **88**, 111, 125
Robinson, J.R., see Park, K. **19**, 107
Robinson, J.R., see Rojanasakul, Y. **61**, 163; **63**, 1; **66**, 131; **68**, 135; **55**, 237
Robinson, J.R., see Scherer, D. **101**, 165
Robinson, J.R., see Wood, R.W. **20**, 285; **23**, 175; **29**, 127
Roblot-Treupel, L., F. Puisieux, Distribution of magnesium stearate on the surface of lubricated particles **31**, 131
Roblot-Treupel, L., see Al Khouri Fallouh, N. **28**, 125
Roblot-Treupel, L., see Seijo, B. **62**, 1
Roca, M., I. Carrio, J. Mones, J. Mora, M. Estorch, L. Berna, Labelling procedure of antacid preparations using ^{99m}Tc-pyrophosphate **69**, 189
Rocco, W.L., C. Morphet, S.M. Laughlin, Solid-state characterization of zanoterone **122**, 17
Rocco, W.L., J.R. Swanson, WIN 63843 polymorphs: Prediction of enantiotropy **117**, 231
Roch-Arveiller, M., see Regnault, C. **132**, 263
Rochard, E., D. Barthes, P. Courtois, Stability and compatibility study of carboplatin with three portable infusion pump reservoirs **101**, 257
Rochat, M.H., see Andres, C. **144**, 141
Rochira, M., M.R. Miglietta, J.L. Richardson, L. Ferrari, M. Beccaro, L. Benedetti, Novel vaginal delivery systems for calcitonin. II. Preparation and characterization of HYAFF^{R} microspheres containing calcitonin **144**, 19
Rochira, M., see Ghezzo, E. **87**, 21
Rochira, M., see Richardson, J.L. **115**, 9
Rodado, P., see Chantres, J.R. **138**, 139
Rodarte, J.A., see Vidgren, M. **115**, 209
Rodley, G.A., H.K. Chan, I. Gonda, Dissolution studies of hexamethylmelamine **95**, 143
Rodman, J.H., see Peck, C.C. **82**, 9
Rodrigues Jr., J.M., H. Fessi, C. Bories, F. Puisieux, -Ph. J. Devissaguet, Primaquine-loaded poly(lactide) nanoparticles: Physicochemical study and acute tolerance in mice **126**, 253
Rodriguez, Barbero J., E.L. Marino, Accelerated stability studies on Rocephin by high-efficiency liquid chromatography **19**, 199
Rodriguez, I.C., see Fernandez, M. **84**, 197; **98**, 29
Rodriguez, I.C., see Margarit, M.V. **81**, 67
Rodriguez, L., see Saettone, M.F. **86**, 159; **126**, 83
Rodriguez Pico, A., see de Bolos, J. **40**, 175
Rodriguez-Hornedo, N., D. Lechuga-Ballesteros, H.J. Wu, Phase transition and heterogeneous/epitaxial nucleation of hydrated and anhydrous theophylline crystals **85**, 149
Rodriguez-Hornedo, N., see Lechuga-Ballesteros, D. **115**, 139, 151
Rodriguez-Hornedo, N., see Zipp, G.L. **51**, 147; **98**, 189
Rodriguez-Puente, B., see Cal, S. **136**, 13
Rodriquez, I.C., see Margarit, M.V. **108**, 101
Rodriquez, L., see Dal Pozzo, A. **50**, 97
Roe, J.M., B.W. Barry, Photon correlation spectroscopy of pharmaceutical systems (sodium dodecyl sulphate, sodium deoxycholate and chlorpromazine hydrochloride micelles and polystyrene lattices) **14**, 159
Roe, J.M., see Buxton, P.C. **42**, 135
Roe, R.L., see Kent, J.S. **7**, 245
Roets, E., see Hendrix, C. **100**, 213
Roets, E., see Naidong, W. **96**, 13
Roets, E., see Paesen, J. **113**, 215
Roets, E., see Van Schepdael, A. **73**, 105
Rogers, D.J., see Nicholls, T.J. **138**, 175
Rogers, D.T., see El-Falaha, B.M.A. **23**, 239; **25**, 329
Rogers, J.A., Solution thermodynamics of phenols **10**, 89
Rogers, J.A., Lipid monolayer interaction behavior of sympathomimetic amines **21**, 75
Rogers, J.A., see Betageri, G.V. **36**, 165
Rogers, J.A., see Habib, M.J. **44**, 235
Rogers, J.A., see Owusu-Ababio, G. **122**, 9
Rogers, J.A., see Vudathala, G.K. **69**, 13
Rogers, J.A., see Yamamura, S. **130**, 65
Rogers, J.D., R.B. Lee, P.R. Souder et al., Pharmacokinetic evaluation of osmotically controlled indomethacin delivery systems in man **16**, 191
Rogers, M.S., see Pickup, M.E. **22**, 311
Rogers, N.J., see Aungst, B.J. **33**, 225; **53**, 227
Rohdewald, P., see Derendorf, H. **15**, 167
Rohdewald, P., see El Samaligy, M. **13**, 23
Rohdewald, P., see Krause, H.J. **27**, 145
Rohr, G., see Djordjevic, N.M. **81**, 21
Rohr, U., see Masada, T. **49**, 57

Rohr, U., see Ruland, A. **107**, 23
Rojanasakul, Y., J. Liaw, J.R. Robinson, Mechanisms of action of some penetration enhancers in the cornea: Laser scanning confocal microscopic and electrophysiology studies **66**, 131
Rojanasakul, Y., S.W. Paddock, J.R. Robinson, Confocal laser scanning microscopic examination of transport pathways and barriers of some peptides across the cornea **61**, 163
Rojanasakul, Y., J.R. Robinson, Transport mechanisms of the cornea: Characterization of barrier permselectivity **55**, 237
Electrophysiological and ultrastructural characterization of the cornea during in vitro perfusion **63**, 1
The cytoskeleton of the cornea and its role in tight junction permeability **68**, 135
Rojanasakul, Y., see Liaw, J. **88**, 111
Rojas, O., see Alvarez-Fuentes, J. **139**, 237
Roksvaag, P.O., see Waehler, K. **97**, 55
Roland, M., see Gaspar, R. **68**, 111
Roland, M., see Tasset, C. **58**, 41; **81**, R5
Rolland, A., Pharmacokinetics and tissue distribution of doxorubicin-loaded polymethacrylic nanoparticles in rabbits **42**, 145
Clinical pharmacokinetics of doxorubicin in hepatoma patients after a single intravenous injection of free or nanoparticle-bound anthracycline **54**, 113
Rolland, A., J.M. Begue, Verge R. Le, A. Guillouzo, Increase of doxorubicin penetration in cultured rat hepatocytes by its binding to polymethacrylic nanoparticles **53**, 67
Rolland, A., A. Brzokewicz, B. Shroot, J.C. Jamoulle, Effect of penetration enhancers on the phase transition of multilammellar liposomes of dipalmitoylphosphatidylcholine. A study by differential scanning calorimetry **76**, 217
Rolland, A., C. Washington, B. Genetet, Verge R. Le, Monoclonal antibodies covalently coupled to polymethacrylic nanoparticles: In vitro specific targeting to human T lymphocytes **39**, 173
Roller, R.G., see Bodor, N. **1**, 189
Rollet, J., see Armand, J.Y. **40**, 33
Rollet, M., see Bogdanova, S. **38**, 99
Rollet, M., see Droin, A. **27**, 233
Rollet, M., see Farah, N. **36**, 81
Rollet, M., see Sebert, P. **99**, 37; **106**, 103; **110**, 285; **122**, 129
Romani, A., see Mulinacci, N. **128**, 81
Rombaut, P., see Talukdar, M.M. **129**, 233
Rombaut, P., see Vertommen, J. **146**, 21
Romeijn, S.G., J.C. Verhoef, E. Marttin, F.W.H.M. Merkus, The effect of nasal drug formulations on ciliary beating in vitro **135**, 137
Romeijn, S.G., see Marttin, E. **141**, 151
Romer Rassing, M., see Jacobsen, J. **125**, 165; **141**, 217
Romero, A.J., L. Savastano, C.T. Rhodes, Monitoring crystal modifications in systems containing ibuprofen **99**, 125
Romeyn, S.G., see Schipper, N.G.M. **64**, 61
Ronchi, M., see Pena Romero, A. **73**, 239
Roomer, A.C.J., see O'Neal, J.S. **12**, 355
Roos, C.F., S. Matsumoto, Y. Takakura et al., Physicochemical and antitumor characteristics of some polyamino acid prodrugs of mitomycin C **22**, 75
Roques, C., see Boulenc, X. **123**, 13
Roques, C., see Caro, I. **116**, 147
Roques, R., see Rambaud, J. **15**, 199
Rork, G., see Urtti, A. **61**, 235, 241
Rork, G.S., see Haslam, J.L. **56**, 227
Rork, G.S., see Merfeld, A.E. **69**, 63
Roseboom, H., see Sorel, R.H.A. **3**, 93
Roseman, T.J., see Kuu, W.Y. **55**, 77
Rosen, A., P. Macheras, K.D. Howe, The use of constant volume ultrafiltration for dissolution studies **32**, 171
Rosen, A., see Macheras, P. **6**, 175; **16**, 353
Rosen, D., see Grynpas, R.S.O. **22**, 357
Rosenberg, E., see Barnwell, S.G. **128**, 145
Rosenberg, E., see Burns, S.J. **141**, 9
Rosenthaler, J., see Nimmerfall, F. **32**, 1
Rosing, H., see Beijnen, J.H. **32**, 111
Rosoff, M., T.M. Serajuddin, Solubilization of diazepam in bile salts and in sodium cholate-lecithin-water phases **6**, 137
Ross, D.L., S.K. Elkinton, S.R. Knaub, C.M. Riley, Physicochemical properties of the fluoroquinolone antimicrobials. VI. Effect of metal-ion complexation on octan-1-ol-water partitioning **93**, 131
Ross, D.L., S.K. Elkinton, C.M. Riley, Physicochemical properties of the fluoroquinolone antimicrobials. III. 1-Octanol/water partition coefficients and their relationships to structure **88**, 379
Erratum: Physicochemical properties of the fluoroquinolone antimicrobials. IV. 1-Octanol-water partition coefficients and their relationships (Int. J. Pharm. (1992) 379–389) **90**, 179
Ross, D.L., C.M. Riley, Aqueous solubilities of some variously substituted quinolone antimicrobials **63**, 237
Physiochemical properties of the fluoroquinolone antimicrobials. II. Acid ionization constants and their relationship to structure **83**, 267
Physicochemical properties of the fluoroquinolone antimicrobials. III. Complexation of lomefloxacin with various metal ions and the effect of metal ion complexation on aqueous solubility **87**, 203
Physicochemical properties of the fluoroquinolone antimicrobials. V. Effect of fluoroquinolone structure and pH on the complexation of various fluoroquinolones with magnesium and calcium ions **93**, 121
Ross, G., see Jahnke, R.W.O. **77**, 47
Ross, M.J., see Moore, J.A. **34**, 35
Ross, S.D., see Leary, J.R. **17**, 193
Rossi, S., see Bonferoni, M.C. **117**, 41
Rossi, S., see Ferrari, F. **117**, 49; **121**, 11; **136**, 71
Rößling, G., see Schneider, T. **117**, 1; **132**, 9
Rößling, G.L., see Zingel, C. **140**, 13
Rostron, C., see Ford, J.L. **71**, 213
Rostron, C., see Mitchell, K. **66**, 233; **100**, 143, 155
Rostron, C., see Perez-Marcos, B. **111**, 251
Roth, H.J., see Muller, C.E. **57**, 41

Roth, J., see Rapaka, R.S. **11**, 237; **12**, 285
Rothlisberger, T., B. Testa, P.A. Carrupt, J.M. Mayer, J.C. Etter, The influence of hydration and structure-promoting effects on the viscosity of binary solvent mixtures **44**, 141
Rothlisberger, T., B. Testa, H. Van de Waterbeemd, P.A. Carrupt, Hydration of substituted benzenes. Experimental studies and relationship with lipophilicity **51**, 73
Rouge, N., P. Buri, E. Doelker, Drug absorption sites in the gastrointestinal tract and dosage forms for site-specific delivery **136**, 117
Rouland, J.C., see Ceolin, R. **74**, 77
Rouland, J.C., see Dubernet, C. **64**, 99
Rousset, M., see Caro, I. **116**, 147
Rowan, M.G., see Kraisintu, K. **10**, 67
Rowan, M.G., see Lacy, J.E. **43**, 111
Rowe, J.S., J.E. Carless, The influence of the microcapsule wall on the assay of indomethacin microcapsules in the presence of antacids—implications for product stability **13**, 313
Rowe, P., see Ford, J.L. **71**, 95
Rowe, P.H., see Nokhodchi, A. **129**, 21
Rowe, R.C., Quantitative opacity measurements on tablet film coatings containing titanium dioxide **22**, 17
The effect of the molecular weight of ethyl cellulose on the drug release properties of mixed films of ethyl cellulose and hydroxypropyl methylcellulose **29**, 37
Tablet-tablet contact and mutual rubbing within a coating drum—An important factor governing the properties and appearance of tablet film coatings **43**, 155
Binder–substrate interactions in granulation: A theoretical approach based on surface free energy and polarity **52**, 149
Interactions in coloured powders and tablet formulations: A theoretical approach based on solubility parameters **53**, 47
Surface free energy and polarity effects in the granulation of a model system **53**, 75
Correlation between predicted binder spreading coefficients and measured granule and tablet properties in the granulation of paracetamol **58**, 209
Polar–non-polar interactions in the granulation of organic substrates with polymer binding agents (Int. J. Pharm. 56 (1988) 117–124) **59**, 83
Calorimetric studies on some nonionic surfactants **79**, 251
Interactions in powders and granules—A reappraisal **79**, 257
Molecular weight dependence of the properties of ethyl cellulose and hydroxypropyl methylcellulose films **88**, 405
Characterization of wet powder masses using a mixer torque rheometer. 4. Effect of blade orientation **133**, 133
Predicting film thickness on film coated tablets **133**, 253
Rowe, R.C., S.F. Forse, Pitting—a defect on film-coated tablets **17**, 347
Rowe, R.C., A.D. Kotaras, E.F.T. White, An evaluation of the plasticizing efficiency of the dialkyl phthalates in ethyl cellulose films using the torsional braid pendulum **22**, 57
Rowe, R.C., A.G. McKillop, D. Bray, The effect of batch and source variation on the crystallinity of microcrystalline cellulose **101**, 169
Rowe, R.C., J. McMahon, R.F. Stewart, The stability of oil-in-water emulsions containing cetrimide and cetostearly alcohol **31**, 281
Rowe, R.C., M.D. Parker, A.G. McKillop, The effect of particle size on the heat of immersion of microcrystalline cellulose **91**, 247
Rowe, R.C., R.J. Roberts, Simulation of crack propagation in tablet film coatings containing pigments **78**, 49
The effect of some formulation variables on crack propagation in pigmented tablet film coatings using computer simulation **86**, 49
Rowe, R.C., N. Washington, The rheology of microcrystalline cellulose powder/water mixes-measurement using a mixer torque rheometer **38**, 227
Rowe, R.C., see Bassam, F. **64**, 55
Rowe, R.C., see Bateman, S.D. **49**, 209
Rowe, R.C., see Davies, M.C. **62**, 97
Rowe, R.C., see Fielden, K.E. **79**, 47; **81**, 205, 225; **97**, 79
Rowe, R.C., see Gibson, S.H.M. **47**, 205; **48**, 63, 113; **50**, 163
Rowe, R.C., see Hancock, B.C. **76**, 239; **83**, 147; **102**, 167; **148**, 1
Rowe, R.C., see Harrison, P.J. **35**, 235
Rowe, R.C., see Landin, M. **104**, 271; **107**, 247; **123**, 143; **133**, 127; **134**, 243
Rowe, R.C., see Louden, J.D. **25**, 179; **63**, 219
Rowe, R.C., see Newton, J.M. **120**, 95, 101
Rowe, R.C., see Parker, M.D. **64**, 207; **72**, 243; **80**, 179
Rowe, R.C., see Patel, H.K. **25**, 13, 237
Rowe, R.C., see Payne, R.S. **145**, 165
Rowe, R.C., see Roberts, R.J. **36**, 205; **37**, 15; **52**, 213; **91**, 173; **99**, 157; **105**, 177; **109**, 83; **125**, 157; **129**, 79
Rowe, R.C., see Sakellariou, P. **27**, 267; **31**, 55, 175; **34**, 93; **102**, 207; **125**, 289
Rowe, R.C., see Tricehurst, M.D. **111**, 241; **141**, 93
Rowe, R.C., see York, P. **66**, 143
Rowland, G.G., see Allsopp, M.A. **69**, 197
Rowley, J., see Adeyeye, C.M. **116**, 65
Rowlings, C.E., D.E. Wurster, P.J. Ramsey, Calorimetric analysis of powder compression. II. The relationship between energy terms measured with a compression calorimeter and tableting behavior **116**, 191
Rowlings, C.E., see Katikaneni, P.R. **117**, 13
Rowlings, C.E., see Sheen, P.C. **118**, 221
Rowlings, C.E., see Upadrashta, S.M. **112**, 173
Rowlings, C.E., see Wurster, D.E. **116**, 179
Roy, A.K., J.K. Guillory, The kinetics and mechanism of the hydrolysis of cyclopentolate hydrochloride in alkaline solutions **120**, 169
The effect of cyclodextrins on the aqueous stability of cyclopentolate hydrochloride **138**, 37
Roy, D., X. Guillon, F. Lescure, P. Couvreur, N. Bru, P. Breton, On shelf stability of freeze-dried: Poly(methylidene malonate 2.1.2) nanoparticles **148**, 165
Roy, M.L., see Pikal, M.J. **60**, 203
Roy, S.D., J.S. De Groot, Percutaneous absorption of nafarelin acetate, an LHRH analog, through human cadaver skin and monkey skin **110**, 137

Royz, M., see Magdassi, S. **88**, 171
Rozier, A., C. Mazuel, J. Grove, B. Plazonnet, Gelrite[R]: a novel, ion-activated, in-situ gelling polymer for ophthalmic vehicles. Effect on bioavailability of timolol **57**, 163
Rozier, A., see Meseguer, G. **95**, 229
Rozier, A., see Thermes, F. **81**, 59
Rubessa, F., see Kristl, A. **100**, 33
Rubessa, F., see Lovrecich, M. **40**, 63; **131**, 247
Rubessa, F., see Moneghini, M. **63**, R1
Rubessa, F., see Mrhar, A. **62**, 15
Rubin, P., see Wilding, I.R. **132**, 81
Rubino, J., see Thomas, E. **130**, 179
Rubino, J.T., Electrostatic and non-electrostatic free energy contributions to acid dissociation constants in cosolvent-water mixtures **42**, 181
Rubino, J.T., L.M. Halterlein, J. Blanchard, The effects of aging on the dissolution of phenytoin sodium capsule formulations **26**, 165
Rubino, J.T., E. Thomas, Influence of solvent composition on the solubilities and solid-state properties of the sodium salts of some drugs **65**, 141
Rubinstein, A., D. Nakar, A. Sintov, Chondrotin sulfate: A potential biodegradable carrier for colon-specific drug delivery **84**, 141
Rubinstein, A., see Bialer, M. **23**, 25
Rubinstein, A., see Liaw, J. **59**, 105
Rubinstein, A., see Pathak, Y.V. **65**, 169
Rubinstein, A., see Sintov, A. **143**, 101
Rubinstein, A., see Touitou, E. **30**, 95
Rubinstein, M.H., I.M. Jackson, Stress relaxation behaviour of compacts of sodium chloride and polyethylene **36**, 99
Rubinstein, M.H., see Bateman, S.D. **49**, 209
Rubinstein, M.H., see Dabbagh, M.A. **140**, 85
Rubinstein, M.H., see Ford, J.L. **8**, 311; **12**, 11; **24**, 327, 339; **40**, 223
Rubinstein, M.H., see Garr, J.S.M. **64**, 223; **72**, 117; **73**, 75; **81**, 187; **82**, 71
Rubinstein, M.H., see Kochhar, S.K. **112**, 225; **115**, 35
Rubinstein, M.H., see Larhrib, H. **147**, 187, 199
Rubinstein, M.H., see Marwaha, S.B. **43**, 249
Rubinstein, M.H., see Nokhodchi, A. **118**, 191; **120**, 13; **126**, 189; **129**, 21
Rubinstein, M.H., see Waring, M.J. **36**, 29; **40**, 15
Rubio, F., see Marino, E.L. **9**, 263
Rubitski, B.A., see Desai, D.S. **91**, 217; **103**, 69; **107**, 141; **110**, 249, 257; **142**, 61
Ruchatz, F., P. Kleinebudde, B.W. Muller, Residual solvents in biodegradable microparticles: Determination by a dynamic headspace gas chromatographic method **142**, 67
Rudant, E., see Bonhomme-Faivre, L. **134**, 99
Ruddy, S.B., see Hadzija, B.W. **67**, 185
Rudnic, E.M., see Chang, R.K. **70**, 261
Rudolph, P., see Michel, C. **98**, 131
Rudraraju, V.S., see Gordon, M.S. **97**, 119
Rudt, S., R.H. Muller, In vitro uptake of polystyrene latex particles and parenteral fat emulsions by human granulocytes **99**, 1
Rue, P.J., see Cutt, T. **33**, 81; **49**, 157
Ruelle, P., M. Buchmann, N.T. Ho, U.W. Kesselring, Comparison of the solubility of polycyclic aromatic hydrocarbons in non-associated and associated solvents: The hydrophobic effect **87**, 47
Ruelle, P., E. Sarraf, U.W. Kesselring, Prediction of carbazole solubility and its dependence upon the solvent nature **104**, 125
Ruff, F., see Efthymiopoulos, C. **95**, 127
Ruhl, D., see Muller, R.H. **144**, 115
Ruiz, J.M., B. Tissier, J.P. Benoit, Microencapsulation of peptide: A study of the phase separation of poly(D,L-lactic acid-*co*-glycolic acid) copolymers 50/50 by silicone oil **49**, 69
Ruizk, S., C. Duru, D. Gaudy, M. Jacob, F. Ferrari, M. Bertoni, C. Caramella, Physico-chemical characterization and tabletting properties of scleroglucan **112**, 125
Ruland, A., J. Kreuter, Transdermal permeability and skin accumulation of amino acids **72**, 149
Influence of various penetration enhancers on the in vitro permeation of amino acids across hairless mouse skin **85**, 7
Ruland, A., J. Kreuter, J.H. Rytting, Transdermal delivery of the tetrapeptide hisetal (melanotropin (6–9)). I. Effect of various penetration enhancers: In vitro study across hairless mouse skin **101**, 57
Ruland, A., J. Kreuter, J.H. Rytting, Transdermal delivery of the tetrapeptide hisetal (melanotropin (6–9)). II. Effect of various penetration enhancers. In vitro study across human skin **103**, 77
Ruland, A., U. Rohr, J. Kreuter, Transdermal delivery of the tetrapeptide Hisetal (melanotropin (6–9)) and amino acids. Their contribution to the elucidation of the existence of an 'acqueous pore' pathway **107**, 23
Runge, S., see Almeida, A.J. **149**, 255
Runge, S.A., see Muller, R.H. **144**, 115
Runge, T.A., see Snider, B.G. **66**, 63
Runkel, R., see Kent, J.S. **7**, 245
Rupena, P., see Moneghini, M. **63**, R1
Rupp, W., see Badian, M. **18**, 9
Rupprecht, H., see Kristl, J. **99**, 13
Rupprecht, H., see Schelenz, G. **113**, 133
Rupprecht, H., see Srcic, S. **73**, 221; **99**, 21
Rush, B.D., K.F. Wilkinson, W.Z. Zhong, S.K. Closson, D.B. Lakings, M.J. Ruwart, Absolute oral bioavailability of ditekiren, a renin inhibitor peptide, in conscious rats **73**, 231
Russell, A.D., Mechanisms of action of chemical sporicidal and sporistatic agents **16**, 127
Russell, A.D., J.R. Furr, The effects of antiseptics, disinfectants and preservatives on smooth, rough and deep rough strains of *Salmonella typhimurium* **34**, 115
Comparative sensitivity of smooth, rough and deep rough strains of *Escherichia coli* to chlorhexidine, quaternary ammonium compounds and dibromopropamidine isethionate **36**, 191
Russell, A.D., J.R. Furr, W.J. Pugh, Susceptibility of porin- and lipopolysaccharide-deficient mutants of *Escherichia coli*

to a homologous series of esters of *p*-hydroxybenzoic acid **27**, 163
Sequential loss of outer membrane lipopolysaccharide and sensitivity of *Escherichia coli* to antibacterial agents **35**, 227
Russell, A.D., B.D. Jones, P. Milburn, Reversal of the inhibition of bacterial spore germination and outgrowth by antibacterial agents **25**, 105
Russell, A.D., see El-Falaha, B.M.A. **23**, 239; **25**, 329
Russell, A.D., see Greenslade, M.D. **26**, 133
Russell, A.D., see Shaker, L.A. **34**, 51
Russell, S.J., see Buckton, G. **73**, 1
Russell-Jones, G.J., see Habberfield, A. **145**, 1
Ruth, H.S., D. Attwood, G. Ktistis, C.J. Taylor, Phase studies and particle size analysis of oil-in-water phospholipid microemulsions **116**, 253
Rutherford, B.S., see Pearlman, R. **18**, 53
Rutt, K.J., see Lloyd, A.W. **131**, 257
Rutten-Kingma, J.J., C.J. de Blaey, J. Polderman, Biopharmaceutical studies of fatty suspension suppositories. II. Influence of particle size and concentration on in vitro release of readily water-soluble compounds **3**, 179
Biopharmaceutical studies of fatty suspension suppositories. III. Influence of particle size and concentration of bioavailability of lithium sulphate in rats **3**, 187
Rutten-Kingma, J.J., J. Polderman, C.J. de Blaey, Biopharmaceutical studies of fatty suspension suppositories. I. Spreading in situ **3**, 39
Ryan, F.M., see Gochoco, C.H. **104**, 187
Ryan, F.M., see Hidalgo, I.J. **98**, 83
Ryden, L., P. Edman, Effect of polymers and microspheres on the nasal absorption of insulin in rats **83**, 1
Ryoo, S.H., see Jang, S.H. **84**, 273
Rytting, J.H., see Acree, Jr. W.E. **8**, 69; **13**, 197
Rytting, J.H., see Anderson, B.D. **1**, 15
Rytting, J.H., see Bhatt, P.P. **72**, 219
Rytting, J.H., see Bhattachar, S.N. **79**, 263
Rytting, J.H., see Buyuktimkin, S. **118**, 113; **127**, 245
Rytting, J.H., see Nishihata, T. **21**, 239
Rytting, J.H., see Ruland, A. **101**, 57; **103**, 77
Rytting, J.H., see Wong, O. **52**, 191
Ryuu, S.A., see Kim, C.K. **147**, 131

S

Saab, A.N., K.B. Sloan, The effect of aminomethyl (N-Mannich base) derivatization on the ability of S^6-pivaloyloxymethyl-6-mercaptopurine prodrug to deliver 6-mercaptopurine through hairless mouse skin **57**, 253
Saarikivi, P., see Lindholm, T. **21**, 119
Saarinen-Savolainen, P., T. Jarvinen, P. Suhonen, A. Urtti, Amphiphilic properties of pilocarpine prodrugs **133**, 171
Saarnivaara, K., see Laine, E. **38**, 33
Sabatier, R., see Delalonde, M. **146**, 159
Sabnis, S.S., see Adeyeye, C.M. **116**, 65
Sabra, K., see Corrigan, O.I. **18**, 195
Sabri, S.M., see Wahbi, A.M. **92**, 15
Sachse, A., see Schneider, T. **117**, 1; **132**, 9
Sachse, A., see Zingel, C. **140**, 13
Sack, R.L., see Lee, B.J. **124**, 119
Sadana, G.S., A.B. Ghogare, Mechanistic studies on photolytic degradation of nifedipine by use of ^1H NMR and ^{13}C NMR spectroscopy **70**, 195
Sadeghnejad, G.R., see Gould, P.L. **38**, 231
Sado, P.A., see Sarciaux, J.M. **120**, 127
Saettone, M.F., P. Chetoni, L.M. Bianchi, B. Giannaccini, U. Conte, M.E. Sangalli, Controlled release of timolol maleate from coated ophthalmic mini-tablets prepared by compression **126**, 79
Saettone, M.F., P. Chetoni, M.T. Torracca, S. Burgalassi, B. Giannaccini, Evaluation of muco-adhesive properties and in vivo activity of ophthalmic vehicles based on hyaluronic acid **51**, 203
Saettone, M.F., B. Giannaccini, P. Chetoni, M.T. Torracca, D. Monti, Evaluation of high- and low-molecular-weight fractions of sodium hyaluronate and an ionic complex as adjuvants for topical ophthalmic vehicles containing pilocarpine **72**, 131
Saettone, M.F., B. Giannaccini, G. Delmonte, V. Campigli, G. Tota, Marca F. La, Solubilization of tropicamide by poloxamers: Physicochemical data and activity data in rabbits and humans **43**, 67
Saettone, M.F., B. Giannacinni, A. Guinducci et al., Polymer effects on ocular bioavailability. II. The influence of benzalkonium chloride on the mydriatic response of tropicamide in different polymeric vehicles **25**, 73
Saettone, M.F., B. Giannaccini, A. Guiducci, P. Savigni, Semisolid ophthalmic vehicles. III. An evaluation of four organic hydrogels containing pilocarpine **31**, 261
Saettone, M.F., B. Giannaccini, S. Ravecca et al., Polymer effects on ocular bioavailability—the influence of different liquid vehicles on the mydriatic response of tropicamide in humans and in rabbits **20**, 187
Saettone, M.F., G. Perini, P. Rijli, L. Rodriguez, M. Cini, Effect of different polymer-plasticizer combinations on 'in vitro' release of theophylline from coated pellets **126**, 83
Saettone, M.F., M.T. Torracca, A. Pagano, B. Giannaccini, L. Rodriguez, M. Cini, Controlled release of pilocarpine from coated polymeric ophthalmic inserts prepared by extrusion **86**, 159
Saettone, M.F., see Burgalassi, S. **133**, 1
Saettone, M.F., see Cavalli, R. **117**, 243
Saettone, M.F., see Chetoni, P. **105**, 147
Saettone, M.F., see Gallarate, M. **100**, 219
Safadi, M., D.S. Bindra, T. Williams, R.C. Moschel, V.J. Stella, Kinetics and mechanism of the acid-catalyzed hydrolysis of O_6-benzylguanine **90**, 239
Safwat, S.M., see Attia, M.A. **47**, 21
Sagara, K., see Masegi, M. **78**, 103
Sagstrom, A., see Alderborn, G. **37**, 155
Sahashi, M., see Bando, H. **135**, 91; **145**, 103
Sahlin, J.J., see Peppas, N.A. **57**, 169
Said, A.M., see Makki, S. **133**, 245
Said, S., H. Al-Shora, Adsorption of certain oral hypoglycaemics on kaolin and charcoal and its relationship to hypoglycaemic effects of drugs **5**, 223

Sustained release from inert matrices. I. Effect of microcrystalline cellulose on aminophylline and theophylline release **6**, 11
Said, S., see Al-Shora, A. **7**, 77
Saidna, M., see Bakhouya, A. **143**, 143; **146**, 225
Saija, A., F. Bonina, D. Trombetta, A. Tomaino, L. Montenegro, P. Smeriglio, F. Castelli, Flavonoid-biomembrane interactions: A calorimetric study on dipalmitoylphosphatidylcholine vesicles **124**, 1
Saija, A., see Bonina, F. **145**, 87
Sailaja, A., see Sastry, C.S.P. **93**, 13
Saint-Germain, J., see Abi, Khalil F. **140**, 1
Sainz, M.C., J.R. Chantres, B. Elorza, M.A. Elorza, DSC study of the action of phenylbutazone on DMPC and DPPC bilayers **89**, 183
DSC study of the action of phenylbutazone on phospholipid phase transitions **91**, 1
Saito, H., see Imai, T. **67**, 11
Saito, K., see Yanai, S. **123**, 237
Saito, Y., see Imai, T. **53**, 7
Saiz, D., see Montero, M.T. **138**, 113
Sakaeda, T., see Takino, T. **114**, 43
Sakane, T., C. Tanaka, A. Yamamoto, M. Hashida, H. Sezaki, H. Ueda, H. Takagi, The effect of polysorbate 80 on brain uptake and analgesic effect of D-kyotorphin **57**, 77
Sakane, T., see Yamamoto, A. **39**, 255; **51**, 137
Sakellariou, P., H. Hassan, R.C. Rowe, Plasticization and interactions of polyethylene glycol 6000 with hydroxypropyl methylcellulose/polyvinyl alcohol blends **102**, 207
Sakellariou, P., R.C. Rowe, The morphology of blends of ethylcellulose with hydroxypropyl methylcellulose as used in film coating **125**, 289
Sakellariou, P., R.C. Rowe, E.F.T. White, The thermomechanical properties and glass transition temperatures of some cellulose derivatives used in film coating **27**, 267
An evaluation of the interaction and plasticizing efficiency of the polyethylene glycols in ethyl cellulose and hydroxypropyl methylcellulose films using the torsional braid pendulum **31**, 55
The solubility parameters of some cellulose derivatives and polyethylene glycols used in tablet film coating **31**, 175
Polymer–polymer interaction in blends of ethyl cellulose with both cellulose derivatives and polyethylene glycol 6000 **34**, 93
Saket, M., see Sallam, E. **67**, 247
Saket, M.M., K.C. James, I.W. Kellaway, Partitioning of some 21-alkyl esters of hydrocortisone and cortisone **21**, 155
The partitioning of some 21-alkyl steroid esters between human stratum corneum and water **27**, 287
Sakiya, Y., S. Ishida, T. Ichikawa, Absorption of benzene derivatives through the body surface and gill membranes of goldfish: Substituent and intramolecular interaction effects **47**, 185
Sakiya, Y., Y. Tsuemura, Y. Sawada et al., Prediction of ftorafur disposition in rats and man by a physiologically based pharmacokinetic model **25**, 347
Sako, K., T. Mizumoto, A. Kajiyama, T. Ohmura, Influence of physical factors in gastrointestinal tract on acetaminophen release from controlled-release tablets in fasted dogs **137**, 225
Sakr, A., see Hutchings, D. **104**, 203
Sakr, A., see Turkoglu, M. **88**, 75; **103**, 115
Sakr, F.M., A new approach for insulin delivery via the pulmonary route: Design and pharmacokinetics in non-diabetic rabbits **86**, 1
The pharmacokinetics of pulmonary nebulized insulin and its effect on glucose tolerance in streptozotocin-induced diabetic rabbits **128**, 215
Nasal administration of glucagon combined with dimethyl-B-cyclodextrin: Comparison of pharmacokinetics and pharmacodynamics of spray and powder formulations **132**, 189
Sakr, F.M., N. Pilpel, The tensile strength and consolidation of lactose coated with non-ionic surfactants. I. Powder **10**, 43
The tensile strength and consolidation of lactose coated with non-ionic surfactants. II. Tablets **10**, 57
Sakuma, S., N. Suzuki, H. Kikuchi, K.I. Hiwatari, K. Arikawa, A. Kishida, M. Akashi, Oral peptide delivery using nanoparticles composed of novel graft copolymers having hydrophobic backbone and hydrophilic branches **149**, 93
Sakurai, H., see Ishimitsu, T. **12**, 271
Salam, M.S., see Gharaibeh, A.E.M. **100**, 253
Salama, R.B., B.M. Ahmed, A.I.H. Omer, A new colorimetric method for the determination of bendrofluazide in pharmaceuctical preparations **17**, 325
Salama, R.B., see A Karim, E.F.I. **142**, 251
Salcedo, J., see Gallardo, V. **71**, 223
Saleh, A.M., A.A. Badwan, Khordagui El, A study of hydrotropic salts, cyclohexanol and water systems **17**, 115
Saleh, A.M., L.K. El-Khordagui, Hydrotropic agents: A new definition **24**, 231
Saleh, A.M., L.K. El-Khordagui, D.A. Robb, A.T. Florence, Effect of some drugs and additives on the cross-linking of bovine serum albumin by glutaraldehyde **57**, 205
Saleh, A.M., S.A. Khalil, L.K. El-Khordagui, Solubility and stability of diazepam in sodium salicylate solution **5**, 161
Saleh, A.M., see Badwan, A.A. **13**, 67
Saleh, A.M., see El-Khordagui, L.K. **7**, 111
Saleh, A.M., see Khalil, S.A. **16**, 271
Saleh, M., see Sheikh, Salem M. **141**, 257
Saleh, S.I., C. Boymond, A. Stamm, Preparation of direct compressible effervescent components: Spray-dried sodium bicarbonate **45**, 19
Saleh, S.I., P. Wehrle, A. Stamm, Improvement of lubrication capacity of sodium benzoate: Effects of milling and spray drying **48**, 149
Saleh, S.I., see Ahmed, S.M. **96**, 5
Saleki-Gerhardt, A., C. Ahlneck, G. Zografi, Assessment of disorder in crystalline solids **101**, 237
Saletti, M.A., see Di Giulio, A. **74**, 183; **89**, 133
Salivirta, J., see Finne, U. **75**, R1

Sallam, E., H. Ibrahim, M. Takieddin, T. Baghal, M. Saket, R. Awad, T. Arafat, Dissolution characteristics of interactive powder mixtures. 4. Effects of additives on the dissolution of griseofulvin from Emcompress carrier **67**, 247
Sallam, E.S., see Ibrahim, H.G. **93**, 111
Sallis, J.D., see Cooper, C.M. **98**, 165
Salminen, L., A. Urtti, L. Periviita, Effect of ocular pigmentation on pilocarpine pharmacology in the rabbit eye. I. Drug distribution and metabolism **18**, 17
Salminen, L., see Urtti, A. **19**, 53; **23**, 147
Salole, E.G., see Al-Dujaili, H. **34**, 67, 75
Salomies, H.E.M., R.M. Heinonen, M.A.I. Toppila, Sorptive loss of diazepam, nitroglycerin and warfarin sodium to polypropylene-lined infusion bags (Softbags) **110**, 197
Salomons-de Vries, M.A., see Bouwstra, J.A. **79**, 141; **84**, 205
Salonen, H., see Sirkia, T. **107**, 179
Salonen, M., see Posti, J. **17**, 225
Salter, J.M., see Zingerman, J.P. **88**, 303
Salvioli, G., see Vandelli, M.A. **118**, 77
Sam, A.P., J.T.M. Van den Heuij, J.J. Tukker, Mucoadhesion of both film-forming and non-film-forming polymeric materials as evaluated with the Wilhelmy plate method **79**, 97
Sam, A.P., see Ijntema, K. **112**, 215
Samadian, T., see Dehpour, A.R. **119**, 133
Samaha, M.W., N.A. El Gindy, The effect of film-inducing substances on the characteristics of binary powder mixtures **19**, 63
Samaha, M.W., V.F. Naggar, Micellar properties of non-ionic surfactants in relation to their solubility parameters **42**, 1
Samaha, M.W., see El Gindy, N.A. **13**, 35
Samaha, M.W., see Gadalla, M.A.F. **38**, 71
Samir Sayed Abu, Zaid, see Morii, M. **17**, 215
Samomons-de, Vries M.A., see Bouwstra, J.A. **144**, 81
Sampath, S.S., K. Garvin, D.H. Robinson, Preparation and characterization of biodegradable poly(L-lactic acid) gentamicin delivery systems **78**, 165
Samsom, M., see De, Vroe C. **65**, 49
Samuel, P.N.K., see Pitt, C.G. **45**, 1
Samuels, G.J., see Ong, J.T.H. **96**, 231
Samyn, C., see Kalala, W. **139**, 187
Samyn, C., see Van den Mooter, G. **87**, 37
Samyn, C., see Van den Mooter, G. **97**, 133
Samyn, C., see Van den Mooter, G. **111**, 127
Samyn, C., W. Kalala, den Mooter Van, R. Kinget, Synthesis and in vitro biodegradation of poly(ether-ester) azo polymers designed for colon targeting **121**, 211
Samyn, N., see Kiekens **146**, 239
Sanathanan, L., see Peck, C.C. **82**, 9
Sanchez, A., J.L. Vila-Jato, M.J. Alonso, Development of biodegradable microspheres and nanospheres for the controlled release of cyclosporin A **99**, 263
Sanchez, B.M., M.M. Cabarga, A.S. Navarro, Hurle A. Dominguez-Gil, A physico-chemical study of the interaction of ciprofloxacin and ofloxacin with polivalent cations **106**, 229
Sanchez Camazano M, M.J. Sanchez, M.T. Vicente, A. Dominguez-Gil, Adsorption of chlorpheniramine maleate by montmorillonite **6**, 243
Sanchez, E., C.M. Evora, M. Llabres, Effect of humidity and packaging on the long-term aging of commercial sustained-release theophylline tablets **83**, 59
Sanchez, J.A., see Lucero, M.J. **104**, 49; **127**, 73
Sanchez, L., S. Torrado, J.L. Lastres, Gelatinized/freeze-dried starch as excipient in sustained release tablets **115**, 201
Sanchez, M.J., see Sanchez, Camazano M. **6**, 243
Sanchez-Soto, P.J., see Gines, J.M. **143**, 247
Sanchika, K., see Katsu, T. **53**, 61
Sanchis-Cortes, A., see Fabra-Campos, S. **109**, 197
Sandal, J.B.P., see Montanari, M.L.C. **85**, 199
Sandberg, A., B. Abrahamsson, J. Sjogren, Influence of dissolution rate on the extent and rate of bioavailability of metoprolol **68**, 167
Sandberg, A., see Abrahamsson, B. **140**, 229
Sandberg, A., see Ragnarsson, G. **79**, 223
Sande, S.A., J. Karlsen, Curve fitting of dissolution data by personal computer **55**, 193
Curve fitting of stability data by personal computer. Software in pharmaceutics. II **73**, 147
Evaluation of reaction order. Software in pharmaceutics. III **98**, 209
Sande, S.A., see Henriksen, I. **119**, 231; **146**, 193
Sandeaux, J., see Raffin, F. **120**, 205
Sanders, L.M., see Gharavi, M. **14**, 333
Sanderson, F.D., see Irwin, W.J. **66**, 193, 243
Sandouk, P., see Dumortier, G. **59**, 1
Saneyoshi, M., see Kawaguchi, T. **58**, R1
Sangalli, M.E., see Colombo, P. **63**, 43
Sangalli, M.E., see Gazzaniga, A. **91**, 167; **108**, 77
Sangalli, M.E., P. Giunchedi, A. Gazzaniga, U. Conte, Erodible perforated coated matrix for extended release of drugs **91**, 151
Sangchantra, R., see Alway, B. **130**, 213
Sangekar, S., W.A. Vadino, I. Chaudry et al., Evaluation of the effect of food and specific gravity of tablets on gastric retention time **35**, 187
Sanghavi, N.M., see Choudhari, K.B. **89**, 207
Sankar, D.G., see Sastry, C.S.P. **39**, 137
Sannan, T., see Ito, R. **61**, 109
Sanniez, W.H.K., see Pilpel, N. **5**, 273
Sansdrap, P., A.J. Moes, Influence of manufacturing parameters on the size characteristics and the release profiles of nifedipine from poly(DL-lactide-*co*-glycolide) microspheres **98**, 157
Santagati, N.A., A. Villari, A. Spadaro, G. Puglisi, Simultaneous determination of antazoline and its degradation product by UV spectrophotometry **65**, 137
Santi, P., P.L. Catellani, P. Colombo, C. Ringard-Lefebvre, C. Barthelemy, A.M. Guyot-Hermann, Partition and transport of verapamil and nicotine through artificial membranes **68**, 43
Santi, P., P.L. Catellani, G. Massimo, G. Zanardi, P. Colombo, Iontophoretic transport of verapamil and melatonin. I. Cellophane membrane as a barrier **92**, 23
Santi, P., see Catellani, P.L. **88**, 285

Santoni, G., A. Tonsini, P. Gratteri, P. Mura, S. Furlanetto, S. Pinzauti, Determination of atropine sulphate and benzalkonium chloride in eye drops by HPLC **93**, 239
Santoni, G., L. Fabbri, P. Gratteri, G. Renzi, S. Pinzauti, Simultaneous determination of aspirin, codeine phosphate and propyphenazone in tablets by reversed-phase high-performance liquid chromatography **80**, 263
Santoni, G., L. Fabbri, P. Mura, G. Renzi, P. Gratteri, S. Pinzauti, Simultaneous determination of otilonium bromide and diazepam by high performance liquid chromatography **71**, 1
Santoni, G., P. Mura, S. Pinzauti, E. Lombardo, P. Gratteri, Simultaneous UV spectrophotometric determination of procaine hydrochloride and phenazone in an otic formulation **64**, 235
Santoni, G., P. Mura, S. Pinzauti, P. Gratteri, Porta E. La, Simultaneous determination of naphazoline and diphenhydramine hydrochlorides in nasal drops by second-order derivative UV spectroscopy **50**, 75
Santoni, G., see Bramanti, G. **49**, 241
Santos Magalhaes, N.S., G. Cave, M. Seiller, S. Benita, The stability and in vitro release kinetics of a clofibride emulsion **76**, 225
Santos Magalhaes, N.S., see Weingarten, C. **75**, 171
Santoyo, S., A. Arellano, P. Ygartua, C. Martin, Penetration enhancer effects on the in vitro percutaneous absorption of piroxicam through rat skin **117**, 219
Santoyo, S., see Arellano, A. **130**, 141
Sanz-Nebot, V., see Barbosa, J. **149**, 213
Sanzgiri, Y.D., E.M. Topp, L. Benedetti, V.J. Stella, Evaluation of mucoadhesive properties of hyaluronic acid benzyl esters **107**, 91
Sanzgiri, Y.D., see Gorecka, B.A. **125**, 55
Sanzgiri, Y.D., see Hume, L.R. **111**, 295
Sarbach, Ch., N. Yagoubi, J. Sauzieres, Ch. Renaux, D. Ferrier, E. Postaire, Migration of impurities from a multilaper plastics container into a parenteral infusion fluid **140**, 169
Sarciaux, J.M., L. Acar, P.A. Sado, Using microemulsion formulations for oral drug delivery of therapeutic peptides **120**, 127
Sarciron, M.E., see Walchshofer, N. **100**, 271
Sarpotdar, P., see Zatz, J.L. **9**, 315
Sarraf, E., see Ruelle, P. **104**, 125
Sasaki, H., Y. Igarashi, K. Nishida, J. Nakamura, Ocular delivery of the β-blocker, tilisolol, through the prodrug approach **93**, 49
Sasaki, H., M. Kojima, Y. Mori, J. Nakamura, J. Shibasaki, Enhancing effect of pyrrolidone derivatives on transdermal drug delivery. I. **44**, 15
Enhancing effect of pyrrolidone derivatives on transdermal drug delivery. II. Effect of application concentration and pre-treatment of enhancer **60**, 177
Sasaki, H., E. Mukai, M. Hashida et al., Development of lipophilic prodrugs of mitomycin C. I. Synthesis and antitumor activity of 1a-*N*-substituted derivatives with aromatic pro-moiety **15**, 49
Development of lipophilic prodrugs of mitomycin C. II. Stability and bioactivation of 1a-*N*-substituted derivatives with aromatic pro-moiety **15**, 61
Sasaki, H., Y. Matsukawa, M. Hashida, H. Sezaki, Characterization of alkylcarbamoyl derivatives of 5-fluorouracil and their application to liposome **36**, 147
Sasaki, H., T. Takahashi, Y. Mori, J. Nakamura, J. Shibasaki, Transdermal delivery of 5-fluorouracil and its alkylcarbamoyl derivatives **60**, 1
Sasaki, H., see Nakamura, J. **87**, 59; **99**, 189
Sasaki, H., see Takeuchi, H. **86**, 25
Sasaki, K., see Sawamoto, T. **146**, 181
Sasaki, Y., see Sugito, K. **47**, 157; **60**, 89
Sastry, C.S.P., S.S. Davis, Reddy M. Narayana, D.G. Sankar, Spectrophotometric determination of menadione and menadione sodium bisulfite in pharmaceutical preparations **39**, 137
Sastry, C.S.P., T. Thirupathi Rao, A. Sailaja, Application of *p*-*N*,*N*-dimethylphenylenediamine dihydrochloride for the determination of some antiarrhythmic agents and anticoagulants **93**, 13
Sathyan, G., W.A. Ritschel, A.S. Hussain, Transdermal delivery of tacrine. I. Identification of a suitable delivery vehicle **114**, 75
Sato, H., see Maitani, Y. **113**, 165
Sato, H., see Miyagawa, Y. **138**, 215
Sato, H., see Miyajima, M. **95**, 153
Sato, H., see Miyamoto, Y. **149**, 25
Sato, K., T. Mine, Analysis of in vitro rat skin permeation and metabolism of SM-10902, prodrug of synthetic prostacyclin analogue **135**, 127
Sato, K., K. Sugibayashi, Y. Morimoto, Effect and mode of action of aliphatic esters on the in vitro skin permeation of nicorandil **43**, 31
Sato, S., S.W. Kim, Macromolecular diffusion through polymer membranes **22**, 229
Satoh, N., see Tashima, T. **42**, 61
Satoh, T., see Imai, T. **53**, 7
Satou, V., J. Chopineau, I. Gremeau, R. Chevrier, F. Bruneaux, Compatibility with medical plastics and stability of continuously and simultaneously infused isosorbide dinitrate and heparin **107**, 111
Sattari, S., see Dwivedi, S.K. **87**, 95
Sau, S.W.J., see Shah, J.C. **112**, 263
Saudemon, P., see Boulenc, X. **123**, 71
Saunders, J., S.S. Davis, C.G. Wilson, A. Smith, Effect of uptake of albumin microspheres on cellular activity of mouse peritoneal macrophages **68**, 265
Saunders, M., see Ferrari, F. **121**, 11
Sautereau, A.M., see Peyrot, M. **102**, 25
Sautou, V., see Levadoux, E. **127**, 255
Sautou-Miranda, V., E. Levadoux, M.T. Groueix, J. Chopineau, Compatibility of propofol diluted in 5% glucose with glass and plastics (polypropylene, polyvinylchloride) containers **130**, 251
Sauvageon-Martre, H., see Cohen, T. **137**, 249
Sauzieres, J., see Sarbach, Ch. **140**, 169

Savastano, L., see Romero, A.J. **99**, 125
Savelyev, V.Y., see Papisov, M.I. **40**, 201
Savigni, P., see Saettone, M.F. **31**, 261
Sawabe, H., see Kurono, Y. **80**, 135
Sawada, M., see Kitagawa, S. **98**, 203
Sawada, M., see Yamauchi, H. **90**, 73
Sawada, Y., see Sakiya, Y. **25**, 347
Sawamoto, T., Y. Kurosaki, K. Sasaki, T. Kimura, T. Nakayama, Biopharmaceutical studies on drug/conjugated-metabolite interactions. II. Effect of acetaminophen sulfate on pharmacokinetics of acetaminophen in rats **146**, 181
Sawamoto, T., see Nakayama, T. **121**, 55
Sawh, D., see Ford, J.L. **71**, 213
Sayalero, M.L., see Lanao, J.M. **86**, 123; **117**, 113
Scabbia, A., see Esposito, E. **142**, 9
Scalia, S., A.S. Tirendi, P. Pazzi, E. Bousquet, Assay of free bile acids in pharmaceutical preparations by HPLC with electrochemical detection **115**, 249
Scalia, S., M. Massaccesi, Simultaneous determination of bendazac and underivatized lysing in ophthalmic preparations by reversed-phase ion-pair high-performance liquid chromatography **82**, 179
Scalia, S., see Giunchedi, P. **130**, 41
Scarnecchia, C., see Cavalli, R. **148**, 47
Scarpellini, A., see Pedersen, M. **90**, 247
Schacht, E., see Morgan, S.M. **122**, 121
Schacht, E., see Vermeersch, H. **60**, 253
Schacht, E.H., see Lehr, C.M. **78**, 43
Schade, A., T. Niwa, H. Takeuchi, T. Hino, Y. Kawashima, Aqueous colloidal polymer dispersions of biodegradable DL-lactide/glycolide copolymer as basis for latex films: A new approach for the development of biodegradable depot systems **117**, 209
Schaefer, H., see Shah, V.P. **74**, 1
Schaefer, T., Melt pelletization in a high shear mixer. VI. Agglomeration of a cohesive powder **132**, 221
Melt pelletization in a high shear mixer. X. Agglomeration of binary mixtures **139**, 149
Schaefer, T., C. Mathiesen, Melt pelletization in a high shear mixer. VII. Effects of product temperature **134**, 105
Melt pelletization in a high shear mixer. VIII. Effects of binder viscosity **139**, 125
Melt pelletization in a high shear mixer. IX. Effects of binder particle size **139**, 139
Schaeffer, H., see Shah, V.P. **82**, 21
Schapoval, E.E.S., see Fratini, L. **127**, 279
Schara, M., see Gabrijelcic, V. **102**, 151
Schara, M., see Kristl, J. **99**, 13
Schaupp, K., see Johnson, O.L. **53**, 237
Scheer, R., see Marz, F. **58**, 155; **61**, 57
Schelenz, G., J. Engel, H. Rupprecht, Sublimation during lyophilization detected by temperature profile and X-ray technique **113**, 133
Schenker, U., see Antonin, K.H. **130**, 33
Schentag, J.J., see Peck, C.C. **82**, 9
Schep, G.P., see Schep, L.J. **105**, 39
Schep, L.J., D.S. Jones, G.P. Schep, M.G. Shepherd, A method to study the adherence of *Candida albicans* to human buccal epithelial cells in vitro using $^{35}SO_4$ radiolabelled blastospores and a Percoll(TM) gradient **105**, 39
Schep, L.J., D.S. Jones, I. Tucker, M.G. Shepherd, A colorimetric ion-pairing analytical method for quaternay ammonium compounds using eosin-Y and Triton X-100 **113**, 263
Schep, L.J., see Jones, D.S. **126**, 281
Scherer, D., J.R. Robinson, J. Kreuter, Influence of enzymes on the stability of polybutylcyanoacrylate nanoparticles **101**, 165
Scherer, P.W., see Waldrep, J.C. **97**, 205
Schilling, R.J., A.K. Mitra, Intestinal mucosal transport of insulin **62**, 53
Schipper, N.G.M., W.A.J.J. Hermens, S.G. Romeyn, J. Verhoef, F.W.H.M. Merkus, Nasal absorption of 17-β-estradiol and progesterone from a dimethyl-cyclodextrin inclusion formulation in rats **64**, 61
Schmalfuss, U., see Bendas, B. **116**, 19
Schmidt, P.C., see Beckert, T.E. **143**, 13
Schmidt, P.C., see Braun, M.A. **135**, 53
Schmidt, P.C., see Karau, C. **128**, 89
Schmidt, P.C., see Maul, K.A. **118**, 103
Schmidtgen, M., see Schneider, T. **132**, 9
Schmitz, L.E., see Welling, P.G. **4**, 135
Schmitz, W., see Wolf, B. **139**, 87
Schneemann, H., see Lippold, B.C. **22**, 31
Schneider, H., see Wolf, B. **139**, 87
Schneider, I.M., B. Dobner, R. Neubert, W. Wohlrab, Evaluation of drug penetration into human skin ex vivo using branched fatty acids and propylene glycol **145**, 187
Schneider, J.A., see Benjamin, E.J. **35**, 73
Schneider, T., A. Sachse, J. Leike, G. Rossling, M. Schmidtgen, M. Drechsler, M. Brandl, Surface modification of continuously extruded contrast-carrying liposomes: Effect on their physical properties **132**, 9
Schneider, T., A. Sachse, G. Rößling, M. Brandl, Generation of contrast-carrying liposomes of defined size with a new continuous high pressure extrusion method **117**, 1
Schnorrenberger, B., see Dongowski, G. **144**, 233
Schnurrer, J., C.M. Lehr, Mucoadhesive properties of the mussel adhesive protein **141**, 251
Schoeffler, P., see Levadoux, E. **127**, 255
Schoen, F.J., see Johnston, T.P. **52**, 139; **59**, 95
Schoen, M.P., B.K. Braxton, L.A. Gatlin, III R.P. Jefferis, A simulation model for the primary drying phase of the freeze-drying cycle **114**, 159
Schoenhard, G.L., see Cook, C.S. **60**, 125
Schoenmaker, R., see Jonkman, J.H.G. **8**, 153; **10**, 177
Schoonen, A.J.M., H. Grijseels, G.W. de Vries-Nijboer, Dissolution of a particle system at the fluid/fluid interface **47**, 249
Schoonen, A.J.M., F. Moolenaar, T. Huizinga, Release of drugs from fatty suppository bases. I. The release mechanism **4**, 141
Schoonen, A.J.M., F. Moolenaar, K.A. Reuvers, T. Huizinga, Release of drugs from fatty suppository bases. II. A rate-limiting interfacial process **7**, 29

Schoonen, A.J.M., G.W. de Vries-Nijboer, Dissolution of acetaminophen in a single pore and its relevance for dissolution of tablets **49**, 261
Schoonen, A.J.M., see Frijlink, H.W. **49**, 91; **64**, 195
Schoonen, A.J.M., see Stuurman-Bieze, A.G.G. **1**, 337
Schoonen, B.J.M., see Moolenaar, F. **45**, 161
Schoutens, A., see Beten, D.B. **123**, 145
Schowen, R.L., see Allsopp, M.A. **69**, 197
Schreiber, B., see Djordjevic, N.M. **81**, 21
Schreier, H., R.M. Abra, J.E. Kaplan, C.A. Hunt, Murine plasma fibronectin depletion after intravenous injection of liposomes **37**, 233
Schreier, H., K.J. McNicol, M. Ausborn, D.M. Soucy, H. Derendorf, A.A. Stecenko, R.J. Gonzalez-Rothi, Pulmonary delivery of amikacin liposomes and acute liposome toxicity in the sheep **87**, 183
Schreier, H., see Gaddipati, N. **128**, 55
Schreier, H., see Wichert, B.V. **78**, 227
Schubert, R., see Hirnle, P. **72**, 259
Schuckler, F., G. Lee, The influence of Azone on monomolecular films of some stratum corneum lipids **70**, 173
Relating the concentration-dependent action of azone and dodecyl-L-pyroglutamate on the structure of excised human stratum corneum to changes in drug diffusivity, partition coefficient and flux **80**, 81
Schulman, S.G., see Lovell, M.W. **11**, 345
Schulman, S.G., see Maruyama, T. **59**, 137
Schulman, S.G., see O'Neal, J.S. **12**, 355
Schultz, R.K., see Randall, C.S. **120**, 235
Schurgers, N., C.J. de Blaey, Effect of pH, buffer concentration and buffer composition on the absorption of theophylline from the small intestine of the rat **19**, 283
Schwartz, J.B., see Herman, J. **42**, 15
Schwartz, K.E., see Gordon, M.S. **115**, 29
Schwartz, M., see Valenta, C. **148**, 131
Schwarz, A., see Krause, H.J. **27**, 145
Schwendener, R.A., R. Wuthrich, S. Duewell, G. Westera, Schulthess Von, Small unilamellar liposomes as magnetic resonance contrast agents loaded with paramagnetic Mn-, Gd- and Fe-DTPA-stearate complexes **49**, 249
Schwinke, D.L., M.G. Ganesan, N.D. Weiner, In situ jejunal uptake from liposomal systems **20**, 119
Scieszka, J.F., T.J. Vidmar, L.C. Haynes, M.J. Cho, Biochemical stability in serum of a lipid-soluble probe molecule entrapped in an o/w emulsion as a carrier for passive drug targeting **45**, 165
Scieszka, J.F., see Cho, M.J. **52**, 79
Sclafani, J., P. Liu, E. Hansen, M.G. Cettina, J. Nightingale, A protocol for the assessment of receiver solution additive-induced skin permeability changes. An example with γ-cyclodextrin **124**, 213
Sclavons, M., see Desager, J.P. **31**, 179
Scott, D.C., R.D. Shah, L.L. Augsburger, A comparative evaluation of the mechanical strength of sealed and unsealed hard gelatin capsules **84**, 49
Scott, E.M., see Gorman, S.P. **4**, 57; **17**, 291
Scott, M.G., see Harron, D.W.G. **57**, 9
Scott, R.C., see Dugard, P.H. **28**, 219
Sebert, P., N. Andrianoff, M. Rollet, Effect of γ irradiation on hydroxypropylmethylcellulose powders: Consequences on physical, rheological and pharmacotechnical properties **99**, 37
Sebert, P., E. Bourny, A. Benghalem, J.M. Vergnaud, M. Rollet, Modelling of release kinetics of drugs from irradiated NaCMC matrix **110**, 285
Modelling of release kinetics of drugs from irradiated NaCMC matrix (Int. J. Pharm. 110 (1994) 285–289) **122**, 129
Sebert, P., E. Bourny, M. Rollet, γ Irradiation of carboxymethylcellulose: Technological and pharmaceutical aspects **106**, 103
Sebhatu, T., C. Ahlneck, G. Alderborn, The effect of moisture content on the compression and bond-formation properties of amorphous lactose particles **146**, 101
Sebhatu, T., M. Angberg, C. Ahlneck, Assessment of the degree of disorder in crystalline solids by isothermal microcalorimetry **104**, 135
Sebhatu, T., see Elamin, A.A. **119**, 25
Sedee, A.G.J., G.M.J.B. Van Henegouwen, H.J.A. Blaauwgeers, Isolation, identification and densitometric determination of norethisterone-4β,5β-epoxide after photochemical decomposition of norethisterone **15**, 149
Sedik, L., see Melendres, J.L. **92**, 243
Seegmuller, E., see Langer, K. **110**, 21
Segura-Bono, M.J., T.M. Garrigues, V. Merino, M.V. Bermejo, Compared effects of synthetic and natural bile acid surfactants on xenobiotic absorption. III. Studies with mixed micelles **107**, 159
Segura-Bono, M.J., see Bermejo, M.V. **69**, 221
Segura-Bono, M.J., see Garrigues, T.M. **101**, 209
Sehgal, P.K., see Vasantha, R. **47**, 95
Seidl, S., see Thiermann, H. **137**, 167
Seijo, B., E. Fattal, L. Roblot-Treupel, P. Couvreur, Design of nanoparticles of less than 50 nm diameter: Preparation, characterization and drug loading **62**, 1
Seijo, B., see Torres, D. **59**, 9; **121**, 239
Seijo, B., see Vila-Jato, J.L. **30**, 229
Seijsener, C.B.J., see Kadir, F. **81**, 193
Seiller, E., see Michel, C. **84**, 93
Seiller, M., see Bonhomme, L. **80**, 271
Seiller, M., see Ferreira, L.A.M. **109**, 251; **121**, 169
Seiller, M., see Mouazen, F. **1**, 275
Seiller, M., see Rabaron, A. **99**, 29
Seiller, M., see Santos Magalhaes, N.S. **76**, 225
Seiller, M., see Tokgoz, N.S. **141**, 27
Seiller, M., see Weingarten, C. **75**, 171
Seki, T., T. Kawaguchi, K. Sugibayashi, K. Juni, Y. Morimoto, Percutaneous absorption of azidothymidine in rats **57**, 73
Seki, T., see Kawaguchi, T. **58**, R1; **77**, 71
Seki, T., see Okumura, K. **88**, 63
Sekine, Y., see Yamaguchi, T. **37**, 127
Sekino, M., see Naito, S. **8**, 253

Sekulovic, D., M. Birmancevic, Investigation of the properties of phenacetin tablets: Dependence on the concentration of binding and disintegration agents **32**, 269
Sekulovic, D., C.A. Gooch, Investigation of the properties of Compactrol tablets relative to the type of disintegrant used **38**, 255
Selk, S., see Bodor, N. **7**, 63
Selkamaa, R., S. Tammilehto, Photochemical decomposition of midazolam. I. Isolation and identification of products **49**, 83
Selkirk, A.B., see Edwards, D. **4**, 21
Selkirk, A.B., see Ojile, J.E. **10**, 99
Selleri, S., see Orzalesi, G. **52**, 225
Seminoff, L.A., J.M. Gleeson, J. Zheng, G.B. Olsen, D. Holmberg, S.F. Mohammad, D. Wilson, S.W. Kim, A self-regulating insulin delivery system. II. In vivo characteristics of a synthetic glycosylated insulin **54**, 251
Seminoff, L.A., G.B. Olsen, S.W. Kim, A self-regulating insulin delivery system. I. Characterization of a synthetic glycosylated insulin derivative **54**, 241
Semla, S., see Qiu, Y. **143**, 179
Sencan, A., see Acarturk, F. **131**, 183
Sencar-Bozic, P., S. Srcic, Z. Knez, Improvement of nifedipine dissolution characteristics using supercritical CO_2 **148**, 123
Sendelbeck, S.L., see Swisher, D.A. **22**, 219
Senderoff, R.I., M. Mahjour, G.W. Radebaugh, Characterization of adsorption behavior by solid dosage form excipients in formulation development acetamide◇ **83**, 65
Senderoff, R.I., T.D. Sokoloski, A practical approach to evaluate the effect of the aqueous stagnant layer on drug release from polymeric devices **91**, 235
Sendo, T., see Urtti, A. **61**, 241
Sener, E., H. Turgut, I. Yalcin, I. Oren, L. Turker, N. Celebi, A. Akin, Structure-activity relationships of some antimicrobial 5-substituted 2-(3-pyridyl)benzoxazoles using quantum-chemical calculations **110**, 109
Sener, E., see Yalcin, I. **98**, 1
Senou, T., see Ishikura, T. **116**, 51
Sentjurc, M., see Gabrijelcic, V. **62**, 75; **102**, 151; **118**, 207
Serafini, M.F., see Buonaguidi, M. **147**, 1
Serafini, M.F., see Carelli, V. **55**, 199; **88**, 89; **89**, 81; **94**, 103
Serafini, M.F., see Di Colo, G. **50**, 27
Serajuddin, A.T.M., see Morris, K.R. **105**, 209; **108**, 195
Serajuddin, T.M., see Rosoff, M. **6**, 137
Sergienko, V.B., see Papisov, M.I. **40**, 201
Serrand, P., see Burgot, G. **63**, 73
Sertin, A., see Bourget, P. **146**, 167
Seta, Y., A.H. Ghanem, W.I. Higuchi, S. Borsadia, C.R. Behl, A.W. Malick, Physical model approach to understanding finite dose transport and uptake of hydrocortisone in hairless guinea-pig skin **81**, 89
Seta, Y., F. Higuchi, Y. Kawahara, K. Nishimura, R. Okada, Design and preparation of captopril sustained-release dosage forms and their biopharmaceutical properties **41**, 245
Seta, Y., F. Higuchi, T. Otsuka, Y. Kawahara, K. Nishimura, R. Okada, H. Koike, Preparation and pharmacological evaluation of captopril sustained-release dosage forms using oily semisolid matrix **41**, 255
Seta, Y., T. Otsuka, H. Tokiwa, H. Naganuma, Y. Kawahara, K. Nishimura, R. Okada, Design of captopril sustained-release preparation with oily semisolid matrix intended for use in human subjects **41**, 263
Setanoians, A., see Azmin, M.N. **10**, 109
Seth, S.C., Jr. L.V. Allen, P. Pinnamaraju, Stability of hydrocortisone salts during iontophoresis **106**, 7
Setti, M., see Ochoa, Machiste E. **126**, 65
Seung Jin Lee, P.J. Stewart, Wan Kim Sung, Ion-paired drug diffusion through polymer membranes **39**, 59
Sevgi, F., see Ozyazici, M. **138**, 25
Sewell, G.J., A.J. Palmer, The chemical and physical stability of three intravenous infusions subjected to frozen storage and microwave thawing **72**, 57
Sewell, G.J., A.J. Palmer, P.J. Tidy, Characterisation of a frozen storage-microwave thawing system for intravenous infusions **70**, 119
Sewell, G.J., see Allsopp, M.A. **69**, 197
Seyda, K., see Brewster, M.E. **68**, 215
Seydel, J.K., see Lehner, S.J. **93**, 201
Seymour, L.W., see Cremers, H.F.M. **110**, 117
Sezaki, H., see Hashida, M. **2**, 245
Sezaki, H., see Masegi, M. **78**, 103
Sezaki, H., see Mukai, E. **25**, 95
Sezaki, H., see Muranishi, S. **2**, 101
Sezaki, H., see Nishikawa, M. **85**, 75
Sezaki, H., see Okamoto, H. **30**, 35
Sezaki, H., see Sakane, T. **57**, 77
Sezaki, H., see Sasaki, H. **36**, 147
Sezaki, H., see Takakura, Y. **37**, 145
Sezaki, H., see Taniguchi, K. **4**, 219
Sezaki, H., see Yamamoto, A. **93**, 91
Sezaki, H., see Yamashita, F. **89**, 199
Sezaki, H., see Yoshikawa, H. **8**, 291; **13**, 321
Sezaki, H., see Yoshioka, T. **8**, 131
Sezakib, H., see Takakura, Y. **105**, 19
Sezer, A.D., J. Akbuga, Controlled release of piroxicam from chitosan-beads **121**, 113
Sforzini, B.V., see Thomsen, K.F. **112**, 143
Shaco-Ezra, N., see Touitou, E. **70**, 159
Shaffer, K.N., see Jones, D.S. **101**, R7
Shah, A.C., see Yamamoto, K. **102**, 133
Shah, D.O., see Miller, II K.J. **98**, 101
Shah, H.S., K. Tojo, Y.W. Chien, Transdermal controlled delivery of verapamil: Characterization of in vitro skin permeation **86**, 167
Shah, J.C., Analysis of permeation data: Evaluation of the lag time method **90**, 161
Shah, J.C., Application of kinetic model toin vitro percutaneous permeation of drugs **133**, 179
Shah, J.C., J.R. Chen, D. Chow, Oral bioavailability and in situ absorption of etoposide in rat **84**, 223
Preformulation study of etoposide. II. Increased solubility and dissolution rate by solid-solid dispersions **113**, 103

Shah, J.C., I. Kaka, S. Tenjarla, S.W.J. Lau, D. Chow, Analysis of percutaneous permeation data. II. Evaluation of the lag time method **109**, 283
Analysis of percutaneous permeation data. II. Evaluation of the lag time method (Int. J. Pharm. 109 (1994) 283–290) **112**, 263
Shah, J.C., see Ramana, V. **145**, 145
Shah, K.P., L. Chafetz, Use of sparingly soluble salts to prepare oral sustained release suspensions **109**, 271
Shah, M.H., see Stewart, J.T. **20**, 65
Shah, N.H., see Felton, L.A. **113**, 17; **127**, 203
Shah, N.H., M.T. Carvajal, C.I. Patel, M.H. Infeld, A.W. Malick, Self-emulsifying drug delivery systems (SEDDS) with polyglycolyzed glycerides for improving in vitro dissolution and oral absorption of lipophilic drugs **106**, 15
Shah, P.K., R.L. Bartel, R.T. Borchardt, Permeability characteristics of cultured mouse keratinocytes compared to hairless mouse skin **68**, 285
Shah, P.K., I.J. Hidalgo, R.T. Borchardt, A simple diffusion device to study transport across cells cultured on microporous membranes **63**, 281
Shah, P.K., see Zheng, H. **129**, 103; **131**, 159
Shah, R., see Buckton, G. **42**, 35
Shah, R.D., see Scott, D.C. **84**, 49
Shah, S., see Pikal, M.J. **17**, 237; **60**, 203; **62**, 165
Shah, V.P., C.R. Behl, G.L. Flynn, W.I. Higuchi, H. Schaeffer, Principles and criteria in the development and optimization of topical therapeutic products **82**, 21
Shah, V.P., J. Elkins, S.Y. Lam, J.P. Skelly, Determination of in vitro drug release from hydrocortisone creams **53**, 53
Shah, V.P., G.L. Flynn, R.H. Guy, H.I. Maibach, H. Schaefer, J.P. Skelly, R.C. Wester, A. Yacobi, In vivo percutaneous penetration/absorption **74**, 1
Shah, V.P., K.K. Midha, S. Dighe, I.J. McGilveray, J.P. Skelly, A. Yacobi, T. Layloff, C.T. Viswanathan, C.E. Cook, R.D. McDowall, K.A. Pittman, S. Spector, Analytical methods validation: Bioavailability, bioequivalence and pharmacokinetics studies **82**, 1
Shah, V.P., A. Noory, C. Noory, B. McCullough, S. Clarke, R. Everett, H. Naviasky, B.N. Srinivasan, D. Fortman, J.P. Skelly, In vitro dissolution of sparingly water-soluble drug dosage forms **125**, 99
Shah, V.P., N.W. Tymes, L.A. Yamamoto, J.P. Skelly, In vitro dissolution profile of transdermal nitroglycerin patches using paddle method **32**, 243
Shah, V.P., see Brain, K.R. **89**, R13
Shah, V.P., see Peck, C.C. **82**, 9
Shah, V.P., see Pershing, L.K. **86**, 201
Shah, V.P., see Prasad, V.K. **13**, 1
Shah, V.P., see Skelly, J.P. **63**, 83
Shaheen, O., S. Othman, I. Jalal, A. Awidi, W. Al-Turk, Comparison of pharmacokinetics and pharmacodynamics of a conventional and a new rapidly dissolving glibenclamide preparation **38**, 123
Shaheen, O., see Othman, S. **41**, 197
Shahjahan, M., R.P. Enever, Solubilities of nitrofurazone and ultraviolet light absorbers in polyethylene glycols and nonionic surfactants **70**, 9
A stability-indicating assay for nitrofurazone by paper chromatography **82**, 215
Some parameters for the solubilization of nitrofurazone and ultraviolet light absorbers by nonionic surfactants **82**, 223
Investigation of the nature of the interaction of nitrofurazone with urea **82**, 229
Photolability of nitrofurazone in aqueous solution. I. Quantum yield studies **143**, 75
Photolability of nitrofurazone in aqueous solution. II. Kinetic studies **143**, 83
Shaker, L.A., A.D. Russell, J.R. Furr, Aspects of the action of chlorhexidine on bacterial spores **34**, 51
Shakhtshneider, T.P., M.A. Vasiltchenko, A.A. Politov, V.V. Boldyrev, The mechanochemical preparation of solid disperse systems of ibuprofen-polyethylene glycol **130**, 25
Shalev, O., see Grinberg, L.N. **132**, 251
Shalmi, M., see Mork, N. **60**, 163
Shamat, M.A., The role of the gastrointestinal microflora in the metabolism of drugs **97**, 1
Shami, E.G., see Aungst, B.J. **38**, 199
Shams-Eldeen, M.A., see Naggar, V.F.B. **28**, 239
Shangraw, R.F., see Pande, G.S. **101**, 71; **124**, 231
Shani, S., M. Friedman, D. Steinberg, Relation between surface activity and antibacterial activity of amine-fluorides **131**, 33
Shankar, M., see Tonnesen, H.H. **38**, 247
Shanker, R.M., I. Ahmed, P.A. Bourassa, K.V. Carola, An in vitro technique for measuring contact angles on the corneal surface and its application to evaluate corneal wetting properties of water soluble polymers **119**, 149
Shankland, K., K.S. Knight, Some observations on the crystal structure of (*R,S*-propranolol hydrochloride **137**, 255
Shankland, K., D.S. Sivia, Unit cell determination of sotalol hydrochloride by powder X-ray diffraction **139**, 9
Shanks, M.L., see Millership, J.S. **28**, 1
Shanks, R.G., see Harron, D.W.G. **12**, 277; **57**, 9
Shanley, D.B., see Collins, A.E.M. **51**, 103
Shao, Z., A.J. Hoffmann, A.K. Mitra, Biodegradation characteristics of acyclovir 2′-esters by respiratory carboxylesterases: Implications in prodrug design for intranasal and pulmonary drug delivery **112**, 181
Shao, Z., see Fort, J.J. **100**, 233
Shao, Z., see Pujara, C.P. **114**, 197
Sharif, S., D.T. O'Hagan, A comparison of alternative methods for the determination of the levels of proteins entrapped in poly(lactide-*co*-glycolide) microparticles **115**, 259
Sharif, S., A.W. Wheeler, D.T. O'Hagan, Biodegradable microparticles as a delivery system for the allergens of *Dermatophagoides pteronyssinus* (house dust mite). I. Preparation and characterization of microparticles **119**, 239
Sharif, S., see O'Hagan, D.T. **108**, 133
Sharkawi, M., see El-Ridy, M. **56**, 23
Sharma, A.K., see Jain, G.K. **130**, 169
Sharma, H., see Ashford, M. **95**, 193
Sharma, H., see Hunter, E. **4**, 175; **17**, 59
Sharma, H., see Kaus, L.C. **22**, 99

Sharma, H.L., see Harris, D. **56**, 97
Sharma, M., see Podczeck, F. **137**, 41
Sharma, P.D., see Wadhwa, L.K. **118**, 31
Sharma, R.K., see Bhise, S.B. **28**, 145
Sharma, S., see Ugwu, S.O. **102**, 193
Shastri, A.S., see Sheorey, D.S. **68**, 19
Shaw, A., see Flinn, N. **138**, 167; **143**, 125
Shaw, P.N., see Nankervis, R. **101**, 249; **119**, 173; **130**, 57
Shawky, Tous S., see Ibrahim, S.A. **61**, 1
Sheen, P.C., V.K. Khetarpal, C.M. Cariola, C.E. Rowlings, Formulation studies of a poorly water-soluble drug in solid dispersions to improve bioavailability **118**, 221
Shefter, E., K.C. Cheng, Drug-polyvinylpyrrolidone (PVP) dispersions. A differential scanning calorimetric study **6**, 179
Shefter, E., R.P. Giannini, Hydrolytic behavior of some aluminum antiperspirant materials in acid **1**, 49
Shefter, E., see Aungst, B.J. **33**, 225; **38**, 199
Shefter, E., see Borka, L. **16**, 93
Shefter, E., see Cheng, K.C. **2**, 81
Shefter, E., see Hussain, M.A. **36**, 127
Sheikh Salem, M., G.K. Pillai, L. Nabulsi, H.N. Al-Kaysi, T.A. Arafat, Malooh A. Abu., M. SalehBadwan A.A., Preparation, characterisation and transformation of terfenadine polymorphic forms **141**, 257
Sheikh Salem, M., see Alkaysi, H.N. **87**, 73
Sheikh Salem, M., see Hassan, M.A. **149**, 227
Sheikh Salem, M., see Sheikh, Salem M. **141**, 257
Sheiner, L.B., see Peck, C.C. **82**, 9
Shek, E., J. Bragonje, E.J. Benjamin et al., A stability indicating high-performance liquid chromatography determination of triple corticoid integrated system in a cream **11**, 257
Shek, P.N., see Barber, R.F. **60**, 219
Shek, P.N., see Jurima-Romet, M. **63**, 227; **88**, 201
Shen, C.C., see Tsai, T.H. **84**, 279
Shen, D.D., see Pollack, G.M. **21**, 3
Sheng, H., P.K. Shah, K.L. Audus, Sucralfate effects on mucus synthesis and secretion by human gastric epithelium in vitro **131**, 159
Shenouda, L.S., K.A. Adams, M.A. Zoglio, A controlled release delivery system using two hydrophilic polymers **61**, 127
Sheorey, D.S., A.S. Shastri, A.K. Dorle, Effect of variables on the preparation of shellac microcapsules by solvent evaporation technique: Part 1 **68**, 19
Shepherd, M.G., see Jones, D.S. **126**, 281
Shepherd, M.G., see Schep, L.J. **105**, 39; **113**, 263
Sher, A., H. Rau, G. Greiner, W. Haubold, Spectroscopic and polarographic investigations of copper(II)–azithromycin interactions under equilibrium conditions **133**, 237
Sher, A., M. Veber, M. Marolt-Gomiscek, Spectroscopic and polarographic investigations: Copper(II)-penicillin derivatives **148**, 191
Sher, A., M. Veber, M.M. Marolt-Gomiscek, S. Gomiscek, Complexation of copper(II) ions with ampicillin. I. Spectroscopic and electrochemical investigation of interactions under equilibrium conditions **90**, 181
The study of complexation of copper(II) ions with ampicillin. II. Dynamics of reactions **99**, 119
Sherertz, E.F., see Sloan, K.B. **44**, 87
Sheridan, P.L., G. Buckton, D.E. Storey, The extent of errors associated with contact angles. II. Factors affecting data obtained using a Wilhelmy plate technique for powders **109**, 155
The use of molecular orbital indices to predict the surface properties of pharmaceutical powders **125**, 141
Sheth, B.B., see Gullapalli, R.P. **140**, 97
Sheth, N.V., D.J. Freeman, W.I. Higuchi, S.L. Spruance, The influence of Azone, propylene glycol and polyethylene glycol on in vitro skin penetration of trifluorothymidine **28**, 201
Sheth, N.V., see Freeman, D.J. **48**, 141
Shetty, B., see Chiang, C.C. **117**, 197
Shetty, B., see Pavliv, L. **105**, 227
Sheu, M.T., H.L. Chou, C.C. Kao, C.H. Liu, T.D. Sokoloski, Dissolution of diclofenac sodium from matrix tablets **85**, 57
Sheu, M.T., T.D. Sokoloski, Entrapment of bioactive compounds within native albumin beads. IV. Characterization of drug release from polydisperse systems **71**, 7
Sheu, M.T., C.M. Yeh, T.D. Sokoloski, Characterization and dissolution of fenofibrate solid dispersion systems **103**, 137
Sheu, M.T., see Ho, H.O. **139**, 223
Shi, F., see Borchard, G. **110**, 29
Shi, W.M., see Liu, J.C. **44**, 197
Shibab, F.A., see Ezzedeen, F.W. **59**, 255
Shibasaki, J., see Sasaki, H. **44**, 15; **60**, 1, 177
Shibata, T., see Natsume, H. **58**, 79
Shichiri, M., see Nishihata, T. **38**, 83
Shihab, F.A., A.R. Ebian, R.M. Mustafa, Effect of polyethylene glycol, sodium lauryl sulfate and polysorbate-80 on the solubility of furosemide **4**, 13
Shihab, F.A., see Ezzedeen, F.W. **28**, 113
Shija, R., V.B. Sunderland, C. McDonald, Alkaline hydrolysis of methyl, ethyl and *n*-propyl 4-hydroxybenzoate esters in the liquid and frozen states **80**, 203
Shim, C.K., B.H. Jung, Inter- and intra-subject variations of multiple saliva peaks of acetaminophen after oral administration of tablets **82**, 233
Noncontribution of enterohepatic recycling to multiple plasma peaks of acetaminophen in rats **83**, 257
Shim, C.K., R. Nishigaki, T. Iga, M. Hanano, Determination of extraction constant, true partition coefficient and formation constant of ion-pair complexes of quaternary ammonium salts, tetrabutylammonium bromide and isopropamide iodide, with some organic anions by a solvent extraction technique **8**, 143
Shim, C.K., S.K. Yun, M.H. Lee, S.K. Kim, Effect of sodium taurodeoxycholate on distribution and elimination of amaranth in rats **56**, 269
Shim, C.K., see Kim, C.K. **99**, 109
Shim, C.K., see Lee, H.J. **121**, 1; **131**, 91
Shim, C.K., see Lee, M.J. **113**, 175
Shim, C.K., see Lee, Y.H. **76**, 71

Shim, C.K., see Young, C.S. **45**, 59
Shim, H.J., E.D. Lee, E.J. Yoon, S.D. Lee, W.B. Kim, J. Yang, M.G. Lee, Pharmacokinetics of FT-ADM after intravenous administration of DA-125, a prodrug of FT-ADM or FT-ADM to rats. A new adriamycin analog containing fluorine **103**, 147
Shim, H.J., see Shim, H.J. **103**, 147
Shim, H.J., see Yoon, E.J. **109**, 181
Shimamori, Y., see Nakano, N.I. **7**, 335
Shimamura, K., see Oguma, T. **63**, 101
Shimazaki, Y., see Hata, T. **110**, 1
Shimizu, T., see Aioi, A. **93**, 1; **113**, 159
Shin, H.J., see Kim, C.K. **124**, 61
Shin, S.C., S.Y. Byun, Controlled release of ethinylestradiol from ethylene-vinyl acetate membrane **137**, 95
Shinkuma, D., T. Hamaguchi, C. Muro et al., Bioavailability of phenytoin from oil suspension and emulsion in dogs **9**, 17
Shinkuma, D., T. Hamaguchi, Y. Yamanaka, N. Mizuno, Correlation between dissolution rate and bioavailability of different commercial mefenamic acid capsules **21**, 187
Shinwari, M.K., see Neau, S.H. **99**, 303
Shiozawa, K., see Fujii, M. **137**, 117
Shiraishi, S., see Imai, T. **67**, 11
Shiu, G.K., see El-Arini, S.K. **55**, 25
Shiu, G.K., see Lootvoet, G. **85**, 113
Shiu, G.K., see Su, S.Y. **67**, 211
Shivanand, P., O.L. Sprockel, Release of propranolol HCl from a tablet coated with a macroporous membranes **92**, 35
Shively, M.L., A.P. Simonelli, The investigation of dual-wavelength spectroscopy for the analysis of dissolved drug in microcapsule suspensions **50**, 39
Shively, M.L., D.C. Thompson, Oral bioavailability of vancomycin solid-state emulsions **117**, 119
Shivers, R.R., see Heiati, H. **146**, 123
Shivji, A.S.H., see Taylor, R.B. **26**, 259
Shoji, H., see Morimoto, Y. **91**, 9
Short, M., see Yuen, K.H. **97**, 61
Short, M.B., see Clarke, G.M. **114**, 1
Short, M.D., see Barker, S.A. **102**, 159
Short, M.D., see Clarke, G.M. **100**, 81
Short, M.D., see Zainudin, B.M.Z. **51**, 67
Short, R.D., see Davies, M.C. **57**, 183; **62**, 97
Shoshan, S., see Magdassi, S. **88**, 171
Shott, M.J., see Matthews, G.P. **50**, 111
Shroot, B., see Martin, B. **49**, 63
Shroot, B., see Rolland, A. **76**, 217
Shtohryn, L., see Pandit, N.K. **50**, 7
Sia, L.L., see Faulkner, R.D. **43**, 53
Sickmueller, A., see Guy, R.H. **19**, 129
Sidahmed, A.M., see McElany, J.C. **26**, 267; **31**, 107
Siddiqui, O., M.S. Roberts, A.E. Polack, Topical absorption of methotrexate: role of dermal transport **27**, 193
Siddiqui, O., see Lelawongs, P. **56**, 13
Siddiqui, O., see Liu, J.C. **44**, 197
Sidhom, M., see Mesiha, M. **114**, 137
Sidhu, D.S., J.K. Sugden, Effect of food dyes on the photostability of aqueous solutions of L-ascorbic acid **83**, 263
Siebert, J.G.H., A.S. Luyt, C. Ackermann, A new transmission electron microscopic (TEM) method to determine differences between cationic polymers in solution **61**, 157
Factors affecting the permeation of urea and water through nude mouse skin in vitro. IV. Polymer carriers **65**, 29
The effect of various polymer applications on the physical structure nude mouse skin **66**, 157
Siekmann, B., see Westesen, K. **93**, 189
Siesler, H.W., see Grunenberg, A. **129**, 147
Sigurdardottir, A.M., see Loftsson, T. **98**, 225
Sigurdsson, P., see Gizurarson, S. **140**, 77; **146**, 133
Siguroardottir, A.M., T. Loftsson, The effect of polyvinylpyrrolidone on cyclodextrin complexation of hydrocortisone and its diffusion through hairless mouse skin **126**, 73
Siguroardottir, A.M., see Loftsson, T. **110**, 169; **115**, 255
Silber, B.M., see Faulkner, R.D. **43**, 53
Silvennoinen, R., P. Raatikainen, J. Ketolainen, P. ketolainen, P. Paronen, Holographic evaluation of bending and integrity of pharmaceutical powder beams **131**, 209
Silvestri, S., G. Gabrielson, Degradation of tragacanth by high shear and turbulent forces during microfluidization **73**, 163
Silvestri, S., G. Gabrielson, L.L. Wu, Effect of terminal block on the microfluidization induced degradation of a model A-B-A block copolymer **71**, 65
Silvestri, S., see Cencia-Rohan, L. **95**, 23
Silvestri, S.L., R.T. Lostritto, Theoretical evaluation of dispersed droplet radii in submicron oil-in-water emulsions **50**, 141
Simamora, P., see Surakitbanharn, Y. **109**, 27
Siman-Tov, A., see Magdassi, S. **59**, 69
Simberg, M., see Sintov, A. **143**, 101
Simeonova, M., T. Ivanova, E. Raikova, M. Georgieva, Z. Raikov, Tissue distribution of polybutylcyanoacrylate nanoparticles carrying spin-labelled nitrosourea **43**, 267
Simeonova, M., see Atcheva, M. **103**, 303
Simmons, J.E., see Storm, J.E. **65**, 265
Simmons, J.E., see Su, S.Y. **67**, 211
Simoes, S., L. Pereira de Almeida, M. Figueiredo, Testing the applicability of classical diffusional models to polydisperse systems **139**, 169
Simoes, S., A. Sousa, M. Figueiredo, Dissolution rate studies of pharmaceutical multisized powders—A practical approach using the Coulter method **127**, 283
Simoes, S., F. Veiga, A. Sousa, M. Figueiredo, Using a particle counter to assess in vitro dissolution studies **131**, 273
Simoes, S., see Lourenco, C. **138**, 1
Simoes, S.I.D., see Martins, M.B.F. **142**, 75
Simon, B.H., H.Y. Ando, P.K. Gupta, Circulation time and body distribution of protein A-coated amino modified polystyrene nanoparticles in mice **121**, 233
Simonelli, A.P., see Shively, M.L. **50**, 39
Simpkins, J., see Kourounakis, A. **141**, 239

Sims, S.M., W.I. Higuchi, V. Srinivasan, Skin alteration and convective solvent flow effects during iontophoresis. I. Neutral solute transport across human skin **69**, 109
Interaction of electric field and electro-osmotic effects in determining iontophoretic enhancement of anions and cations **77**, 107
Sims, E.E., R. Withington, Description and evaluation of a semi-automatic system for particle size analysis by microscopy **17**, 167
Sims, E.E., H.E.C. Worthington, Formulation studies on certain oily injection products **24**, 287
Sims, E.E., see Harnor, K.J. **95**, 111
Sims, E.E., see Wilson, C.G. **72**, 79
Sinchaipanid, N., see Mitrevej, A. **140**, 61
Singh, A., see Baveja, S.K. **19**, 229; **41**, 55
Singh, A., see Singh, M. **76**, R5; **85**, R5
Singh, A.K., see Morris, K.R. **105**, 209; **108**, 195
Singh, K., M. Mezei, Liposomal ophthalmic drug delivery system. I. Triamcinolone acetonide **16**, 339
Liposomal ophthalmic drug delivery system. II. Dihydrostreptomycin sulfate **19**, 263
Singh, M., R. Rathi, A. Singh, J. Heller, G.P. Talwar, J. Kopecek, Controlled release of LHRH-DT from bioerodible hydrogel microspheres **76**, R5
Singh, M., O. Singh, A. Singh, G.P. Talwar, Immunogenicity studies on diphtheria toxoid loaded biodegradable microspheres **85**, R5
Singh, O., see Singh, M. **85**, R5
Singh, P., R.H. Guy, M.S. Roberts, H.I. Maibach, What is the transport-limiting barrier in iontophoresis? **101**, R5
Singh, P., see Patil, S.M. **110**, 147
Singh, R., see Vyas, S.P. **118**, 23
Singh, S., M. Gulati, R.L. Gupta, Complexation behaviour of azathioprine with metal ions **68**, 105
Singh, S., R.L. Gupta, A critical study on degradation of azathioprine in aqueous solutions **42**, 263
Singh, S.K., J. Dodge, M.J. Durrani, M.A. Khan, Optimization and characterization of controlled release pellets coated with an experimental latex. I. Anionic drug **125**, 243
Singh, S.K., I.K. Reddy, M.A. Khan, Optimization and characterization of controlled release pellets coated with an experimental latex. II. Cationic drug **141**, 179
Singh, S.K., M. Suurkuusk, C. Eldsater, Kinetics of decomposition of hydroperoxides formed during the oxidation of soya phosphatidylcholine by analysis of chemiluminescence generated **142**, 215
Singh, S.K., M. Suurkuusk, C. Eldsater, S. Karlsson, A.C. Albertsson, Chemiluminescence is a rapid and sensitive method to assess phosphatidylcholine oxidation **142**, 199
Singhvi, S.M., see Morrison, R.A. **143**, 265
Singla, A.K., D.K. Mediratta, K. Pathak, Bioavailability of indomethacin from zinc-indomethacin complex **60**, 27
Singla, A.K., H. Wadhwa, Zinc-aspirin complex:Synthesis, physicochemical and biological evaluation **108**, 173
Zinc-indomethacin complex: Synthesis, physicochemical and biological evaluation in the rat **120**, 145
Sinko, C.M., D.P. Smith, P.R. Nixon, Mechanical characterization of hydroxypropyl methylcellulose: Modulus determination from indentation loading profiles **81**, 243
Sinko, C.M., G.L. Amidon, Plasticizer-induced changes in the mechanical rate of response of film coatings: An approach to quantitating plasticizer effectiveness **55**, 247
Sinko, C.M., G.T. Carlson, D.S. Gierer, The identification of percolation and mechanical thresholds during the compaction of hydroxypropyl methylcellulose: Comparison to thresholds determined from out-of-die indentation experiments **114**, 85
Sinko, P.J., N.R. Patel, P. Hu, Site-specific oral absorption of didanosine: In situ characterization and correlation with extent of absorption in vivo **109**, 125
Sinko, P.J., see Oh, D.M. **85**, 181
Sintov, A., R.J. Levy, Polymeric drug delivery of enzymatically degradable pendant agents: Peptidyl-linked procainamide model system studies **146**, 55
Sintov, A., M. Simberg, A. Rubinstein, Absorption enhancement of captopril in the rat colon as a putative method for captopril delivery by extended release formulations **143**, 101
Sintov, A., see Rubinstein, A. **84**, 141
Sirkia, T., H. Salonen, P. Veski, H. Jurjenson, M. Marvla, Biopharmaceutical evaluation of new prolonged-release press-coated ibuprofen tablets containing sodium alginate to adjust drug release **107**, 179
Sistat, P., see Raffin, F. **120**, 205
Sitaram, B.R., M. Tsui, H.B. Rawicki, S. Lam, P. Courage, M. Sitaram, C.B. Chapman, Stability and compatibility of baclofen and morphine admixtures for use in an implantable infusion pump **118**, 181
Sitek, S., see Francoeur, M. **16**, 203
Sitek, S.J., see Francoeur, M.L. **25**, 275
Sitges, R., see Montero, M.T. **138**, 113
Siver, K.G., K.B. Sloan, The effect of structure of Mannich base prodrugs of 6-mercaptopurine on their ability to deliver 6-mercaptopurine through hairless mouse skin **48**, 195
Siver, K.G., see Sloan, K.B. **21**, 251
Siver, K.G., see Waranis, R.P. **36**, 211
Sivia, D.S., see Shankland, K. **139**, 9
Sizer, T., see Washington, C. **83**, 227
Sjoberg, H., K. Karami, P. Beronius, L.O. Sundelof, Ionization conditions for iontophoretic drug delivery. A revised pK_a of lidocaine hydrochloride in aqueous solution at 25°C established by precision conductometry **141**, 63
Sjoberg, M., see Lindstedt, B. **67**, 21
Sjodin, L., T. Nederman, M. Prahl, K. Montelius, Radioreceptor assay for formulations of salmon calcitonin **63**, 135
Sjogren, J., see Holzer, A.W. **2**, 145; **7**, 269
Sjogren, J., see Nystrom, C. **10**, 209
Sjogren, J., see Ragnarsson, G. **3**, 127; **12**, 163; **16**, 349; **79**, 223
Sjogren, J., see Sandberg, A. **68**, 167
Sjoholm, I., see Artursson, P. **32**, 165
Sjoholm, I., see Larhammar, H. **23**, 13
Sjoholm, I., see Stjarnkvist, P. **40**, 215

Sjokvist, E., C. Nystrom, Physicochemical aspects of drug release. VI. Drug dissolution rate from solid particulate dispersions and the importance of carrier and drug particle properties **47**, 51
Physicochemical aspects of drug release. XI. Tableting properties of solid dispersions, using xylitol as carrier material **67**, 139
Sjokvist, E., C. Nystrom, M. Alden, Physicochemical aspects of drug release. IX. Investigation of some factors that impair dissolution of drugs from solid particulate dispersion systems **54**, 161
Physicochemical aspects of drug release. XIII. The effect of sodium dodecyl sulphate additions on the structure and dissolution of a drug in solid dispersions **69**, 53
Sjokvist, E., C. Nystrom, M. Alden, N. Caram-Lelham, Physicochemical aspects of drug release. XIV. The effects of some ionic and non-ionic surfactants on properties of a sparingly soluble drug in solid dispersions **79**, 123
Sjokvist, E., see Alden, M. **83**, 47
Sjokvist, Saers E., D.Q.M. Craig, An investigation into the mechanisms of dissolution of alkyl *p*-aminobenzoates from polyethylene glycol solid dispersions **83**, 211
Sjokvist, Saers E., C. Nystrom, M. Alden, Physicochemical aspects of drug release. XVI. The effect of storage on drug dissolution from solid dispersions and the influence of cooling rate and incorporation of surfactant **90**, 105
Sjokvist, Saers E., see Alden, M. **94**, 31
Sjostrom, B., B. Bergenstahl, Preparation of submicron drug particles in lecithin-stabilized o/w emulsions. I. Model studies of the precipitation of cholesteryl acetate **84**, 107
Preparation of submicron drug particles in lecithin-stabilized o/w emulsions. I. Model studies of the precipitation of cholesteryl acetate **88**, 53
Sjostrom, B., K. Westesen, B. Bergenstahl, Preparation of submicron drug particles in lecithin-stabilized o/w emulsions. II. Characterization of cholesteryl acetate particles **94**, 89
Skalko, N., M. Cajkovac, I. Jalsenjak, Liposomes with clindamycin hydrochloride in the therapy of *Acne vulgaris* **85**, 97
Skalko, N., see Filipovic-Grcic, J. **135**, 183
Skauen, D.M., G.M. Zentner, Phonophoresis **20**, 235
Skaugrud, O., see Aspden, T.J. **122**, 69
Skaugrud, O., see Henriksen, I. **98**, 181
Skelley, J.P., see El-Arini, S.K. **55**, 25
Skelly, J.P., G.L. Amidon, W.H. Barr, L.Z. Benet, J.E. Carter, J.R. Robinson, V.P. Shah, A. Yacobi, Report of the workshop on: In vitro and in vivo testing and correlation for oral controlled/modified release dosage forms **63**, 83
Skelly, J.P., see Peck, C.C. **82**, 9
Skelly, J.P., see Shah, V.P. **32**, 243; **53**, 53; **82**, 1; **125**, 99
Skelly, J.P., see Su, S.Y. **67**, 211
Skelton-Stroud, P., see Hastewell, J. **101**, 115
Skerratt, M.C.L., see Washington, C. **77**, 57
Skiba, M., D. Duchene, F. Puisieux, D. Wouessidjewe, Development of a new colloidal drug carrier from chemically-modified cyclodextrins: Nanospheres and influence of physicochemical and technological factors on particle size **129**, 113
Skiba, M., C. Morvan, D. Duchene, F. Puisieux, D. Wouessidjewe, Evaluation of gastrointestinal behaviour in the rat of amphiphilic β-cyclodextrin nanocapsules, loaded with indomethacin **126**, 275
Skiba, M., F. Nemati, F. Puisieux, D. Duchene, D. Wouessidjewe, Spontaneous formation of drug-containing amphiphilic β-cyclodextrin nanocapsules **145**, 241
Skiba, M., F. Puisieux, D. Duchene, D. Wouessidjewe, Direct imaging of modified β-cyclodextrin nanospheres by photon scanning tunnelling and scanning force microscopy **120**, 1
Skiba, M., D. Wouessidjewe, F. Puisieux, D. Duchene, A. Gulik, Characterization of amphiphilic β-cyclodextrin nanospheres **142**, 121
Skinner, M., I. Kanfer, Intrinsic dissolution rate and solubility studies on josamycin, a macrolide antibiotic **88**, 151
Skjetne, T., see Kristensen, S. **100**, 15
Skoog, B., see Larhammar, H. **23**, 13
Skoug, J.W., see Sung, K.C. **142**, 53
Skytte, S., see Pedersen, M. **90**, 247
Slaats, N., see Crommelin, D.J.A. **16**, 79
Slack, J.A., see Baker, N.D. **52**, 231
Slattery, J.T., see Lau, S.W.J. **54**, 171
Slegel, P., see Valko, K. **83**, 123
Sloan, K., see Beall, H. **111**, 223
Sloan, K.B., H.D. Beall, W.R. Weimar, R. Villanueva, The effect of receptor phase composition on the permeability of hairless mouse skin in diffusion cell experiments **73**, 97
Sloan, K.B., N. Bodor, Hydroxymethyl and acyloxymethyl prodrugs of theophylline: Enhanced delivery of polar drugs through skin **12**, 299
Sloan, K.B., J.J. Getz, H.D. Beall, Transdermal delivery of 5-fluorouracil (5-FU) through hairless mouse skin by 1-alkylaminocarbonyl-5-FU prodrugs: Physicochemical characterization of prodrugs and correlations with transdermal delivery **93**, 27
Sloan, K.B., S.A.M. Koch, K.G. Siver, Mannich base derivatives of theophylline and 5-fluorouracil: Syntheses, properties and topical delivery characteristics **21**, 251
Sloan, K.B., E.F. Sherertz, R.G. McTiernan, The effect of structure of Mannich base prodrugs on their ability to deliver theophylline and 5-fluorouracil through hairless mouse skin **44**, 87
Sloan, K.B., see Beall, H.D. **93**, 37; **129**, 203
Sloan, K.B., see Bodor, N. **10**, 307; **15**, 235
Sloan, K.B., see Koch, S.A.M. **35**, 243
Sloan, K.B., see Moelands, D. **86**, 263
Sloan, K.B., see Prankerd, R.J. **88**, 189
Sloan, K.B., see Saab, A.N. **57**, 253
Sloan, K.B., see Siver, K.G. **48**, 195
Sloan, K.B., see Waranis, R.P. **36**, 211
Slok, F., see Nielsen, L.S. **102**, 231
Slop, D., see Bottenberg, P. **49**, 147; **58**, 203
Smail, G.A., see Davidson, A.G. **69**, 1
Smal, J., see Veski, P. **111**, 171

Small, D., M. Dais, M. Wong, D. Tang-Liu, Influence of pH and buffer concentration on the ocular bioavailability of ophthalmic AGN 191103 formulations in albino rabbits **149**, 195
Small, D., M. Dais, H. Zolezio, T. Nguyen, J. Bushong, M. Wong, D. Tang-Liu, Increasing pH of ophthalmic AGN 191103 formulation increases ocular but not systemic bioavailability in albino rabbits **149**, 203
Small, L.E., see Botzolakis, J.E. **12**, 341
Smart, J.D., I.W. Kellaway, Pharmaceutical factors influencing the rate of gastrointestinal transit in an animal model **53**, 79
Smart, J.D., An in vitro assessment of some mucosa-adhesive dosage forms **73**, 69
Smart, J.D., see Henriksen, I. **145**, 231
Smart, J.D., see Mortazavi, S.A. **83**, 221; **94**, 195; **116**, 223
Smedley, S.I., see Friend, D.R. **97**, 39
Smedsrod, B., see Laakso, T. **36**, 253
Smeets, B.P.G.M., see Vendrig, D.E.M.M. **43**, 131
Smets, P., see Fortier, M. **52**, 27
Smeyers-Verbeke, J., see Baert, L. **96**, 225
Smid-Korbar, J., see Kerc, J. **68**, 25; **81**, R1
Smid-Korbar, J., see Kristl, J. **99**, 13
Smid-Korbar, J., see Srcic, S. **87**, 1; **99**, 21
Smid-Korbar, J., see Vrecer, F. **68**, 35
Smirnov, V.N., see Kosykh, V.A. **57**, 223
Smistad, G., see Henriksen, I. **101**, 227; **119**, 231; **145**, 231; **146**, 193
Smistad, G., see Tonneson, H.H. **90**, 221
Smith, A., I.M. Hunneyball, Evaluation of poly(lactic acid) as a biodegradable drug delivery system for parenteral administration **30**, 215
Smith, A., J.F. Lampard, K.M. Carruthers, P. Regan, The filling of molten ibuprofen into hard gelatin capsules **59**, 115
Smith, A., see Saunders, J. **68**, 265
Smith, A., see Woolfrey, S.G. **26**, 35
Smith, B.V., see Beezer, A.E. **38**, 251
Smith, C.M., see Bratty, J.R. **115**, 119
Smith, D., see Dey, M. **49**, 121
Smith, D.P., see Hiestand, E.N. **67**, 231
Smith, D.P., see Sinko, C.M. **81**, 243
Smith, E.W., J.M. Haigh, I. Kanfer, A stability-indicating HPLC assay with on-line clean-up for β methasone 17-valerate in topical dosage forms **27**, 185
Smith, G., see Neil, J.M. **22**, 105
Smith, G.A., see Miller, L.L. **49**, 15
Smith III, W.J., see Kipp, J.E. **74**, 215
Smith, J.C., see Irwin, W.J. **76**, 151
Smith, P.L., J. Marcello, D.C. Chiossone, D. Orner, I.J. Hidalgo, Absorption of an RGD peptide (SK and F 106760) following intratracheal administration in rats amide]◆ **106**, 95
Smith, P.L., see Gochoco, C.H. **104**, 187
Smith, P.L., see Hidalgo, I.J. **98**, 83
Smith, R.B., see Chungi, V.S. **4**, 27
Smith, W.M., see Ackermann, C. **36**, 67
Smyth, W.F., C. Yarnitzky, An analytical study of drug adsorption at mercury electrodes using square wave polarography **42**, 235
Snider, B.G., T.A. Runge, P.E. Fagerness, R.H. Robins, B.D. Kaluzny, Identification of degradation products occurring in acidic solutions of a 21-aminosteroid (tirilazad) methylpregna 1,4,9(11) triene 3,20 dione,B,A **66**, 63
Snyder, G.A., see Jay, M. **14**, 343
Soebagyo, S.S., P.J. Stewart, The characterization of drug redistribution in a ternary interactive mixture of diazepam **91**, 227
Sokoloski, T., see Wu, C.C. **8**, 323
Sokoloski, T.D., C.C. Wu, A.M. Burkman, Rapid adsorptive loss of nitroglycerin from aqueous solution to plastic **6**, 63
Sokoloski, T.D., see Duddu, S.P. **114**, 247
Sokoloski, T.D., see Liu, C.H. **100**, 241
Sokoloski, T.D., see Senderoff, R.I. **91**, 235
Sokoloski, T.D., see Sheu, M.T. **71**, 7; **85**, 57; **103**, 137
Solanki, P., D.J. Morton, Comparative bioavailability of adrenaline-containing ophthalmic solutions: Use of an in vitro method of evaluation **42**, 47
Solans, G., see Garcia-Celma, M.J. **105**, 77
Sole, N., see Alsina, M.A. **70**, 111
Soli, M., see Gatti, R. **115**, 113
Soliman, O.A., K. Kimura, F. Hirayama, K. Uekama, H.M. El-Sabbagh, A.E. El-Gawad, F.M. Hashim, Amorphous spironolactone-hydroxypropylated cyclodextrin complexes with superior dissolution and oral bioavailability **149**, 73
Somaratne, K.D., see Niven, R.W. **101**, 81
Sonanglio, D., B. Bataille, C. Ortigosa, M. Jacob, Factorial design in the feasibility of producing Microcel MC 101 pellets by extrusion/spheronization **115**, 53
Song, H.M., see Lee, K.C. **106**, 167
Song, H.M., see Oh, I.J. **106**, 135
Song, Y., H. Onishi, T. Nagai, Conjugate of mitomycin C with *N*-succinyl-chitosan: In vitro drug release properties, toxicity and antitumor activity **98**, 121
Sontum, P.C., A. Naevestad, A.K. Fahlvik, H.G. Gundersen, Adsorption of poly(sodium(4)styrenesulfonate) on colloidal iron oxide particles **128**, 269
Soozandehfar, S.H., see Lloyd, A.W. **106**, 255
Sorel, R.H.A., H. Roseboom, Rapid in vitro and in vivo conversion of hydroxymethyl-nitrofurantoin into nitrofurantoin as measured by HPLC **3**, 93
Sorensen, A.R., see Bechgaard, E. **128**, 287
Sorgi, F.L., L. Huang, Large scale production of DC-Chol cationic liposomes by microfluidization **144**, 131
Soriano, I., C. Evora, M. Llabres, Preparation and evaluation of insulin-loaded poly(DL-lactide) microspheres using an experimental design **142**, 135
Soriano, I., M. Llabres, C. Evora, Release control of albumin from polylactic acid microspheres **125**, 223
Soti, F., see Pop, E. **140**, 33
Soucy, D.M., see Schreier, H. **87**, 183
Souder, P.R., see Rogers, J.D. **16**, 191
Sousa, A., see Simoes, S. **127**, 283; **131**, 273
Sousa, A., see Sousa, J.J. **144**, 159

Sousa, A., see Veiga, F. **129**, 63
Sousa, A.T., see Pina, M.E. **133**, 139; **148**, 73
Sousa, J.J., A. Sousa, F. Podczeck, J.M. Newton, Influence of process conditions on drug release from pellets **144**, 159
Souter, A., see Hung, C.T. **44**, 117
Southard, M.Z., see Neervannan, S. **96**, 167
Southwell, D., B.W. Barry, Penetration enhancement in human skin: Effect of 2-pyrrolidone, dimethylformamide and increased hydration on finite dose permeation of aspirin and caffeine **22**, 291
Southwell, D., B.W. Barry, R. Woodford, Variations in permeability of human skin within and between specimens **18**, 299
Souto, C., see Cal, S. **129**, 253; **136**, 13
Souto, C., see Doldan, C. **124**, 69
Souto, C., see Landin, M. **91**, 123, 133, 143; **103**, 9; **109**, 1; **123**, 143
Souto, C., see Martinez-Pacheco, R. **32**, 99; **47**, 37
Souto, C., see Mosquera, M.J. **135**, 147
Souto, C., see Perez-Marcos, B. **67**, 113; **100**, 207
Sowell Sr., J.W., see Michniak, B.B. **91**, 85; **116**, 201
Spadaro, A., see Giammona, G. **57**, 55
Spadaro, A., see Santagati, N.A. **65**, 137
Spagnuolo, C.J., see Morris, K.R. **105**, 209
Spalitto, A.J., see Hileman, G.A. **100**, 71
Spampinato, S., see Giammona, G. **127**, 165
Spancake, C.W., A.K. Mitra, D.O. Kildsig, Kinetics of aspirin hydrolysis in aqueous solutions and gels of poloxamines (Tetronic 1508)—Influence of microenvironment **75**, 231
Sparrow, R.A., see Wilding, I.R. **83**, 155; **84**, 59; **101**, 263
Specht, F., see Muller, R.H. **138**, 85
Spector, S., see Shah, V.P. **82**, 1
Speiser, P., see Harmia, T. **33**, 45, 187
Speiser, P.P., see Gipps, E.M. **40**, 23
Spenlehauer, G., see Husson, I. **86**, 113
Spies, F., see Hoogstraate, A.J. **76**, 37
Spies, F., see Jousma, H. **35**, 263
Spiller, R.C., see Watts, P.J. **87**, 215
Spiro, S.G., see Biddiscombe, M.F. **91**, 111
Spittler, J., see Marchal-Heussler, L. **58**, 115
Spohrer, U., see Horst, Thiermann **114**, 125
Spohrer, U., see Thiermann, H. **109**, 35
Sponton, E., D. Drouin, J. Delattre et al., Design of sterile muramyldipeptide-containing oligolamellar liposomes suitable for i.v. injection. Effect of liposome size and lipid composition on their ability to render peritoneal macrophages antitumoral **23**, 299
Spooner, P., see Kanaya, Y. **16**, 171
Spooner, P.J., Y. Kanaya, J.L. Fox, W.I. Higuchi, Novel topical fluoride-delivery system for remineralization of dental enamel: Optimization studies **15**, 177
Spring, E., see Mannermaa, J.P. **103**, 125
Spring, M.S., The effects of some process variables on the concentration of minor components in tablet granules **7**, 261
Spring, M.S., see Cutt, T. **33**, 81; **49**, 157
Spring, M.S., see Femi-Oyewo, M.N. **112**, 17
Sprockel, O.L., see Shivanand, P. **92**, 35
Sprockel, O.L., W. Prapaitrakul, Effect of eluant properties on drug release from cellulose acetate butyrate-coated drug resin complexes **48**, 217
A comparison of microencapsulation by various emulsion techniques **58**, 123
Spruance, S.L., see Freeman, D.J. **48**, 141
Spruance, S.L., see Sheth, N.V. **28**, 201
Srcic, S., J. Kerc, U. Urleb, I. Zupancic, G. Lahajnar, B. Kofler, J. Smid-Korbar, Investigation of felodipine polymorphism and its glassy state **87**, 1
Srcic, S., H. Rupprecht, J. Daca, J. Smid-Korbar, Chemisorbates of *p*-hydroxybenzoic acid acid methyl ester on silica as a new type of pro-drug. IV. Drug release from chemisorbates dispersed in lipophilic vehicles **99**, 21
Srcic, S., H. Rupprecht, A. Mrhar, R. Karba, Chemiadsorbates of *p*-hydroxybenzoic acid methyl ester on silica as a new type of pro-drug. III. Modeling and simulation of drug release from chemiadsorbates to acid aqueous solution **73**, 221
Srcic, S., see Kerc, J. **68**, 25; **81**, R1
Srcic, S., see Kristl, A. **139**, 231
Srcic, S., see Sencar-Bozic, P. **148**, 123
Srcic, S., see Vrecer, F. **68**, 35
Sreejayan, M.N.A. Rao, Curcumin inhibits iron-dependent lipid peroxidation **100**, 93
Sreenivasa, Reddy M., see Subrahmanyam, C.V.S. **78**, 17
Srikrishnan, T., see Cheng, K.C. **2**, 81
Srinivasan, B.N., see Shah, V.P. **125**, 99
Srinivasan, V., W.I. Higuchi, A model for iontophoresis incorporating the effect of convective solvent flow **60**, 133
Srinivasan, V., see Inamori, T. **105**, 113
Srinivasan, V., see Masada, T. **49**, 57
Srinivasan, V., see Peck, K.D. **98**, 141
Srinivasan, V., see Sims, S.M. **69**, 109; **77**, 107
Srivastava, R.C., S.J. Holland, A. Tandon, A.N. Nagappa, Liquid membranes phenomnena in anticancer drugs. Studies on 5-fluorouracil and its derivatives **38**, 239
Srivastava, R.C., see Bhise, S.B. **17**, 263; **21**, 27; **24**, 297; **28**, 145
Srivastava, R.C., see Tandon, A. **32**, 39
Staack, H., see Foster, T.M. **134**, 193
Stabholz, A., see Friedman, M. **44**, 243
Stachowicz, M., see Aignasse, M.F. **113**, 241
Stahl, E., see Huselton, C.A. **134**, 157
Stainmesse, S., see Ezpeleta, I. **131**, 191; **142**, 227
Stalker, D.J., see MacLean, D.S. **131**, 243
Stamm, A., see Brochart, H. **91**, 95
Stamm, A., see Bury, M. **124**, 183
Stamm, A., see Goldbach, P. **117**, 225; **123**, 33
Stamm, A., see Saleh, S.I. **45**, 19; **48**, 149
Stanford, J.B., see Parkinson, H. **51**, 189
Staniforth, J.N., Relationship between vibration produced during powder handling and segregation of pharmaceutical powder mixes **12**, 199
Staniforth, J.N., A.R. Baichwal, J.P. Hart, Interpretation of creep behaviour of microcrystalline cellulose powders and granules during compaction **40**, 267

Staniforth, J.N., S.R. Iveson, An investigation of the use of UV fluorescence microscopy as a method for quantifying the homogeneity of powder mixes **31**, 145
Staniforth, J.N., S.M. Quincey, Granulation monitoring in a planetary mixer using a probe vibration analysis technique **32**, 177
Staniforth, J.N., S. Walker, P. Flanders, Granulation monitoring in a high speed mixer/processor using a probe vibration analysis technique **31**, 277
Staniforth, J.N., see Allwood, M.C. **39**, 183
Staniforth, J.N., see D'Emanuele, A. **92**, 225
Staniforth, J.N., see Grosvenor, M.P. **135**, 103
Staniforth, J.N., see Tobyn, M.J. **128**, 113
Stanislaus, F., see Weiss, G. **124**, 87, 97
Stanley, P., see Carlucci, G. **93**, 245
Stanley, T.H., see Zhang, J. **101**, 15
Stanski, D.R., see Peck, C.C. **82**, 9
Stapleton, K.W., see Finlay, W.H. **149**, 63
Stark, E., see Kendall, A.T. **57**, 217
Stark, G., J.P. Fawcett, I.G. Tucker, I.L. Weatherall, Instrumental evaluation of color of solid dosage forms during stability testing **143**, 93
Stavchansky, S., see Gowan, W.G. **28**, 193
Stavchansky, S., see Reyderman, L. **124**, 75
Stavchansky, S.A., see Gudipati, R.M. **118**, 41
Stead, J.A., M. Freeman, E.G. John et al., Ibuprofen tablets: Dissolution and bioavailability studies **14**, 59
Steadman, B.L., see Burke, C.J. **86**, 89
Stearns, J.F., see Pena, L.E. **113**, 89
Steed, K.P., G. Hooper, J. Brickwell, S.P. Newman, The oropharyngeal and lung deposition patterns of a fusafungine MDI spray delivered by HFA 134a propellant or by CFC 12 propellant **123**, 291
Steed, K.P., G. Hooper, P. Ventura, R. Musa, I.R. Wilding, The in vivo behaviour of a colonic delivery system: A pilot study in man **112**, 199
Steed, K.P., see Watts, P.J. **87**, 215
Steed, K.P., see Wilding, I.R. **101**, 263
Steerenberg, P.A., G. Storm, Groot G. de, J.J. Bergers, A. Claessen, Jong W.H. de, Liposomes as a drug carrier system for *cis*-diamminedichloroplatinum(II). I. Binding capacity, stability and tumor cell growth inhibition in vitro **40**, 51
Stefansson, E., see Loftsson, T. **104**, 181
Steffansen, B., P. Ashton, A. Buur, Intraocular drug delivery. In vitro release studies of 5-fluorouracil from N_1-alkoxycarbonyl prodrugs in silicone oil **132**, 243
Steffansen, B., H. Bundgaard, Erythromycin prodrugs: Kinetics of hydrolysis of erythromycin and various erythromycin 2′-esters in aqueous solution and human plasma **56**, 159
Steffansen, B., see Taub, M.E. **146**, 205
Steger, G.G., see Mader, R.M. **68**, 151
Stein, M., E. Hamacher, Degradation of polybutyl 2-cyanoacrylate microparticles **80**, R11
Steinbach, D., H. Moeller, Investigations into the accuracy of dosage and release of active drug from sustained release preparations of isosorbide dinitrate **1**, 197
Steinbach, D., see Moeller, H. **7**, 157
Steinberg, D., see Shani, S. **131**, 33
Steinijans, V.W., see Jonkman, J.H.G. **43**, 139
Stella, V., P. Vallee, P. Albrecht, E. Postaire, Gliadin films. I. Preparation and in vitro evaluation as a carrier for controlled drug release **121**, 117
Stella, V., see Bundgaard, H. **10**, 181
Stella, V.J., W.K. Anderson, A. Benedetti, W.A. Waugh, Jr. R.B. Killion, Stability of carmethizole hydrochloride (NSC-602668), an experimental cytotoxic agent **71**, 157
Stella, V.J., H.K. Lee, D.O. Thompson, The effect of SBE4-β-CD on i.v. methylprednisolone pharmacokinetics in rats: Comparison to a co-solvent solution and two water-soluble prodrugs **120**, 189
The effect of SBE4-β-CD on i.m. prednisolone pharmacokinetics and tissue damage in rabbits: Comparison to a co-solvent solution and a water-soluble prodrug **120**, 197
Stella, V.J., K. Umprayn, W.N. Waugh, Development of parenteral formulations of experimental cytotoxic agents. I. Rhizoxin (NSC-332598) **43**, 191
Stella, V.J., see Al-Razzak, L.A. **60**, 53
Stella, V.J., see Anderson, B.D. **45**, 27
Stella, V.J., see Antipas, A.S. **109**, 261
Stella, V.J., see Charman, W.N.A. **33**, 155, 165, 173; **34**, 175
Stella, V.J., see Gorecka, B.A. **125**, 55
Stella, V.J., see Kennedy, P.E. **48**, 179
Stella, V.J., see Killion, Jr. R.B. **66**, 149
Stella, V.J., see Muller, D.G. **75**, 201; **86**, 175; **102**, 109
Stella, V.J., see Myers, R.A. **78**, 217; **80**, 51
Stella, V.J., see Neervannan, S. **96**, 167
Stella, V.J., see Noguchi, T. **24**, 173, 185
Stella, V.J., see O'Driscoll, C.M. **73**, 177
Stella, V.J., see Paborji, M. **42**, 243
Stella, V.J., see Phillips, E.M. **94**, 1
Stella, V.J., see Pipkin, J.D. **14**, 263
Stella, V.J., see Pochopin, N.L. **105**, 169; **121**, 157
Stella, V.J., see Prankerd, R.J. **52**, 71
Stella, V.J., see Rajewski, L.G. **82**, 205
Stella, V.J., see Safadi, M. **90**, 239
Stella, V.J., see Sanzgiri, Y.D. **107**, 91
Stella, V.J., see Umprayn, K. **66**, 253
Stella, V.J., see Williams, D.B. **14**, 113
Stelliou, I., G. Venthoye, K.M.G. Taylor, Manipulating the temperature of pentamidine isethionate solutions in jet nebulisers **99**, R1
Stepanek, J., see Hampl, J. **144**, 107
Stephenson, G.A., R.R. Pfeiffer, S.R. Byrn, Solid-state investigation of the tautomerism of acetohexamide **146**, 93
Stephenson, P.J., see Davies, M.C. **66**, 223
Sterling, E.J., see Surendran, N. **135**, 81
Sternagel, H., see Blanque, D. **119**, 203
Sternberg, B.B., see Yoshioka, A.T. **105**, 1
Sternson, L.A., see Kristjansson, F. **41**, 67
Sternson, L.A., see Long, D.F. **6**, 167
Sternson, L.A., see Patton, T.F. **10**, 77
Sternson, L.A., see Repta, A.J. **18**, 277
Stevens, H.N.E., see Farraj, N.F. **43**, 93; **52**, 11, 19
Stevens, J., see Taylor, K.M.G. **58**, 49, 57

Stevens, N.E., see Farraj, N.F. **43**, 83
Stevenson, J.M., see Ip, D.P. **28**, 183
Steward, A., D.L. Bayley, C. Howes, The effect of enhancers on the buccal absorption of hybrid (BDBB) α-interferon **104**, 145
Steward Jr., D.L., see Streng, W.H. **61**, 265
Stewart, A.F., see Ford, J.L. **28**, 11
Stewart, J.T., E.H. Parks, Colorimetric determination of hydralazine hydrochloride with 9-methoxyacridine **17**, 161
Stewart, J.T., M.H. Shah, Amperometric determination of guanethidine sulfate in a flowing stream at the glassy carbon electrode **20**, 65
Stewart, J.T., see Vallner, J.J. **16**, 47
Stewart, P.J., see Alway, B. **130**, 213
Stewart, P.J., see Kulvanich, P. **35**, 111; **36**, 243
Stewart, P.J., see Padmadisastra, Y. **104**, R1; **112**, 55
Stewart, P.J., see Seung Jin Lee **39**, 59
Stewart, P.J., see Soebagyo, S.S. **91**, 227
Stewart, P.J., see Sulihtyowati Soebagyo, S. **25**, 225; **66**, 263
Stewart, R.F., see Patel, H.K. **25**, 13, 237
Stewart, R.F., see Rowe, R.C. **31**, 281
Stieneker, F., see Langer, K. **137**, 67
Stinnett, A.A., see Rajewski, L.G. **82**, 205
Stjarnkvist, P., Biodegradable microspheres. XIV. Effect of microparticle-bound primaquine on *Leishmania donovani* in mice **96**, 23
Stjarnkvist, P., P. Artursson, A. Brunmark, T. Laakso, I. Sjoholm, Biodegradable microspheres. VIII. Killing of *Leishmania donovani* in cultured macrophages by microparticle-bound primaquine **40**, 215
Stobaugh, J.F., see Monnot, E.A. **60**, 41
Stoeva, S., see Kalinkova, G.N. **135**, 111
Stohs, S.J., see Ezzedeen, F.W. **28**, 113; **59**, 255
Stojanoski, K., see Dimitrovska, A. **115**, 175; **134**, 213
Stone, H.W., see Moelands, D. **86**, 263
Stone, H.W., see Prankerd, R.J. **88**, 189
Stor-Pellinen, J., see Ketolainen, J. **125**, 45
Storey, D.E., see Craig, D.Q.M. **96**, 147
Storey, D.E., see Matuszewska, B. **136**, 147
Storey, D.E., see Sheridan, P.L. **109**, 155; **125**, 141
Storm, G., see Bekers, O. **56**, 103
Storm, G., see Janssen, M.J.H. **23**, 1
Storm, G., see Steerenberg, P.A. **40**, 51
Storm, J.E., R.L. Bronaugh, A.S. Carlin, J.E. Simmons, Cutaneous metabolism of nitroglycerin in viable rat skin in vitro **65**, 265
Story, M.J., see Barnwell, S.G. **88**, 423; **89**, 245
Story, M.J., see Cole, S.K. **80**, 63; **88**, 211
Story, M.J., see White, D.G. **69**, 169
Stout, S.A., C.M. Riley, The hydrolysis of L-phenylalanine mustard (melphalan) **24**, 193
Hydrolysis of L-phenylalanine mustard (melphalan). II. Further observations on the effects of pH, chloride ions and buffers on the rate of reaction **37**, 257
Stratford Jr., R.E., V.H.L. Lee, Aminopeptidase activity in homogenates of various absorptive mucosae in the albino rabbit: Implications in peptide delivery **30**, 73
Stratford Jr., R.E., D.C. Yang, M.A. Redell, V.H.L. Lee, Effects of topically applied liposomes on disposition of epinephrine and inulin in the albino rabbit eye **13**, 263
Stratford Jr., R.E., see Kuhfeld, M.T. **133**, 47
Stratford Jr., R.E., see Lee, V.H.L. **13**, 183
Straub, L.E., see Wichert, B.V. **78**, 227
Streng, W.H., D.H.S. Yu, C. Zhu, Determination of solution aggregation using solubility, conductivity, calorimetry, and pH measurements **135**, 43
Streng, W.H., H.G.H. Tan, General treatment of pH solubility profiles of weak acids and bases. II. Evaluation of thermodynamic parameters from the temperature dependence of solubility profiles applied to a zwitterionic compound **25**, 135
Streng, W.H., Jr. D.L. Steward, Ionization constant of an amino acids as a function of temperature **61**, 265
Streng, W.H., see Zhu, C. **130**, 159
Strickley, R., see Brandl, M. **93**, 75
Strickley, R.G., see Gu, L. **45**, 129; **60**, 99
Strid, S., see Gjellan, K. **102**, 213
Strittmatter, T., see Ali, S.L. **1**, 185; **4**, 111
Strittmatter, Th., see Ali, S.L. **6**, 183
Strohbeck, L., see Won, C.M. **121**, 95
Strolin-Benedetti, M., see Efthymiopoulos, C. **95**, 127
Strom, F., see Thomsen, K.F. **112**, 143
Strom, P., see Juppo, A.M. **88**, 141
Strom, R., see Di Giulio, A. **89**, 133
Struc, E., see Kristl, J. **99**, 13
Struijke, A., see Grit, M. **50**, 1
Strykowski, J.M., see Pandit, N.K. **50**, 7
Stuart, J.F.B., see Halbert, G.W. **21**, 219
Stubberud, L., H.G. Arwidsson, C. Graffner, Water–solid interactions. I. A technique for studying moisture sorption/desorption **114**, 55
Stubberud, L., H.G. Arwidsson, A. Larsson, C. Graffner, Water–solid interactions. II. Effect of moisture sorption and glass transition temperature on compactibility of microcrystalline cellulose alone or in binary mixtures with polyvinyl pyrrolidone **134**, 79
Stubbs, C., see Tandt, L.A.G.L. **104**, 11
Stubbs, C., I. Kanfer, A stability-indicating high-performance liquid chromatographic assay of erythromycin estolate in pharmaceutical dosage forms **63**, 113
Stupar, M., see Vulovic, N. **55**, 123
Sturesson, C., J. Carlfors, K. Edsman, M. Andersson, Preparation of biodegradable poly(lactic-*co*-glycolic) acid microspheres and their in vitro release of timolol maleate **89**, 235
Sturesson, C., see Ghaderi, R. **141**, 205
Stuurman-Bieze, A.G.G., F. Moolenaar, A.J.M. Schoonen et al., Biopharmaceutics of rectal administration of drugs in man. II. Effect of particle size on absorption rate and bioavailability **1**, 337
Stuurman-Bieze, A.G.G., see Moolenaar, F. **1**, 323
Su, H.L., see Ho, H.O. **139**, 223
Su, M.H., see Lee, P.H. **93**, 139

Su, S.Y., G.K. Shiu, J.E. Simmons, J.P. Skelly, Some considerations on the analytical method for dissolution of conjugated estrogen tablets **67**, 211
Su, T.J., see Hu, O.Y.P. **53**, 85
Su, X.Y., C. Mattern, R. Hacker, A. Li Wan Po, Does sea-water made isotonic affect ciliary beat frequency? **123**, 47
Subr, V., see Morgan, S.M. **128**, 99
Subrahmanyam, C.V.S., Reddy M. Sreenivasa, Rao J. Venkata, Rao P. Gundu, Irregular solution behaviour of paracetamol in binary solvents **78**, 17
Subrahmanyam, C.V.S., see Bhise, S.B. **17**, 263; **21**, 27; **24**, 297; **28**, 145
Sucker, H., see Lerk, P.C. **92**, 197, 203
Suda, M., see Watanabe, Y. **99**, 337
Suda, Y., see Yamashita, S. **108**, 241
Sudho, M., see Nishihata, T. **33**, 181
Sudoh, M., see Ahmed, I. **38**, 9
Suedee, R., see Heard, C.M. **139**, 15
Sueishi, T., see Tamura, M. **131**, 263
Sueliman, M.S., see Hassan, M.A. **149**, 227
Suen, K.O., see Lee, K.C. **130**, 103
Suenaga, A., O. Bekers, J.H. Beijnen, W.J.M. Underberg, T. Tanimoto, K. Koizumi, M. Otagari, The effect of heptakis (2,6-di-*O*-methyl)-γ-cyclodextrin on mitomycin C stability in aqueous solution **59**, 121
Stabilization of daunorubicin and 4-demethoxydaunorubicin on complexation with octakis(2,6-di-*O*-methyl)-γ-cyclodextrin in acidic aqueous solution **82**, 29
Suenaga, A., see Bekers, O. **72**, 123
Sugawara, I., see Inoue, N. **137**, 75
Sugden, J.K., see Allman, J. **115**, 241
Sugden, J.K., see Barnes, A.R. **44**, 31
Sugden, J.K., see Billany, M.R. **121**, 107; **124**, 279; **137**, 143
Sugden, J.K., see Ho, A.H.L. **104**, 247; **107**, 199
Sugden, J.K., see Kendall, A.T. **57**, 217
Sugden, J.K., see Parkash, A.K. **92**, 151
Sugden, J.K., see Parkinson, H. **51**, 189
Sugden, J.K., see Sidhu, D.S. **83**, 263
Sugden, K., see Hessell, P.G. **52**, 159
Sugibayashi, K., D. Kobayashi, E. Nakagaki, T. Hatanaka, N. Inoue, S. Kusumi, M. Kobayashi, M. Kimura Y. Morimoto, Differences in enhancing effect of 1-menthol, ethanol and their combination between hairless rat and human skin **113**, 189
Sugibayashi, K., see Hatanaka, T. **79**, 21
Sugibayashi, K., see Inoue, N. **137**, 75
Sugibayashi, K., see Kubo, H. **103**, 27
Sugibayashi, K., see Manabe, E. **129**, 211
Sugibayashi, K., see Morimoto, Y. **32**, 31; **54**, 27; **91**, 9
Sugibayashi, K., see Nakamura, H. **141**, 71
Sugibayashi, K., see Natsume, H. **58**, 79
Sugibayashi, K., see Sato, K. **43**, 31
Sugibayashi, K., see Seki, T. **57**, 73
Sugibayashi, K., see Sugibayashi, K. **113**, 189
Sugibayashi, K., see Tamura, M. **131**, 263
Sugibayashi, K., see Ueda, H. **137**, 217; **143**, 37
Sugibayashi, K., see Yamaguchi, H. **58**, 135
Sugihara, M., see Watanabe, A. **117**, 23
Sugimori, K., see Kakee, A. **111**, 15
Sugimoto, I., see Akimoto, K. **3**, 109
Sugimoto, I., see Matsuda, Y. **54**, 211
Suginaka, A., see Unezaki, S. **126**, 41
Sugita, E.T., see Maderich, A.B. **137**, 85
Sugito, K., H. Ogata, H. Goto, M. Noguchi, T. Kogure, M. Takano, Y. Maruyama, Y. Sasaki, Gastrointestinal transit of non-disintegrating solid formulations in humans **60**, 89
Sugito, K., H. Ogata, M. Noguchi, T. Kogure, M. Takano, Y. Maruyama, Y. Sasaki, The spreading of radiolabelled fatty suppository bases in the human rectum **47**, 157
Suh, H., H.W. Jun, Physicochemical and release studies of naproxen in poloxamer gels **129**, 13
Suhonen, P., T. Jarvinen, K. Lehmussaari, T. Reunamaki, A. Urtti, Rate control of ocular pilocarpine delivery with bispilocarpic acid diesters **127**, 85
Suhonen, P., T. Jarvinen, P. Peura, A. Urtti, Permeability of pilocarpic acid diesters across albino rabbit cornea in vitro **74**, 221
Suhonen, P., see Jarho, P. **137**, 209
Suhonen, P., see Jarvinen, T. **75**, 249, 259; **79**, 233, 243
Suhonen, P., see Saarinen-Savolainen, P. **133**, 171
Sujja-Areevath, J., D.L. Munday, P.J. Cox, K.A. Khan, Release characteristics of diclofenac sodium from encapsulated natural gum mini-matrix formulations **139**, 53
Suleiman, M., see Najib, N.M. **32**, 229
Suleiman, M.S., M.E. Abdulhameed, N.M. Najib, H.Y. Muti, Effect of ultraviolet radiation on the stability of diltiazem **50**, 71
Suleiman, M.S., N.M. Najib, Isolation and physicochemical characterization of solid forms of glibenclamide **50**, 103
Suleiman, M.S., N.M. Najib, M.A. Hassan, M.E. Abdel-Hamid, Kinetics of the acid-catalyzed hydrolysis of famotidine **54**, 65
Suleiman, M.S., see Hassan, M.A. **67**, 131; **58**, 19
Suleiman, M.S., see Najib, N.M. **24**, 165; **45**, 139; **51**, 225; **57**, 197; **61**, 173
Suleiman, M.S., see Tabba, H.D. **54**, 57
Suleimans, M.S., see Najib, N.M. **59**, 83
Sulihtyowati, Soebagyo S, P.J. Stewart, The effect of cohesive and non-cohesive ternary components on the homogeneity and stability of a prednisolone interactive mixture **25**, 225
Factors influencing the homogeneity and drug redistribution of disintegrant-diazepam-carrier ternary mixtures **66**, 263
Sumi, M., see Yamashita, S. **108**, 241
Summers, M.P., see Hilton, J.E. **31**, 157; **32**, 13; **33**, 219
Summers, R.S., B. Summers, S. Rawnsley, The bioavailability, absorption characteristics and formulation of four commercially-available controlled-release theophylline products **30**, 83
Summers, R.S., see Bluhm, D.P. **68**, 277, 281
Sun, C., see Aoki, H. **115**, 183
Sun, Y., see Liu, J.C. **25**, 265; **44**, 197
Sundaresan, G.M., see Goehl, T.J. **11**, 181; **15**, 115
Sundelof, L.O., see Caram-Lelham, N. **115**, 103

Sundelof, L.O., see Orbe, A. **41**, 49
Sundelof, L.O., see Sjoberg, H. **141**, 63
Sundelof, L.O., see Nicklasson, M. **15**, 87; **23**, 97
Sunderland, V.B., D.W. Watts, Kinetics of the degradation of methyl, ethyl and *n*-propyl 4-hydroxybenzoate esters in aqueous solution **19**, 1
Alkaline ethanolysis of methyl 4-hydroxybenzoate and hydrolysis of methyl and ethyl 4-hydroxybenzoates in ethanol-water systems **27**, 1
Sunderland, V.B., see Shija, R. **80**, 203
Sundholm, F., see Hyppola, R. **133**, 161
Sung, K.C., P.R. Nixon, J.W. Skoug, T.R. Ju, P. Gao, E.M. Topp, M.V. Patel, Effect of formulation variables on drug and polymer release from HPMC-based matrix tablets **142**, 53
Sung Wan Kim, see Seung Jin Lee **39**, 59
Supico, A., see Martins, M.B.F. **142**, 75
Surakitbanharn, Y., P. Simamora, G.H. Ward, S.H. Yalkowsky, Precipitation of pH solubilized phenytoin **109**, 27
Surendran, N., R. Bhardwaj, S.O. Ugwu, E.J. Sterling, J. Blanchard, Partitioning properties and degradation kinetics of the [Nle^4- DPhe^7]α-MSH analog Melanotan-I (MT-I) **135**, 81
Suriol, M., see Calpena, A.C. **103**, 179
Suryanarayanan, R., C.S. Herman, Quantitative analysis of the active ingredient in a multi-component tablet formulation by powder X-ray diffractometry **77**, 287
Suryanarayanan, R., A.G. Mitchell, Evaluation of two concepts of crystallinity using calcium gluceptate as a model compound **24**, 1
Phase transitions of calcium gluceptate **32**, 213
Suryanarayanan, R., S. Venkatesh, L. Hodgin, P. Hanson, Determination of the physical state of salicylic acid in hydrogel formulations by X-ray diffractometry **78**, 77
Sustar, B., see Kristl, A. **139**, 231
Sutananta, W., D.Q.M. Craig, R.M. Hill, J.M. Newton, The use of low frequency dielectric spectroscopy as a novel means of investigating the structure of pharmaceutical glyceride bases **125**, 123
Sutananta, W., D.Q.M. Craig, J.M. Newton, An investigation into the effect of preparation conditions on the structure and mechanical properties of pharmaceutical glyceride bases **110**, 75
The effects of ageing on the thermal behaviour and mechanical properties of pharmaceutical glycerides **111**, 51
The use of dielectric analysis as a means of characterising the effects of moisture uptake by pharmaceutical glyceride bases **132**, 1
Sutinen, R., O. Bilbao-Revoredo, A. Urtti, P. Paronen, Effect of sodium chloride on the mechanical and osmotic properties of silicone matrices **57**, 155
Sutinen, R., A. Kovanen, A. Urtti, P. Paronen, Diffusion kinetics of alkyl *p*-aminobenzoates in silicone polymers and their release from silicone reservoir devices **57**, 149
Sutinen, R., A. Urtii, P. Raatikainen, P. Paronen, Dissolution medium independent release of propranolol from silicone reservoir devices **92**, 177
Sutinen, R., A. Urtti, R. Miettunen, P. Paronen, Water-activated and pH-controlled release of weak bases from silicone reservoir devices **62**, 113
Sutinen, R., see Hirvonen, J. **99**, 253
Sutter, B.K., see Lippold, B.H. **54**, 15
Sutton, L.E., see Grynpas, R.S.O. **22**, 357
Suurkuusk, M., see Singh, S.K. **142**, 199, 215
Suwa, T., see Akimoto, M. **100**, 133; **117**, 31
Suwa, T., see Yamada, K. **119**, 1
Suzuki, H., see Fujii, M. **137**, 117
Suzuki, K., see Asano, M. **67**, 67
Suzuki, K., see Swarbrick, J. **32**, 21
Suzuki, N., see Sakuma, S. **149**, 93
Suzuki, T., see Nakamura, Y. **145**, 29
Sveinsson, S.J., W.P. Holbrook, Oral mucosal adhesive ointment containing liposomal corticosteroid **95**, 105
Sveinsson, S.J., T. Kristmundsdottir, Naproxen microcapsules: Preparation and in vitro characterization **82**, 129
Sveinsson, S.J., T. Kristmundsdottir, K. Ingvarsdottir, The effect of tableting on the release characteristics of naproxen and ibuprofen microcapsules **92**, 29
Sveinsson, S.J., see Arnardottir, H.B. **117**, 237; **134**, 71
Svenheden, A., see Abrahamsson, B. **140**, 229
Svensson, C.K., Effect of single and short-term multiple exposure to honeybee venom on the disposition of antipyrine in the rat **37**, 163
Sviridov, D.D., M.Y. Pavlov, V.S. Repin, Effect of the inhibitors of different steps of cholesterol biosynthesis pathway on high density lipoprotein binding to isolated human enterocytes **96**, 1
Swanson, J.R., see Rocco, W.L. **117**, 231
Swanson, P.D., F.J. Muzzio, A. Annapragada, A. Adjei, Numerical analysis of motion and deposition of particles in cascade impactors **142**, 33
Swarbrick, J., S. Prakongpan, K. Suzuki, Sustained release delivery systems. III. Preparation and in vitro dissolution of surface-reacted dapsone particles **32**, 21
Swarbrick, J., see Chattaraj, S.C. **120**, 119
Swarbrick, J., see Lee, G. **23**, 43
Swarbrick, J., see Lee, V.H.L. **16**, 163
Swart, P.J., F.A.M. Toulouse, R.A. de Zeeuw, The influence of azone on the transdermal penetration of the dopamine D_2 agonist N-0923 in freely moving rats **88**, 165
Swart, P.J., W.L. Weide, R.A. de Zeeuw, In vitro penetration of the dopamine D_2 agonist N-0923 with and without Azone **87**, 67
Swisher, D.A., S.L. Sendelbeck, J.W. Fara, Adherence of various oral dosage forms to the esophagus **22**, 219
Symillides, M., see Macheras, P. **52**, 249
Szasz, G., see Takacs-Novak, K. **79**, 89; **89**, 261; **113**, 47
Szefler, S.J., see Pollack, G.M. **21**, 3
Szeitz, A., see Chow, A.H.L. **126**, 11
Szejtli, J., see Csabai, K. **91**, 15
Szejtli, J., see Cserhati, T. **141**, 1
Szeptycki, P., see Bhatt, P.P. **50**, 197
Sznitowska, M., The influence of ethanol on permeation behaviour of the porous pathway in the stratum corneum **137**, 137

Sznitowska, M., B. Berner, H.I. Maibach, In vitro permeation of human skin by multipolar ions **99**, 43
Percutaneous penetration of multipolar ions: Evidence for porous transport **123**, 41
Sznitowska, M., S. Janicki, T. Gos, The effect of sorption promoters on percutaneous permeation of a model zwitterion—Baclofen **137**, 125
Sznitowska, M., see Kubota, K. **96**, 105
Szogyi, M., see Cserhati, T. **61**, 189
Szpunar, G.J., see Chiang, C.M. **49**, 109; **50**, 21; **55**, 229
Szwarocka, A., T. Robak, E. Krykowski, Z. Jozwiak, Interaction of anthracyclines with human erythrocytes at hyperthermic temperature **135**, 167

T

Tabata, Y., see Yamaoka, T. **113**, 149
Tabatabay, C., see Bernatchez, S.F. **106**, 161
Tabba, H.D., M.E. Abdel-Hamid, M.S. Suleiman, M.M. Al-Arab, M.M. Hasan, S. Abu-Lafi, N.M. Najib, Synthesis, identification and preliminary evaluation of esters and amide derivatives of diflunisal **54**, 57
Tabbara, K.F., see El-Sayed, Y.M. **121**, 181
Tabernero, J.M., see Marino, E.L. **9**, 263
Tabibi, S.E., R.G. Hollenbeck, Interaction of water vapor and compressible sugar **18**, 169
Tabosa de Egito, E.S., H. Fessi, M. Appel, G. Barratt, P. Legrand, J. Bolard, J.Ph. Devissaguet, A morphological study of an amphotericin B emulsion-based delivery system **145**, 17
Tabosa de Egito, E.S., see Tabosa, de Egito E. **145**, 17
Tabosa de Egito, E.S., see Yu, W. **89**, 139
Tada, N., see Yoshimura, Y. **103**, 1
Tagami, C., see Nakamura, J. **99**, 189
Taggart, A.J., see McElnay, J.C. **33**, 195
Tahara, K., M. Mikawa, S. Yokohama, T. Nishihata, Characteristics on intestinal absorption of adenazolam and in vivo evaluation of oral sustained release tablets of adinazolam in beagle dogs **99**, 311
Tahara, K., K. Yamasmoto, T. Nishihata, Application of model-independent and model analysis for the investigation of effect of drug solubility on its release rate from hydroxypropyl methylcellulose sustained release tablets **133**, 17
Tait, C.J., see Attwood, D. **26**, 25
Tajana, A., see Dal Pozzo, A. **50**, 97
Tak, K.K., see Ha, I.S. **144**, 91
Takacs, M., see Volford, O. **137**, 111
Takacs-Novak, K., M. Jozan, I. Hermecz, G. Szasz, Lipophilicity of antibacterial fluoroquinolones **79**, 89
Takacs-Novak, K., M. Jozan, G. Szasz, Lipophilicity of amphoteric molecules expressed by the true partition coefficient **113**, 47
Takacs-Novak, K., B. Noszal, M. Tokes-Kovesdi, G. Szasz, Acid-base properties and proton-speciation of vancomycin **89**, 261
Takada, K., Y. Furuya, H. Yoshikawa, S. Muranishi, T. Yasumura, T. Oka, Increased systemic availability of cyclosporin A by formulation design: Pharmacokinetic consideration on its transport **44**, 107
Takada, K., Y. Nakahigashi, T. Tanaka, S.I. Kishimoto, Y. Ushirogawa, T. Takaya, Pharmacological activity of tablets containing recombinant human granulocyte colony-stimulating factor (rhG-CSF) in rats **101**, 89
Takada, K., H. Yoshikawa, S. Muranishi, Partial avoidance of first-pass elimination of azathioprine in rats by rectal dosing **25**, 155
Pharmacokinetic analysis of plasma drug level data obtained from a transdermal therapeutic system with a complex absorption model **65**, 159
Takada, K., see Fukui, H. **31**, 239
Takada, K., see Katayama, N. **110**, 155
Takada, K., see Murakami, M. **31**, 231; **79**, 159
Takada, K., see Ohtani, T. **77**, 141
Takada, K., see Takaya, T. **110**, 47
Takada, K., see Ushirogawa, Y. **81**, 133
Takada, M., see Miyazaki, S. **118**, 257
Takada, S., T. Kurokawa, K. Miyazaki, S. Iwasa, Y. Ogawa, Sustained release of a water-soluble GP IIb/IIIa antagonist from copoly(DL-lactic/glycolic) acid microspheres **146**, 147
Takagi, H., see Sakane, T. **57**, 77
Takagi, K., see Hasegawa, T. **58**, 129
Takagi, T., see Bando, H. **135**, 91
Takahashi, K., T. Katagi, S. Tamagawa, T. Nishihata, Effect of pH on the dissolution of drug particle dispersed in solid matrix of hydrogenated soya phospholipid **49**, 179
Takahashi, K., see Motozono, H. **108**, 39
Takahashi, K., see Nishihata, T. **42**, 257
Takahashi, K., see Oguma, T. **63**, 101
Takahashi, M., see Kikuchi, S. **42**, 193
Takahashi, M., see Kohri, N. **81**, 49
Takahashi, T., K. Takayama, Y. Machida, T. Nagai, Characteristics of polyion complexes of chitosan with sodium alginate and sodium polyacrylate **61**, 35
Takahashi, T., see Sasaki, H. **60**, 1
Takahata, H., see Nashihata, T. **33**, 89
Takakura, Y., T.F. A, H. Furitsu, M. Nishikawa, H. Sezakib, M. Hashida, Pharmacokinetics of succinylated proteins and dextran sulfate in mice: Implications for hepatic targeting of protein drugs by direct succinylation via scavenger receptors **105**, 19
Takakura, Y., R. Atsumi, M. Hashida, H. Sezaki, Development of a novel polymeric prodrug of mitomycin C, mitomycin C–dextran conjugate with anionic charge. II. Disposition and pharmacokinetics following intravenous and intramuscular administration **37**, 145
Takakura, Y., M. Kitajima, S. Matsumoto et al., Development of a novel polymeric prodrug of mitomycin C, mitomycin C–dextran conjugate with anionic charge. I. Physicochemical characteristics and in vivo and in vitro antitumor activities **37**, 135
Takakura, Y., see Bando, H. **135**, 91; **145**, 103
Takakura, Y., see Nishida, K. **65**, 7
Takakura, Y., see Roos, C.F. **22**, 75
Takakura, Y., see Takino, T. **114**, 43
Takakura, Y., see Yamashita, F. **117**, 173
Takano, M., see Sugito, K. **47**, 157

Takashima, Y., see Tanaka, S. **27**, 29
Takaya, T., K. Niwa, K. Takada, Pharmacological effect of recombinant human colony-stimulating factor (rhG-CSF) after administration into rat large intestine **110**, 47
Takayama, K., T. Nagai, Simultaneous optimization for several characteristics concerning percutaneous absorption and skin damage of ketoprofen hydrogens containing D-limonene **74**, 115
Takayama, K., H. Okabe, Y. Obata, T. Nagai, Formulation design of indomethacin gel ointment containing D-limonene using computer optimization methodology **61**, 225
Takayama, K., see Akimoto, J. **128**, 251; **140**, 257
Takayama, K., see Maitani, Y. **117**, 129; **126**, 89; **146**, 11
Takayama, K., see Morishita, I. **78**, 9; **91**, 29
Takayama, K., see Morishita, M. **78**, 1
Takayama, K., see Nakamura, Y. **145**, 29
Takayama, K., see Obata, Y. **89**, 191
Takayama, K., see Ohara, N. **105**, 31; **119**, 125; **123**, 281
Takayama, K., see Oribe, T. **124**, 27
Takayama, K., see Quan, D. **68**, 239
Takayama, K., see Takahashi, T. **61**, 35
Takayama, K., see Watanabe, Y. **99**, 337
Takechi, S., see Kohri, N. **81**, 49
Takeda, Y., see Izutsu, K. **71**, 137
Takeeda, T., Y. Hatakeyama, I. Kitaura, Y. Iwamoto, K. Morimoto, M. Yasuda, Y. Nakamoto, K. Morisaka, K. Danjo, A. Otsuka, Lymphatic and plasma transport of 1-(2-tetrahydrofuryl)-5-fluorouracil by polyacrylic acid aqueous gel after rectal administration and its antitumor effect on Yoshida sarcoma implanted in rats **41**, 21
Takeeda, T., see Kamiya, E. **17**, 273
Takeeda, T., see Morimoto, K. **12**, 107; **14**, 149
Takeeda, T., see Takeeda, T. **41**, 21
Takemoto, K.A., see Lee, V.H.L. **29**, 43
Takenaka, H., see Kawashima, Y. **18**, 335
Takenaka, H., see Yamahira, Y. **3**, 23
Takeuchi, H., H. Sasaki, T. Niwa, T. Hino, Y. Kawashima, K. Uesugi, H. Ozawa, Improvement of photostability of ubidecarenone in the formulation of a novel powdered dosage form termed redispersible dry emulsion **86**, 25
Takeuchi, H., see Kawashima, Y. **72**, 65; **75**, 25; **89**, 9; **99**, 229; **119**, 139
Takeuchi, H., see Morishima, K. **105**, 11
Takeuchi, H., see Murakami, H. **149**, 43
Takeuchi, H., see Niwa, T. **121**, 45
Takeuchi, H., see Schade, A. **117**, 209
Takeuchi, H., see Takeuchi, H. **86**, 25
Takeuchi, K., see Oribe, T. **124**, 27
Takeuchi, M., see Higaki, K. **132**, 165
Takeuchi, T., see Yamaguchi, T. **85**, 87
Takieddin, M., see Sallam, E. **67**, 247
Takino, T., E. Nagahama, T. Sakaeda, F. Yamashita, Y. Takakura, M. Hashida, Pharmacokinetic disposition analysis of lipophilic drugs injected with various lipid carriers in the single-pass rat liver perfusion system **114**, 43
Takino, T., see Nishida, K. **80**, 101
Takla, P.G., C.J. Dakas, A study of interactions of chloroquine with ethanol, sugars and glycerol using ultraviolet spectrophotometry **43**, 225
Takla, P.G., see Gassim, A.E.H. **34**, 23
Talaee, N., see Newman, S.P. **74**, 203, 209
Talsma, H., H. Jousma, K. Nicolay, D.J.A. Crommelin, Multilamellar or multivesicular vesicles? **37**, 171
Talsma, H., M.J. Van Steenbergen, D.J.A. Crommelin, The cryopreservation of liposomes: 3. Almost compete retention of a water-soluble marker in small liposomes in a cryoprotectant containing dispersion after a freezing/thawing cycle **77**, 119
Talsma, H., see Jousma, H. **35**, 263
Talsma, H., see Koster, V.S. **134**, 235
Talsma, H., see Ozer, A.Y. **55**, 185
Talukdar, M.M., R. Kinget, Swelling and drug release behaviour of xanthan gum matrix tablets **120**, 63
Talukdar, M.M., A. Michoel, P. Rombaut, R. Kinget, Comparative study on xanthan gum and hydroxypropylmethyl cellulose as matrices for controlled-release drug delivery. I. Compaction and in vitro drug release behaviour **129**, 233
Talwar, G.P., see Singh, M. **76**, R5; **85**, R5
Tamagawa, S., see Takahashi, K. **49**, 179
Tamai, R., see Naito, S. **8**, 253
Tamaki, H., see Kurono, Y. **80**, 135; **81**, 179
Tamburic, S., D.Q.M. Craig, G. Vuleta, J. Milic, A comparison of electrical and rheological techniques for the characterisation of creams **137**, 243
Tammilehto, S., K. Torniainen, Photochemical stability of dothiepin in aqueous solutions **52**, 123
Tammilehto, S., see Andersin, R. **56**, 175; **123**, 229
Tammilehto, S., see Malkki, L. **63**, 17
Tammilehto, S., see Malkki-Laine, L. **117**, 189
Tammilehto, S., see Selkamaa, R. **49**, 83
Tammilehto, S., see Torniainen, K. **132**, 53
Tamura, M., T. Sueishi, K. Sugibayashi, Y. Morimoto, K. Juni, T. Hasegawa, T. Kawaguchi, Metabolism of testosterone and its ester derivatives in organotypic coculture of human dermal fibroblasts with differentiated epidermis **131**, 263
Tamura, S., see Hata, T. **110**, 1
Tan, E.L., J.C. Liu, Y.W. Chien, Controlled drug release from silicone coated tablets: Preliminary evaluation of coating techniques and characterization of membrane permeation kinetics **42**, 161
Tan, H.G.H., see Streng, W.H. **25**, 135
Tan, S.B., J.M. Newton, Powder flowability as an indication of capsule filling performance **61**, 145
Minimum compression stress requirements for arching and powder retention within a dosator nozzle during capsule filling **63**, 275
Influence of capsule dosator wall texture and powder properties on the angle of wall friction and powder-wall adhesion **64**, 227
Capsule filling performance of powders with dosator nozzles of different wall texture **66**, 207

Influence of compression setting ratio on capsule fill weight and weight variability **66**, 273
Observed and expected powder plug densis obtained by a capsule dosator nozzle system **66**, 283
Tan, X., S. Lindenbaum, Studies on complexation between β-cyclodextrin and bile salts **74**, 127
Tan, Y.T.F., see Heng, P.W.S. **143**, 107; **138**, 57
Tanaka, C., see Sakane, T. **57**, 77
Tanaka, H., see Fujita, T. **134**, 47
Tanaka, H., see Naito, S. **8**, 253
Tanaka, I., see Yamauchi, H. **113**, 141
Tanaka, R., see Katayama, N. **115**, 87
Tanaka, S., Y. Takashima, H. Murayama, S. Tsuchiya, Studies on drug release from ointments. V. Release of hydrocortisone butyrate propionate from topical dosage forms to silicone rubber **27**, 29
Tanaka, S., see Fujimori, J. **119**, 47
Tanaka, S., see Naito, S. **18**, 117
Tanaka, S., see Naito, S.I. **17**, 109
Tanaka, T., see Kaneo, Y. **57**, 263
Tanaka, T., see Kaneo, Y. **44**, 265
Tanaka, T., see Takada, K. **101**, 89
Tandon, A., A.N. Nagappa, R.C. Srivastava, Liquid membrane phenomenon in vitamin E: Studies on α-tocopherol **32**, 39
Tandon, A., see Srivastava, R.C. **38**, 239
Tandt, L.A.G.L., C. Stubbs, I. Kanfer, The use of dissolution rate data to account for differences in the absorption profiles of two controlled/modified-release capsule dosage forms of indomethacin in human volunteers **104**, 11
Tang, S.Y., see Won, C.M. **121**, 95
Tang-Liu, D., see Small, D. **149**, 195, 203
Tanigawara, Y., see Heya, T. **69**, 69
Tanigawara, Y., see Yamaoka, K. **10**, 291
Taniguchi, K., S. Muranishi, H. Sezaki, Enhanced intestinal permeability to macromolecules. II. Improvement of the large intestinal absorption of heparin by lipid-surfactant mixed micelles in rat **4**, 219
Taniguchi, M., see Maitani, Y. **64**, 139
Tanimoto, T., see Suenaga, A. **59**, 121; **82**, 29
Tanino, T., see Ogiso, T. **98**, 113; **102**, 141
Tanira, M.O.M., I.A. Wasfi, A.K. Bashir, D.W.G. Harron, Chronic verapamil administration alters phenytoin pharmacokinetics in the rat **102**, 267
Tanizawa, T., see Kurosaki, Y. **67**, 1
Tanninen, V.P., J. Yliruusi, X-ray powder diffraction profile fitting in quantitative determination of two polymorphs from their powder mixture **81**, 169
Tantishaiyakul, V., N. Kawnopparat, S. Ingkatawornwong, Properties of solid dispersions of piroxicam in polyvinylpyrrolidone K-30 **143**, 59
Tanzawa, K., see Tokui, T. **110**, 277
Tapia, C., G. Buckton, J.M. Newton, Factors influencing the mechanism of release from sustained release matrix pellets, produced by extrusion/spheronisation **92**, 211
Tarimci, N., D. Ermis, Sustained release characteristics and pharmacokinetic parameters of ketoprofen suppositories using chitosan **147**, 71
Tarimci, N., see Ermis, D. **113**, 65
Tariq, M., see Moustafa, M.A. **30**, 225
Tarpila, S., see Kivinen, A. **108**, 109
Tartar, A., see Blanco, Prieto M.J **111**, 137
Tashima, T., U. Kawakami, M. Harada, N. Satoh, T. Nakagawa, H. Tanaka, Relationship between precipitation in aqueous solution of glutaraldehyde for chemosterilization and impurities detected by gas chromatography **42**, 61
Tasset, C., F. Goethals, V. Preat, M. Roland, Effect of polyoxyethyleneglycol (24) cholesterol on the solubility, toxicity and activity of amphotericin B **58**, 41
Tasset, C., V. Preat, M. Roland, Comparison of the in vivo antifungal activity of amphotericin B-Solulan C24, amphotericin B-Myrj 59 and amphotericin B-Synperonic A50 with fungizone **81**, R5
Tasset, C., see Thysman, S. **101**, 105
Tasyurek, M., see Gursoy, A. **52**, 101
Tatsuishi, K., see Aoki, S. **85**, 29, 65; **95**, 67
Tatsumi, E., see Matsuda, Y. **60**, 11
Taub, M.E., Larsen B. Due, B. Steffansen, S. Frokjaer, β-Carboxylic acid esterified D-Asp-Ala retains a high affinity for the oligopeptide transporter in Caco-2 monolayers **146**, 205
Tavakoli-Saberi, M.R., K.L. Audus, Physicochemical factors affecting β-adrenergic antagonist permeation across cultured hamster pouch buccal epithelium **56**, 135
Tavakoli-Saberi, M.R., see Audus, K.L. **76**, 247
Taverdet, J.L., see Armand, J.Y. **40**, 33
Taverdet, J.L., see Eddine, A. **32**, 143
Taverdet, J.L., see Laghoueg, N. **50**, 133
Taverdet, J.L., see Magron, P. **38**, 91
Tavipatana, W., J.W. Ayres, Etophylline and theophylline pharmacokinetics when administered concomitantly in rabbits **30**, 143
Proxyphylline and theophylline pharmacokientics when administered concomitantly in rabbits **38**, 139
Taw, J., see Ma, X. **142**, 115
Tawashi, R., see Akbarieh, M. **35**, 81
Tawashi, R., see El-Ridy, M. **56**, 23
Tawashi, R., see Heiati, H. **146**, 123
Tawashi, R., see Kassem, M. **47**, 179
Tawashi, R., see Laurin, P. **28**, 177
Tawashi, R., see Thibert, R. **47**, 171
Tawfiq, F.A., A. Hussain, L. Dittert, P.A. Crooks, Kinetics of decomposition of *O*-acyl derivatives of salicylamide and related compounds **64**, 45
Taylor, A.J., see Biddiscombe, M.F. **91**, 111
Taylor, A.J., see McCallion, O.N.M. **129**, 123; **130**, 1; **133**, 203
Taylor, C.J., see Attwood, D. **84**, R5; **88**, 417
Taylor, C.J., see Ruth, H.S. **116**, 253
Taylor, D.C., see Harris, D. **56**, 97
Taylor, D.C., see Kaus, L.C. **20**, 315; **22**, 99
Taylor, G., see Cabral Marques, H. **77**, 297, 303
Taylor, G., see Davies, N.M. **105**, 97
Taylor, G., see Dennis, A.B. **65**, 85
Taylor, G., see Dickinson, P.A. **116**, 231; **148**, 55

Taylor, G., see Donnelly, A. **135**, 191
Taylor, G., see Greenslade, M.D. **26**, 133
Taylor, G., see Gumblteon, M. **38**, 261
Taylor, G., see Taylor, K.M.G. **58**, 49, 57
Taylor, G., see Williams, A.S. **83**, 233
Taylor, G., see Williams, A.W. **87**, 255
Taylor, G., see Woolfrey, S.G. **26**, 35
Taylor, K.M.G., C. Hoare, Ultrasonic nebulisation of pentamidine isethionate **98**, 45
Taylor, K.M.G., G. Taylor, I.W. Kellaway, J. Stevens, Drug entrapment and release from multilamellar and reverse-phase evaporation liposomes **58**, 49
The stability of liposomes to nebulisation **58**, 57
Taylor, K.M.G., G. Venthoye, A. Chawla, Pentamidine isethionate delivery from jet nebulisers **85**, 203
Taylor, K.M.G., see Ah-Fat, N.M.W. **107**, 239
Taylor, K.M.G., see Barker, S.A. **102**, 159
Taylor, K.M.G., see Chawla, A. **108**, 233
Taylor, K.M.G., see Craig, D.Q.M. **114**, 124 + 128
Taylor, K.M.G., see Leung, K.K.M. **145**, 95
Taylor, K.M.G., see McCallion, O.N.M. **129**, 123; **130**, 1; **133**, 203
Taylor, K.M.G., see Stelliou, I. **99**, R1
Taylor, M.J., see Davis, S.S. **21**, 167, 331
Taylor, M.J., see Lilley, J.C. **107**, 149
Taylor, P.J., see de Meere, A.L. **31**, 209
Taylor, P.J., see Jones, M.F. **4**, 1
Taylor, P.J., see Leahy, D.E. **50**, 117
Taylor, R.B., D.G. Durham, A.S.H. Shivji, A kinetic study of tetracycline decomposition in acid solution **26**, 259
Taylor, R.B., R.R. Moody, N.A. Ochekpe, A.S. Low, M.I.A. Harper, A chemical stability study of proguanil hydrochloride **60**, 185
Taylor, R.B., R.M.E. Richards, A.S. Low, L. Hardie, Chemical stability of polymyxin B in aqueous solution **102**, 201
Taylor, R.B., see Durham, D.G. **12**, 31
Taylor, R.B., see Edwards, D. **4**, 21
Taylor, R.B., see Moody, R.R. **7**, 177
Taylor, W.F., see Johnson, D.M. **31**, 125
Tchoreloff, P., see Ribardiere, A. **144**, 195
Tcnnesen, H.H., J.V. Greenhill, Studies on curcumin and curcuminoids. XXII. Curcumin as a reducing agent and as a radical scavenger **87**, 79
Tcnnesen, H.H., D.E. Moore, Photochemical stability of biologically active compounds. III. Mefloquine as a photosensitizer **70**, 95
Te Wierik, G.H, J. Bergsma, A.W. Arends-Scholte, T. Boersma, A.C. Eissens, C.F. Lerk, A new generation of starch products as excipient in pharmaceutical tablets. I. Preparation and binding properties of high surface area potato starch products **134**, 27
Te Wierik, G.H, A.C. Eissens, C.F. Lerk, Preparation, characterization and pharmaceutical application of linear dextrins. IV. Drug release from capsules and tablets containing amylodextrin **98**, 219
Preparation, characterization and pharmaceutical application of linear dextrins. V. Study on the binding properties of amylodextrin, metastable amylodextrin and metastable amylose **102**, 81
Teeuwse, J., see Van der Houwen, O.A.G.J. **105**, 249
Tegenfeldt, J., see Alden, M. **83**, 47; **94**, 31; **110**, 267
Teh, G.W., see Law, S.L. **43**, 257
Teixeira, M., see Lourenco, C. **138**, 1
Tekle, A., see Al-Hassan, M.I. **50**, 35; **93**, 7
Tempera, see Puglisi, G. **118**, 65
Temple, D.J., D.W.G. Harron, P.S. Collier, Utilisation of digitalis glycosides: The relevance of their biotransformation **2**, 127
Temple, D.J., see Karim, E.F.I.A. **52**, 155
Temple, R.J., see Peck, C.C. **82**, 9
Tenjarla, S., see Mandal, T.K. **137**, 187
Tenjarla, S., see Shah, J.C. **109**, 283; **112**, 263
Ter-Minassian-Saraga, L., G. Albrecht, Vinblastine sulphate encapsulation in phospholipid vesicles and liposomes **24**, 149
Terada, K., see El-Gendy, G.A. **31**, 25
Terada, K., see Yamamura, S. **133**, 117
Teramura, Y., see Masegi, M. **78**, 103
Terao, K., see Kurosaki, Y. **82**, 165
Terao, T., see Aso, Y. **93**, 153; **115**, 133
Terao, T., see Izutsu, K.I. **90**, 187
Teraoka, R., Y. Matsuda, Stabilization-oriented preformulation study of photolabile menatetrenone (vitamin K_2) **93**, 85
Teraoka, R., see Matsuda, Y. **26**, 289; **54**, 211
Terasaki, T., Y. Deguchi, Y. Kasama, W.M. Partridge, A. Tsuji, Determination of in vivo steady-state unbound drug concentration in the brain interstitial fluid by microdialysis **81**, 143
Terepolsky, S.A., I. Kanfer, Stability of erythromycin and some of its esters in methanol and acetonitrile **115**, 123
Testa, B., see Gryllaki, M. **51**, 95
Testa, B., see Pannatier, A. **8**, 167
Testa, B., see Repond, C. **38**, 47
Testa, B., see Rothlisberger, T. **44**, 141; **51**, 73
Testa, B., see Tsai, R.S. **80**, 39
Testa, B., see Van de Waterbeemd, H. **14**, 29
Testa, B., see Wernly-Chung, G.N. **63**, 129
Texeira-Dias, J.J.C., see Veiga, F. **129**, 63
Thacharodi, D., K.P. Rao, Transdermal absorption of nifedipine from microemulsions of lipophilic skin penetration enhancers **111**, 235
Collagen-chitosan composite membranes for controlled release of propranolol hydrochloride **120**, 115
Collagen membrane controlled transdermal delivery of propranolol hydrochloride **131**, 97
Thacharodi, D., K. Panduranga Rao, Release of nifedipine through crosslinked chitosan membranes **96**, 33
Collagen-chitosan composite membranes controlled transdermal delivery of nifedipine and propranolol hydrochloride **134**, 239
Thakur, A.B., see Morris, K.R. **105**, 209
Theeuwes, F., see McMahon, F.G. **91**, 75

Thermes, F., A. Rozier, B. Plazonnet, J. Grove, Bioadhesion: The effect of polyacrylic acid on the ocular bioavailability of timolol **81**, 59
Thevenin, M.A., J.L. Grossiord, M.C. Poelman, Sucrose esters/cosurfactant microemulsion systems for transdermal delivery: Assessment of bicontinuous structures **137**, 177
Thibert, R., M. Akbarieh, R. Tawashi, Morphic features variation of solid particles after size reduction: Sonification compared to jet mill grinding **47**, 171
Thibert, R., H. Mach, S.D. Clas, D.R. Meisner, E.B. Vadas, Characterization of the self-association properties of a leukotriene D_4 receptor antagonist, MK-0476 **134**, 59
Thiel, Z., see Regosz, A. **88**, 437
Thiermann, H., S. Seidl, P. Eyer, HI 6 dimethanesulfonate has better dissolution properties than HI 6 dichloride for application in dry/wet autoinjectors **137**, 167
Thiermann, H., U. Spohrer, R. Klimmek, P. Eyer, Operational evaluation of wet/dry autoinjectors containing atropine in solution and powdered HI 6 or HLo 7 ether◇ ether diiodide◇ **109**, 35
Thies, C., see Benoit, J.P. **29**, 95
Thioune, O., H. Fessi, J.P. Devissaguet, F. Puisieux, Preparation of pseudolatex by nanoprecipitation: Influence of the solvent nature on intrinsic viscosity and interaction constant **146**, 233
Thirupathi Rao, T., see Sastry, C.S.P. **93**, 13
Thoma, K., R. Klimek, Photostabilization of drugs in dosage forms without protection from packaging materials **67**, 169
Thoma, K., T. Zimmer, Retardation of weakly basic drugs with diffusion tablets **58**, 197
Thomas, C., see Calvo, P. **103**, 283
Thomas, E., J. Rubino, Solubility, melting point and salting-out relationships in a group of secondary amine hydrochloride salts **130**, 179
Thomas, E., see Rubino, J.T. **65**, 141
Thomas, J., see Hendrix, C. **100**, 213
Thomas, M., see Aboofazeli, R. **125**, 107
Thomas, M., see Davies, N.M. **105**, 97
Thomas, M., see Durrani, A.M. **88**, 409
Thomas, M., see McCallion, O.N.M. **129**, 123; **130**, 1; **133**, 203
Thomas, M., I.W. Kellaway, B.E. Jones, A comparative study of gelatin gelation using photon correlation spectroscopy and a conventional gelation timer **27**, 299
Gelatin film formation at the air–water interface **71**, 147
Gelatin film formation at the solid–aqueous interface **73**, 185
A study by photon correlation spectroscopy of the influence of gelatin source, ionic strength and temperature on the gelation process **83**, 139
Thomas, N.W., L.J. Lack, B.A. Woodhouse, K.J. Palin, P.L. Gould, Formulation of Fenbufen suppositories. III. Histology of the rectal mucosa of rats following repeat dosing of Fenbufen in Witepsol H12 and PEG vehicles **44**, 261
Thomas, N.W., K.J. Palin, D.A. Waller, The application of an in situ loop technique to the study of rectal absorption **20**, 163
Thomas, N.W., see Chandler, S.G. **70**, 19; **76**, 61; **117**, 139
Thomas, N.W., see Jenkins, P.G. **102**, 261
Thomas, N.W., see Reid, A.S. **40**, 181
Thomas, N.W., see Richardson, J.L. **56**, 29; **77**, 75
Thomas, N.W., see Whiston, R.J. **23**, 247
Thomas, N.W., see Young, C. **40**, 187
Thomas, W.H., see Hui, L.K. **10**, 133
Thomasin, C., H.P. Merkle, B.A. Gander, Physico-chemical parameters governing protein microencapsulation into biodegradable polyesters by coacervation **147**, 173
Thombre, A.G., A.R. DeNoto, F.C. Falkner, J.D. Lazar, In vitro–in vivo correlations of sustained-release coated multiparticulate formulations of doxazosin **111**, 181
Thompson, D.C., see Shively, M.L. **117**, 119
Thompson, D.O., see Stella, V.J. **120**, 189, 197
Thompson, G., see Toth, I. **106**, 85
Thompson, S.P., see White, D.A. **40**, 151
Thomsen, K., see Pridal, L. **136**, 53
Thomsen, K.F., F. Strom, B.V. Sforzini, M. Begtrup, N. Mork, Evaluation of phenyl carbamates of ethyl diamines as cyclization-activated prodrug forms for protecting phenols against first-pass metabolism **112**, 143
Thomsen, M., see Helboe, P. **2**, 317
Thomsen, M.K., see Vermehren, C. **128**, 239
Thorbek, P., see Bundgaard, H. **18**, 67
Thorisdottir, S., see Loftsson, T. **104**, 181
Thornton, D., see Chen, T.K. **59**, 211
Thorvaldsson, T., see Gizurarson, S. **140**, 77; **146**, 133
Thouvenot, P., see Zimmer, A.K. **110**, 211
Throne, O., see McElnay, J.C. **7**, 83
Thysman, S., C. Tasset, V. Preat, Transdermal iontophoresis of fentanyl: Delivery and mechanistic analysis **101**, 105
Thysman, S., see Jadoul, A. **120**, 221
Thysman, S., see Preat, V. **96**, 189
Ticehurst, M.D., R.C. Rowe, P. York, Determination of the surface properties of two batches of salbutamol sulphate by inverse gas chromatography **111**, 241
Tidy, P.J., see Sewell, G.J. **70**, 119
Tiedt, L.R., see Du Plessis, J. **34**, 173
Tiemessen, H.L.G.M., H.E. Bodde, H.E. Junginger, A silicone membrane sandwich method to measure drug transport through isolated human stratum corneum having a fixed water content **56**, 87
Tiemessen, H.L.G.M., H.E. Bodde, H. Mollee, H.E. Junginger, A human stratum corneum-silicone membrane sandwich to simulate drug transport under occlusion **53**, 119
Tighe, B.J., see Gould, P.L. **38**, 231
Timmermans, J., A.J. Moes, How well do floating dosage forms float? **62**, 207
Timmermans, J., see Ingani, H.M. **35**, 157
Timmins, P., I.M. Jackson, Y.J. Wang, Factors affecting captopril stability in aqueous solution **11**, 329
Timmins, P., see Hussain, M.S.H. **42**, 89; **70**, 103; **78**, 203
Timmins, P., see Nicklin, P.L. **140**, 175
Timoney, R.F., Structural requirements of muscarine, 1,3-dioxolane and their analogues for agonist activity at cholinergic receptors **15**, 223

Timoney, R.F., see Corrigan, O.I. **1**, 299; **4**, 67
Timoney, R.F., see Harte, V.J. **1**, 165
Timoney, R.F., see Irwin, V.P. **34**, 241; **43**, 187
Timoney, R.F., see Jordan, C.G.M. **84**, 175; **141**, 125
Timoney, R.F., see Quigley, J.M. **58**, 107; **101**, 145
Timsina, M.P., G.P. Martin, C. Marriott, D. Ganderton, M. Yianneskis, Drug delivery to the respiratory tract using dry powder inhalers **101**, 1
Tincer, T., see Yuksel, N. **140**, 145
Tio, F., see Gowan, W.G. **29**, 169
Tirendi, A.S., see Scalia, S. **115**, 249
Tirendi, S., see Bonina, F.P. **77**, 21
Tirendi, S., see Ventura, C.A. **149**, 1
Tirkkonen, S., P. Paronen, Enhancement of drug release from ethylcellulose microcapsules using solid sodium chloride in the wall **88**, 39
Release of indomethacin from tabletted ethylcellulose microcapsules **92**, 55
Tirkkonen, S., A. Urtti, P. Paronen, Buffer controlled release of indomethacin from ethylcellulose microcapsules **124**, 219
Tissier, B., see Ruiz, J.M. **49**, 69
Titulaer, H.A.C., J. Zuidema, C.B. Lugt, Formulation and pharmacokinetics of artemisinin and its derivatives **69**, 83
Titulaer, H.A.C., see Zuidema, J. **105**, 189
Tjornelund, J., see Jensen, J. **88**, 177
Tobyn, M.J., J.N. Staniforth, A.R. Baichwal, T.W. McCall, Prediction of physical properties of a novel polysaccharide controlled release system. I. **128**, 113
Tocanne, J.F., see Peyrot, M. **102**, 25
Toddywala, R.D., K. Ulman, P. Walters, Y.W. Chien, Effect of physicochemical properties of adhesive on the release, skin permeation and adhesiveness of adhesive-type transdermal drug delivery systems (a-TDD) containing silicone-based pressure-sensitive adhesives **76**, 77
Toguchi, H., see Heya, T. **69**, 69; **72**, 199
Toguchi, H., see Iga, K. **57**, 241
Toguchi, H., see Okada, H. **54**, 231
Tohmakhchi, V.N., see Feldstein, M.M. **131**, 229
Toikkanen, E., see Turakka, L. **17**, 99
Tojo, K., Concentration profile in plasma after transdermal drug delivery **43**, 201
Tojo, K., see Liu, J.C. **25**, 265
Tojo, K., see Shah, H.S. **86**, 167
Tokes-Kovesdi, M., see Takacs-Novak, K. **89**, 261
Tokgoz, N.S., J.L. Grossiord, A. Fructus, M. Seiller, P. Prognon, Evaluation of two fluorescent probes for the characterization of w/o/w emulsions **141**, 27
Tokiwa, H., see Seta, Y. **41**, 263
Tokui, T., Y. Tokui, M. Ishigami, K. Tanzawa, T. Ikeda, T. Komai, Targeting of an antitumor agent, RS-1541 (palmitoyl-rhizoxin), via low-density lipoprotein receptor **110**, 277
Tokui, Y., see Tokui, T. **110**, 277
Tolfree, S.E.J., see Zainudin, B.M.Z. **51**, 67
Tolley, J.A., J. Rassing, A new ultrasonic velocity scanning technique giving concentration-distance profiles and diffusion coefficient of drugs in gels **14**, 223
Tolley, J.A., see Attwood, D. **9**, 285
Tolli, J., see Abrahamsson, B. **140**, 229
Tom, J., see Benjamin, E.J. **14**, 209
Tomaino, A., see Bonina, F. **145**, 87
Tomaino, A., see Saija, A. **124**, 1
Tomarchio, V., see Giammona, G. **127**, 165
Tome, D., see Benkhelifa, S. **145**, 115
Tome, D., see Crauste-Manciet, S. **149**, 241
Tomenson, J.A., see Leahy, D.E. **50**, 117
Tomer, R., A.T. Florence, Photo-responsive hydrogels for potential responsive release applications **99**, R5
Tomikawa, M., see Yamauchi, H. **90**, 73
Tominaga, H., see Naito, S. **24**, 115
Tomka, I., see Beck, M.I. **141**, 137
Tomlinson, E., see Buist, H.P.A. **15**, 347
Tomlinson, E., see Burger, J.J. **23**, 333
Tomlinson, E., see Burgess, D.J. **39**, 129
Tomlinson, E., see de Meere, A.L. **17**, 331; **22**, 177; **31**, 209
Tomlinson, E., see Grunbauer, H.J.M. **21**, 61
Tomlinson, E., see Hafkenscheid, T.L. **16**, 225; **17**, 1; **19**, 349
Tomlinson, E., see Kinkel, J.F.M. **6**, 261
Tomlinson, E., see O'Mullane, J.E. **63**, 177
Tomlinson, E., J.A.M. Van Dooremalen, H.H. Van Rooij, H.J.A. Wynne, Ion-pair and complex-coacervate effects on large ion flux through polyamide-6 membrane **12**, 87
Tomlinson, E., Enthalpy–entropy compensation analysis of pharmaceutical, biochemical and biological systems **13**, 115
Tonegawa, C., see Nishida, K. **65**, 7
Tonnesen, H.H., Studies on curcumin and curcuminoids. XIII. Catalytic effect of curcumin on the peroxidation of linoleic acid by 15-lipoxygenase **50**, 67
Studies on curcumin and curcuminoids. XIV. Effect of curcumin on hyaluronic acid degradation in vitro **50**, 91
Studies on curcumin and curcuminoids. XV. Catalytic effect of demethoxy- and bisdemethoxycurcumin on the peroxidation of linoleic acid by 15-lipoxygenase **51**, 179
Studies on curcumin and curcuminoids. XVI. Effect of curcumin analogs on hyaluronic acid degradation in vitro **51**, 259
Tonnesen, H.H., A.L. Grislingaas, Photochemical stability of biologically active compounds. II. Photochemical decomposition of mefloquine in water **60**, 157
Tonnesen, H.H., A.L. Grislingaas, S.O. Woo, J. Karlsen, Photochemical stability of antimalarials. I. Hydroxychloroquine **43**, 215
Tonnesen, H.H., J. Karlsen, Studies on curcumin and curcuminoids. XI. Stabilization of photolabile drugs in serum samples by addition of curcumin **41**, 75
Tonnesen, H.H., S. Kristensen, L.N. Grinberg, Studies on curcumin and curcuminoids. XXV. Inhibition of primaquine-induced lysis of human red blood cells by curcumin **110**, 161
Tonnesen, H.H., M. Shankar, Studies on curcumin and curcuminoids. X. The use of curcumin as a formulation aid to protect light-sensitive drugs in soft gelatin capsules **38**, 247

Tonneson, H.H., G. Smistad, T. Agren, J. Karlsen, Studies on curcumin and curcuminoids. XXIII. Effects of curcumin on liposomal lipid peroxidation **90**, 221
Tonnesen, H.H., see Bjerga, Bjaen A.K. **92**, 183
Tonnesen, H.H., see Grinberg, L.N. **132**, 251
Tonnesen, H.H., see Nord, K. **72**, 11
Tonsini, A., see Santoni, G. **93**, 239
Toomanian, M., see Barzegar-Jalali, M. **12**, 351
Topp, E., see Kyyronen, K. **80**, 161
Topp, E.M., see Bhatt, P.P. **72**, 219
Topp, E.M., see Hume, L.R. **111**, 295
Topp, E.M., see Joshi, H.N. **51**, 19; **80**, 213
Topp, E.M., see Rajewski, L.G. **82**, 205
Topp, E.M., see Sanzgiri, Y.D. **107**, 91
Toppila, M.A.I., see Salomies, H.E.M. **110**, 197
Torchilin, V.P., see Papisov, M.I. **40**, 201, 207
Torniainen, K., S. Tammilehto, V. Ulvi, The effect of pH, buffer type and drug concentration on the photodegradation of ciprofloxacin **132**, 53
Torniainen, K., see Tammilehto, S. **52**, 123
Toro, I., see Barbosa, J. **149**, 213
Torracca, M.T., see Saettone, M.F. **51**, 203; **72**, 131; **86**, 159
Torrado, J.J., L.L. Augsburger, Effect of different excipients on the tableting of coated particles **106**, 149
Torrado, J.J., L. Illum, S.S. Davis, Particle size and size distribution of albumin microspheres produced by heat and chemical stabilization **51**, 85
Torrado, J.J., see Torrado, S. **86**, 147; **140**, 45, 247; **133**, 65
Torrado, S., R. Cadorniga, J.J. Torrado, Effect of drug release rate on bioavailability of different aspirin tablets **133**, 65
Torrado, S., J.J. Torrado, R. Cadorniga, Topical application of albumin microspheres containing vitamin A. Drug release and availability **86**, 147
Torrado, S., S. Torrado, R. Cadorniga, J.J. Torrado, Formulation parameters of albendazole solution **140**, 45
Torrado, S., S. Torrado, J.J. Torrado, R. Cadorniga, Preparation, dissolution and characterization of albendazole solid dispersions **140**, 247
Torrado, S., see Sanchez, L. **115**, 201
Torrado, S., see Torrado, S. **140**, 45, 247
Torre, M.L., see Maggi, L. **135**, 73
Torres, D., G. Garcia-Encina, B. Seijo, Jato J.L. Vila, Formulation and in vitro evaluation of HPMCP-microencapsulated drug-resin complexes for sustained release of diclofenac **121**, 239
Torres, D., B. Seijo, G. Garcia-Encina, M.J. Alonso, J.L. Vila-Jato, Microencapsulation of ion-exchange resins by interfacial nylon polymerization **59**, 9
Torres, F., V. Girona, M. Puiol, J. Prat, Bolos J. De, Stability of carboplatin in 5% glucose solution exposed to light **129**, 275
Torres-Labandeira, J., see Pitha, J. **80**, 253
Torres-Labandeira, J.J., see Esclusa-Diaz, M.T. **142**, 183; **143**, 203
Torresi, M.C., see Bramanti, G. **49**, 241
Torri, G., see Ahmed, S.M. **109**, 99
Torricelli, C., A. Martini, L. Muggetti, R. de Ponti, Stability studies on a steroidal drug/β-cyclodextrin compound mixture **71**, 19
Torricelli, C., A. Martini, L. Muggetti, M. Eli, R. de Ponti, Stability studies on steroidal drug/β-cyclodextrin kneaded systems **75**, 147
Torricelli, C., see Martini, A. **75**, 141
Torstensson, A., see Gottfries, J. **106**, 141
Tota, G., see Saettone, M.F. **43**, 67
Toth, I., M. Christodoulou, K. Bankowsky, N. Flinn, W.A. Gibbons, G. Godeau, E. Moczar, W. Hornebeck, Design of potent lipophilic-peptide inhibitors of human neutrophil elastase: In vitro and in vivo studies **125**, 117
Toth, I., E. Del Olmo Fernandez, R.A. Hafeez, J.L. Holley, I.P. Wood, P. Ward, W.A. Gibbons, Lipidic peptides. XII. Cellular uptake studies of a lipidic amino acid, its oligomers and highly lipophilic drug conjugates on Ehrlich ascites tumour cells **83**, 131
Toth, I., N. Flinn, A. Hillery, W.A. Gibbons, P. Artursson, Lipidic conjugates of luteinizing hormone releasing hormone $(LHRH)^+$ and thyrotropin releasing hormone $(TRH)^+$ that release and protect the native hormones in homogenates of human intestinal epithelial (Caco-2) cells **105**, 241
Toth, I., I.P. Griffith, E. Del Olmo Fernandez, R.A. Hafeez, J.L. Holley, P. Ward, W.A. Gibbons, Lipidic peptides. VIII. Cellular uptake studies of lipidic amino acid, its oligomers and highly lipophilic drug conjugates **79**, 39
Toth, I., A.M. Hillery, I.P. Wood, C. Magnusson, P. Artursson, Oral absorption of lipidic amino acid conjugates **102**, 223
Toth, I., R.A. Hughes, M.R. Munday, C.A. Murphy, P. Mascagni, W.A. Gibbons, Lipidic peptides. II. Synthesis, activity and transport of anti-inflammatory benzoquinolizine-lipidic peptide conjugates **68**, 191
Toth, I., R.A. Hughes, P. Ward, M.A. Baldwin, K.J. Welham, A.M. McColm, D.M. CoxGibbons W.A., Lipidic peptides. IV. Penicillin and cephalosporin amide conjugates with lipidic amino acids and their oligomers **73**, 259
Toth, I., R.A. Hughes, P. Ward, A.M. McColm, D.M. Cox, G.J. Anderson, W.A. Gibbons, Fatty peptides. VI. Penicillin and cephalosporin esters with increased lipophilic character **77**, 13
Toth, I., G. Thompson, P. Ward, Lipophilic peptide inhibitors of ribonucleotide reductase enzyme of herpes simplex virus **106**, 85
Toth, I., see Del Olmo, E. **81**, 267
Toth, I., see Flinn, N. **137**, 33
Toth, I., see Valko, K. **83**, 123
Toth, I., see Wood, I.P. **87**, 141
Touchard, F., see Gadalla, M.A.F. **38**, 71
Touitou, E., Transdermal delivery of anxiolytics: In vitro skin permeation of midazolam maleate and diazepam **33**, 37
Skin permeation enhancement by *n*-decyl methyl sulfoxide: Effect of solvent systems and insights on mechanism of action **43**, 1

Touitou, E., L. Abed, Effect of propylene glycol, azone and *n*-decylmethyl sulphoxide on skin permeation kinetics of 5-fluorouracil **27**, 89
Touitou, E., M. Alkabes, A. Memoli, F. Alhaique, Glutathione stabilizes ascorbic acid in aqueous solution **133**, 85
Touitou, E., D.D. Chow, J.R. Lawter, Chiral β-blockers for transdermal delivery **104**, 19
Touitou, E., J. Deutsch, S. Matar, Iodine-polyurethane matrices: Antimicrobial activity vs. method of preparation **103**, 199
Touitou, E., M. Donbrow, Deviation of dissolution behaviour of benzoic acid from theoretical predictions with lowering of temperature: Limitations as a model dissolution substance **9**, 97
Influence of additives on (hydroxyethyl) methylcellulose properties: Relation between gelation temperature change, compressed matrix integrity and drug release profile **11**, 131
Drug release from non-disintegrating hydrophilic matrices: Sodium salicylate as a model drug **11**, 355
Promoted rectal absorption of insulin: Formulative parameters involved in the absorption from hydrophilic bases **15**, 13
Touitou, E., B. Fabin, Altered skin permeation of a highly lipophilic molecule: Tetrahydrocannabinol **43**, 17
Touitou, E., B. Fabin, S. Dany, S. Almog, Transdermal delivery of tetrahydrocannabinol **43**, 9
Touitou, E., D. Friedman, The release mechanism of drugs from polyurethane transdermal delivery systems **19**, 323
Touitou, E., D. Gilhar, F. Alhaique, A. Memoli, F.M. Riccieri, E. Santucci, Ascorbic acid in aqueous solution: Bathochromic shift in dilution and degradation **78**, 85
Touitou, E., F. Levi-Schaffer, N. Dayan, F. Alhaique, F. Riccieri, Modulation of caffeine skin delivery by carrier design: Liposomes versus permeation enhancers **103**, 131
Touitou, E., F. Levi-Schaffer, N. Shaco-Ezra, R. Ben-Yossef, B. Fabin, Enhanced permeation of theophylline through the skin and its effect on fibroblast proliferation **70**, 159
Touitou, E., A. Rubinstein, Targeted enteral delivery of insulin to rats **30**, 95
Touitou, E., see Fabin, B. **74**, 59
Toulouse, F.A.M., see Swart, P.J. **88**, 165
Tous, S.S., see Hamed, M.Y. **69**, 93
Toyama, M., see Inoue, N. **137**, 75
Tozer, T.N., see McLeod, A.D. **92**, 105
Trapani, G., C. Altomare, M. Franco, A. Latrofa, G. Liso, Determination of hydrophile-lipophile balance of some polyethoxylated non-ionic surfactants by reversed-phase thin layer chromatography **116**, 95
Trapani, G., A. Lopedota, M. Franco, A. Latrofa, G. Liso, Effect of 2-hydroxypropyl-β-cyclodextrin on the aqueous solubility of the anaesthetic agent propofol (2,6-diisopropylphenol) **139**, 215
Trapani, G., see Bonina, F.P. **124**, 45
Travagli, V., see Memoli, A. **117**, 159
Treiner, C., see Jansen, J. **103**, 19
Trendovska, G., see Panzova, B. **70**, 187
Trenk, D., see Oie, S. **48**, 223
Trenktrog, T., B.W. Muller, Preparation and characterization of a peptide containing w/o emulsion **123**, 199
Treuheit, M.J., see Niven, R.W. **127**, 191
Treupel, L., see Gallardo, M. **100**, 55
Treupel-Acar, L., see Nemati, F. **102**, 55
Triccas, I.M., see Davis, S.S. **30**, 29
Tricehurst, M.D., P. York, R.C. Rowe, S.K. Dwivedi, Characterisation of the surface properties of α-lactose monohydrate with inverse gas chromatography, used to detect batch variation **141**, 93
Trier, L., see Buur, A. **129**, 223
Tripathi, M., D.V. Kohli, R.K. Uppadhyay, Effect of cholic and deoxycholic acid conjugates on solubility and dissolution of indomethacin and phenylbutazone **67**, 207
Triqueneaux, C., see Burgess, D.J. **39**, 129
Trivedi, J.S., J.J. Fort, Kinetics of degradation in aqueous solution of Abbott-79175, a potent second generation 5-lipoxygenase inhibitor **123**, 217
Trnovec, T., see Piotrovskij, V.K. **119**, 213
Troche, G., see Rieutord, A. **119**, 57
Trofast, E., see Newman, S.P. **74**, 209
Trombelli, L., see Esposito, E. **142**, 9
Trombetta, D., see Saija, A. **124**, 1
Troster, S.D., J. Kreuter, Contact angles of surfactants with a potential to alter the body distribution of colloidal drug carriers on poly(methyl methacrylate) surfaces **45**, 91
Troster, S.D., U. Muller, J. Kreuter, Modification of the body distribution of poly(methyl methacrylate) nanoparticles in rats by coating with surfactants **61**, 85
Trotta, M., R. Cavalli, E. Ugazio, M.R. Gasco, Phase behaviour of microemulsion systems containing lecithin and lysolecithin as surfactants **143**, 67
Trotta, M., see Cavalli, R. **148**, 47
Trotta, M., see Gallarate, M. **100**, 219
Trotta, M., see Gasco, M.R. **18**, 235; **29**, 267
Truelove, J., R. Bawarshi-Nassar, N.R. Chen, A. Hussain, Solubility enhancement of some developmental anti-cancer nucleoside analogs by complexation with nicotinamide **19**, 17
Truman, C.A., see Ali, H.M. **42**, 155
Tsai, H.T., C.F. Chen, Ultraviolet spectral identification of ferulic acid in rabbit plasma by HPLC and its pharmacokinetic application **80**, 75
Tsai, J.C., M.J. Cappel, N.D. Weiner, G.L. Flynn, J. Ferry, Solvent effects on the harvesting of stratum corneum from hairless mouse skin through adhesive tape stripping in vitro **68**, 127
Tsai, J.C., G.L. Flynn, N. Weiner, J.J. Ferry, Effect of minoxidil concentration on the deposition of drug and vehicle into the skin **96**, 111
Tsai, J.C., N.D. Weiner, G.L. Flynn, J. Ferry, Properties of adhesive tapes used for stratum corneum stripping **72**, 227
Tsai, P.K., see Burke, C.J. **86**, 89
Tsai, R.S., Tayar N. El, P.A. Carrupt, B. Testa, Physicochemical properties and transport behaviour of piribedil: Considerations on its membrane-crossing potential **80**, 39

Tsai, T., R.C. Mehta, P.P. DeLuca, Adsorption of peptides to poly(D,L-lactide-*co*-glycolide). 1. Effect of physical factors on the adsorption **127**, 31
Adsorption of peptides to poly(D,L-lactide-*co*-glycolide). 2. effect of solution properties on the adsorption **127**, 43
Tsai, T., see Ho, H.O. **139**, 223
Tsai, Y.H., L.R. Hsu, S.I. Naito, Percutaneous absorption of piroxicam from ointment bases in rabbits **24**, 61
Simultaneous determination of piroxicam and its main metabolite in plasma and urine by high-performance liquid chromatography **24**, 101
Tsai, Y.H., S.I. Naito, Simultaneous determination of indomethacin and its metabolites in rabbit plasma by high-pressure liquid chromatography **8**, 203
Changes in site and size of application of indomethacin ointment and percutaneous absorption in rabbits **10**, 125
Percutaneous absorption of phenylbutazone from ointment bases in rabbits **23**, 315
Tsai, T.H., C.C. Shen, C.F. Chen, Determination and UV spectral identification of 18α-glycyrrhetinic acid and 18β-glycyrrhetinic acid for stability studies **84**, 279
Tsai, Y.H., M.Y. Yang, S.I. Naito, Stability, anti-inflammatory effect and percutaneous absorption of indomethacin in absorption ointment formulations **28**, 47
Tsai, T.R., see Chuo, W.H. **134**, 247; **144**, 241
Tsai, Y.H., see Fang, J.Y. **126**, 119; **128**, 169; **130**, 121; **131**, 143; **143**, 47; **145**, 175; **149**, 183
Tsai, Y.H., see Hsu, L.R. **71**, 193; **106**, 1
Tsai, Y.H., see Huang, Y.B. **126**, 111; **131**, 137; **134**, 183
Tsai, Y.H., see Lin, H.H. **112**, 165
Tsai, Y.H., see Wu, P.C. **143**, 119; **145**, 215; **148**, 41
Tsang, S., see Hunt, C.A. **8**, 101
Tsantili-Kakoulidou, A., see Wernly-Chung, G.N. **63**, 129
Tsaprounis, C., see Koupparis, M. **27**, 349
Tsaprounis, C., see Macheras, P. **30**, 123; **33**, 125
Tsardaka, K.D., see Rees, J.E. **92**, 137
Tsau, J., J.W. Poole, Stoichiometric analysis by chromatographic techniques. I. Analysis of some antioxidants and penicillins by HPLC methods **12**, 185
Tsuchiya, S., Y. Aramaki, S. Ozawa, H. Matsumaru, Effects of polyoxyethylene cetyl ether on the absorption of 3′,4′-dideoxykanamycin B from rat rectum **14**, 279
Tsuchiya, S., see Hara, T. **42**, 69; **67**, 123
Tsuchiya, S., see Kikuchi, S. **42**, 193
Tsuchiya, S., see Niibuchi, J.J. **30**, 181
Tsuchiya, S., see Tanaka, S. **27**, 29
Tsuchiya, S., see Unezaki, S. **144**, 11
Tsuemura, Y., see Sakiya, Y. **25**, 347
Tsuge, H., see Gonsho, A. **65**, 183
Tsui, M., see Sitaram, B.R. **118**, 181
Tsuji, A., see Terasaki, T. **81**, 143
Tsujimoto, C., see Ogiso, T. **98**, 113
Tsunekawa, Y., see Hasegawa, T. **56**, 235
Tsunematsu, H., E. Ishida, S. Yoshida, M. Yamamoto, Synthesis and enzymatic hydrolysis of aspirin-basic amino acid ethyl esters **68**, 77
Tsunemi, S., see Oribe, T. **124**, 27
Tsunenari, Y., see Irie, T. **43**, 41
Tsuzuki, N., see Akazawa, M. **50**, 53
Tsuzuki, N., see Wong, O. **52**, 191
Tu, Y.H., L.V. Allen Jr., D.P. Wang, Stability of loperamide hydrochloride in aqueous solutions as determined by high performance liquid chromatography **51**, 157
Tu, Y.H., Y. Wang, L.V. Allen Jr., D.D. Albers, M.K. Gorgin, Pharmacokinetics of metronidazole administered intravenously to male rats **61**, 119
Tucker, I., see Schep, L.J. **113**, 263
Tucker, I.G., see Bunt, C.R. **99**, 93; **113**, 257
Tucker, I.G., see Jones, D.S. **126**, 281
Tucker, I.G., see Khan, M.Z.I. **76**, 161; **90**, 255
Tucker, I.G., see Koh, G.L. **34**, 183
Tucker, I.G., see Medlicott, N.J. **84**, 85; **143**, 25
Tucker, I.G., see Stark, G. **143**, 93
Tucker, S.A., see Acree, W.E. **101**, 199
Tudball, N., see Donnelly, A. **135**, 191
Tukker, J.J., see Lehr, C.M. **70**, 235
Tukker, J.J., see Poelma, F.G.J. **64**, 161
Tukker, J.J., see Sam, A.P. **79**, 97
Tumer, A., see Onur, M.A. **88**, 313
Tung, I.C., Rheological behavior of poloxamer 407 aqueous solutions during sol-gel and dehydration processes **107**, 85
Tuominen, V., see Laine, E. **20**, 307
Turakka, L., E. Toikkanen, Release of salicylic acid from aqueous triglyceride vehicles containing surfactants **17**, 99
Turco Liveri, M.L, see Turco, Liveri V. **99**, 285
Turco Liveri, V., G. Cavallaro, G. Giammona, G. Pitarresi, Liveri M.L Turco, Calorimetric and viscosimetric investigation of the interaction between α,β-polyasparthydrazide and sodium dodecyl sulfate micelles **99**, 285
Turgut, H., see Sener, E. **110**, 109
Turker, L., see Sener, E. **110**, 109
Turkistani, A.A.A., M. Gaber, M.A. Al-Meshal, H.I. Al-Shora, M.W. Gouda, Effect of sucralfate on procainamide absorption **59**, R1
Turkoglu, M., W.A. Ritschel, A. Sakr, In vivo evaluation of fluidized-bed coated pellets **103**, 115
Turkoglu, M., A. Sakr, Mathematical modelling and optimization of a rotary fluidized-bed coating process **88**, 75
Turkyilmaz, A., see Celebi, N. **136**, 89
Tushuizen, E., see Zanen, P. **81**, 199
Tyle, P., see Chiang, C.C. **117**, 197
Tyle, P., see Wilke, T.L. **103**, 187
Tymes, N.W., see Shah, V.P. **32**, 243

U

Uchegbu, I.F., see Gianasi, E. **148**, 139
Uchida, N., Y. Maitani, Y. Machida, M. Nakagaki, T. Nagai, Influence of bile salts on the permeability of insulin through the nasal mucosa of rabbits in comparison with dextran derivatives **74**, 95
Uchida, N., see Maitani, Y. **64**, 139; **69**, 21
Uchiyama, K., A. Nagayasu, Y. Yamagiwa, T. Nishida, H. Harashima, H. Kiwada, Effects of the size and fluidity of liposomes on their accumulation in tumors: A presumption of their interaction with tumors **121**, 195

Udagawa, C., see Hodoshima, N. **146**, 81
Udeani, G.O., see Wang, P.L. **114**, 177
Udupa, N., see Chandraprakash, K.S. **61**, R1
Ueda, C., see Katayama, N. **115**, 87
Ueda, H., M. Ogihara, K. Sugibayashi, Y. Morimoto, Change in the electrochemical properties of skin and the lipid packing in stratum corneum by ultrasonic irradiation **137**, 217
Ueda, H., R. Isshiki, M. Ogihara, K. Sugibayashi, Y. Morimoto, Combined effect of ultrasound and chemical enhancers on the skin permeation of aminopyrine **143**, 37
Ueda, H., see Loftsson, T. **110**, 169
Ueda, M., see Kawashima, Y. **18**, 335
Ueda, S., see Hata, T. **110**, 1
Uekama, K., T. Fujinaga, F. Hirayama et al., Inclusion complexations of steroid hormones with cyclodextrins in water and in solid phase **10**, 1
Uekama, K., S. Narisawa, F. Hirayama et al., Improvement of thermal and photochemical stability of benzaldehyde by cyclodextrin complexation **13**, 253
Uekama, K., S. Narisawa, F. Hirayama, M. Otagiri, Improvement of dissolution and absorption characteristics of benzodiazepines by cyclodextrin complexation **16**, 327
Uekama, K., K. Oh, T. Irie et al., Stabilization of isosorbide 5-mononitrate in solid state by β-cyclodextrin complexation **25**, 339
Uekama, K., M. Otagiri, T. Irie et al., Improvement of dissolution and absorption characteristics of phenytoin by a water-soluble β-cyclodextrin-epichlorohydrin polymer **23**, 35
Uekama, K., see Abe, K. **123**, 103
Uekama, K., see Arima, H. **57**, 107
Uekama, K., see Hirayama, F. **35**, 193
Uekama, K., see Hoshino, T. **38**, 265; **50**, 45; **98**, 239
Uekama, K., see Irie, T. **43**, 41; **84**, 129
Uekama, K., see Kaji, Y. **24**, 79
Uekama, K., see Kikuchi, M. **38**, 191
Uekama, K., see Soliman, O.A. **149**, 73
Uekama, K., see Yamamoto, M. **49**, 163
Uematsu, T., see Oguma, T. **63**, 101
Ueno, M., see Liu, H.X. **118**, 55
Uenoyama, S., see Aioi, A. **93**, 1
Uesugi, K., see Aoki, S. **95**, 67; **85**, 29, 65
Ugazio, E., see Morel, S. **132**, 259
Ugazio, E., see Trotta, M. **143**, 67
Ugwoke, M.I., N. Verbeke, R. Kinget, Microencapsulation of apomorphine HCl with gelatin **148**, 23
Ugwu, S.O., E.L. Lan, S. Sharma, V. Hruby, J. Blanchard, Kinetics of degradation of a cyclic lactam analog of α-melanotropin (MT-II) in aqueous solution **102**, 193
Ugwu, S.O., see Surendran, N. **135**, 81
Ukai, M., see Ohta, N. **124**, 165
Ulbrich, K., see Morgan, S.M. **128**, 99
Ulman, K., see Toddywala, R.D. **76**, 77
Ulrich, K.K., see Reynolds, D.L. **9**, 67
Ulutin, S., see Gursoy, A. **52**, 101
Ulvi, V., see Torniainen, K. **132**, 53
Umadevi, P., see Chandraprakash, K.S. **61**, R1
Umapathi, P., Determination of atenolol, nifedipine, aspirin and dipyridamole in tablet preparations by second-order derivative spectrophotometry **108**, 11
Umapathi, P., see Parimoo, P. **100**, 227
Umprayn, K., see Stella, V.J. **43**, 191
Umprayn, K., W. Waugh, V.J. Stella, Mechanism of degradation of the investigational cytotoxic agent, cyclodisone (NSC-348948) **66**, 253
Underberg, W.J.M., U.K. Underberg-Chitoe, O. Bekers, P.L. Meenhorst, J.H. Beijnen, A rapid, simple and accurate method for the bioanalysis of zidovudine **50**, 175
Underberg, W.J.M., see Beijnen, J.H. **31**, 75; **32**, 111, 123; **24**, 219; **41**, 169
Underberg, W.J.M., see Bekers, O. **56**, 103; **68**, 271; **72**, 123
Underberg, W.J.M., see de Vries, J.D. **100**, 181
Underberg, W.J.M., see Suenaga, A. **59**, 121; **82**, 29
Underberg, W.J.M., see Van der Houwen, O.A.G.J. **67**, 155; **89**, R5; **109**, 191
Underberg-Chitoe, U.K., see Underberg, W.J.M. **50**, 175
Unezaki, S., K. Maruyama, J.I. Hosoda, I. Nagae, Y. Koyanagi, M. Nakata, O. Ishida, M. Iwatsuru, S. Tsuchiya, Direct measurement of the extravasation of polyethyleneglycol-coated liposomes into solid tumor tissue by in vivo fluorescence microscopy **144**, 11
Unezaki, S., K. Maruyama, O. Ishida, A. Suginaka, J.I. Hosoda, M. Iwatsuru, Enhanced tumor targeting and improved antitumor activity of doxorubicin by long-circulating liposomes containing amphipathic poly(ethylene glycol) **126**, 41
Unlu, N., see Ludwig, A. **61**, 15
Uno, T., see Murai, Y. **15**, 309
Uno, T., see Yamaoka, K. **10**, 291
Upadhyay, A.H., see Patel, N.K. **110**, 203
Upadhyay, P., see Misra, A. **132**, 267
Upadrashta, S.M., P.R. Katikaneni, G.A. Hileman, S.H. Neau, C.E. Rowlings, Compressibility and compactibility properties of ethylcellulose **112**, 173
Upadrashta, S.M., D.E. Wurster, Equilibrium binding studies of the interaction between anthralin and bovine serum albumin **49**, 103
Upadrashta, S.M., see Goskonda, S.R. **111**, 89
Upadrashta, S.M., see Hileman, G.A. **100**, 71
Upadrashta, S.M., see Katikaneni, P.R. **117**, 13; **123**, 119
Upadrashta, S.M., see Wurster, D.E. **55**, 221
Uppadhyay, R.K., see Tripathi, M. **67**, 207
Uppoor, S., see Ramana, V. **145**, 145
Upton, R.A., see Collier, P.S. **26**, 99
Uriarte, E., see Fernandez, B. **79**, 199
Urien, S., see Bonhomme-Faivre, L. **134**, 99
Urleb, U., see Srcic, S. **87**, 1
Urtti, A., Pilocarpine release from matrices of alkyl half-esters of poly(vinyl methyl ether/maleic anhydride) **26**, 45
Urtti, A., M. Juslin, O. Miinalainen, Pilocarpine release from hydroxypropyl-cellulose-polyvinylpyrrolidone matrices **25**, 165
Urtti, A., J.D. Pipkin, G. Rork, A.J. Repta, Controlled drug delivery devices for experimental ocular studies with timolol. 1. In vitro release studies **61**, 235

Urtti, A., J.D. Pipkin, G. Rork, T. Sendo, U. Finne, A.J. Repta, Controlled drug delivery devices for experimental ocular studies with timolol. 2. Ocular and systemic absorption in rabbits **61**, 241
Urtti, A., L. Salminen, H. Kujari, V. Jantti, Effect of ocular pigmentation on pilocarpine pharmacology in the rabbit eye. II. Drug response **19**, 53
Urtti, A., L. Salminen, O. Miinalainen, Systemic absorption of ocular pilocarpine is modified by polymer matrices **23**, 147
Urtii, A., see Sutinen, R. **92**, 177
Urtti, A., see Finne, U. **65**, 19; **75**, R1; **78**, 237; **84**, 217
Urtti, A., see Hirvonen, J. **99**, 253
Urtti, A., see Jarho, P. **137**, 209
Urtti, A., see Jarvinen, T. **75**, 249, 259; **79**, 233, 243
Urtti, A., see Kyyronen, K. **80**, 161
Urtti, A., see Marjukka, Turunen T. **92**, 89
Urtti, A., see Monkkonen, J. **107**, 189
Urtti, A., see Saarinen-Savolainen, P. **133**, 171
Urtti, A., see Salminen, L. **18**, 17
Urtti, A., see Suhonen, P. **74**, 221; **127**, 85
Urtti, A., see Sutinen, R. **57**, 149, 155; **62**, 113
Urtti, A., see Tirkkonen, S. **124**, 219
Ushio, Y., see Oguma, T. **63**, 101
Ushirogawa, Y., Y. Nakahigashi, A. Kiriyama, K. Takada, Effect of organic acids, trypsin inhibitors and dietary protein on the pharmacological activity of recombinant human granulocyte colony-stimulating factor (rhG-CSF) in rats **81**, 133
Ushirogawa, Y., see Takada, K. **101**, 89
Usteri, M., H. Leuenberger, Agglomeration of binary mixtures in a high-speed mixer **55**, 135
Utsumi, E., see Marszall, L. **39**, 263
Utsumi, E., see Yamamoto, A. **51**, 137

V

Vachon, M.G., D.J.W. Grant, Enthalpy–entropy compensation in pharmaceutical solids **40**, 1
Vadas, E.B., see Clas, S.D. **121**, 73
Vadas, E.B., see Cotton, M.L. **40**, 129; **109**, 237
Vadas, E.B., see Kwong, E.C. **103**, 259
Vadas, E.B., see Thibert, R. **134**, 59
Vadiei, K., G. Lopez-Berestein, R. Perez-Soler, D.R. Luke, In vitro evaluation of liposomal cyclosporine **57**, 133
Vadiei, K., R. Perez-Soler, G. Lopez-Berestein, D.R. Luke, Pharmacokinetic and pharmacodynamic evaluation of liposomal cyclosporine **57**, 125
Vadino, W.A., see Sangekar, S. **35**, 187
Vadnere, M., G. Amidon, S. Lindenbaum, J.L. Haslam, Thermodynamic studies on the gel-sol transition of some pluronic polyols **22**, 207
Vadnere, M., see Adjei, A. **90**, 141
Vagen, S.R., see Henriksen, I. **146**, 193
Vaidyanathan, R., M.G. Chaubal, R.C. Vasavada, Effect of pH and solubility on in vitro skin penetration of methotrexate from a 50% v/v propylene glycol-water vehicle **25**, 85
Vaillant, L., see Machet, L. **72**, R1
Vaillant, L., see Pelucio-Lopes, C. **96**, 249
Vainio, P., see Vidgren, M. **64**, 1; **96**, 183; **73**, 131
Vaisanen, V., see Finne, U. **65**, 19
Vakaet, L., see Bouckaert, S. **130**, 257
Valencia, G., see Reig, F. **44**, 257, 269
Valenta, C., A. Bernkop-Schnurch, M. Schwartz, J. Kerc, Modification of lysozyme with cinnamaldehyde: A strategy for constructing novel preservatives for dermatics **148**, 131
Valiere, C., P. Arnaud, E. Caroff, J.F. Dauphin, G. Clement, F. Brion, Stability and compatibility study of a carboplatin solution in syringes for continuous ambulatory infusion **138**, 125
Valjakka, R., see Monkkonen, J. **107**, 189
Valko, K., I. Toth, P. Ward, P. Slegel, W.A. Gibbons, Lipidic peptides. XI. Quantitative structure–activity relationships of a series of lipidic amino acid conjugates of β-lactam antibiotics **83**, 123
Vallee, P., see Stella, V. **121**, 117
Vallner, J.J., I.L. Honigberg, J.A. Kotzan, J.T. Stewart, A proposed general protocol for testing bioequivalence of controlled-release drug products **16**, 47
Vallner, J.J., see Bayoumi, S.M. **30**, 57
Valls, O., see Egea, M.A. **67**, 103
Valls, O., see Hernandez, J. **57**, 211
Valls, O., see Montero, M.T. **62**, 21
Vamos, J., see Volford, O. **137**, 111
Van Aerde, P., J.P. Remon, In vitro evaluation of modified starches as matrices for sustained release dosage forms **45**, 145
Van Amelsfoort, J.G.M, see Fokkens, J.G. **14**, 79
Van Amerongen, I.A., H.A. de Ronde, N.T.M. Klooster, Physical-chemical characterization of semisolid topical dosage form using a new dissolution system **86**, 9
Van Bennekom, W.P., see Leeuwenkamp, O.R. **24**, 27
Van Bloois, L., see Crommelin, D.J.A. **16**, 79; **17**, 135
Van Bloois, L., see Grijseels, H. **14**, 299
Van Bommel, E.M.G., D.J.A. Crommelin, Stability of doxorubicin-liposomes on storage: As an aqueous dispersion, frozen or freeze-dried **22**, 299
Van Bommel, E.M.G., R.F.R. Dezentje, D.J.A. Crommelin, J.G. Fokkens, Drug release kinetics from the gradient matrix system: Mathematical modelling **72**, 19
Van Bommel, E.M.G., see Baert, L. **99**, 7
Van Bork, L.E., see Jonkman, J.H.G. **3**, 55
Van Criekinge, W., see Van der Meeren, P. **106**, 89
Van Damme, M., see Abi Khalil, F. **140**, 1
Van Damme, M., see Verbiese-Genard, N. **2**, 155; **9**, 295
Van de Donk, H., S. Jadoenath, J. Zuidema, F.W.H.M. Merkus, The effects of drugs on ciliary motility. I. Decongestants **12**, 57
Van de Donk, H., A.L Van Egmond, A.G.M. Van den Heuvel et al., The effects of drugs on ciliary motility. III. Local anaesthetics and anti-allergic drugs **12**, 77
Van de Donk, H., A.L Van Egmond, J. Zuidema, F.W.H.M. Merkus, The effects of drugs on ciliary motility. II. Antimicrobial agents **12**, 67
Van de Griend, J., see Beijersbergen, van Henego **35**, 259

Van de Waterbeemd, H., B. Testa, The development of a hydration factor omega and its relation to correction terms in current hydrophobic fragmental systems **14**, 29
Van de Waterbeemd, H., see Camenisch, G. **147**, 61
Van de Waterbeemd, H., see de Haan, F.H.N **13**, 75
Van de Waterbeemd, H., see El Taylor, N. **19**, 271
Van de Waterbeemd, H., see Gryllaki, M. **51**, 95
Van de Waterbeemd, H., see Repond, C. **38**, 47
Van de Waterbeemd, H., see Rothlisberger, T. **51**, 73
Van de Zijde, H.J., see Beijersbergen van Henegouwen, G.M.J. **35**, 259
Van den Bergh, B.A.I., see Bouwstra, J.A. **144**, 81
Van den Heuij, J.T.M., see Sam, A.P. **79**, 97
Van den Heuvel, A.G.M., see Van de Donk H.J.M. **12**, 77
Van den Mooter, G., C. Samyn, R. Kinget, Azo polymers for colon-specific drug delivery **87**, 37
Azo polymers for colon-specific drug delivery. II. Influence of the type of azo polymer on the degradation by intestinal microflora **97**, 133
Characterization of colon-specific azo polymers. A study of the swelling properties and the permeability of isolated polymer films **111**, 127
Van den Mooter, G., see Cachet, T. **55**, 59
Van den Mooter, G., see Kalala, W. **139**, 187
Van den Mooter, G., see Samyn, C. **121**, 211
Van den Mooter, G., see Vinckier, C. **55**, 67
Van der Boon, W.J.V., see Jonkman, J.H.G. **25**, 113
Van der Giesen, W., see O'Neal, J.S. **12**, 355
Van der Giesen, W.F., L.H.M. Janssen, Influence of ionization and ion-pair formation on lipophilicity of some 4-hydroxycoumarin derivatives in the octanol-water system **12**, 231
Van der Griend, J., J. Juffermans, G.M.J. Beijersbergen van Henegouwen, K.W. Gerritsma, Determination of diphenhydramine in photochemically decomposed solutions **1**, 257
Van der Houwen, O.A.G.J., J.H. Beijen, A. Bult, O. Bekers, R.M.J. Goossen, W.J.M. Underberg, Degradation kinetics of antitumour drug nimustine (ACNU) in aqueous solution **99**, 73
Van der Houwen, O.A.G.J., J.H. Beijnen, A. Bult, W.J.M. Underberg, A general approach to the interpretation of pH buffer catalyzed degradation profiles **109**, 191
Van der Houwen, O.A.G.J., O. Bekers, A. Bult, J.H. Beijnen, W.J.M. Underberg, Computation of mitomycin C-γ-cyclodextrin complex stability constant **89**, R5
Van der Houwen, O.A.G.J., O. Bekers, J.H. Beijnen, A. Bult, S.C.M. Lubbers, W.J.M. Underberg, Degradation kinetics of the investigational antitumor compound LL-D49194α_1 in the presence of buffers **112**, 97
Van der Houwen, O.A.G.J., O. Bekers, J.H. Beijnen, A. Bult, W.J.M. Underberg, A general approach to the interpretation of pH degradation profiles in the presence of ligands **67**, 155
Van der Houwen, O.A.G.J., J. Teeuwse, O. Bekers, J.H. Beijnen, A. Bult, W.J.M. Underberg, Degradation kinetics of 7-*N*-(*p*-hydroxyphenyl)mitomycin C (M-83) in aqueous solution in the presence of γ-cyclodextrin **105**, 249
Van der Houwen, O.A.G.J., see Beijnen, J.H. **31**, 75; **32**, 123; **41**, 169
Van der Houwen, O.A.G.J., see Vendrig, D.E.M.M. **43**, 131; **50**, 189
Van der Kolk, H., see Zanen, P. **81**, 199
Van der Mark E.J., see Leeuwenkamp, O.R. **24**, 27; **33**, 1
Van der Meeren, P., W. Van Criekinge, J. Vanderdeelen, L. Baert, Phospholipid composition of r-DNA hepatitis B surface antigens **106**, 89
Van der Merwe, E., C. Ackermann, Wyk C.J. Van, Factors affecting the permeability of urea and water through nude mouse skin in vitro. I. Temperature and time of hydration **44**, 71
Van der Spek, J.A., see Bouwstra, J.A. **84**, 205
Van der Veen, J., H.H. Buitendijk, C.F. Lerk, Dissolution kinetics of ketanserin tartrate, the salt of a weakly basic drug **87**, 161
Van der Voet, H., P. de Haan, D.A. Doornbos, The use of the Durbin–Watson statistic for testing the validity of kinetic models for dissolution **14**, 291
Van der Voort Maarschalk, K., K. Zuurman, H. Vromans, G.K. Bolhuis, C.F. Lerk, Porosity expansion of tablets as a result of bonding and deformation of particulate solids **140**, 185
Van der Watt, J.G., The effect of the particle size of microcrystalline cellulose on tablet propers in mixtures with magnesium stearate **36**, 51
Van der Watt, J.G., see Danckwerts, M.P. **123**, 85
Van der Watt, J.G., see de Villiers M.M. **88**, 275
Van der Watt, J.G., see Gerber, J.J. **69**, 265
Van Deurs, B., see Jacobsen, J. **125**, 165
Van Dooremalen, J.A.M., see Tomlinson, E. **12**, 87
Van Dooren, A.A, B.W. Muller, Purity determinations of drugs with differential scanning calorimetry (DSC)—a critical review **20**, 217
Van Doorne, H., E.H. Bosch, Stability and in vitro activity of nystatin and its γ-cyclodextrin complex against *Candida albicans* **73**, 43
Van Doorne, H., see Bootsma, H.P.R. **51**, 213
Van Doorne, H., see Bos, C.E. **55**, 175
Van Egmond, A.L, see Van de Donk H. **12**, 67, 77
Van Gansbeke, B, see Beten, D.B. **123**, 145
Van Helden, S.P, see Jansen, A.C.A. **75**, 193
Van Henegouwen, G.M.J.B., see Cornelissen, P.J.G. **1**, 173
Van Henegouwen, G.M.J.B., see Van der Griend, J. **1**, 257
Van Houtven, C., see Hendrix, C. **100**, 213
Van Kamp, H.V., G.K. Bolhuis, K.D. Kussendrager, C.F. Lerk, Studies on tableting properties of lactose. IV. Dissolution and disintegration properties of different types of crystalline lactose **28**, 229
Van Koevering, C.L., see Gordon, R.E. **21**, 99
Van Laarhoven, J.A.H., see Horsthuis, G.J.B. **92**, 143
Van Laarhoven, J.A.H., see Vromans, H. **79**, 301
Van Ooijen, R.D, see Beijnen, J.H. **34**, 247
Van Ooteghem, M., see de Beukelaer, P. **16**, 345
Van Ooteghem, M., see Ludwig, A. **54**, 95; **61**, 15
Van Rooij, H.H., see Grunbauer, H.J.M. **32**, 187

Van Rooij, H.H., see Tomlinson, E. **12**, 87
Van Rooij, R.C., see Horsthuis, G.J.B. **92**, 143
Van Savage, G., see Yang, S.T. **86**, 247
Van Schepdael, A., N. Ossembe, L. Jie, P. Herdewijn, E. Roets, J. Hoogmartens, Kinetics of the hydrolysis of 2′,3′-dideoxyguanosine: A potent anti-HIV agent **73**, 105
Van Schie, D.M., J.G. Winchester, Stabilisation of a protein prodrug formulation: A novel approach **93**, 177
Van Scoik, K.G., J.T. Carstensen, Nucleation phenomena in amorphous sucrose systems **58**, 185
Van Spiegel, P.I., see Zanen, P. **81**, 199
Van Steenbergen, M.J., see Talsma, H. **77**, 119
Van Tonder, E.C., M.R. Caira, S.A. Botha, A.P. Lotter, Polymorphism of doxylamine succinate and X-ray structural study of Form I and of doxylamine succinate 0.5 succinic acid **63**, 35
Van Vooren, J.P, see Benameur, H. **89**, 157
Van Vugt, M., see Cremers, H.F.M. **110**, 117
Van Wyk, C.J., see Du Plessis, J. **34**, 173
Van Wyk, C.J., see Van der Merwe, E. **44**, 71
Vandamme, Th.F., Mukendi J. Ngombo, Controlled release of levamisole from poly-(ε-caprolactone) matrices. II. Effects of water-soluble polymer and iron powder incorporated into the matrices **132**, 153
Controlled release of levamisole from poly-(ε-caprolactone) matrices. III. Effects of molecular weight and polymer coating on drug release **145**, 77
Vandelli, M.A., G. Salvioli, A. Mucci, R. Panini, L. Malmusi, F. Forni, 2-Hydroxypropyl-β-cyclodextrin complexation with ursodeoxycholic acid **118**, 77
Vandelli, M.A., R. Cameroni, Selective coating of cylindrical matrices with a central hole. I. An interpretation of the swelling process **100**, 107
Vandelli, M.A., see Forni, F. **60**, 83
Vandenbossche, G., see Voorspoels, J. **133**, 9
Vandenbossche, G.M.R., E. Vanhaecke, C. de Muynck, J.P. Remon, Stability of topical erythromycin formulations **67**, 195
Vander Velde, D., see Antipas, A.S. **109**, 261
Vanderdeelen, J., see Van der Meeren, P. **106**, 89
VanderWatt, J.G., see Gerber, J.J. **73**, 137
Vanelli, M.A., G. Coppi, R. Cameroni, Selective coating of cylindrical matrices with a central hole. II. An interpretation of the release process **100**, 115
Vanhaecke, E., see Vandenbossche, G.M.R. **67**, 195
Vargas, R., see McMahon, F.G. **91**, 75
Vargha-Butler, E.I., see Chow, A.H.L. **126**, 21
Vargha-Butler E.I, see Krishnan, T.R. **80**, 277
Varia, S.A., see Desai, D.S. **91**, 217; **103**, 69; **107**, 141; **110**, 249, 257; **142**, 61; **144**, 153
Varia, S.A., see Morris, K.R. **108**, 195
Varia, S.A., see Williams, D.B. **14**, 113
Vasantha, R., P.K. Sehgal, K.P. Rao, Collagen ophthalmic inserts for pilocarpine drug delivery system **47**, 95
Vasavada, R.C., see Chou, C.K.H. **25**, 199
Vasavada, R.C., see Iyer, B.V. **3**, 247
Vasavada, R.C., see Mummaneni, V. **66**, 71
Vasavada, R.C., see Rao Jasti, B. **118**, 161
Vasavada, R.C., see Vaidyanathan, R. **25**, 85
Vasiliev, A.E., see Feldstein, M.M. **131**, 229
Vasiltchenko, M.A., see Shakhtshneider, T.P. **130**, 25
Vauthier, C., see Belbella, A. **129**, 95
Vauthier, C., see Pense, A.M. **81**, 111
Vaution, C., J. Paris, F. Puisieux, J.T. Carstensen, Solubilization of phenobarbital in sodium paraffin sulfonate **1**, 349
Vaution, C., see Jansen, J. **103**, 19
Vayumhasuwan, P., see Chen, Y. **137**, 149
Vayumhasuwan, P., see Matheson, L.E. **77**, 163
Vazquez, J.L., see Montero, M.T. **138**, 113
Veber, M., see Sher, A. **90**, 181; **99**, 119; **148**, 191
Veenstra, J., see Riepma, K.A. **76**, 9
Veiga, F., J.J.C. Texeira-Dias, F. Kedzierewicz, A. Sousa, P. Maincent, Inclusion complexation of tolbutamide with β-cyclodextrin and hydroxypropyl-β-cyclodextrin **129**, 63
Veiga, F., see Simoes, S. **131**, 273
Veiga, M., F. Gutierrez, Mathematical transformation of data, fit, and graphic representation of regression curves through computer programming **79**, 1
Veiga, M.D., C. Escobar, M.J. Bernad, Dissolution behaviour of drugs from binary and ternary systems **93**, 215
Veiga, M.D., J. Alvarez de Eulate, Dissolution study of spiramycin: Influence of agitation intensity and addition of several substances to the dissolution medium **110**, 223
Veiga, M.D., M.J. Bernad, C. Escobar, Thermal behaviour of drugs from binary and ternary systems **89**, 119
Veiga, M.D., see Frances, C. **77**, 193
Veiga, M.D., see Gines, J.M. **126**, 287
Veillard, M., see Coutel-Egros, A. **84**, 117
Veillard, M., see Husson, I. **86**, 113
Veje, J., see Hansen, J. **16**, 141
Vejosoth, S., see Mesiha, M. **111**, 213
Vela, M.T., see Fernandez-Hervas, M.J. **113**, 39; **130**, 115
Vela, M.T., see Holgado, M.A. **142**, 143
Velasco Antequera, M.V., A. Munoz-Ruiz, Perales M. Monedero, Munoz N. Munoz, Ballestero Jimenez-Castellanos, Evaluation of an adequate method of estimating flowability according to powder characteristics **103**, 155
Velasco Antequera, M.V., see Monedero Perales, M.C. **132**, 183
Velasco Antequera, M.V., see Munoz-Ruiz, A. **95**, 201
Velasco, M.V., A. Munoz, M.R. Jimenez-Castellanos, I. Castellano, I. Goni, M. Gurrachaga, In vitro evaluation of sustained-release matrix tablets prepared with new modified polymeric carbohydrates **136**, 107
Velasco, M.V., see Castellano, I. **146**, 71
Velasco, M.V., see Ferrero, C. **147**, 11
Velasco, M.V., see Ferrero, M.C. **149**, 233
Velasco, M.V., see Velasco, M.V. **136**, 107
Velasco, V., see Munoz-Ruiz, A. **144**, 147
Velissaratou, A.S., G. Papaioannou, In vitro release of chlorpheniramine maleate from ointment bases **52**, 83
Vendrig, D.E.M.M., J.H. Beijnen, der Houwen Van, J.J.M. Holthuis, Degradation kinetics of vincristine sulphate and vindesine sulphate in aqueous solutions **50**, 189

Vendrig, D.E.M.M., B.P.G.M. Smeets, J.H. Beijnen, O.A.G.J. Van der Houwen, J.J.M. Holthuis, Degradation kinetics of vinblastine sulphate in aqueous solutions **43**, 131
Venho, V.M.K., H. Eriksson, The rabbit: An animal model for comparative bioavailability studies of drugs **30**, 91
Venkata Rao, J., see Subrahmanyam, C.V.S. **78**, 17
Venkataram, S., see Joseph, I. **126**, 161
Venkataram, S., see Kaul, D. **88**, 345
Venkataramanan, R., see D'Souza, M. **31**, 165
Venkatesh, S., see Suryanarayanan, R. **78**, 77
Venkatraghavan, V., see Bodor, N. **53**, 195
Venthoye, G., see Stelliou, I. **99**, R1
Venthoye, G., see Taylor, K.M.G. **85**, 203
Ventimiglia, A., see Montenegro, L. **133**, 89
Ventouras, K., see Gander, B. **27**, 117
Ventura, C., see Giammona, G. **105**, 57
Ventura, C.A., S. Tirendi, G. Puglisi, E. Bousquet, L. Panza, Improvement of water solubility and dissolution rate of ursodeoxycholic acid and chenodeoxycholic acid by complexation with natural and modified β-cyclodextrins **149**, 1
Ventura, C.A., see Castelli, F. **88**, 1
Ventura, C.A., see Puglisi, G. **125**, 283
Ventura, P., see Pitcairn, G. **102**, 11
Ventura, P., see Redenti, E. **129**, 289
Ventura, P., see Steed, K.P. **112**, 199
Vepsalainen, J., see Jarvinen, T. **75**, 249; **79**, 233
Verbeke, N., see Ugwoke, M.I. **148**, 23
Verbiese-Genard, N., M. Hanocq, Damme M. Van, L. Molle, Separation, identification and quantitative analysis of substituted benzamides in pharmaceutical dosage forms by high pressure liquid chromatography **2**, 155
High performance liquid chromatographic capacity factor and partition coefficient relationship in a benzamide series **9**, 295
Vergnaud, J.M., Liquid transport controlled release processes in polymeric materials: Applications to oral dosage forms **90**, 89
Vergnaud, J.M., see Bakhouya, A. **143**, 143; **146**, 225
Vergnaud, J.M., see Bidah, D. **58**, 215; **72**, 35; **77**, 81; **80**, 145
Vergnaud, J.M., see Chafi, N. **52**, 203; **67**, 265
Vergnaud, J.M., see Eddine, A. **32**, 143
Vergnaud, J.M., see Kolli, M. **81**, 103
Vergnaud, J.M., see Laghoueg, N. **50**, 133
Vergnaud, J.M., see Laghoueg-Derriche, N. **67**, 51, 163; **71**, 229
Vergnaud, J.M., see Magron, P. **38**, 91
Vergnaud, J.M., see Nia, B. **119**, 165
Vergnaud, J.M., see Ouriemchi, E.M. **102**, 47; **113**, 231; **127**, 177
Vergnaud, J.M., see Sebert, P. **110**, 285; **122**, 129
Verheijden, J.H.M., see Kadir, F. **64**, 171
Verhoef, J., see Hoogstraate, A.J. **76**, 37
Verhoef, J., see Schipper, N.G.M. **64**, 61
Verhoef, J.C., see Akiyama, Y. **136**, 155; **138**, 13
Verhoef, J.C., see de Vries, M. **76**, 25
Verhoef, J.C., see Luessen, H.L. **141**, 39
Verhoef, J.C., see Marttin, E. **141**, 151
Verhoef, J.C., see Romeijn, S.G. **135**, 137
Verhoeven, J., L.J.C. Peschier, M. Danhof, H.E. Junginger, A controlled-release matrix tablet of furosemide: Design, in vitro evaluation, pharmacological and pharmacodynamic evaluation **45**, 65
Verma, N., see Buckton, G. **72**, 157
Verma, P.K., see Razdan, B. **79**, 83
Vermeersch, H., J.P. Remon, D. Permentier, E. Schacht, In vitro antitrichomonal activity of water-soluble prodrug esters of metronidazole **60**, 253
Vermeersch, H., see Baert, L. **96**, 225
Vermeersch, H., see Inghelbrecht, S. **148**, 103
Vermeersch, H., see Vervaet, C. **106**, 157
Vermehren, C., H.S. Hansen, M.K. Thomsen, Time dependent effects of two absorption enhancers on the nasal absorption of growth hormone in rabbits **128**, 239
Vermeij, P., see Blok, D. **73**, 171
Vermeire, A., see Kiekens, A. **146**, 239
Vermeire, A., J.P. Remon, The solubility of morphine and the stability of concentrated morphine solutions in glass, polypropylene syringes and PVC containers **146**, 213
Veronico, M., G. Ragno, G. Carlucci, C. Vetuschi, Roxatidine estimation in human fluids by UV derivative spectrophotometry **119**, 109
Veronico, M., see Ragno, G. **99**, 351; **119**, 115
Verrijk, R., see Cremers, H.F.M. **120**, 51
Verstraeten, A.E., see Vincke, B.J. **54**, 181
Vert, M., see Illum, L. **26**, 113
Vert, M., see Le Ray, A.M. **106**, 201
Vertommen, J., P. Rombaut, R. Kinget, Shape and surface smoothness of pellets made in a rotary processor **146**, 21
Vertommen, J., see Hoa Ngo, T. **146**, 271
Vervaet, C., L. Baert, J.P. Remon, Enhancement of in vitro drug release by using polyethylene glycol 400 and PEG-40 hydrogenated castor oil in pellets made by extrusion/spheronisation **108**, 207
Extrusion-spheronisation. A literature review **116**, 131
Vervaet, C., L. Baert, P.A. Risha, J.P. Remon, The influence of the extrusion screen on pellet quality using an instrumented basket extruder **107**, 29
Vervaet, C., H. Vermeersch, M.S. Khotz, L. Massart, J.P. Remon, Parameters influencing granule quality using a continuous granulator **106**, 157
Vervaet, C., J.P. Remon, Influence of impeller design, method of screen perforation and perforation geometry on the quality of pellets made by extrusion-spheronisation **133**, 29
Vervaet, C., see Ntawukuliyayo, J.D. **139**, 79
Vervaet, C., see Zhou, F. **133**, 155; **147**, 23
Vervoort, L., R. Kinget, In vitro degradation by colonic bacteria of inulinHP incorporated in Eudragit RS films **129**, 185
Vervoort, L., see de Vos, A. **92**, 191
Veski, P., M. Marvola, J. Smal, I. Heiskanen, H. Jurjenson, Biopharmaceutical evaluation of pseudoephedrine hydrochloride capsules containing different grades of sodium alginate **111**, 171
Veski, P., see Sirkia, T. **107**, 179

Vesterhus, L., see Ostberg, T. **97**, 183
Vetuschi, C., G. Ragno, Fourth UV derivative spectrophotometry for the simultaneous assay of atenolol and chlorthalidone in pharmaceuticals **65**, 177
Vetuschi, C., see Carlucci, G. **49**, 79; **102**, 271
Vetuschi, C., see Ragno, G. **99**, 351; **119**, 115
Vetuschi, C., see Veronico, M. **119**, 109
Vezin, W.R., A.T. Florence, The determination of dissociation constants and partition coefficients of phenothiazine derivatives **3**, 231
Vialard, A.P., see Contreras Claramonte, M.D. **94**, 23
Vialard, A.P., see Morell, J.L.P. **137**, 49
Viana, B., see Vila-Jato, J.L. **30**, 229
Vianna, R.F., see Bentley, M.V.L.B. **146**, 255
Vicente, M.T., see Calvo, M.B. **67**, 259
Vicente, M.T., see Lanao, J.M. **86**, 123; **117**, 113
Vicente, M.T., see Marino, E.L. **16**, 23
Vicente, M.T., see Sanchez Camazano, M. **6**, 243
Vickery, R.D., see Maurin, M.B. **94**, 11; **144**, 99
Victorov, A.V., see Kosykh, V.A. **57**, 223
Vidgren, M., A. Karkkainen, P. Karjalainen, P. Paronen, J. Nuutinen, Effect of powder inhaler design on drug deposition in the respiratory tract **42**, 211
Vidgren, M., P. Paronen, P. Vainio, K. Bergstrom, P. Pikkarainen, Gastrointestinal transit of ^{99m}Tc-labelled oral dosage forms of sucralfate in healthy volunteers **96**, 183
Vidgren, M., P. Paronen, P. Vidgren, P. Vainio, J. Nuutinen, Radiotracer evaluation of the deposition of drug particles inhaled from a new powder inhaler **64**, 1
Vidgren, M., J.C. Waldrep, J. Arppe, M. Black, J.A. Rodarte, W. Cole, V. Knight, A study of ^{99m}technetium-labelled beclomethasone dipropionate dilauroylphosphatidylcholine liposome aerosol in normal volunteers **115**, 209
Vidgren, M., see Vidgren, P. **73**, 131
Vidgren, M.T., A. Karkkainen, A. Karkkainen, P. Karjalainen, Effect of extension devices on the drug deposition from inhalation aerosols **39**, 107
Vidgren, M.T., A. Karkkainen, P. Karjalainen, T.P. Paronen, A novel labelling method for measuring the deposition of drug particles in the respiratory tract **37**, 239
Vidgren, M.T., P. York, T.P. Paronen, P. Karjalainen, Respiratory tract deposition of ^{99m}Tc-labelled drug particles administered via a dry powder inhaler **39**, 101
Vidgren, M.T., P.A. Vidgren, T.P. Paronen, Comparison of physical and inhalation properties of spray-dried and mechanically micronized disodium cromoglycate **35**, 139
Vidgren, P., M. Vidgren, P. Vainio, J. Nuutinen, P. Paronen, Double-labelling technique in the evaluation of nasal mucoadhesion of disodium cromoglycate microspheres **73**, 131
Vidgren, P., see Vidgren, M. **64**, 1
Vidgren, P.A., see Vidgren, M.T. **35**, 139
Vidmar, T.J., see Scieszka, J.F. **45**, 165
Vierling, P., see Privitera, N. **120**, 73
Vigo, J., see Lucero, M.J. **104**, 49; **106**, 125; **110**, 241; **111**, 261; **116**, 31; **127**, 73; **140**, 251
Vikholm, I., see Kivinen, A. **108**, 109
Vila-Dorrio, B., see Blanco-Fuente, H. **138**, 103
Vila-Jato, J.L., A. Concheiro, B. Seijo, B. Viana, Aging of nitrofurantoin tablets containing Carbopol 934 as a binder **30**, 229
Vila-Jato, J.L., see Alonso, M.J. **42**, 27; **68**, 69
Vila-Jato, J.L., see Anguiano-Igea, S. **135**, 161
Vila-Jato, J.L., see Calvo, P. **103**, 283
Vila-Jato, J.L., see Delgado Charro, M.B. **78**, 35
Vila-Jato, J.L., see Esclusa-Diaz, M.T. **142**, 183; **143**, 203
Vila-Jato, J.L., see Ganza-Gonzalez, A. **106**, 179
Vila-Jato, J.L., see Martinez-Pacheco, R. **32**, 99; **47**, 37
Vila-Jato, J.L., see Otero-Espinar, F.J. **70**, 35; **75**, 37; **79**, 149
Vila-Jato, J.L., see Remunan, C. **80**, 151
Vila-Jato, J.L., see Sanchez, A. **99**, 263
Vila-Jato, J.L., see Torres, D. **59**, 9; **121**, 239
Vilchez, F.G., see Contreras Claramonte, M.D. **94**, 23
Villanueva, R., see Sloan, K.B. **73**, 97
Villar, T.P., see Munoz-Ruiz, A. **144**, 147
Villari, A., M. Fresta, N. Micali, G. Puglisi, Potential application of UV reflection spectroscopy on solid pharmaceutical formulation analysis **127**, 185
Villari, A., see Cavallaro, G. **111**, 31
Villari, A., see Fresta, M. **99**, 145
Villari, A., see Panico, A.M. **98**, 19
Villari, A., see Santagati, N.A. **65**, 137
Villaverde, C., see Del Estal, J.L. **91**, 105; **102**, 257
Villieras, F., see Kedzierewicz, F. **117**, 247
Vincent-Ballereau, F., A. Fisch, J.E. Malkin, Prevention of transmission of AIDS and hepatitis viruses in health care workers: Evaluation of waste containers for injection, blood sampling and incision materials after use **58**, 25
Vincent-Ballereau, F., C. Merville, M.T. Lafleuriel, Sodium hypochlorite as a disinfectant for injection materials in third world rural dispensaries **50**, 87
Vincentelli, J., J.C. Vire, C. Paul, Y. Looze, Poly(ethylene glycol) derivatized prodrugs through mixed disulfide bond formation: Preliminary report on captopril **134**, 147
Vincieri, F.F., N. Mulinacci, G. Mazzi, S.A. Coran, M. Bambagiotti-Alberti, Stabilization of tinctures with cyclodextrins: A tincture from the fruits of *Oenanthe aquatica* L. **48**, 119
Vincieri, F.F., see Mulinacci, N. **90**, 35; **128**, 81
Vincke, B.J., A.E. Verstraeten, D.I.D El Eini, T.M. McCarthy, Extended stability of 5-fluorouracil and methotrexate solutions in PVC containers **54**, 181
Vincke, B.J., M.J. Devleeschouwer, J. Dony, G.J. Patriarche, Analytical determination of nicotinamide using bacterial electrodes **21**, 265
Efficiency of bioconversion of steroids by Fusarium oxysporum into Δ1,4-androstadiene-3,17-dione using gas chromatography **32**, 61
Vinckier, C., R. Hauchecorne, T. Cachet, G. Van den Mooter, J. Hoogmartens, A new mechanism for the decomposition of erythromycin A in acidic aqueous solutions **55**, 67
Vinckier, C., see Cachet, T. **55**, 59
Vinocur, M., see Matzek, K.M. **28**, 151
Vire, J.C., see Vincentelli, J. **134**, 147

Vis, B.J., see Bekers, O. **72**, 123
Visavarungroj, N., J. Herman, J.P. Remon, Crosslinked starch as binding agent. I. Conventional wet granulation **59**, 73
Visavarungroj, N., J.P. Remon, Crosslinked starch as a disintegrating agent **62**, 125
Crosslinked starch as a binding agent. II. Granulation in a high shear mixer **65**, 43
Crosslinked starch as binding agent. III. Granulation of an insoluble filler **69**, 43
Visavarungroj, N., see Herman, J. **42**, 15; **55**, 143
Vishnuvajjala, B.R., see Oh, I. **73**, 23
Visor, G.C., see Gu, L. **45**, 129
Visser, J., see Gould, P.L. **19**, 149
Visser, J., see Moolenaar, F. **1**, 323; **4**, 195; **5**, 127; **9**, 353; **33**, 249; **45**, 161; **114**, 117
Viswanathan, C.T., see Peck, C.C. **82**, 9
Viswanathan, C.T., see Shah, V.P. **82**, 1
Viswanathan, K., see Bhaskar, R. **28**, 59
Vitale, D., see Di Colo, G. **50**, 27
Vlachou, M.D., D.M. Rekkas, P.P. Dallas, N.H. Choulis, Development and in vitro evaluation of griseofulvin gels using Franz diffusion cells **82**, 47
Vlasak, J., see Pop, E. **84**, 39
Voges, R., see Franz, J.M. **7**, 19
Vogt, P., see Gonsho, A. **65**, 183
Vojnovic, D., D. Chicco, Zenary H. El, Doehlert experimental design applied to optimization and quality control of a granulation process in a high shear mixer **145**, 203
Vojnovic, D., see Kristl, A. **100**, 33; **139**, 231
Volford, O., J. Vamos, M. Takacs, The significance of different chromatographic methods in the European Pharmacopoeia **137**, 111
Volgushev, S.A., see Kosykh, V.A. **57**, 223
Volkin, D.B., see Burke, C.J. **86**, 89
Vollmer, U., B.W. Muller, J. Mesens, B. Wilffert, T. Peters, In vivo skin pharmacokinetics of liarozole: Percutaneous absorption studies with different formulations of cyclodextrin derivatives in rats **99**, 51
Volpe, P.L.O., see Beezer, A.E. **29**, 237
Von Schulthess, G.K., see Schwendener, R.A. **49**, 249
Vonderscher, J.P., see Franz, J.M. **7**, 19
Voorspoels, J., J.P. Remon, H. Nelis, G. Vandenbossche, Validation of filter sterilisation in autoclaves **133**, 9
Voskuilen, M.C.H., see Beijnen, J.H. **31**, 75
Vraka, V., see Loukas, Y.L. **144**, 225
Vrecer, F., S. Srcic, J. Smid-Korbar, Investigation of piroxicam polymorphism **68**, 35
Vrecer, F., see Kristl, A. **139**, 231
Vrijhof, W.P., see Kok, W.J.J. **86**, 259
Vrkljan, M., see Foster, T.M. **134**, 193
Vromans, H., G.K. Bolhuis, C.F. Lerk et al., Studies on tableting properties of lactose. VII. The effect of variations in primary particle size and percentage of amorphous lactose in spray dried lactose products **35**, 29
Vromans, H., G.K. Bolhuis, C.F. Lerk, K.D. Kussendrager, Studies of tableting properties of lactose. IX. The relationship between particle structure and compactibility of crystalline lactose **39**, 207
Vromans, H., H. Jorgensen, C.F. Lerk, K.D. Kussendrager, Studies on tableting properties of lactose. VIII. The effect of variations in primary particle size, percentage of amorphous lactose and addition of a disintegrant on the disintegration of spray-dried lactose tablets **39**, 201
Vromans, H., F. Moolenaar, Effect of solvents on rectal absorption rate of paracetamol in man: An in vitro approach **26**, 5
Vromans, H., J.A.H. Van Laarhoven, A study on water permeation through rubber closures of injection vials **79**, 301
Vromans, H., see Bos, C.E. **67**, 39
Vromans, H., see Horsthuis, G.J.B. **92**, 143
Vromans, H., see Leuenberger, H. **52**, 91
Vromans, H., see Piepma, K.A. **76**, 9; **97**, 29, 195
Vromans, H., see Van der Voort Maarschalk, K. **140**, 185
Vromans, H., see Zuurman, K. **102**, 1; **119**, 65
Vudathala, G.K., J.A. Rogers, Effect of cholesterol on the aging of griseofulvin-phospholipid coprecipitates **69**, 13
Vuleta, G., see Tamburic, S. **137**, 243
Vulovic, N., M. Primorac, M. Stupar, J.L. Ford, Some studies into the properties of indomethacin suspensions intended for ophthalmic use **55**, 123
Vulto, A., see Kadir, F. **64**, 171
Vyas, S.P., S. Bhatnagar, P.J. Gogoi, N.K. Jain, Preparation and characterization of HSA-propranolol microspheres for nasal administration **69**, 5
Vyas, S.P., S.K. Goswami, R. Singh, Liposomes based nasal delivery system of nifedipine: Development and characterization **118**, 23

W

Waaler, P.J., B.W. Muller, Solid-phase extraction of naftidrofuryl from human plasma for high-performance liquid chromatography analysis **87**, 223
Waaler, P.J., C. Graffner, B.W. Muller, Biopharmaceutical studies of naftidrofuryl in hydrocolloid matrix tablets **87**, 229
Waaler, T., see Waehler, K. **97**, 55
Wada, H., see Nishihata, T. **27**, 245
Wadhwa, H., see Singla, A.K. **108**, 173; **120**, 145
Wadhwa, L.K., P.D. Sharma, Glycolamide esters of 6-methoxy-2-naphthylacetic acid as potential prodrugs—physicochemical properties, chemical stability and enzymatic hydrolysis **118**, 31
Waehler, K., P.O. Roksvaag, T. Waaler, Determination of pyridostigmine and the main degradation product in tablets by high-performance liquid chromatography **97**, 55
Wafik Gouda, H., M.A. Moustafa, Shora H.I. Al, Effect of storage on nitrofurantoin solid dosage forms **18**, 213
Wafik Gouda, M., Effect of an antidiarrhoeal mixture on the bioavailability of tetracycline **89**, 75
Wafik Gouda, M., see Al-Meshal, M.A. **64**, 155
Wafik Gouda, M., see El-Sayed, Y.M. **64**, 109
Wagner, D.L., J.D. Nesbitt, J.D. Conklin, F.J. Hailey, Absorption of orally, intraduodenally, and intraileally administered nitrofurantoin in man **57**, 177

Wagner, D.L., V.S. Patel, Steady-state human pharmacokinetics and bioavailability of guaifenesin and pseudoephedrine in a sustained-release tablet relative to immediate-release liquids **114**, 171
Wagner, Z.G., see Olsson, B. **119**, 223
Wahbi, A.M., F.A. El-Yazbi, M.H. Barary, S.M. Sabri, Application of orthogonal functions to spectrophotometric analysis. Determination of dissociation constants **92**, 15
Waigh, R.D., see Anie, S.J. **69**, 181; **76**, 183
Wait, A.R., see Leahy, D.E. **50**, 117
Wakabayashi, N., see Masegi, M. **78**, 103
Wakamatsu, K., see Irie, T. **84**, 129
Wakerly, Z., J.T. Fell, D. Attwood, D.A. Parkins, In vitro evaluation of pectin-based colonic drug delivery systems **129**, 73
Wakiyama, N., A. Kusai, K. Nishimura, Mechanism of caking of granules containing oily materials **78**, 95
Wakusawa, S., see Hasegawa, T. **58**, 129
Walbaum, S., see Walchshofer, N. **100**, 271
Walchshofer, N., P. Delatour, J. Paris, Pharmacokinetics of an anthelmintic potential prodrug of a new benzimidazole in gerbil, mouse and sheep **77**, 265
Walchshofer, N., M.E. Sarciron, C. Arsac, S. Walbaum, J. Paris, A.F. Petavy, Biological effects of a dipeptide methyl ester on *Echinococcus multilocularis* metacestodes in vivo **100**, 271
Walczak, B., see Inghelbrecht, S. **148**, 103
Waldrep, J.C., see Vidgren, M. **115**, 209
Walker, C.V., J.I. Wells, Rheological synergism between ionic and non-ionic cellulose gums **11**, 309
Walker, C.V., see Wells, J.I. **15**, 97
Walker, I., see Davis, S.S. **17**, 203
Walker, I., D. Nicholls, W.J. Irwin, S. Freeman, Drug delivery via active transport at the blood-brain barrier: Affinity of a prodrug of phosphonoformate for the large amino acid transporter **104**, 157
Drug delivery via active transport at the blood-brain barrier. II. Investigation of monocarboxylic acid transport in vitro **108**, 225
Walker, I.M., see Davis, S.S. **38**, 133
Walker, M., L.A. Chambers, D.A. Hollingsbee, J. Hadgraft, Significance of vehicle thickness to skin penetration of Halcinonide **70**, 167
Walker, M., J. Hadgraft, Oleic acid—A membrane 'fluidiser' or fluid within the membrane? **71**, R1
Walker, M., see Edwardson, P.A.D. **91**, 51
Walker, S., see Staniforth, J.N. **31**, 277
Walker, S.E., see McTaggart, C.M. **19**, 139
Walkling, W.D., H. Almond, V. Paragamian et al., Difenoxin hydrochloride polymorphism **4**, 39
Walkow, J.C., J.W. McGinity, The effect of physicochemical properties on the in vitro diffusion of drug through synthetic membranes and pigskin. I. Methyl salicylate **35**, 91
Walkow, J.C., J.W. McGinity, The effect of physicochemical properties on the in vitro diffusion of drug through synthetic membranes and pigskin. II. Salicylic acid **35**, 103
Wall, D.A., A.T. Lanutti, High levels of exopeptidase activity are present in rat and canine bronchoalveolar lavage fluid **97**, 171
Wallace, S., see Li, C. **94**, 143
Waller, D.A., see Thomas, N.W. **20**, 163
Waller, I.L., see Akhter, S.A. **21**, 17
Waller, I.L., see Michniak-Mikolajczak, B.B. **31**, 247
Wallin, R., see Engstrom, S. **86**, 137
Wallis, K.H., see Muller, R.H. **89**, 25
Walmsley, A.D., see Meidan, V.M. **118**, 129
Walson, P.D., see Blanchard, J. **18**, 259
Walters, D.L., see Cefali, E.A. **127**, 147
Walters, J.M., see Prankerd, R.J. **78**, 59
Walters, K.A., A.T. Florence, P.H. Dugard, Non-ionic surfactants and the membrane transport of barbiturates in goldfish **10**, 153
Walters, K.A., see Ashton, P. **87**, 261, 265
Walters, K.A., see Bialik, W. **92**, 219
Walters, K.A., see Brain, K.R. **89**, R13
Walters, K.A., see Hadgraft, J. **32**, 257
Walters, P., see Toddywala, R.D. **76**, 77
Walters, V., see Alpar, O.H. **7**, 349
Walz, C., see Groning, R. **119**, 127
Wan, L.S.C., P.W.S. Heng, L.W. Chan, Influence of hydrophile-lipophile balance on alginate microspheres **95**, 77
Surfactant effects on alginate microspheres **103**, 267
Wan, L.S.C., P.W.S. Heng, C.G.H. Chia, Preparation of coated particles using a spray drying process with an aqueous system **77**, 183
Wan, L.S.C., P.W.S. Heng, C.V. Liew, Spheronization conditions on spheroid shape and size **96**, 59
The influence of liquid spray rate and atomizing pressure on the size of spray droplets and spheroids **118**, 213
Wan, L.S.C., P.W.S. Heng, B.L. Ling, Effect of polyvinylpyrrolidone solutions containing dissolved drug on characteristics of lactose fluidized bed granules **141**, 161
Wan, L.S.C., P.W.S. Heng, G. Muhuri, Incorporation and distribution of a low dose drug in granules **88**, 159
Wan, L.S.C., P.W.S. Heng, L.F. Wong, Matrix swelling: A simple model describing extent of swelling of HPMC matrices **116**, 159
Wan, L.S.C., P.W.S. Heng, L.F. Wongh, The effect of hydroxypropylmethylcellulose on water penetration into a matrix system **73**, 111
Wan, L.S.C., W.F. Lai, Factors affecting drug release from drug-coated granules prepared by fluidized-bed coating **72**, 163
Multilayer drug-coated cores: A system for controlling drug release **81**, 75
The influence of antitack additives on drug release from film-coated granules **94**, 39
Wan, L.S.C., K.P.P. Prasad, Effect of microcrystalline cellulose and cross-linked sodium carboxymethylcellulose on the properties of tablets with methylcellulose as a binder **41**, 159

Uptake of water by excipients in tablets **50**, 147
Uptake of water into tablets with low-substituted carboxymethyl cellulose sodium as disintegrant **55**, 115
Wan, L.S.C., see Heng, P.W.S. **116**, 119; **138**, 57; **143**, 107
Wan Po, A.L., see Irwin, W.J. **21**, 35
Wang, C.Y., see Huang, Y.Y. **129**, 41
Wang, D.P., see Tu, Y.H. **51**, 157
Wang, J.Z.Y., R.H. Bogner, Techniques to monitor the UV curing of potential solvent-free film-coating polymers **113**, 113
Solvent-free film coating using a novel photocurable polymer **119**, 81
Wang, L.H., Z.T. Chowhan, Drug–excipient interactions resulting from powder mixing. V. Role of sodium lauryl sulfate **60**, 61
Wang, L.H., see Imamura, Y. **5**, 25; **8**, 277
Wang, P., T.P. Johnston, Sustained-release interleukin-2 following intramuscular injection in rats **113**, 73
Wang, P.L., G.O. Udeani, T.P. Johnston, Inhibition of granulocyte colony stimulating factor (G-CSF) adsorption to polyvinyl chloride using a nonionic surfactant **114**, 177
Wang, P.L., T.P. Johnston, Thermal-induced denaturation of two model proteins: Effect of poloxamer 407 on solution stability **96**, 41
Wang, P.Y., Lipids as excipient in sustained release insulin implants One-month release injectable microspheres of leuprolide acetate inhibit steroidogenesis and genital organ growth in rats **54**, 223
Wang, Y., see Tu, Y.H. **61**, 119
Wang, Y.J., see Timmins, P. **11**, 329
Wang, Z., see Chow, K.Y. **25**, 41
Wannebroucq, J., see Fournier, C. **106**, 41
Wanwimolruk, S., S. Lipschitz, M.S. Roberts, Pharmacokinetics and bioavailability of Naprosyn CR 500 mg tablet, a new controlled-release formulation of naproxen, after single and multiple dosing **75**, 55
Waranis, R.P., K.G. Siver, K.B. Sloan, The solubility parameter of vehicles as a predictor of relative vehicle effects on the diffusion of 6-mercaptopurine **36**, 211
Warchol, M.P., see Duncan, M.R. **120**, 179
Ward, A.J.I., Reau C. Du, The essential role of lipid bilayers in the determination of stratum corneum permeability **74**, 137
Ward, G.H., see Surakitbanharn, Y. **109**, 27
Ward, P., see Toth, I. **73**, 259; **77**, 13; **106**, 85
Ward, P., see Valko, K. **83**, 123
Wargin, W.A., D.E. Wurster, Determination of the rearrangement kinetics of chloramphenicol-3-monosuccinate using an automated HPLC system **15**, 37
Waring, M.J., see Ferrari, F. **112**, 29; **117**, 49
Waring, M.J., L. Monger, D.A. Hollingsbee, G.P. Martin, C. Marriott, Assessment of corticosteroid-induced skin blanching: Evaluation of the Minolta Chromameter CR200 **94**, 211
Waring, M.J., M.H. Rubinstein, J.R. Howard, Acoustic emission of pharmaceutical materials during compression **36**, 29
Acoustic emission of pharmaceutical materials: The effect of compression speed, ejection, lubrication and tablet weight **40**, 15
Waring, M.J., see Wong, D.Y.T. **72**, 233
Warrack, J.K., see Holland, S.J. **65**, 225
Warren, S.J., S.J. Farr, Formulation of solution metered dose inhalers and comparison with aerosols emitted from conventional suspension systems **124**, 195
Wartewig, S., see Wegener, M. **128**, 203
Waser, P.G., G.K. Bolhuis, J. Kreuter, S. Berger, K. Munz, E. Kaiser, B. Pfluger, Localization of colloidal particles (liposomes, hexylcyanoacrylate nanoparticles and albumin nanoparticles) by histology and autoradiography in mice **39**, 213
Wasfi, I.A., see Tanira, M.O.M. **102**, 267
Washington, C., Evaluation of non-sink dialysis methods for the measurement of drug release from colloids: effects of drug partition **56**, 71
Drug release from microdisperse systems: A critical review **58**, 1
The electrokinetic properties of phospholipid-stabilized fat emulsions. II. Droplet mobility in mixed electrolytes **58**, 13
The electrokinetic properties of phospholipid-stabilized fat emulsions. III. Interdroplet potentials and stability ratios in monovalent electrolytes **64**, 67
The stability of intravenous fat emulsions in total parenteral nutrition mixtures **66**, 1
The electrokinetic propers of phospholipid stabilized fat emulsions. VI. Zeta potentials of Intralipid 20% in TPN mixtures **87**, 167
Washington, C., A. Athersuch, D.J. Kynoch, The electrokinetic properties of phospholipid stabilized fat emulsions. IV. The effect of glucose and of pH **64**, 217
Washington, C., C.J. Briggs, Reduction of absorption of drugs into TPN plastic containers by phospholipids and fat emulsions **48**, 133
Washington, C., A. Chawla, N. Christy, S.S. Davis, The electrokinetic properties of phospholipid-stabilized fat emulsions **54**, 191
Washington, C., M.A. Connolly, R. Manning, M.C.L. Skerratt, The electrokinetic properties of phospholipid stabilized fat emulsions. V. The effect of amino acids on emulsion stability **77**, 57
Washington, C., S.S. Davis, The production of parenteral feeding emulsions by Microfluidizer **44**, 169
Washington, C., S.P. Denyer, Ageing effects in parenteral fat emulsions: The role of fatty acids **39**, 33
Washington, C., F. Koosha, Drug release from microparticulates; deconvolution of measurement errors **59**, 79
Washington, C., T. Sizer, Stability of TPN mixtures compounded from lipofundin S and Aminoplex amino-acid solutions: Comparison of laser diffraction and Coulter counter droplet size analysis **83**, 227
Washington, C., see Beastall, J.C. **43**, 207
Washington, C., see Bower, C. **118**, 229
Washington, C., see Davis, S.S. **82**, 99
Washington, C., see Gilbert, J.C. **40**, 93

Washington, C., see Illum, L. **54**, 41
Washington, C., see Johnson, O.L. **53**, 237; **59**, 131; **63**, 65
Washington, C., see Manning, R.J. **81**, 1
Washington, C., see Rolland, A. **39**, 173
Washington, C., see Washington, N. **28**, 139; **29**, 253; **34**, 105; **108**, 125
Washington, C., see Wilson, C.G. **72**, 79
Washington, N., G. Lamont, C.G. Wilson, C. Washington, R. Withington, The effect of high and low fat meals on the absorption of rifampicin from the gastrointestinal tract in volunteer subjects **108**, 125
Washington, N., G. Ridout, C.G. Wilson, Gastric distribution and residence time of two anti-reflux formulations **39**, 163
Washington, N., C.G. Wilson, Antacids: Physiology versus pharmaceutics **28**, 249
Washington, N., C.G. Wilson, S.S. Davis, Evaluation of 'raft-forming' antacid neutralizing capacity: In vitro and in vivo correlations **27**, 279
Washington, N., C.G. Wilson, J.L. Greaves, S. Norman, J.M. Peach, K. Pugh, A γ scintigraphic study of gastric coating by Expidet, tablet and liquid formulations **57**, 17
Washington, N., C. Washington, C.G. Wilson, S.S. Davis, The effect of inclusion of aluminium hydroxide in alginate-containing raft-forming antacids **28**, 139
The neutralization capacity of magnesium trisilicate BP and magnesium trisilicate mixture BP **29**, 253
What is 'Liquid Gaviscon'? A comparison of four international formulations **34**, 105
Washington, N., see Davis, S.S. **35**, 253; **60**, 191
Washington, N., see Rowe, R.C. **38**, 227
Washington, N., see Wilson, C.G. **40**, 119; **50**, 155; **72**, 79
Wassermann, K., H. Bundgaard, Kinetics of the acid-catalyzed hydrolysis of doxorubicin **14**, 73
Wassmus, W., see Bleich, J. **97**, 111
Wastie, M., see Harnor, K.J. **95**, 111
Watanabe, A., T. Hanawa, M. Sugihara, K. Yamamoto, Development and pharmaceutical properties of a new oral dosage form of theophylline using sodium caseinate for the possible use in elderly patients **117**, 23
Watanabe, H., see Hodoshima, N. **146**, 81
Watanabe, Y., T. Kogoshi, Y. Amagai, M. Matsumoto, Preparation and evaluation of enteric granules of aspirin prepared by acylglycerols **64**, 147
Watanabe, Y., Y. Matsumoto, R. Kikuchi, M. Kiriyama, R. Ito, M. Matsumoto, H. Nomura, K. Maruyama, Absorption and blood leukocyte dynamics of recombinant human granulocyte colony-stimulating factor (rhG-CSF) from intranasally administered preparations containing rhG-CSF and cyclodextrins in rabbits **110**, 93
Watanabe, Y., W. Zhao, Y. Matsumoto, M. Suda, K. Takayama, Y. Ohgawara, Y. Kobayashi, M. Matsumoto, Increasing bioavailability of nifedipine from matrix granules prepared using macrogol and trilaurin **99**, 337
Watari, N., N. Kaneniwa, Dissolution of slightly soluble drugs. VIII. In vivo drug dissolution in human subjects and rabbits **7**, 307
Watari, N., N. Kaneniwa, M. Hanano, Comparison of in vivo dissolution of sulfa drugs calculated by deconvolution to absorption rates calculated by multicompartment model method **6**, 155
Watari, N., T. Funaki, K. Aizawa, N. Kaneniwa, A flip-flop model for nitrofurantoin disposition in the rabbit following oral administration **21**, 85
Watari, N., see Funaki, T. **43**, 59
Watari, N., see Kaneniwa, N. **31**, 33; **34**, 231
Watkinson, A.C., A.L. Bunge, J. Hadgraft, Aspects of the transdermal delivery of prostaglandins II **83**, 171
Watkinson, A.C., A.L. Bunge, J. Hadgraft, A. Naik, Computer simulation of penetrant concentration-depth profiles in the stratum corneum **87**, 175
Watkinson, A.C., H. Joubin, D.M. Green, K.R. Brain, J. Hadgraft, The influence of vehicle on permeation from saturated solutions **121**, 27
The influence of vehicle on permeation from saturated solutions. (Int. J. Pharm. 121 (1995) 27–36) **123**, 303
Watkinson, A.C., J. Hadgraft, A. Bye, Aspects of the transdermal delivery of prostaglandins **74**, 229
Watkinson, A.C., see Diez-Sales, O. **129**, 33
Watkinson, A.C., see Heard, C.M. **90**, R5
Watkinson, A.C., see Lafforgue, C. **121**, 89
Watkinson, A.C., see Roberts, M.S. **126**, 219
Watson, D., see Davidson, A.G. **63**, 29
Watts, D.W., see Sunderland, V.B. **19**, 1; **27**, 1
Watts, J.F., see Davies, M.C. **57**, 183
Watts, O., see Martin, B. **49**, 63
Watts, P.J., B.P. Atkin, C.G. Wilson, M.C. Davies, C.D. Melia, Radiolabelling of polymer microspheres for scintigraphic investigations by neutron activation. 1. Incorporation of samarium oxide and its effects on the properties of Eudragit RS: Sulphasalazine microspheres **76**, 55
Watts, P.J., L. Barrow, K.P. Steed, C.G. Wilson, R.C. Spiller, C.D. Melia, M.C. Davies, The transit rate of different-sized model dosage forms through the human colon and the effects of a lactulose-induced catharis **87**, 215
Watts, P.J., C.G. Wilson, M.C. Davies, C.D. Melia, Radiolabelling of polymer microspheres for scintigraphic investigations by neutron activation. 2. Effects of irradiation on the propers of Eudragit RS-sulphasalazine microspheres **98**, 63
Radiolabelling of polymer microspheres for scintigraphic investigations by neutron activation. 3. Changes in the physical properties of Eudragit RS-sulphapyridine microspheres from incorporating samarium oxide **98**, 75
Radiolabelling of polymer microspheres for scintigraphic investigations by neutron activation. 4. A pharmacoscintigraphic study of colon-targeted Eudragit RS-sulphapyridine microspheres in human volunteers **102**, 101
Waugh, A., see Bowtle, W. **9**, 305
Waugh, W., see Umprayn, K. **66**, 253
Waugh, W.A., see Anderson, B.D. **45**, 27
Waugh, W.A., see Stella, V.J. **71**, 157
Waugh, W.N., see Stella, V.J. **43**, 191
Waysek, E., see Beck, M.I. **141**, 137

Wearley, L.L., Y.W. Chien, Iontophoretic transdermal permeation of verapamil (III): Effect of binding and concentration gradient on reversibility of skin permeation rate **59**, 87
Weatherall, I.L., see Stark, G. **143**, 93
Webb, A.I., see Estes, K.S. **65**, 101
Wegener, M., R. Neubert, W. Rettig, S. Wartewig, Structure of stratum corneum lipids characterized by FT-Raman spectroscopy and DSC. I. Ceramides **128**, 203
Wehrle, P., see Goldbach, P. **117**, 225
Wehrle, P., see Saleh, S.I. **48**, 149
Wei, S., V. Kumar, G.S. Banker, Phosphoric acid mediated depolymerization and decrystallization of cellulose: Preparation of low crystallinity cellulose—A new pharmaceutical excipient **142**, 175
Wei, S.L., see Qi, X.R. **146**, 31
Weide, W.L., see Swart, P.J. **87**, 67
Weierstall, R., see Dey, M. **49**, 121
Weimar, W.R., see Sloan, K.B. **73**, 97
Weiner, N., see Addicks, W.J. **56**, 243
Weiner, N., see Chiang, C. **40**, 143
Weiner, N., see Chiang, C.M. **37**, 75
Weiner, N., see Du Plessis, J. **103**, 277, R1; **127**, 273
Weiner, N., see Tsai, J.C. **96**, 111
Weiner, N.D., see Chiang, C.M. **49**, 109; **50**, 21; **55**, 229
Weiner, N.D., see Ganesan, M.G. **20**, 139
Weiner, N.D., see Lin, S.Y. **95**, 57
Weiner, N.D., see Ma, L. **70**, 209; **77**, 127
Weiner, N.D., see Schwinke, D.L. **20**, 119
Weiner, N.D., see Tsai, J.C. **68**, 127; **72**, 227
Weiner, N.D., see Williams, N.A. **50**, 261; **51**, 241
Weingarten, C., Santos Magalhaes, A. Baszkin, S. Benita, M. Seiller, Interactions of a non-ionic ABA copolymer surfactant with phospholipid monolayers: Possible relevance to emulsion stabilization **75**, 171
Weingarten, C., A. Moufti, J. Delattre et al., Protection of insulin from enzymatic degradation by its assocation to liposomes **26**, 251
Weinstock, M., see Benita, S. **30**, 47
Weiss, G., A. Knoch, A. Laicher, F. Stanislaus, R. Daniels, Simple coacervation of hydroxypropyl methylcellulose phthalate (HPMCP). I. Temperature and pH dependency of coacervate formation **124**, 87
Simple coacervation of hydroxypropyl methylcellulose phthalate (HPMCP). II. Microencapsulation of ibuprofen **124**, 97
Weiss, J., see Yang, S.T. **86**, 247
Weissinger, J., see Peck, C.C. **82**, 9
Welham, K.J., see Toth, I. **73**, 259
Welling, P.G., L.E. Schmitz, R.J. Wills, R.K. Bush, Plasma levels of phenobarbital resulting from sustained release tablets containing theophylline, phenobarbital and ephedrine **4**, 135
Wells, J.I., C.V. Walker, The influence of granulating fluids upon granule and tablet properties: The role of secondary binding **15**, 97
Wells, J.I., see Larhrib, H. **147**, 187, 199
Wells, J.I., see Walker, C.V. **11**, 309
Welshman, I.R., see Antal, E.J. **54**, 33
Wennergren, B., J. Lindberg, M. Nicklasson, G. Nilsson, G. Nyberg, R. Ahlgren, C. Persson, B. Palm, A collaborative in vitro dissolution study: Comparing the flow-through method with the USP paddle method using USP prednisone calibrator tablets **53**, 35
Wennergren, B., see Nicklasson, M. **37**, 195
Wepierre, J., see Bernard, E. **126**, 235
Wepierre, J., see Ferreira, L.A.M. **109**, 251
Wepierre, J., see Garcia, B. **4**, 205
Werner, I.A., H. Altorfer, X. Perlia, The effectiveness of various scavengers on the γ-irradiated methanolic solution of medazepam **63**, 155
Wernly-Chung, G.N., J.M. Mayer, A. Tsantili-Kakoulidou, B. Testa, Structure-reactivity relationships in the chemical hydrolysis of prodrug esters of nicotinic acid **63**, 129
Wesdyk, R., Y.M. Joshi, J. de Vincentis, A.W. Newman, N.B. Jain, Factors affecting differences in film thickness of beads coated in fluidized bed units **93**, 101
Wesdyk, R., Y.M. Joshi, N.B. Jain, K. Morris, A. Newman, The effect of size and mass on the film thickness of beads coated in fluidized bed equipment **65**, 69
Wesemeyer, H., B.W. Muller, R.H. Muller, Adsorption of ethoxylated surfactants on nanoparticles. II. Determination of adsorption enthalpy by microcalorimetry **89**, 33
Wesoluch, F., A.T. Florence, F. Puisieux, J.T. Carstensen, Solubilization of chlorhexidine diacetate and dichloride in aqueous decaoxyethylene oleic ether systems **2**, 343
Wesseling, M.M., see Cremers, H.F.M. **120**, 51
West, D.P., see Bonina, F.P. **98**, 93
West, P., see Illum, L. **54**, 41
Wester, R.C., P.K. Noonan, Relevance of animal models for percutaneous absorption **7**, 99
Wester, R.C., see Shah, V.P. **74**, 1
Westera, G., see Schwendener, R.A. **49**, 249
Westerberg, M., B. Jonsson, C. Nystrom, Physicochemical aspects of drug release. IV. The effect of carrier particle properties on the dissolution rate from ordered mixtures **28**, 23
Westerberg, M., C. Nystrom, Physicochemical aspects of drug release. XII. The effect of some carrier particle properties and lubricant admixture on drug dissolution from tableted ordered mixtures **69**, 129
Physicochemical aspects of drug release. XVII. The effect of drug surface area coverage to carrier materials on drug dissolution from ordered mixtures **90**, 1
Westerberg, M., see Nilsson, P. **45**, 111
Westerhuis, J.A., see Westerhuis, J.A. **143**, 151
Westerhuis, J.A., Haan P. De, J. Zwinkels, W.T. Jansen, P.J.M. Coenegracht, C.F. Lerk, Optimisation of the composition and production of mannitol/microcrystalline cellulose tablets **143**, 151
Westermann-Clark, G.B., see Miller, II K.J. **98**, 101
Westermarck, E., see Heinamaki, J. **42**, 105; **48**, 51
Westermarck, E., see Marvola, M. **44**, 159
Westesen, K., Phase diagram of tyloxapol and water—I **102**, 91

Westesen, K., H. Bunjes, Do nanoparticles prepared from lipids solid at room temperature always possess a solid lipid matrix? Letter to the editor [1] **115**, 129
Westesen, K., M.H.J. Koch, Phase diagram of tyloxapol and water—II **103**, 225
Westesen, K., B. Siekmann, M.H.J. Koch, Investigations on the physical state of lipid nanoparticles by synchrotron radiation X-ray diffraction **93**, 189
Westesen, K., see Bunjes, H. **129**, 159
Westesen, K., see Sjostrom, B. **94**, 89
Westrom, B.R., see Lundin, S. **59**, 263
Westrup, J., see Wilding, I.R. **101**, 263
Westwood, S.W., see Habberfield, A. **145**, 1
Weyhers, H., see Muller, R.H. **138**, 85
Whalley, D.R., see Palin, K.J. **12**, 315
Whateley, T.L., see Law, T.K. **21**, 277; **33**, 65
Whateley, T.L., see Omotosho, J.A. **61**, 51
Whateley, T.L., see Ping, Q. **61**, 79
Whateley, T.L., see Wood, D.A. **8**, 35
Wheeler, A.W., see Sharif, S. **119**, 239
Whelan, I., see Barnwell, S.G. **128**, 145
Whelan, I., see Burns, S.J. **110**, 291; **121**, 37; **134**, 223; **141**, 9
Whelpton, R., see Aburawi, S. **22**, 327
Whiston, R.J., N.W. Thomas, K.J. Palin, C.G. Wilson, A comparison of prednisolone absorption and the effect of sodium salicylate and mannitol in normal and damaged rat bowel **23**, 247
Whitaker, M., see Zainudin, B.M.Z. **51**, 67
White, D.A., S.P. Thompson, C.J. Wilson, G.D. Bell, A pharmacokinetic comparision of two delayed-release peppermint oil preparations, Colpermin and Mintec, for treatment of the irritable bowel syndrome **40**, 151
White, D.G., M.J. Story, S.G. Barnwell, An experimental animal model for studying the effects of a novel lymphatic drug delivery system for propranolol **69**, 169
White, E.F.T., see Gibson, S.H.M. **47**, 205; **48**, 63, 113; **50**, 163
White, E.F.T., see Rowe, R.C. **22**, 57
White, E.F.T., see Sakellariou, P. **27**, 267; **31**, 55, 175; **34**, 93
Whitehill, D., see Florence, A.T. **11**, 277
Whitmore, J., see Davis, S.S. **1**, 85
Whitworth, C.W., see Pongpaibul, Y. **33**, 243
Wichert, B.M., see Wichert, B.V. **78**, 227
Wichert, B.V., R.J. Gonzalez-Rothi, L.E. Straub, B.M. Wichert, H. Schreier, Amikacin liposomes: Characterization, aerosolization, and in vitro activity against *Mycobacterium avium*-intracellulare in alveolar macrophages **78**, 227
Wichman, A., see Larhammar, H. **23**, 13
Wicks, S.R., see Aboofazeli, R. **111**, 63
Wicks, S.R., see Clarke, J.G. **80**, R1; **93**, 221
Wiechers, J.W., B.F.H. Drenth, J.H.G. Jonkman, R.A. de Zeeuw, Percutaneous absorption, metabolism, and elimination of the penetration enhances azone in humans after prolonged application under occlusion **47**, 43
Percutaneous absorption of triamcinolone acetonide from creams with and without Azone^{R} in humans in vivo **66**, 53
Wiechers, J.W., R.E. Herder, B.F.H. Drenth, R.A. de Zeeuw, Percutaneous absorption, disposition, metabolism, and excretion of ^{14}C-labelled cyoctol in humans after a single dermal application **65**, 77
Wiggins, J., see Leigh, T.R. **67**, 275
Wigmore, A.J., see Landin, M. **133**, 127; **134**, 243
Wijsbeek, J., see Jonkman, J.H.G. **3**, 55
Wikberg, M., G. Alderborn, Compression characteristics of granulated materials. II. Evaluation of granule fragmentation during compression by tablet permeability and porosity measurements **62**, 229
Compression characteristics of granulated materials. III. The relationship between air permeability and mechanical strength of tablets of some lactose granulations **63**, 23
Compression characteristics of granulated materials. IV. The effect of granule porosity on the fragmentation propensity and the compactibility of some granulations **69**, 239
Compression characteristics of granulated materials. VI. Pore size distributions, assessed by mercury penetration, of compacts of two lactose granulations with different fragmentation propensities **84**, 191
Wikberg, M., see Johansson, B. **117**, 57
Wikman, A., see Karlsson, J. **99**, 209
Wilding, E.A., see Davis, S.S. **94**, 235
Wilding, I.R., S.S. Davis, F. Pozzi, P. Furlani, A. Gazzaniga, Enteric coated timed release systems for colonic targeting **111**, 99
Wilding, I.R., S.S. Davis, G.H. Rimoy, P. Rubin, T. Kurihara-Bergstrom, V. Tipnis, B. Berner, J. Nightingale, Pharmacokinetic evaluation of transdermal buprenorphine in man **132**, 81
Wilding, I.R., S.S. Davis, R.A. Sparrow, J.R. Bloor, G. Hayes, G.T. Ward, The effect of food on the in vivo behaviour of a novel sustained release formulation of tiaprofenic acid **83**, 155
Wilding, I.R., S.S. Davis, K.P. Steed, R.A. Sparrow, J. Westrup, J.M. Hempenstall, Gastrointestinal transit of a drug-resinate administered as an oral suspension **101**, 263
Wilding, I.R., J.G. Hardy, M. Maccari, V. Ravelli, S.S. Davis, Scintigraphic and pharmacokinetic assessment of a multiparticulate sustained release formulation of diltiazem **76**, 133
Wilding, I.R., R.A. Sparrow, S.S. Davis, R.J. Horton, The role of gastric emptying in the absorption and metabolism of nifedipine given in a modified release pellet formulation **84**, 59
Wilding, I.R., see Christrup, L.L. **66**, 169
Wilding, I.R., see Coupe, A.J. **78**, 69; **92**, 167
Wilding, I.R., see Davies, M.C. **57**, 183; **62**, 97
Wilding, I.R., see Davis, S.S. **68**, 199; **94**, 235
Wilding, I.R., see Kenyon, C.J. **112**, 207
Wilding, I.R., see Price, J.M.C. **76**, 123
Wilding, I.R., see Steed, K.P. **112**, 199
Wilding, I.R., see Wilson, C.G. **72**, 79
Wilffert, B., see Vollmer, U. **99**, 51
Wilhelm, F., see Engel, A. **107**, 99

Wilisch, I.L., C.C. Muller-Goymann, Correlation of colloidal microstructure, drug release and permeation through excised human skin **96**, 79
Wilke, T., see Pavliv, L. **105**, 227
Wilke, T.L., T.H. Nguyen, P. Tyle, HPLC method development for AG-85 and benzyl alcohol in anhydrous topical formulations **103**, 187
Wilkins, K.M., G.W. Hanlon, G.P. Martin, C. Marriott, The role of adhesion in the migration of bacteria along intrauterine contraceptive device polymer monofilaments **58**, 165
Wilkins, K.M., G.P. Martin, G.W. Hanlon, C. Marriott, The influence of critical surface tension and microrugosity on the adhesion of bacteria to polymer monofilaments **57**, 1
Wilkinson, K.F., see Rush, B.D. **73**, 231
Williams, A.C., B.W. Barry, Urea analogues in propylene glycol as penetration enhancers in human skin **56**, 43
The enhancement index concept applied to terpene penetration enhancers for human skin and model lipophilic (oestradiol) and hydrophilic (5-fluorouracil) drugs **74**, 157
Williams, A.C., P.A. Cornwell, B.W. Barry, On the non-Gaussian distribution of human skin permeabilities **86**, 69
Williams, A.C., H.G.M. Edwards, B.W. Barry, Fourier transform Raman spectroscopy. A novel application for examining human stratum corneum **81**, R11
Williams, A.C., see Anigbogu, A.N.C. **125**, 265
Williams, A.C., see Megrab, N.A. **116**, 101; **131**, 103, 117; **145**, 37, 49; **146**, 41
Williams, A.C., see Yamane, M.A. **116**, 237
Williams, A.S., W.G. Love, B.D. Williams, Synthesis of methotrexate-dimyristoylphosphatidylethanolamine analogs and characterization of methotrexate release in vivo **85**, 189
Williams, A.S., G. Taylor, Synthesis, characterization and release of cromoglycate from dextran conjugates **83**, 233
Williams, A.W., G. Taylor, Release of cromoglycate from dextran and inulin conjugates **87**, 255
Williams, B.D., see Williams, A.S. **85**, 189
Williams, D.B., S.A. Varia, V.J. Stella, I.H. Pitman, Evaluation of the prodrug potential of the sulfate esters of acetaminophen and 3-hydroxymethyl-phenytoin **14**, 113
Williams, D.G., see Hadgraft, J. **141**, 17
Williams, D.R., see Hall, S.B. **70**, 251
Williams, E.J., see Davis, M. **28**, 157
Williams, J.D., see Hendriksen, B.A. **69**, 175
Williams, M.D., see Johnson, J.L. **90**, 151
Williams, N.A., G.P. Polli, Differential scanning calorimetric studies on frozen cephalosporin solutions **44**, 205
Williams, N.A., N.D. Weiner, Interactions of small polypeptides with dimyristoylphosphatidylcholine monolayers: Effect of size and hydrophobicity **50**, 261
Interaction of recombinant leukocyte A interferon with lipid monolayers **51**, 241
Williams, N.A., see Cannon, J.B. **114**, 65
Williams, P.L., see Riviere, J.E. **127**, 213
Williams, R.D., see Porter, C.J.H. **141**, 227
Williams, R.L., see Pradhan, S. **138**, 207
Williams, S.O., see Jozwiakowski, M.J. **91**, 195
Williams, T., see Safadi, M. **90**, 239
Williamson, I., see Hastewell, J. **101**, 115
Williamson, I., see Nicklin, P. **83**, 197
Willmott, N., P.J. Harrison, Characterisation of freeze-dried albumin microspheres containing the anti-cancer drug adriamycin **43**, 161
Wills, R.J., see Welling, P.G. **4**, 135
Willson, R.J., A.E. Beezer, J.C. Mitchell, Solid state reactions studied by isothermal microcalorimetry; the solid state oxidation of ascorbic acid **132**, 45
Wilson, A.P.R., see Podczeck, F. **142**, 235
Wilson, C.G., C. Marriott, J.G. Hardy, S.W. Bond, The influence of food on the absorption of acyclovir: A pharmacokinetic and scintigraphic assessment **38**, 221
Wilson, C.G., G.D. Parr, J.W. Kennerley, Pharmacokinetics and in vivo scintigraphic monitoring of a sustained release acetylsalicylic acid formulation **18**, 1
Wilson, C.G., N. Washington, J.L. Greaves, F. Kamali, J.A. Rees, A.K. Sempik, J.F. Lampard, Bimodal release of ibuprofen in a sustained-release formulation: A scintigraphic and pharmacokinetic open study in healthy volunteers under different conditions of food intake **50**, 155
Wilson, C.G., N. Washington, J.L. Greaves, C. Washington, I.R. Wilding, T. Hoadley, E.E. Sims, Predictive modelling of the behaviour of a controlled release buflomedil HCl formulation using scintigraphic and pharmacokinetic data **72**, 79
Wilson, C.G., N. Washington, J. Peach, G.R. Murray, J. Kennerley, The behaviour of a fast-dissolving dosage form (Expidet) followed by γ-scintigraphy **40**, 119
Wilson, C.G., see Bennett, C.E. **21**, 341
Wilson, C.G., see Davis, S.S. **32**, 85; **35**, 253; **60**, 191
Wilson, C.G., see Fitzgerald, P. **40**, 81; **83**, 177
Wilson, C.G., see Harnor, K.J. **95**, 111
Wilson, C.G., see Illum, L. **12**, 135
Wilson, C.G., see Jenkins, J.R.F. **14**, 143
Wilson, C.G., see Kreuter, J. **16**, 105
Wilson, C.G., see May, H.A. **19**, 169
Wilson, C.G., see O'Reilly, S. **34**, 213
Wilson, C.G., see Palin, K.J. **12**, 315
Wilson, C.G., see Pennington, A.K. **43**, 221
Wilson, C.G., see Rao, L.S. **17**, 283
Wilson, C.G., see Saunders, J. **68**, 265
Wilson, C.G., see Washington, N. **27**, 279; **28**, 139, 249; **29**, 253; **34**, 105; **39**, 163; **57**, 17; **108**, 125
Wilson, C.G., see Watts, P.J. **76**, 55; **87**, 215; **98**, 63, 75; **102**, 101
Wilson, C.G., see Whiston, R.J. **23**, 247
Wilson, C.G., see Wood, E. **25**, 191
Wilson, C.J., see White, D.A. **40**, 151
Wilson, D., see Seminoff, L.A. **54**, 251
Wilson, D.E., see Hovgaard, L. **132**, 115
Wilson, G., see Eddy, E.P. **115**, 79
Winchcomb, K.N., see Davis, S.S. **30**, 29
Winchester, J.G., see Van, Schie D.M. **93**, 177
Winfield, A.J., Saidan S.M. Al, Compound formation in phenobarbitone-urea systems **8**, 211
Winfield, A.J., see Cumming, K.I. **108**, 141

Winfield, A.J., see Ritchie, K. **18**, 89
Wingstrand, K., B. Abrahamsson, B. Edgar, Bioavailability from felodipine extended-release tablets with different dissolution properties **60**, 151
Winkelhorst, J., see De, Vries J.D. **100**, 181
Winkins, J.H.M., see Busker, R.W. **36**, 113
Winter, H.M.I., see de Boer, A.H. **130**, 231
Winterle, J.S., see Chen, S.H. **72**, 89
Winwood, D., see Bodor, N. **53**, 195
Wissens, A.C., see Frijlink, H.W. **64**, 195
Withington, R., see Farraj, N.F. **38**, 39
Withington, R., see Sims, E.E. **17**, 167
Withington, R., see Washington, N. **108**, 125
Wohlrab, W., see Neubert, R. **75**, 89
Wohlrab, W., see Schneider, I.M. **145**, 187
Wolf, B., W. Schmitz, H. Schneider, Composites of bead cellulose and hydrophilic solubilizers **139**, 87
Wolf, W., see Leh, F.K.V. **1**, 41
Wolff, H.M., see Hadgraft, J. **73**, 125; **89**, R1; **123**, 285
Wolff, M., see Green, P.G. **55**, 265
Wolff, M., see Hadgraft, J. **64**, 187
Wolfson, A.D., D.F. McCafferty, K.E. McGowan, V. Boston, Non-invasive monitoring of percutaneous local anaesthesia using laser-Doppler volocimetry **51**, 183
Wolowacz, S., see Nicholls, T.J. **138**, 175
Won, C.M., Kinetics and mechanism of hydrolysis of tyrphostins **104**, 29
Won, C.M., A.K. Iula, Kinetics of hydrolysis of diltiazem **79**, 183
Won, C.M., S.Y. Tang, L. Strohbeck, Photolytic and oxidative degradation of an anmetic agent, RG 12915 **121**, 95
Wondergem, J., see Cremers, H.F.M. **120**, 51
Wong, A., see Zatz, J.L. **9**, 315
Wong, D.Y.T., M.J. Waring, P. Wright, M.E. Aulton, Elucidation of the compressive deformation behaviour of α-lactose monohydrate and anhydrous α-lactose single crystals by mechanical strength and acoustic emission analyses **72**, 233
Wong, F.N., see Conard, G.J. **10**, 259
Wong, J., W.Y. Kuu, H. Kajimoto, H. Kazama, Correlation of the intravenous in-line drug delivery kinetics with the diluent flow rate, angle of internal flow, wettability, solubility and particle surface area **144**, 177
Wong, J., see Kuu, W.Y. **124**, 241
Wong, K.T., see Nixon, J.R. **50**, 205; **58**, 31
Wong, L.F., see Wan, L.S.C. **116**, 159
Wong, L.P., C.A. Gilligan, A. Li Wan Po, Preparation and characterisation of sustained-release ibuprofen-cetostearyl alcohol spheres **83**, 95
Wong, L.P., see Li Wan Po, A. **66**, 111
Wong, L.W., N. Pilpel, The effect of particle shape on the mechanical properties of powders **59**, 145
Wong, M., see Small, D. **149**, 195
Wong, M.W.Y., A.G. Mitchell, Physicochemical characterization of a phase change produced during the wet granulation of chlorpromazine hydrochloride and its effects on tableting **88**, 261
Wong, O., C. Anderson, L. Allaben, R. Pabmanabhan, G. Lattin, Stability-indicating assay method for pilocarpine nitrate in reservoirs used in the cystic fibrosis indicator system **76**, 171
Wong, O., N. Tsuzuki, B. Nghiem, J. Kuehnhoff, T. Itoh, K. Masaki, J. Huntington, R. Konishi, J.H. Rytting, T. Higuchi, Unsaturated cyclic ureas as new non-toxic biodegradable transdermal penetration enhancers. II. Evaluation study **52**, 191
Wong, W., see Zhang, X. **137**, 199
Wongh, L.F., see Wan, L.S.C. **73**, 111
Wongpoowarak, W., N. Worakul, Mathematical aspect of pH-zero point of charge relationship **75**, R9
Alternative statistical method to determine pH-zero point of charge **80**, R21
Effect of titrant concentration on pH-zero point of charge: Verification by bootstrap technique **97**, 141
Woo, B.H., see Ha, I.S. **144**, 91
Woo, B.H., see Lee, K.C. **114**, 215
Woo, S.O., see Tonnesen, H.H. **43**, 215
Wood, B., see Anie, S.J. **76**, 183
Wood, C., see Eddy, E.P. **115**, 79
Wood, D.A., Biodegradable drug delivery systems **7**, 1
Wood, D.A., T.L. Whateley, A.T. Florence, Formation of poly(butyl 2-cyanoacrylate)microcapsules and the microencapsulation of aqueous solutions of [^{125}I]-labelled proteins **8**, 35
Wood, E., C.G. Wilson, J.G. Hardy, The spreading of foam and solution enemas **25**, 191
Wood, E., see Hardy, J.G. **38**, 103
Wood, I.P., I. Toth, J.L. Holley, W.A. Gibbons, Lipidic peptides. XIII. Synthesis, structure elucidation and in vitro toxicity assessment of chlorambucil conjugates with lipidic acids, lipidic amino acids and their oligomers **87**, 141
Wood, I.P., see Toth, I. **83**, 131; **102**, 223
Wood, J.M., D. Attwood, J.H. Collett, The swelling properties of poly(2-hydroxyethyl methacrylate) hydrogels polymerized by γ-irradiation and chemical initiation **7**, 189
Wood, R.W., V.H.K. Li, J. Kreuter, J.R. Robinson, Ocular disposition of poly-hexyl-2-cyano[3-^{14}C]acrylate nanoparticles in the albino rabbit **23**, 175
Wood, R.W., M.J. Mulski, Methodology for the determination of water vapor transport across plastic films **50**, 61
Wood, R.W., J.R. Robinson, High-performance chromatographic determination of pilocarpine and pilocarpic acid in ocular tissues **20**, 285
Ocular disposition of pilocarpine in the pigmented rabbit: Site(s) of metabolism **29**, 127
Wood, R.W., see Hansen, L.D. **135**, 31; **137**, 1
Wood, R.W., see Kuu, W.Y. **55**, 77
Woodage, T., see Bowtle, W. **9**, 305
Woodcock, P.M., see Meakin, B.J. **119**, 91, 103
Woodford, R., B.W. Barry, Activity and bioavailability of a new steroid (Timobesone acetate) in cream and ointment compared with Lidex and Dermovate creams and ointments and Betnovate cream **26**, 145
Woodford, R., see Southwell, D. **18**, 299

Woodhead, P.J., see Ashford, M. **91**, 241; **95**, 193
Woodhouse, B.A., see Thomas, N.W. **44**, 261
Woodle, M.C., M.S. Newman, F.J. Martin, Liposome leakage and blood circulation: Comparison of adsorbed block copolymers with covalent attachment of PEG **88**, 327
Woodley, J., see Naisbett, B. **107**, 223; **110**, 127; **114**, 227; **120**, 247
Woodley, J.F., see Bridges, J.F. **44**, 213
Woodley, J.F., see Morgan, S.M. **122**, 121; **128**, 99
Woodley, J.F., see Pato, A.J. **104**, 227
Woodward, L., see Lu, M.F. **71**, 25
Woolfrey, S.G., G. Taylor, I.W. Kellaway, A. Smith, Disposition of carboxyfluorescein in the rat **26**, 35
Woolfrey, S.G., see Farr, S.J. **26**, 303
Woolfson, A.D., D.F. McCafferty, Percutaneous local anaesthesia: Drug release characteristics of the amethocaine phase-change system **94**, 75
Woolfson, A.D., D.F. McCafferty, S.P. Gorman, D.S. Jones, Polarographic analysis of taurolidine, a non-antibiotic antimicrobial agent **48**, 167
Woolfson, A.D., D.F. McCafferty, S.P. Gorman, P.A. McGarron, J.H. Price, Design of an apparatus incorporating a linear variable differential transformer for the measurement of type III bioadhesion to cervical tissue **84**, 69
Woolfson, A.D., D.F. McCafferty, A.P. Launchbury, Stabilisation of hydrotropic temazepam parenteral formulations by lyophilisation **34**, 17
Woolfson, A.D., D.F. McCafferty, K.E. McGowan, The metabolism of amethocaine by porcine and human skin extracts: Influence on percutaneous anaesthesia **62**, 9
Percutaneous penetration characteristics of amethocaine through porcine and human skin **78**, 209
Woolfson, A.D., S.P. Gorman, D.F. McCafferty, D.S. Jones, Assay procedures for Taurolin solutions using pre-column derivatisation and high-performance liquid chromatography with fluorescence detection **49**, 135
Woolfson, A.D., see Almeida, A.J. **149**, 255
Woolfson, A.D., see Ballantine, J. **3**, 239
Woolfson, A.D., see Jones, D.S. **64**, R1
Woolfson, A.D., see Karim, E.F.I.A. **52**, 155
Woolfson, A.D., see McCafferty, D.F. **31**, 9
Woolfson, A.D., see McGovern, J.G. **149**, 251
Woolfson, A.D., see Ponchel, G. **38**, 65
Woolfson, A.D., see Richardson, P.J. **78**, 189
Worakul, N., see Wongpoowarak, W. **75**, R9; **80**, R21; **97**, 141
Wormald, P., see Ek, R. **125**, 257
Worthen, D.B., see Dansereau, R.J. **103**, 97
Worthington, H.E.C., see Sims, E.E. **24**, 287
Wotton, P.K., B. Mollgaard, J. Hadgraft, A. Hoelgaard, Vehicle effect on topical drug delivery. III. Effect of azone on the cutaneous permeation of metronidazole and propylene glycol **24**, 19
Wotton, P.K., see Hadgraft, J. **32**, 257
Wotton, P.K., see Johansen, M. **32**, 199
Wouessidjewe, D., see Lemesle-Lamache, V. **141**, 117
Wouessidjewe, D., see Leroy-Lechat, F. **101**, 97
Wouessidjewe, D., see Lin, S.Z. **69**, 211; **106**, 63
Wouessidjewe, D., see Ponchel, G. **38**, 65
Wouessidjewe, D., see Skiba, M. **120**, 1; **126**, 275; **129**, 113; **142**, 121; **145**, 241
Wout, Z.G., see Johnston, T.P. **100**, 279
Wright III, C.E., see Antal, E.J. **54**, 33
Wright, J., see Illum, L. **52**, 221
Wright, P., see Wong, D.Y.T. **72**, 233
Wu, A.B., see Chen, C.Y. **141**, 171
Wu, A.T., see Pradhan, S. **138**, 207
Wu, C.C., T. Sokoloski, M.F. Blanford, A.M. Burkman, Absence of metabolite in the disappearance of nitroglycerin following incubation with red blood cells **8**, 323
Wu, C.C., see Sokoloski, T.D. **6**, 63
Wu, D.W., see Cotton, M.L. **40**, 129
Wu, H.J., see Rodriguez-Hornedo, N. **85**, 149
Wu, H.L.S., S.C. Miller, In vitro release of nicotinic acid alkyl esters from poloxamer vehicles **66**, 213
Wu, H.J., see Khor, S.P. **23**, 109; **30**, 189
Wu, L.L., see Silvestri, S. **71**, 65
Wu, P.C., Y.B. Huang, J.Y. Fang, Y.H. Tsai, In vitro percutaneouus absorption of captopril **148**, 41
Wu, P.C., Y.B. Huang, H.H. Lin, Y.H. Tsai, In vitro percutaneous absorption of captopril through excised rabbit skin **143**, 119
Percutaneous absorption of captorpil from hydrophilic cellulose gel^{R} through excised rabbit skin and human skin **145**, 215
Wu, P.C., see Fang, J.Y. **126**, 119; **128**, 169; **130**, 121; **131**, 143; **143**, 47; **145**, 175
Wu, P.C., see Hsu, L.R. **106**, 1
Wu, P.C., see Huang, Y.B. **126**, 111; **131**, 137; **134**, 183
Wu, S.M., see Huang, Y.Y. **129**, 41; **131**, 19
Wu-Pong, S., J. Bard, Oligonucleotide adsorption to artificial surfaces **143**, 259
Wulff, M., M. Alden, D.Q.M. Craig, An investigation into the critical surfactant concentration for solid solubility of hydrophobic drug in different polyethylene glycols **142**, 189
Wurster, D.E., J.M. Patel, Catalysis of α-naphthyl acetate hydrolysis by *o*-iodosobenzoic acid in micellar solutions of cetyltrimethylammonium bromide **78**, 157
Wurster, D.E., C.E. Rowlings, J.R. Creekmore, Calorimetric analysis of powder compression. I. Design and development of a compression calorimeter **116**, 179
Wurster, D.E., S.M. Upadrashta, Equilibrium binding studies of the 1,8-dihydroxy-9,10-anthraquinone–bovine serum albumin interaction **55**, 221
Wurster, D.E., see Matheson, Jr. L.E. **4**, 309
Wurster, D.E., see Patel, J.M. **86**, 43
Wurster, D.E., see Rowlings, C.E. **116**, 191
Wurster, D.E., see Upadrashta, S.M. **49**, 103
Wurster, D.E., see Wargin, W.A. **15**, 37
Wuthrich, R., see Schwendener, R.A. **49**, 249
Wygant, M.B., see Anderson, B.D. **45**, 27
Wygant, M.B., see Carlisle, M.R. **80**, 227
Wynne, H.J.A., see Tomlinson, E. **12**, 87

X

Xiang, T.X., B.D. Anderson, Inclusion complexes of purine nucleosides with cyclodextrins. II. Investigation of inclusion complex geometry and cavity microenvironment **59**, 45
Xiang, T.X., see Anderson, B.D. **45**, 27
Xiang, T.X., see Darrington, R.T. **59**, 35
Xing, D.K.L., see Richards, R.M.E. **75**, 81; **82**, 107; **87**, 195; **116**, 217
Xing, J.Z., see Richards, R.M.E. **92**, 237

Y

Yachi, K., see Yamauchi, H. **90**, 73
Yacobi, A., see Shah, V.P. **74**, 1
Yacobi, A., see Skelly, J.P. **63**, 83
Yagi, T., see Nishihata, T. **38**, 83
Yagoubi, N., see Sarbach, Ch. **140**, 169
Yahya, A.M., J.C. McElnay, P.F. D'Arcy, Binding of chloroquine to glass **25**, 217
Photodegradation of frusemide during storage in burette administration sets **31**, 65
Investigation of chloroquine binding to plastic materials **34**, 137
Yahya-Saeb, H.H., see Farsam, H. **7**, 343
Yakatan, G.J., see Cruz, J.E. **2**, 275
Yalcin, I., E. Sener, QSARs of some novel antibacterial benzimidazoles, benzoxazoles, and oxazolopyridines against an enteric gram-negative rod; *K. pneumoniae* **98**, 1
Yalcin, I., see Sener, E. **110**, 109
Yalkowsky, S.H., see Neau, S.H. **49**, 223
Yalkowsky, S.H., see Surakitbanharn, Y. **109**, 27
Yamada, H., see Oguma, T. **63**, 101
Yamada, H., see Yamauchi, H. **113**, 141
Yamada, K., A. Furuya, M. Akimoto, T. Maki, T. Suwa, H. Ogata, Evaluation of gastrointestinal transit controlled-beagle dog as a suitable animal model for bioavailability testing of sustained-release acetaminophen dosage form **119**, 1
Yamada, K., see Akimoto, M. **100**, 133; **117**, 31
Yamada, K., see Yamada, K. **119**, 1
Yamada, M., see Oribe, T. **124**, 27
Yamagiwa, Y., see Uchiyama, K. **121**, 195
Yamaguchi, H., K. Nakamura, S. Numajiri, K. Sugibayashi, Y. Miromoto, Swelling and mechanical properties of poly(vinyl alcohol) hydrogels **58**, 135
Yamaguchi, I., see Yoshida, M. **115**, 61
Yamaguchi, S., see Nakano, N.I. **7**, 335
Yamaguchi, T., C. Ikeda, Y. Sekine, Intestinal absorption of a β-adrenergic blocking agent nadolol. Enhancement of in situ and in vivo absorption of nadolol in rats **37**, 127
Yamaguchi, T., M. Nishimura, R. Okamoto, T. Takeuchi, K. Yamamoto, Glass formation of 4′-*O*-(4-methoxyphenyl) acetyltylosin and physicochemical stability of the amorphous solid **85**, 87
Yamaguchi, Y., see Miyagawa, Y. **138**, 215
Yamahira, T., see Miyazaki, S. **8**, 303
Yamahira, Y., T. Noguchi, H. Takenaka, T. Maeda, Biopharmaceutical studies of lipid-containing oral dosage forms: Relationship between drug absorption rate and digestibility of vehicles **3**, 23
Yamamoto, A., M. Islasse, T. Sakane, T. Kawaratani, M. Hashida, H. Sezaki, Enhanced absorption of salicylic acid and sulfanilamide by the synthetic immunomodulator levamisole in rats **39**, 255
Yamamoto, A., T. Morita, M. Hashida, H. Sezaki, Effect of absorption promoters on the nasal absorption of drugs with various molecular weights **93**, 91
Yamamoto, A., E. Utsumi, T. Kawaratani, T. Sakane, M. Hashida, H. Sezaki, Effect of immunomodulator on drug absorption from the gastrointestinal tract: The mechanism whereby levamisole enhanced the intestinal absorption of sulfanilamide in the rat **51**, 137
Yamamoto, A., see Masegi, M. **78**, 103
Yamamoto, A., see Ohtani, T. **77**, 141
Yamamoto, A., see Sakane, T. **57**, 77
Yamamoto, K., S. Yokohama, H. Emori, A.C. Shah, T. Nishihata, Bioavailability of itazigrel in dogs after oral administration of soft elastic capsules **102**, 133
Yamamoto, K., see El-Gendy, G.A. **31**, 25
Yamamoto, K., see Matsumoto, K. **108**, 167
Yamamoto, K., see Oguchi, T. **61**, 27; **113**, 97
Yamamoto, K., see Watanabe, A. **117**, 23
Yamamoto, K., see Yamaguchi, T. **85**, 87
Yamamoto, L.A., see Shah, V.P. **32**, 243
Yamamoto, M., A. Yoshida, F. Hirayama, K. Uekama, Some physicochemical properties of branched β-cyclodextrins and their inclusion characteristics pyr 1989 **49**, 163
Yamamoto, M., see Tsunematsu, H. **68**, 77
Yamamoto, T., see Maitani, Y. **117**, 129; **126**, 89
Yamamura, K., T. Yotsuyanagi, Adsorption and irreversible binding of adriamycin incorporated into hydroxyapatite beads **79**, R1
Yamamura, S., Y. Momose, K. Terada, E. Fukuoka, Evaluation of paracrystalline lattice distortion by profile analysis using a single X-ray diffraction peak **133**, 117
Yamamura, S., J.A. Rogers, Characterization and dissolution behavior of nifedipine and phosphatidylcholine binary systems **130**, 65
Yamana, T., Y. Mizukami, F. Ichimura et al., Stability kinetics of piperacillin in aqueous solutions **11**, 71
Yamana, T., see Ichimura, F. **15**, 321; **19**, 75; **27**, 45
Yamanaka, H., see Asano, M. **67**, 67
Yamanaka, H., see Imasaka, K. **68**, 87; **81**, 31
Yamanaka, Y., see Shinkuma, D. **21**, 187
Yamane, H., see Kakee, A. **111**, 15
Yamane, M.A., A.C. Williams, B.W. Barry, Effects of terpenes and oleic acid as skin penetration enhancers towards 5-fluorouracil as assessed with time; permeation, partitioning and differential scanning calorimetry **116**, 237
Yamanouchi, S., see Fujii, M. **137**, 117
Yamaoka, K., Y. Tanigawara, T. Nakagawa, T. Uno, Capacity-limited elimination of cefmetazole in rat **10**, 291
Yamaoka, K., see Murai, Y. **15**, 309
Yamaoka, T., M. Kuroda, Y. Tabata, Y. Ikada, Body distribution of dextran derivatives with electric charges after intravenous administration **113**, 149
Yamasaki, M., see Hoshino, T. **50**, 45

Yamashita, C., H. Matsuo, K. Akiyama, H. Kiwada, Enhancing effect of cetylmannoside on targeting of liposomes to Kupffer cells in rats **70**, 225
Yamashita, F., Y. Koyama, M. Kitano, Y. Takakura, M. Hashida, Analysis of in vivo skin penetration enhancement by oleic acid based on a two-layer diffusion model with polar and nonpolar routes in the stratum corneum **117**, 173
Yamashita, F., Y. Koyama, H. Sezaki, M. Hashida, Estimation of a concentration profile of acyclovir in the skin after topical administration **89**, 199
Yamashita, F., see Bando, H. **135**, 91; **145**, 103
Yamashita, F., see Nishida, K. **80**, 101
Yamashita, F., see Takino, T. **114**, 43
Yamashita, S., see Araki, M. **81**, 39
Yamashita, S., T. Nadai, M. Sumi, Y. Suda, Possible role of serum protein binding to improve drug disposition **108**, 241
Yamasmoto, K., see Tahara, K. **133**, 17
Yamauchi, H., H. Kikuchi, K. Yachi, M. Sawada, M. Tomikawa, S. Hirota, Effects of glycophorin and ganglioside GM_3 on the blood circulation and tissue distribution of liposomes in rats **90**, 73
Yamauchi, H., T. Yano, T. Kato, I. Tanaka, S. Nakabayashi, K. Higashi, H. Yamada, Effects of sialic acid derivative on long circulation time and tumor concentration of liposomes **113**, 141
Yamauchi, H., see Maruyama, K. **111**, 103
Yamazaki, J.I., see Morimoto, Y. **91**, 9
Yamazaki, S., see Maitani, Y. **64**, 139
Yamazi, A., see Nishihata, T. **42**, 257
Yanai, S., H. Okada, K. Saito, Y. Kuge, Y. Ogawa, H. Toguchi, Optimal formulation of an angiogenesis inhibitor, TNP-470, for arterial injection determined by in vitro drug release and stability, and in vivo antitumor activity **123**, 237
Yang, A.Y.S., see Zografi, G. **18**, 99
Yang, D.C., see Lee, V.H.L. **16**, 163
Yang, D.C., see Redell, M.A. **17**, 299
Yang, D.C., see Stratford, Jr. R.E. **13**, 263
Yang, D.J., see Li, C. **94**, 143
Yang, I.C., see Chowhan, Z.T. **14**, 231
Yang, J., see Yoon, E.J. **109**, 181
Yang, M.Y., see Tsai, Y.H. **28**, 47
Yang, P.C., see Kararli, T.T. **102**, 177
Yang, S.T., I. Ghebre-Sellassie, The effect of product bed temperature on the microstructure of Aquacoat-based controlled-release coatings **60**, 109
Yang, S.T., G. Van Savage, J. Weiss, I. Ghebre-Sellassie, The effect of spray mode and chamber geometry of fluid-bed coating equipment and other parameters on an aqueous-based ethylcellulose coating **86**, 247
Yang, W., see Chen, Y. **94**, 81
Yang, W.H., D. Brooke, Rate equation for solid state decomposition of aspirin in the presence of moisture **11**, 271
Yano, T., see Yamauchi, H. **113**, 141
Yarnitzky, C., Smyth W. Franklin, Square wave polarographic and voltammetric analysis of selected electroreducible drugs **75**, 161
Yarnitzky, C., see Smyth, W.F. **42**, 235
Yashiki, T., see Hirai, S. **7**, 317; **9**, 165, 173
Yashiki, T., see Iga, K. **6**, 43
Yashiki, T., see Yoshimura, Y. **23**, 117; **26**, 317; **38**, 179
Yasuda, T., see Kitamura, S. **59**, 217
Yasumura, T., see Takada, K. **44**, 107
Yata, N., see Kadir, S. **33**, 235
Yata, N., see Murakami, T. **49**, 173
Yatabe, H., see Kohri, N. **68**, 255
Yatomi, M., see Ogiso, T. **98**, 113
Yazdanian, M., The effect of freezing on cattle skin permeability **103**, 93
Yeh, C.M., see Sheu, M.T. **103**, 137
Ygartua, P., see Arellano, A. **130**, 141
Ygartua, P., see Santoyo, S. **117**, 219
Yi-Hung, Tsai, see Naito, S. **8**, 263
Yianneskis, M., see Lee, K.C. **130**, 103
Yianneskis, M., see Timsina, M.P. **101**, 1
Yin, G., see Zhan, X. **115**, 161, 167; **126**, 29
Yiv, S.H., see Constantinides, P.P. **115**, 225
Yliruusi, J., see Eerikainen, S. **71**, 201; **80**, 259
Yliruusi, J., see Hellen, L. **96**, 197, 205, 217
Yliruusi, J., see Juppo, A.M. **88**, 141
Yliruusi, J., see Juslin, L. **123**, 247, 257
Yliruusi, J., see Kervinen, L. **92**, 115
Yliruusi, J., see Kiesvaara, J. **92**, 81; **97**, 101
Yliruusi, J., see Mannermaa, J.P. **103**, 125
Yliruusi, J., see Merkku, P. **102**, 117
Yliruusi, J., see Murtoniemi, E. **108**, 155; **110**, 101
Yliruusi, J., see Tanninen, V.P. **81**, 169
Yliruusi, J.K., see Heinamaki, J.T. **109**, 9; **112**, 191
Yogev, A., see Amselem, S. **125**, 31
Yogev, A., see Brewster, M.E. **125**, 17
Yokochi, N., see Kitagawa, S. **126**, 49
Yokogawa, K., see Ichimura, F. **15**, 321; **27**, 45; **19**, 75
Yokohama, S., see Tahara, K. **99**, 311
Yokohama, S., see Yamamoto, K. **102**, 133
Yokomizo, Y., Effect of phosphatidylglycerol on the in vitro percutaneous drug penetration through the dorsal skin of guinea pigs, and analysis of the molecular mechanism, using (ATR-FTIR) spectroscopy **147**, 219
Yonemochi, E., see Matsumoto, K. **108**, 167
Yonemochi, E., see Oguchi, T. **61**, 27; **113**, 97
Yongxin, Z., see Hendrix, C. **100**, 213
Yoon, E.J., E.J. Bae, Y.N. Jeong, M.M. Kim, B. Kim, S.H. Lee, M.G. Lee, Dose-dependent pharmacokinetics of human granulocyte/macrophage colony-stimulating factor in rabbits **97**, 213
Yoon, E.J., H.W. Chang, M.G. Lee, H. Lee, M.K. Park, C.K. Kim, Pharmacokinetics of methotrexate after intravenous infusion of methotrexate-rabbit serum albumin conjugate to rabbits **67**, 177
Yoon, E.J., S.H. Kim, M.G. Lee, Effects of sodium thiosulfate on the pharmacokinetics of total platinum after intravenous adminstration of cisplatin to rabbits **99**, 343

Yoon, E.J., E.D. Lee, W.H. Yoon, H.J. Shim, S.D. Lee, W.B. Kim, J. Yang, M.G. Lee, Pharmacokinetics, tissue distribution and biliary excretion of FT-ADM after intravenous administration of DA-125, a prodrug of FT-ADM to dogs. A new adriamycin analog containing fluorine **109**, 181
Yoon, E.J., see Chung, S.M. **68**, 61
Yoon, E.J., see Lee, H.J. **121**, 1
Yoon, E.J., see Shim, H.J. **103**, 147
Yoon, E.J., see Yoon, W.H. **102**, 249
Yoon, J.Y., see Lee, K.C. **114**, 215
Yoon, W.H., E.J. Yoon, S.H. Jang, S.H. Lee, H.J. Lee, J.M. Park, K.E. Choi, M.G. Lee, Pharmacokinetics of ciprofloxacin after intravenous administration of ciprofloxacin-TOF in rabbits **102**, 249
Yoon, W.H., see Lee, S.H. **90**, 81
Yoon, W.H., see Yoon, E.J. **109**, 181
Yoon, Y.S., see Kim, C.K. **106**, 25; **120**, 21
York, P., Solid-state properties of powders in the formulation and processing of solid dosage forms **14**, 1
York, P., F. Bassam, R.C. Rowe, R.J. Roberts, Fracture mechanics of microcrystalline cellulose powders **66**, 143
York, P., D.J.W. Grant, A disruption index for quantifying the solid state disorder induced by additives or impurities. I. Definition and evaluation from heat of fusion **25**, 57
York, P., see Agbada, C.O. **106**, 33
York, P., see Bassam, F. **64**, 55
York, P., see Doherty, C. **34**, 197; **47**, 141; **50**, 222
York, P., see Forbes, R.T. **126**, 199
York, P., see Grant, D.J.W. **28**, 103; **30**, 161
York, P., see Hancock, B.C. **76**, 239; **83**, 147; **102**, 167; **148**, 1
York, P., see Hendriksen, B.A. **118**, 1
York, P., see Hussain, M.S.H. **42**, 89; **70**, 103; **78**, 203
York, P., see Landin, M. **104**, 271; **133**, 127; **134**, 243
York, P., see Marshall, P.V. **55**, 257; **67**, 59
York, P., see Miller, T.A. **23**, 55; **41**, 1
York, P., see Okhamafe, A.O. **22**, 265, 273
York, P., see Parker, M.D. **64**, 207; **72**, 243; **80**, 179
York, P., see Roberts, R.J. **91**, 173; **105**, 177; **125**, 157
York, P., see Tricehurst, M.D. **111**, 241; **141**, 93
York, P., see Vidgren, M.T. **39**, 101
York, P., see Yusuff, N. **73**, 9
Yoshida, A., see Yamamoto, M. **49**, 163
Yoshida, M., M. Asano, H. Omichi, Y. Hayashi, I. Yamaguchi, Y. Matsuda, Study of biodegradable copoly(L-lactic acid/glycolic acid) formulations with controlled release of Z-100 for application in radiation therapy **115**, 61
Yoshida, M., see Asano, M. **67**, 67
Yoshida, M., see Imasaka, K. **68**, 87; **81**, 31
Yoshida, M., see Miyajima, M. **95**, 153
Yoshida, M., see Yoshida, M. **115**, 61
Yoshida, S., see Tsunematsu, H. **68**, 77
Yoshida, T., see Okumura, K. **88**, 63
Yoshikawa, H., S. Muranishi, C. Kato, H. Sezaki, Bifunctional delivery system for selective transfer of bleomycin into lymphatics via enteral route **8**, 291
Yoshikawa, H., H. Sezaki, S. Muranishi, Mechanism for selective transfer of bleomycin into lymphatics by a bifunctional delivery system via the lumen of the large intestine **13**, 321
Yoshikawa, H., see Kaji, Y. **24**, 79
Yoshikawa, H., see Takada, K. **25**, 155; **44**, 107; **65**, 159
Yoshikawa, T., see Oguma, T. **63**, 101
Yoshimura, Y., N. Hamaguchi, N. Kakeya, T. Yashiki, Synthesis and relation between physicochemical propers and oral absorption of 7-*O*-acylmandelamido-3-methyl-3-cephem-4-carboxylic acids **26**, 317
Yoshimura, Y., N. Hamaguchi, T. Yashiki, Synthesis and relationship between physicochemical properties and oral absorption of pivaloyloxymethyl esters of parenteral cephalosporins **23**, 117
Yoshimura, Y., N. Kakeya, Structure-gastrointestinal absorption relationship of penicillins **17**, 47
Yoshimura, Y., N. Tada, K. Kubo, M. Miyamoto, Y. Inada, K. Nishikawa, T. Naka, Evaluation and improvement of bioavailability of a new angiotensin II receptor antagonist, 2-butyl-1-[2′-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl-1H-benzimidazole-7-carboxylic acid by making prodrug **103**, 1
Yoshimura, Y., R.H. Reuning, T. Yashiki, Synthesis and oral absorption of acyloxymethyl esters of 7β-(2-(2-aminothiazol-4-yl)acetamido)-3-(((1-(2-dimethylaminoethyl)-H-tetrazol-5-yl)thio)-methyl)ceph-3-em-4-carboxylic acid (cefotiam) **38**, 179
Yoshino, H., see Narisawa, S. **104**, 95; **148**, 85
Yoshioka, A.T., B.B. Sternberg, A.T. Florence, Preparation and properties of vesicles (niosomes) of sorbitan monoesters (Span 20, 40, 60 and 80) and a sorbitan triester (Span 85) **105**, 1
Yoshioka, M., see Aioi, A. **93**, 1
Yoshioka, S., see Aso, Y. **93**, 153; **115**, 133
Yoshioka, S., see Izutsu, K. **71**, 137
Yoshioka, S., see Izutsu, K.I. **90**, 187
Yoshioka, T., M. Hashida, S. Muranishi, H. Sezaki, Specific delivery of mitomycin C to the liver, spleen and lung: nano- and micro-spherical carriers of gelatin **8**, 131
Yoshioka, T., see Hirai, S. **15**, 261
Yoshioks, T., A.T. Florence, Vesicle (niosome)-in-water-in-oil (v/w/o) emulsions: An in vitro study **108**, 117
Yoshitomi, H., see Nishihata, T. **42**, 257
Yotsuyanagi, T., see Chen, D. **106**, 249
Yotsuyanagi, T., see Ohta, N. **29**, 137; **85**, 39; **118**, 85
Yotsuyanagi, T., see Yamamura, K. **79**, R1
Young, C., K.J. Palin, A.S. Reid, N.W. Thomas, P.L. Gould, Formulation of fenbufen suppositories. II. Selection of a suppository base using dissolution studies and histological studies in rats **40**, 187
Young, C.S., C.K. Shim, M.H. Lee, S.K. Kim, Effect of sodium salicylate on in vitro percutaneous penetration of isopropamide iodide through mouse skin **45**, 59
Young, J.W.M., see Chow, A.H.L. **126**, 11, 21
Young, S.A., G. Buckton, Particle growth in aqueous suspensions: The influence of surface energy and polarity **60**, 235

Young-Harvey, J.A., I.D. Rae, I.H. Pitman, Oxazolidines: Prodrugs for the delivery of β-aminoalcohols through human skin from aqueous solution **30**, 151
Youssef, A.F., see Elshabouri, S.R. **2**, 257
Yska, J.P., see Jonkman, J.H.G. **43**, 139
Yu, C.D., see Fox, J.L. **2**, 41
Yu, D.H.S., see Streng, W.H. **135**, 43
Yu, H.Y., H.M. Liao, Triamcinolone permeation from different liposome formulations through rat skin in vitro **127**, 1
Yu, J.Y., see Lu, S.M. **112**, 117
Yu, K., Y.W. Chien, Spermicidal activity-structure relationship of nonoxynol oligomers: Physicochemical basis **125**, 81
Yu, L.X., J.R. Crison, G.L. Amidon, Compartmental transit and dispersion model analysis of small intestinal transit flow in humans **140**, 111
Yu, W., E.S. Tabosa do Egito, G. Barratt, H. Fessi, J.P. Devissaguet, F. Puisieux, A novel approach to the preparation of injectable emulsions by a spontaneous emulsification process **89**, 139
Yucan, Z., see Parkash, A.K. **92**, 151
Yuen, C., see Zhu, H. **135**, 151
Yuen, K.H., A.A. Deshmukh, J.M. Newton, M. Short, R. Melchor, Gastrointestinal transit and absorption of theophylline from a multiparticulate controlled release formulation **97**, 61
Yuen, K.H., see Peh, K.K. **139**, 95
Yuk, S.H., S.J. Lee, T. Okano, B. Berner, S.W. Kim, One-way membrane for transdermal drug delivery systems. I. Membrane preparation and characterization **77**, 221
One-way membrane for transdermal drug delivery system. II. System optimization **77**, 231
Yuk, S.H., see Lim, C.R. **132**, 175
Yuksel, N., T. Tincer, T. Baykara, Interaction between nicardipine hydrochloride and polymeric microspheres for a controlled release system **140**, 145
Yun, D.S., see Chun, I.K. **96**, 91
Yun, S.K., see Shim, C.K. **56**, 269
Yusuff, N., P. York, Spironolactone-cyclodextrin complexes: Phase solubility and ultrafiltration studies **73**, 9

Z

Zaghloul, I.A., see Khalil, S.A. **19**, 307
Zaghloul, I.Y., Z.H. Aly, M.A. Radwan, Effect of ciprofloxacin on the hypoprothrombinemic activity of warfarin in rats **131**, 131
Zahirul, M., I. Khan, Dissolution testing for sustained or controlled release oral dosage forms and correlation with in vivo data: Challenges and opportunities **140**, 131
Zainudin, B.M.Z., S.E.J. Tolfree, M. Biddiscombe, M. Whitaker, M.D. Short, S.G. Spiro, An alternative to direct labelling of pressurised bronchodilator aerosol **51**, 67
Zajic, L., G. Buckton, The use of surface energy values to predict optimum binder selection for granulations **59**, 155
Zajic, L., see Murphy, A. **38**, 257
Zaki, I., see Hardy, J.G. **37**, 245
Zalisz, R., see Fortier, M. **52**, 27
Zanardi, G., see Santi, P. **92**, 23
Zanen, P., L.T. Go, J.W.J. Lammers, The optimal particle size for β-adrenergic aerosols in mild asthmatics **107**, 211
The optimal particle size for parasympathicolytic aerosols in mild asthmatics Rectal absorption of morphine from controlled release suppositories **114**, 111
Zanen, P., P.I. Van Spiegel, H. Van der Kolk, E. Tushuizen, R. Enthoven, The effect of the inhalation flow on the performance of a dry powder inhalation system **81**, 199
Zaniboni, H.C., J.T. Fell, J.H. Collett, Production and characterisation of enteric beads **125**, 151
Zanic-Grubisic, T., see Juretic, D. **61**, 219
Zanol, M., see Redenti, E. **129**, 289
Zapf, R., see Badian, M. **18**, 9
Zatz, J.L., Effect of formulation on flocculation of dispersions stabilized by a non-ionic surfactant **4**, 83
Zatz, J.L., P. Sarpotdar, G. Gergich, A. Wong, Effect of surfactants on the flocculation of magnesium carbonate suspensions by xanthan gum **9**, 315
Zatz, J.L., see Kushla, G.P. **71**, 167
Zazzo, J.F., see Rieutord, A. **119**, 57
Zeevi, A., S. Klang, V. Alard, F. Brossard, S. Benita, The design and characterization of a positively charged submicron emulsion containing a sunscreen agent **108**, 57
Zeng, X.M., G.P. Martin, C. Marriott, Albumin microspheres as a means of drug delivery to the lung: Analysis of the effects of process variables on particle sizes using factorial design methodology **107**, 205
Tetrandrine delivery to the lung: The optimisation of albumin microsphere preparation by central composite design **109**, 135
The controlled delivery of drugs to the lung **124**, 149
Zentner, G.M., see Skauen, D.M. **20**, 235
Zhan, X., J. Jiang, S. Liu, G. Yin, Computer-controlled heating system and new computation for reciprocal heating stability experiment **115**, 167
Zhan, X., J. Jiang, B. Ma, G. Yin, A computer-controlled temperature recording system and a new computation system for flexible heating stability experiments **126**, 29
Zhan, X., G. Yin, B. Ma, New heating controller and computation for linear heating stability experiment **115**, 161
Zhang, G., see Felton, L.A. **113**, 17; **127**, 203
Zhang, J., S. Niu, C. Ebert, T.H. Stanley, An in vivo dog model for studying recovery kinetics of the buccal mucosa permeation barrier after exposure to permeation enhancers: Apparent evidence of effective enhancement without tissue damage **101**, 15
Zhang, X., J.K. Jackson, H.M. Burt, Development of amphiphilic diblock copolymers as micellar carriers of taxol **132**, 195
Zhang, X., J.K. Jackson, W. Wong, W. Min, T. Cruz, W.L. Hunter, H.M. Burt, Development of biodegradable polymeric paste formulations for taxol: An in vitro and in vivo study **137**, 199
Zhang, Z., G. Liao, T. Nagai, S. Hou, Mitoxantrone polybutyl cyanoacrylate nanoparticles as an anti-neoplastic targeting drug delivery system **139**, 1

Zhao, W., see Watanabe, Y. **99**, 337
Zheng, H., K.L. Audus, Cytotoxic effects of chlorhexidine and nystatin on cultured hamster buccal epithelial cells **101**, 121
Zheng, H., P.K. Shah, K.L. Audus, Evaluation of antiulcer agents with a human adenocarcinoma cell line (AGS) **129**, 103
Zheng, J., see Seminoff, L.A. **54**, 251
Zhong, W.Z., see Rush, B.D. **73**, 231
Zhongshan, W., see Chow, A.H.L. **24**, 239
Zhou, F., C. Vervaet, J.P. Remon, Matrix pellets based on the combination of waxes, starches and maltodextrins **133**, 155
Influence of processing on the characteristics of matrix pellets based on microcrystalline waxes and starch derivatives **147**, 23
Zhou, M., M.D. Donovan, Recovery of the nasal mucosa following laureth-9 induced damage **130**, 93
Intranasal mucociliary clearance of putative bioadhesive polymer gels **135**, 115
Zhou, M., see Donovan, M.D. **116**, 77
Zhou, W., D.K. Moore, Photochemical decomposition of sulfamethoxazole **110**, 55
Zhou, X.H., A. Li Wan Po, Comparison of enzymic activities of tissues lining portals of drug absorption, using the rat as a model **62**, 259
Effects of cholic acid and other enhancers on the bioavailability of insulin from a subcutaneous site **69**, 29
Comparison of enzyme activities of tissues lining portals of absorption of drugs: Species differences **70**, 271
Peptide and protein drugs. I. Therapeutic applications, absorption and parenteral administration **75**, 97
Peptide and protein drugs. II. Non-parenteral routes of delivery **75**, 117
Zhu, C., W.H. Streng, Investigation of drug self-association in aqueous solution using calorimetry, conductivity, and osmometry **130**, 159
Zhu, C., see Hirate, J. **65**, 149
Zhu, C., see Streng, W.H. **135**, 43
Zhu, H., D.J.W. Grant, Influence of water activity in organic solvent + water mixtures on the nature of the crystallizing drug phase. 2. Ampicillin **139**, 33
Zhu, H., C. Yuen, D.J.W. Grant, Influence of water activity in organic solvent + water mixtures on the nature of the crystallizing drug phase. 1. Theophylline **135**, 151
Zia, H., see Dondeti, P. **122**, 91; **127**, 115
Zia, H., see Dua, R. **147**, 233
Zia, H., see Jimenez-Castellanos, M.R. **89**, 223; **105**, 65
Zietlow, D.C., see Jenke, D.R. **78**, 115
Zimmer, A., see Langer, K. **110**, 21
Zimmer, A., see Pirker, S. **128**, 189
Zimmer, A.K., P. Maincent, P. Thouvenot, J. Kreuter, Hydrocortisone delivery to healthy and inflamed eyes using a micellar polysorbate 80 solution or albumin nanoparticles **110**, 211
Zimmer, T., see Thoma, K. **58**, 197
Zimmermann, I., Determination of pK_a values from solubility data **13**, 57
Determination of overlapping pK_a values from solubility data **31**, 69
Zingel, C., A. Sachse, G.L. Rossling, R.H. Muller, Lyophilization and rehydration of iopromide-carrying liposomes **140**, 13
Zingerman, J.P., S.C. Mehta, J.M. Salter, G.W. Radebaugh, Validation of a computerized image analysis system for particle size determination. Pharmaceutical applications **88**, 303
Zingone, G., see Lovrecich, M. **131**, 247
Zingone, G., see Moneghini, M. **63**, R1
Zinutti, C., F. Kedzierewicz, M. Hoffman, J.P. Benoit, P. Maincent, Influence of the casting solvent on the physicochemical properties of 5-fluorouracil loaded microspheres **133**, 97
Zinutti, C., see Kedzierewicz, F. **94**, 69; **117**, 247
Zipp, G.L., N. Rodriguez-Hornedo, Determination of crystal growth kinetics from desupersaturation measurements **51**, 147
The mechanism of phenytoin crystal growth **98**, 189
Zivanovic, L., see Jovanovic, M. **75**, 155
Zmaily, E., see Jalal, I. **52**, 63
Zoglio, M.A., see Carstensen, J.T. **1**, 65
Zoglio, M.A., see Shenouda, L.S. **61**, 127
Zografi, G., G.P. Grandolfi, M.J. Kontny, D.W. Mendenhall, Prediction of moisture transfer in mixtures of solids: Transfer via the vapor phase **42**, 77
Zografi, G., B.A. Johnson, Effects of surface roughness on advancing and receding contact angles **22**, 159
Zografi, G., M.J. Kontny, A.Y.S. Yang, G.S. Brenner, Surface area and water vapor sorption of microcrystalline cellulose **18**, 99
Zografi, G., see Ahlneck, C. **62**, 87
Zografi, G., see Saleki-Gerhardt, A. **101**, 237
Zolezio, H., see Small, D. **149**, 203
Zolfaghari, M.E., see Dehpour, A.R. **119**, 133
Zorc, B., D. Maysinger, I. Kalcic, I. Butula, Macromolecular prodrugs. V. Polymer–broxuridine conjugates **123**, 65
Zorc, B., M. Ljubic, S. Antolic, J. Filipovic-Grcic, D. Maysinger, T. Alebic-Kolbah, I. Jalsenjak, Macromolecular prodrugs. II. Esters of L-dopa and α-methyldopa **99**, 135
Zorc, B., see Filipovic-Grcic, J. **116**, 39
Zorc, B., see Jaksic, P. **135**, 177
Zorc, B., see Kalcic, I. **136**, 31
Zorc, B., see Maysinger, D. **63**, 149
Zuber, G.E., see Lindenbaum, S. **26**, 123
Zuber, M., B. Pellion, P. Arnaud, J.C. Chaumeil, Kinetics of theophylline release from suppositories in vitro: Influence of physicochemical parameters **47**, 31
Zuber, M., see Dumortier, G. **59**, 1
Zuberbuhler, P., see Finlay, W.H. **149**, 63
Zuidam, N.J., D.J.A. Crommelin, Differential scanning calorimetric analysis of dipalmitoylphosphatidylcholine-liposomes upon hydrolysis **126**, 209
Zuidema, J., F. Kadir, H.A.C. Titulaer, C. Oussoren, Release and absorption rates of intramuscularly and subcutaneously injected pharmaceuticals (II) **105**, 189

Zuidema, J., F.A.J.M. Pieters, G.S.M.J.E. Duchateau, Release and absorption rate aspects of intramuscularly injected pharmaceuticals **47**, 1
Zuidema, J., see Duchateau, G.S.M.J.E. **31**, 193, 275; **34**, 131
Zuidema, J., see Kadir, F. **64**, 171
Zuidema, J., see Patel, C.I. **39**, 93
Zuidema, J., see Pieters, F.A.J.M. **39**, 267
Zuidema, J., see Titulaer, H.A.C. **69**, 83
Zuidema, J., see Van de Donk, H. **12**, 57, 67
Zuideman, J., see Kadir, F. **81**, 193
Zulliger, H.W., see May, H.A. **19**, 169
Zupan, J., see Bodor, N. **7**, 63
Zupancic, I., see Srcic, S. **87**, 1
Zurlo, M., see Muggetti, L. **136**, 81
Zuurman, K., G.K. Bolhuis, H. Vromans, Effect of binder on the relationship between bulk density and compactibility of lactose granulations **119**, 65
Zuurman, K., K.A. Riepma, G.K. Bolhuis, H. Vromans, C.F. Lerk, The relationship between bulk density and compactibility of lactose granulations **102**, 1
Zuurman, K., see Riepma, K.A. **76**, 9; **85**, 121; **97**, 29
Zuurman, K., see Van der Voort Maarschalk, K. **140**, 185
Zwart, P., see Marttin, E. **141**, 151
Zweibaum, A., see Caro, I. **116**, 147
Zwinkels, J., see Westerhuis, J.A. **143**, 151

ELSEVIER

International Journal of Pharmaceutics Vols. 1–149 (1997) 201–271

international journal of pharmaceutics

Subject Index

Volumes 1–149

A

A-tab **110**, 37
Abamectin **103**, 93
Abbott-79175 **115**, 95; **123**, 217
Abietic acid **137**, 11
Abietic acid **137**, 11
Abil-B-8842 **75**, 155
Abil-B-8843 **75**, 155
Abil-B-8851 **75**, 155
Abil-B-9950 **75**, 155
Abil-WE-09 **107**, 51
Absorption **1**, 323, 337; **2**, 101, 215; **3**, 13, 23, 143, 247, 299; **4**, 27, 99, 195, 205, 219, 249, 271, 281, 291; **5**, 25, 127; **6**, 1, 43, 155, 175, 243, 285; **7**, 19, 83, 99, 137, 145, 205, 317; **8**, 193, 253, 263, 277; **9**, 17, 107, 165, 173, 221, 353; **10**, 125, 143, 153, 219, 259, 339; **11**, 119; **12**, 107, 153, 351; **13**, 99, 103, 345; **14**, 149, 251, 279; **15**, 13, 125, 167, 185, 235; **16**, 23, 47, 163, 181, 191, 241, 255, 285, 327, 339, 353; **17**, 23, 47; **34**, 35; **100**, 249
ABT-077 **140**, 119
Acacia **6**, 277; **23**, 265; **32**, 207; **50**, 205; **51**, 161; **89**, 177
Accurel **45**, 65
Acebutolol **31**, 179; **36**, 165; **46**, 95; **49**, 141; **63**, 73; **76**, 25; **93**, 13; **119**, 139; **120**, 215; **123**, 149; **130**, 179
Acedapsone **32**, 21
Acefylline **50**, 205
Acemetacin **129**, 147
Acenocoumarol **12**, 231, 355**17**, 1; **19**, 307; **59**, 137; **92**, 1; **93**, 13
Acepromazine **35**, 129; **43**, 59
Acetal **123**, 95
Acetamide **7**, 189
Acetazolamide **6**, 205; **7**, 129; **51**, 27; **110**, 169; **121**, 181; **127**, 293; **133**, 117
Acetiamine **35**, 129; **43**, 59
Acetic acid **7**, 189; **11**, 131; **23**, 87; **32**, 187; **33**, 235; **42**, 181; **49**, 15; **75**, R1; **78**, 237; **86**, 43; **105**, 103; **132**, 53
Acetic acid derivative **38**, 147, 161
Acetic acid ethyl ester **29**, 177; **55**, 17; **58**, 93; **78**, 17; **101**, 63; **126**, 129; **129**, 51, 283; **140**, 237; **141**, 71; **142**, 251; **145**, 247; **148**, 41
Acetoacetic acid ethyl ester **33**, 235; **40**, 257
Acetohexamide **5**, 223; **146**, 93
Acetone **17**, 77; **20**, 273; **21**, 119; **23**, 163; **34**, 217; **87**, 239; **87**, 247; **126**, 129; **129**, 51; **129**, 283; **133**, 97; **142**, 251; **145**, 247
Acetonitrile **34**, 217; **38**, 71; **72**, 123; **113**, 97; **115**, 123; **126**, 129; **140**, 237; **141**, 71
Acetophenone **2**, 295; **9**, 29; **100**, 189
2-Acetotoluidide **36**, 223
3-Acetotoluidide **36**, 223
4-Acetotoluidide **36**, 223
4-Acetoxyacetanilide **24**, 239; **25**, 57; **30**, 161; **57**, 117
1[4(2-Acetoxyethyl)phenoxy]-3-isopropylamino-2-propanol **66**, 23
1-Acetoxymethyl-5-fluorouracil **24**, 43
7-(Acetyloxymethyl)theophylline **12**, 299
2-Acetoxymethylbenzamide derivative **29**, 9
α-Acetoxystyrene **9**, 29
Acetrizoic acid **6**, 205
N-(3-Acetyl-5-ethyl-2-hydroxyphenyl)-1H-tetrazole-5-carboxamide **30**, 101
N-Acetyl-β-glucosaminidase **46**, 103
N-Acetyl-L-cysteine **40**, 85
O-Acetyl-*n*,*n*-diethylsalicylamide **64**, 45
4[4[3(4-Acetyl-3-hydroxy-2-propylphenoxy)propylthio]phenyl]-4-hydroxy-3-methylbutyric acid **109**, 237
O-Acetyl-*n*-methylsalicylamide **64**, 45
N-Acetyl-*n*-methyl-4-toluenesulfonamide **37**, 87
Acetylacetone **22**, 197; **108**, 95
1-Acetylallopurinol **23**, 223
N-Acetylampicillin **17**, 47
Acetylated calcitonin **134**, 47
Acetylcholine **39**, 263
Acetylcholinesterase **60**, 219
Acetylcysteine **7**, 99; **23**, 175; **79**, 97; **124**, 9
Acetylcysteine derivative **62**, 193
Acetyldapsone **32**, 21; **47**, 1
Acetyldigoxin **2**, 127
α-Acetyldigoxin **2**, 127
N-Acetylgalactosamine-4-sulfatase **88**, 293
N-Acetylglutamic-acid-5[*n*-2(5-butyl-2-pyridyl)-hydrazide] **46**, 203
7-*N*-Acetylmitomycin-C **52**, 239
N-Acetylmuramylalanyl-dextro-isoglutamine **23**, 299
N-Acetylneurotensin[8–13] **112**, 133
6′-Acetylpapaverine **9**, 29; **10**, 29, 239

0378-5173/97/$17.00 © 1997 Elsevier Science B.V. All rights reserved.

PII S0378-5173(97)00262-7

6′-Acetylpapaverine-enol-acetate **10**, 29
6′-Acetylpapaverine-enol-isobutyrate **10**, 29
6′-Acetylpapaverine-enol-pivalate **10**, 29
O-Acetylsalicylamide **13**, 273; **30**, 111
O-Acetylsalicylsalicylamide **13**, 273
Acetylsalicylic acid **2**, 91, 145, 195, 283; **4**, 127, 171; **5**, 167; **6**, 205; **7**, 269, 343; **10**, 143; **11**, 271; **13**, 35; **14**, 1, 123, 243; **15**, 97, 167; **16**, 241, 349; **18**, 1, 287, 299; **22**, 291; **25**, 255; **26**, 15, 329; **28**, 133; **29**, 195; **31**, 25, 201; **32**, 257; **34**, 35, 75, 259; **37**, 15, 203; **44**, 177, 235; **45**, 181; **47**, 1, 121, 261, 263; **52**, 265; **54**, 131; **55**, 47; **61**, 67, 219; **64**, 147; **68**, 77; **74**, 49; **75**, 231; **80**, 263; **83**, 163; **84**, 261; **91**, 173; **91**, 75, 123, 151; **100**, 257; **107**, 247; **108**, 11, 109, 173, 213; **114**, 205; **119**, 37; **121**, 107; **123**, 143; **125**, 157; **127**, 177, 185; **133**, 65; **137**, 41; **140**, 61; **144**, 187; **145**, 165; **148**, 1, 73
Acetylsalicylic acid derivative **55**, 1
Acetylsalicyloylphenylalanine ethyl ester **26**, 15
N-Acetylsulfafurazole **23**, 345
N-4-Acetylsulfafurazole **7**, 179
N-4-Acetylsulfanilamide **88**, 437
2-Acetylthiomethyl-3(4-methylbenzoyl)propionic acid **124**, 129
O-Acetyltimolol **33**, 15
Acetyl tributyl citrate **103**, 47, 293; **111**, 89
Acetyl triethyl citrate **103**, 47
Aciclovir **38**, 221; **57**, 229; **65**, 183; **89**, 199; **93**, 139; **99**, 79; **111**, 73; **112**, 181; **120**, 119; **123**, 159; **125**, 215; **133**, 59; **135**, 91; **139**, 231; **145**, 103
Aciclovir derivative **112**, 181; **139**, 231; **145**, 103
Aciclovir isovalerate **145**, 103
Aciclovir pivalate **145**, 103
Aciclovir valerate **145**, 103
Acid **31**, 69; **87**, 11
Acid base balance **80**, 135; **87**, 11; **89**, 103, 261
Acidification **84**, R1
Acinetobacter glutaminasificans **34**, 225
Acipimox **95**, 127
Aclarubicin **116**, 1; **135**, 167
ACN-58548 **113**, 123
ACN-84530 **113**, 123
Acne **85**, 97
Acoustic emission **40**, 15
Acoustics **125**, 45
Acquired immune deficiency syndrome **50**, 175; **67**, 275; **95**, 1
Acrivastine **104**, 253; **136**, 117
Acrolein **90**, 119
Acronine **5**, 149
Acrylamide **32**, 165; **84**, 23
Acrylic acid **84**, 23; **95**, 153; **135**, 1
Acrylic acid butyl ester **84**, 23
Acrylic acid copolymer **145**, 157
Acrylic acid resin **103**, 293; **127**, 203; **131**, 183; **142**, 143
Acryloyl chloride **21**, 277
Activated carbon **5**, 223; **64**, 109
Acute heart failure **140**, 61
Acute lymphoblastic leukemia **59**, 27
N-Acyl-5-fluorouracil derivative **21**, 349
1-Acylallopurinol **23**, 223
Acylation **93**, 177
Acylglycerol **39**, 235; **110**, 75; **111**, 51
1-Acyloxymethyl-5-fluorouracil **12**, 153; **24**, 43
3-Acyloxymethyl-5-fluorouracil **24**, 43
1-Acyloxymethylallopurinol **24**, 307
5-Acyloxymethylallopurinol **24**, 307
Adamantyl-*n*-alkanamide derivative **126**, 35
4[[2[2[[(2-Adamantyloxy)carbonyl]amino]-3-(1H-indol-3-*yl*)-2-methylpropionamido]-1-phenylethyl]amino]-4-oxobutyric-acid-meglumine **92**, 63
Addamel **13**, 9
Adenine **59**, 45; **145**, 145
Adenosine **59**, 45; **145**, 145
Adenosine deaminase **65**, 57
Adenosine derivative **65**, 57
Adenosine receptor **145**, 145
Adenosine triphosphate **22**, 137
S-Adenosylhomocysteine **81**, 267
S-Adenosylmethionine **72**, 97; **81**, 267
Adhesion **19**, 107; **51**, 203; **52**, 149; **53**, 47, 75; **59**, 205; **64**, 227; **73**, 69, 75; **78**, 43; **79**, 97, 257; **80**, 1; **81**, 59; **83**, 221; **84**, 69; **94**, 223; **107**, 91; **113**, 223; **116**, 223; **118**, 257; **124**, 173; **129**, 137; **130**, 257; **134**, 173; **138**, 67, 103; **141**, 251; **142**, 169; **145**, 65, 231
Adhesive agent **64**, 117; **72**, 227; **76**, 77; **79**, 97; **133**, 1; **135**, 1; **145**, 137
Adinazolam mesilate **142**, 53
Adinazolam **99**, 311
Adipic acid **20**, 3; **25**, 41; **28**, 75, 103; **57**, 117; **84**, 167; **125**, 157
Adipic acid dibutyl ester **98**, 37
Adipose tissue **47**, 111
Adrenalin **1**, 165; **3**, 239; **13**, 263; **17**, 299; **29**, 43; **33**, 201; **42**, 47; **101**, R5; **123**, 159
Adrenalone **1**, 189
Adrenalone derivative **1**, 189
α-Adrenergic receptor blocking agent **99**, 85; **130**, 179
β-Adrenergic receptor blocking agent **5**, 167; **6**, 205; **39**, 39; **46**, 95; **93**, 49; **104**, 19; **105**, 189; **120**, 215; **123**, 149; **125**, 165; **130**, 137; **141**, 125, 217; **142**, 103
Adrenergic receptor stimulating agent **28**, 1; **92**, 1; **123**, 181; **142**, 125; **149**, 85
α-Adrenergic receptor stimulating agent **130**, 179
β-Adrenergic receptor stimulating agent **107**, 211
Adsorbent agent **27**, 17; **36**, 185
Adsorption **18**, 157; **31**, 137; **34**, 137; **58**, 229; **63**, 251; **67**, 29; **72**, 243; **73**, 185; **77**, 47; **78**, 95; **79**, R1; **81**, 21; **83**, 65; **99**, 59; **103**, 19, 87; **110**, 197; **120**, 179; **127**, 31, 43, 255; **128**, 129, 269; **132**, 143; **139**, 15; **140**, 155; **143**, 259; **145**, 231
Adverse drug reaction **12**, 277; **13**, 353; **15**, 149; **16**, 23; **19**, 307; **20**, 99; **22**, 137; **26**, 113; **30**, 215; **31**, 275; **34**, 75
Aerosol **1**, 71, 213; **3**, 335; **5**, 287; **8**, 1; **9**, 173; **27**, 99, 255; **35**, 81, 139; **36**, 55; **37**, 239; **41**, 147; **42**, 211, 267; **51**, 39; **61**, 135; **64**, 1; **78**, 227; **80**, R1; **85**, 203; **88**, 63; **91**, 111; **95**, 111; **97**, 111, 205; **98**, 45; **101**, 1, 81; **102**, 159; **104**, 73; **106**, 69; **107**, 57, 211; **108**, 233; **113**, 123; **114**, 111, 117; **115**, 69, 209;

116, 11, 169; **118**, 229; **119**, 91, 103; **123**, 257; **123**, 247, 291; **124**, 149, 195; **125**, 133; **128**, 55; **129**, 123; **130**, 1, 13, 103, 231; **133**, 203; **134**, 137; **135**, 53; **136**, 1; **137**, 261; **138**, 45, 129; **139**, 255; **142**, 33, 129
Aerosil-200 **31**, 169; **35**, 157; **50**, 147
Aerosil R 972 **81**, 67; **114**, 117
AG-85 **103**, 187; **105**, 227
Agar **3**, 13; **112**, 17
Agarose **12**, 19
Age **16**, 203
Aggregation **27**, 99; **52**, 101; **61**, 79; **63**, 177; **105**, 57, 235; **115**, 183; **124**, 165; **125**, 133; **127**, 191; **135**, 43; **138**, 57; **140**, 155; **144**, 47; **146**, 193
Airborne infection **1**, 133
Airborne particle **42**, 267
Airflow **130**, 231
Airway resistance **130**, 231
Alafosfalin **75**, 117
Alamethicin **75**, 117
Alanine **72**, 149; **115**, 139, 151; **121**, 157
β-Alanine **132**, 63
Alanine-4-methoxy-2-naphthylamide **30**, 73
Alaproclate **15**, 87; **23**, 97
Albendazole **91**, 105; **102**, 257; **113**, 47; **140**, 45, 247
Albendazole sulfoxide **102**, 257
Albumin **2**, 283; **7**, 335; **15**, 273; **15**, 25; **16**, 215; **19**, 107; **23**, 333; **25**, 147; **27**, 17; **29**, 137; **30**, 123; **32**, 237; **33**, 137; **33**, 147; **33**, 1; **36**, 113; **37**, 103; **39**, 213; **39**, 129; **43**, 167; **43**, 161; **44**, 49; **46**, 69; **47**, 163; **47**, 265; **48**, 133; **51**, 85; **51**, 253; **55**, 221; **56**, 235; **57**, 211; **57**, 205; **58**, 79; **59**, 137; **59**, 57; **62**, 21; **68**, 265; **68**, 61; **70**, 219; **75**, 117; **75**, 97; **77**, 31; **79**, 191; **80**, 101; **85**, 75; **85**, 39; **85**, 1; **86**, 153; **86**, 147; **90**, 95; **95**, 171; **96**, 85; **101**, 63; **105**, 19; **107**, 205; **109**, 135; **110**, 117; **110**, 211; **110**, 127; **112**, 215; **114**, 177; **115**, 259; **118**, 85; **119**, 57; **119**, 149; **120**, 51; **124**, 149; **124**, 213; **125**, 223; **130**, 49
Albumin blood level **7**, 335
Albumin-I^{125} **8**, 35
Albumin microsphere **22**, 63; **23**, 333
Albumin tannate **147**, 11
Alclofenac **126**, 95
Alcohol **1**, 15; **43**, 17, 225; **54**, 51; **55**, 17; **58**, 93; **63**, 57; **67**, 51; **68**, 285; **72**, R1; **74**, 105, 115; **76**, 91; **78**, 17, 123, 137; **79**, 47, 61; **80**, 17, 243, 253; **81**, R1; **82**, 145; **84**, 117; **89**, 191; **90**, 171, 203; **91**, 9; **92**, 203; **93**, 161; **95**, 23, 51, 57, 161; **98**, 37; **101**, 127; **104**, 145; **105**, 113, 189, 227; **106**, 25, 51; **110**, 137; **111**, 63, 235; **113**, 97, 165, 189; **114**, 75; **116**, 183, 253; **119**, 139; **120**, 21, 189, 197; **121**, 27; **123**, 41; **124**, 261; **125**, 107; **126**, 219; **129**, 103; **131**, 137; **132**, 23, 81; **133**, 71, 97; **137**, 125, 137; **140**, 25, 69, 237; **141**, 71; **143**, 37, 67; **147**, 51; **148**, 41
Alcohol dehydrogenase **74**, 195
Alcohol derivative **1**, 127; **87**, 47; **137**, 177; **138**, 149; **141**, 71; **148**, 1, 41
Aldehyde **23**, 163
Aldosterone **132**, 23; **138**, 149
Alexidine **55**, 209
Alfaxalone **65**, 101
Alfentanil **127**, 255
Algaroba **139**, 53
Alginic acid **19**, 169; **21**, 341; **28**, 249; **34**, 105; **45**, 39; **56**, 207; **61**, 35; **62**, 207; **79**, 11; **96**, 139; **105**, 97; **107**, 179; **111**, 171, 271; **112**, 241; **116**, 39, 125; **118**, 257; **122**, 107, 121; **126**, 161; **129**, 137; **138**, 237; **139**, 25; **143**, 195; **143**, 187; **144**, 37; **148**, 117
Algorithm **31**, 69
Alimemazine **12**, 219; **13**, 227; **35**, 129; **43**, 59; **94**, 135
Alinidine **12**, 277
Aliphatic dicarboxylic acid **123**, 209
Alizapride **123**, 285
Alkali **135**, 13
Alkaline phosphatase **46**, 103; **104**, 157
Alkaloid **83**, 131; **109**, 135
Alkane **14**, 333; **90**, 203
Alkanediol derivative **111**, 63
Alkannin **120**, 137; **141**, 171
Alkanoic acid **57**, 117; **111**, 63; **126**, 35
Alkanol **14**, 325, 333; **36**, 67; **44**, 225; **46**, 19; **80**, 17
3-Alkoxyphenol **29**, 237
3-Alkylaminobenzoic acid derivative **2**, 239
Alkylaniline **37**, 49
Alkylating agent **90**, 239
Allergen **119**, 239
Allergic rhinitis **136**, 181
Allobarbital **3**, 341; **20**, 73; **26**, 157
N-Alkylbromocresotamide c 14 **15**, 273
Allomuscarine **15**, 223
Allopurinol **5**, 167; **18**, 259; **23**, 223; **24**, 307; **25**, 27; **27**, 71; **64**, 75; **80**, 129; **136**, 117
Allopurinol derivative **23**, 223; **24**, 307; **27**, 71; **64**, 75
Allopyrabital **127**, 185
Allyltriphenylphosphonium chloride **30**, 101
Almagate **52**, 173
Almagcit **52**, 173
Alphacetylmethadol **45**, 1
Alprazolam **61**, 189; **88**, 89; **110**, 169
Alprenolol **7**, 269; **34**, 131; **36**, 165; **39**, 39; **46**, 95; **47**, 1, 133; **56**, 135; **63**, 73; **64**, 171; **104**, 19; **120**, 215; **123**, 149
Alteplase **129**, 191
Althesin **65**, 101
Altretamine **27**, 255; **78**, 89; **95**, 143
Aluminum chloride **1**, 49, 141; **45**, 249; **87**, 239; **93**, 131
Aluminum derivative **144**, 37
Aluminum glycinate **19**, 307; **34**, 189
Aluminum hydroxide **1**, 49; **19**, 169, 307; **20**, 99; **21**, 341; **22**, 257; **28**, 151, 239, 249; **34**, 189, 207; **36**, 185; **52**, 173; **68**, 11; **73**, 171; **75**, R9; **80**, R21; **97**, 141; **105**, 97; **129**, 103; **148**, 117
Aluminum hydroxychloride **1**, 49
Aluminum ion **113**, 257
Aluminum magnesium hydroxide **22**, 257; **28**, 249; **69**, 189
Aluminum magnesium hydroxide **69**, 189
Aluminum magnesium sodium silicate **18**, 157; **119**, 71
Aluminum oxide **27**, 17; **83**, 197
Aluminum sodium silicate **127**, 147

Aluminum **33**, 261; **54**, 143; **87**, 73, 203; **106**, 229; **113**, 133; **127**, 147
Alverine citrate **35**, 129; **43**, 59
Amantadine **126**, 35
Amaranth **56**, 269; **83**, 263
Amastatin **95**, 171; **121**, 217
Ambenyl **7**, 343
Amberlite-cg-400 **25**, 191
Americium-241 **131**, 243
AMF-297 **131**, 33
AMF-333 **131**, 33
Amfebutamone **104**, 253
Amidal **71**, 31; **123**, 95
Amide **71**, 31; **110**, 231
Amifloxacin **63**, 237; **79**, 89; **83**, 267; **88**, 379; **93**, 121
Amikacin **30**, 181; **42**, 193; **67**, 103; **68**, 69; **78**, 227; **87**, 183; **105**, 189; **124**, 149
Amiloride **34**, 75; **51**, 169; **88**, 111; **91**, 195; **98**, 83**101**, R7
Amine derivative **40**, 235, 249
Amine **29**, 9, 19; **111**, 63; **116**, 201; **131**, 33
Amino acid **2**, 325; **13**, 9; **19**, 207; **61**, 189; **68**, 77; **72**, 149; **77**, 57; **79**, 77; **79**, 39; **81**, 1, 253, 267; **83**, 123, 227; **85**, 7; **87**, 167; **90**, 187; **102**, 223; **103**, 253; **105**, 169; **106**, 85; **107**, 23; **111**, 7; **117**, 181; **121**, 157; **123**, 41; **132**, 63
Amino acid derivative **46**, 159; **83**, 131; **146**, 205
Amino acid transport **104**, 157
2-Amino-5-chlorobenzophenone **5**, 85
4-Amino-4-deoxy-10-methylpteroic acid **16**, 141
7-Amino-1,3-dihydro-5-phenyl-2H-1,4-benzodiazepin-2-one **61**, 189
1-*N*-(3-Amino-2-hydroxypropionyl)gentamicin B **37**, 113
[*n*(3-Amino-3-methylbutyryl)-4-methoxyphenylalanyl]-*n*(1-cyclohexylmethyl) **63**, 167
2-Amino-2-methylpropanol **114**, 205
3-Amino-1-naphthoic acid **88**, 15
4-Amino-2-naphthoic acid **88**, 15
4-Amino-1,2,4-triazole **81**, 103
Aminoacid amide **20**, 273
Aminoacid transport **32**, 39
2-Aminobenzamide **91**, 39
3-Aminobenzeneboronic acid **80**, 9
Aminobenzoic acid **82**, 107
3-Aminobenzoic acid **88**, 15
4-Aminobenzoic acid **24**, 231; **87**, 195; **116**, 217; **133**, 47
4-Aminobenzoic acid butyl ester **49**, 91
Aminobenzoic acid derivative **70**, 209; **77**, 127, 239
4-Aminobenzoic acid derivative **144**, 127
4-Aminobenzoic acid ester **57**, 149
4-Aminobenzoic acid methyl ester **6**, 179; **127**, 27
4-Aminobenzoic acid pentyl ester **49**, 91
4-Aminobutyric acid **24**, 297; **132**, 63
(2-Aminoethyl)carbamate derivative **112**, 143
Aminoglycolipid **115**, 183
Aminoglycoside **26**, 133; **37**, 113
Aminoglycoside antibiotic agent **81**, 1; **90**, 59
4-Aminohippuric acid **38**, 261
1-Aminohydantoin **8**, 111; **17**, 357
Aminolysis **9**, 273
6-Aminopenicillanic acid **12**, 185
Aminopeptidase **30**, 73; **48**, 189; **70**, 271; **71**, 45; **76**, 113, 247; **81**, 119; **97**, 171; **105**, 7; **117**, 181; **121**, 157
Aminophenazone **6**, 179; **28**, 67; **79**, 21; **81**, 143; **143**, 37; **148**, 155
Aminophylline **3**, 299; **6**, 11; **14**, 1; **24**, 339; **30**, 143; **33**, 55; **36**, 73; **37**, 97; **38**, 139; **40**, 223; **41**, 55; **49**, 183; **50**, 205; **51**, 245; **59**, 105; **60**, 125; **66**, 111, 233; **72**, 175; **81**, 1; **83**, 59; **86**, 231; **129**, 279; **137**, 75; **141**, 109; **148**, 155
Aminoplex **83**, 227
Aminopropanol derivative **8**, 193; **36**, 205
8-Aminoquinoline **99**, 67
Aminosalicylic acid **88**, 177; **126**, 179, 199; **134**, 15
Aminosalicylic acid derivative **122**, 121; **126**, 199
Aminothiazole **67**, 265; **81**, 103
Aminoxid-WS-35 **75**, 155
Amiodarone **122**, 1
Amitriptyline **4**, 47; **73**, 125; **94**, 135; **115**, 103
Amlodipine **133**, 71
Amlodipine besylate **133**, 71
Ammonium chloride **87**, 239; **135**, 31
Ammonium glycyrrhizate **122**, 91
Ammonium hydroxide **80**, 253
Ammonium sulfate **57**, 205; **134**, 193; **135**, 31
Amobarbital **20**, 73; **26**, 157; **41**, 139; **42**, 97, 181; **45**, 1; **60**, 235; **65**, 141; **103**, 19
Amodiaquine **56**, 213
Amoxapine **94**, 135
Amoxicillin **4**, 159; **9**, 273; **18**, 157; **19**, 233; **26**, 133; **30**, 181; **31**, 137; **78**, 35; **82**, 157; **85**, 181; **86**, 79; **110**, 109; **115**, 79; **148**, 191
Ampholyte **45**, 157; **111**, 89; **113**, 47; **137**, 125, 137
Amphophile **86**, 175; **115**, 103
Amphotericin B **29**, 161; **44**, 117; **46**, 25, 169, 223; **54**, 41; **58**, 41; **75**, 45; **80**, 253; **81**, R5; **89**, 139; **118**, 129; **137**, 249; **145**, 17, 241
Amphotericin B deoxycholate **145**, 17
Ampicillin derivative **27**, 175
Ampicillin **1**, 95; **3**, 1, 151; **4**, 159; **6**, 225, 301; **7**, 55, 301; **9**, 91, 233, 239, 273; **12**, 67, 185; **14**, 1; **17**, 47; **18**, 157; **19**, 233; **22**, 147; **23**, 163; **26**, 133; **27**, 175; **30**, 181; **31**, 137; **35**, 121; **37**, 113; **41**, 241; **55**, 77; **62**, 1; **73**, 33; **74**, 183; **75**, 117; **77**, 13; **82**, 117; **85**, 181; **90**, 181; **97**, 219; **99**, 119; **104**, 187; **106**, 33; **110**, 109; **115**, 79; **137**, 1; **139**, 33; **144**, 177; **148**, 191
Amsacrine **42**, 41
Amylase **101**, 165; **102**, 81; **134**, 223
Amylopectin **79**, 97; **134**, 27
Amylose **134**, 27; **136**, 43
Analbuminemia **65**, 149
Analgesia **38**, 171; **42**, 251
Analgesic activity **57**, 55, 77
Analgesic agent **21**, 35; **92**, 1, 15; **148**, 1
Analysis **59**, 211
Analytic method **76**, 171
Anandamide **137**, 209
Anaplastic carcinoma **123**, 237

Androgenic agent **6**, 195; **14**, 325, 333
Androstanediol **131**, 263
Androstanolone **80**, 253; **131**, 263
Androstenediol **80**, 253
Androstenedione **80**, 253; **131**, 263
Androstenedione H 3 **4**, 249
Anemia **111**, 197
Anesthesia **38**, 261; **65**, 101
Anesthetic agent **9**, 353; **92**, 1; **123**, 1; **139**, 215; **149**, 123
Angina pectoris **131**, 229
Angiogenesis **123**, 237
Angiogenesis inhibitor **123**, 237
Angiopeptin **99**, 165; **113**, 83
Angiotensin **121**, 195
Angiotensin-II-antagonist **103**, 1
Anhydroepitetracycline **26**, 259
Anhydroerythromycin A **30**, 199
Anhydrotetracycline **1**, 185; **26**, 259
Aniline **17**, 331; **37**, 49; **55**, 165; **57**, 189; **77**, 1; **100**, 1; **129**, 33, 283
Aniline derivative **1**, 127; **44**, 187; **55**, 165; **77**, 1; **85**, 199; **129**, 33
N-(Anilinomethyl)succinimide **9**, 7
Animal cell **68**, 191
Animal model **30**, 91
Anion **77**, 107; **137**, 217; **145**, 129, 175
Anionic surfactant **129**, 113
Anipamil **64**, 117
Anistreplase **93**, 177
4-Anisyl-curcuminoid **131**, 1
Antacid agent **7**, 83; **13**, 313; **19**, 169, 307; **20**, 99; **21**, 341; **27**, 279; **28**, 151, 239, 249; **29**, 253; **30**, 225; **34**, 105, 189, 207; **36**, 185; **69**, 189; **75**, R9; **106**, 229; **148**, 117
Antazoline **12**, 77; **65**, 137; **86**, 59
Antazyme **28**, 239
Anthelmintic agent **77**, 265; **132**, 153
Anthra[1,9cd]pyrazol-6(2H)one derivative **71**, 73
Anthracene **87**, 47; **140**, 237
Anthracycline antibiotic agent **135**, 167
2-(9-Anthroyloxy)palmitic acid **126**, 49
Antiandrogen **65**, 77; **126**, 235
Antiarrhythmic agent **22**, 345; **28**, 145; **93**, 13; **103**, 103; **122**, 1
Antibacterial activity **75**, 131; **77**, 13; **83**, 123; **85**, 97; **87**, 73, 195; **98**, 1; **101**, R7; **116**, 217; **119**, 183; **126**, 261; **131**, 33; **147**, 51
Antibiotic agent **5**, 167; **6**, 205; **9**, 305; **21**, 239; **25**, 105, 245; **28**, 1; **30**, 181; **36**, 55; **75**, 97, 117; **81**, 1; **83**, 267; **84**, 77; **90**, 181; **92**, 1, 15, 47; **105**, 189; **115**, 79; **122**, 9; **123**, 1, 291; **124**, 149; **143**, 143
Antibiotic resistance **75**, 81
Antibiotic sensitivity **34**, 241; **35**, 227; **75**, 81
Antibody **108**, 133
Antibody production **103**, 303
Antibody response **76**, 161; **144**, 107
Anticoagulant agent **6**, 185, 205; **12**, 355; **18**, 39; **19**, 307; **93**, 13; **131**, 131
Anticonvulsive agent **74**, 77; **126**, 65
Antidepressive agent **6**, 205; **21**, 51
Antidiabetic agent **15**, 261; **128**, 155; **146**, 93
Antidiarrheal agent **30**, 225; **34**, 207; **36**, 185; **89**, 75; **119**, 1
Antidote **46**, 247; **109**, 35; **137**, 167
Antidote to organophosphorus poisoning **40**, 157
Antiemetic agent **9**, 353; **121**, 95; **123**, 285
Antifungal activity **58**, 41; **90**, 247; **117**, 123
Antifungal agent **4**, 57; **66**, 107; **94**, 11; **105**, 77
Antigen **59**, 27; **75**, 1; **90**, 255
Antiglaucoma agent **132**, 165
Antihistaminic agent **9**, 353
Antihypersensitivity agent **12**, 77
Antihypertensive agent **66**, 23; **142**, 61
Antiinfective agent **64**, R1; **84**, 77; **86**, 193; **98**, 1; **110**, 109; **118**, 41; **131**, 33; **147**, 127
Antiinflammatory activity **106**, 63; **120**, 145; **145**, 29
Antiinflammatory agent **28**, 47; **68**, 191; **84**, 93; **91**, 1; **104**, 169; **123**, 127
Antilipemic agent **15**, 261; **108**, 85
Antimalarial activity **110**, 189
Antimalarial agent **128**, 163; **132**, 71
Antimicrobial activity **56**, 169; **60**, 147; **72**, 51; **93**, 201; **103**, 199; **110**, 109; **118**, 65; **148**, 131
Antimicrobial agent **1**, 127; **12**, 67, 295; **16**, 127; **17**, 291; **22**, 283; **34**, 115; **35**, 227
Antimony tartrate **56**, 23
Antineoplastic activity **128**, 139
Antineoplastic agent **13**, 303; **22**, 75; **23**, 299, 333; **25**, 347; **28**, 1; **55**, 1; **65**, 57; **66**, 253; **71**, 157; **72**, 251; **78**, 217; **80**, 51; **81**, 1; **92**, 1; **95**, 85, 143; **100**, 181; **102**, 63; **103**, 87; **105**, 189; **107**, 149; **108**, 69; **109**, 73; **110**, 277; **112**, 97; **113**, 103; **118**, 161; **123**, 1, 237; **124**, 205, 261; **125**, 55; **127**, 229; **128**, 261; **131**, 1, 73; **139**, 1; **143**, 255
Antineoplastic antibiotic **112**, 97
Antioxidant **90**, 19; **110**, 241; **117**, 189; **124**, 149; **127**, 73; **131**, 1; **140**, 33, 169, 251; **147**, 31
Antioxidant activity **145**, 87
Antipsoriasis agent **65**, 115; **103**, 187; **105**, 227
Antirheumatic agent **124**, 129, 279
Antisense oligonucleotide **133**, 257; **136**, 23; **141**, 1
α-1-Antitrypsin **148**, 93
Antiulcer agent **73**, 23; **77**, 65, 247; **84**, 39; **91**, 217; **95**, 1; **105**, 189; **106**, 85; **117**, 231; **123**, 159; **124**, 149; **129**, 103
Anxiety **33**, 37
Apalcillin **77**, 47
Aphthous ulcer **95**, 105
Apomorphine **29**, 73; **146**, 213; **148**, 23
Apparatus **1**, 71; **31**, 247; **121**, 129; **122**, 79; **133**, 59, 127, 133; **134**, 137, 243
Appendix carcinoma **2**, 245
Apricot kernel oil **106**, 15
Aprobarbital **42**, 181
Aprotinin **8**, 253; **69**, 29; **75**, 97; **78**, 1, 9; **81**, 119; **91**, 29; **95**, 101; **113**, 1; **121**, 245; **123**, 199; **126**, 245; **129**, 263
Aqoat **126**, 1
Aquateric **126**, 1

Aqueous acidic solution **40**, 157
Aqueous humor **11**, 21; **66**, 23
Aqueous solution **3**, 319; **37**, 11; **130**, 159, 245; **141**, 197
Arachidic acid **146**, 239
Arachidonate-15-lipoxygenase **50**, 67; **51**, 179
Arachidonic acid metabolism **46**, 149
Arachis oil **11**, 277; **12**, 315; **24**, 173; **38**, 133; **44**, 159; **106**, 15; **117**, 129; **119**, 173; **126**, 89
Arachis oil-I^{123} **12**, 315
Arachis oil-I^{131} **16**, 31
Arecaidine derivative **13**, 89
Arecoline **88**, 251
Arginine-4-methoxy-2-naphthylamide **30**, 73
Arginine **72**, 149; **85**, 7; **105**, 209
Argipressin **46**, 111; **56**, 13; **61**, 179; **75**, 117; **81**, 119; **86**, 35
Argipressin[deglycinamide] **76**, 37; **141**, 39
Argon **63**, 155
Aromatase inhibitor **71**, 19; **75**, 141, 147
Aromatic carboxylic acid **88**, 15
Aromatic compound **15**, 347
Arteether **69**, 83; **128**, 163
Artelinic acid **69**, 83; **105**, 189
Artemether **69**, 83
Artemisinin **69**, 83; **105**, 189; **138**, 185; **146**, 271
Artesunate **69**, 83
Arthritis **22**, 137; **24**, 101; **42**, 251
Artificial embolism **106**, 41
Artificial membrane **68**, 43, 151; **79**, 21
Artificial neural network **148**, 103
Arylaliphatic carboxylic acid **69**, 221; **107**, 159
Arylazide derivative **52**, 231
Arylbutyric acid derivative **139**, 205
Asbestos **27**, 17
Ascites tumor **131**, 1
Ascorbic acid **2**, 145; **7**, 269; **10**, 209; **12**, 185; **22**, 147; **29**, 195; **31**, 83; **35**, 111; **36**, 233, 243; **49**, 1; **63**, 155; **78**, 85; **81**, 1; **83**, 263; **88**, 39; **90**, 19; **100**, 93; **107**, 199; **110**, 241; **111**, 261; **115**, 161, 167; **126**, 29; **127**, 73; **132**, 45, 251; **133**, 85; **140**, 251; **145**, 9
Ascorbic acid derivative **22**, 147
Asialofetuin **42**, 69; **67**, 123
Asiaticoside **146**, 63
Asparaginase **22**, 137; **34**, 225; **75**, 97; **96**, 67; **142**, 75
Asparagine **34**, 225; **72**, 149
Aspartame **75**, R5; **86**, 263; **88**, 189; **107**, 199
Aspartic acid **38**, 239; **72**, 149; **99**, 43; **123**, 41
Aspartic acid derivative **64**, 239
Aspergillus niger **10**, 193; **30**, 241; **46**, 141; **54**, 51
Assay **55**, 99
Association **130**, 159
Asthma **3**, 55; **4**, 135; **8**, 1; **25**, 113; **51**, 67; **74**, 209; **97**, 47; **101**, 1; **107**, 211; **114**, 111; **130**, 231; **138**, 45
Atenolol plus chlortalidone **65**, 177
Atenolol **36**, 165; **46**, 95; **56**, 135; **63**, 73; **64**, 171; **65**, 177; **81**, 235; **89**, 245; **95**, 161; **100**, 257; **104**, 19; **105**, 189; **108**, 11; **120**, 215; **123**, 149; **125**, 165; **130**, 137, 179; **137**, 133; **141**, 217; **142**, 103
Atherosclerosis **98**, 165
Atmosphere **103**, 103
Atrial natriuretic factor **77**, 309
Atrolactic acid **81**, 31
Atropine **1**, 165; **5**, 25; **31**, 33; **43**, 59; **55**, 105; **93**, 239; **105**, 189; **109**, 35; **114**, 125; **138**, 149
Attapulgite **18**, 157
Auranofin **26**, 123
Aurin **9**, 29
Aurin sulfoacetate **10**, 239
Aurothiomalate **8**, 217
Autoclave **12**, 31; **92**, 233
Automation **30**, 123; **34**, 1; **88**, 303
Autooxidation **45**, 129; **65**, 261
Autoradiography **16**, 105; **21**, 341
Avenin **126**, 103
Avicel PH 101 **107**, 141; **110**, 203
Avicel PH 102 **37**, 203; **46**, 35; **48**, 51; **56**, 51; **57**, 95; **71**, 201; **75**, 181; **83**, 65
Avicel PH 103 **101**, 169; **128**, 123; **131**, 201
Avicel PH 105 **101**, 169; **131**, 201
Avicel PH 200 **101**, 169
Azathioprine **11**, 209; **25**, 155; **28**, 119; **35**, 165; **42**, 263; **46**, 267; **68**, 105
Azelaic acid **69**, 193
Azelastine **123**, 47; **136**, 181
Azide **69**, 197
3′-Azidopyrimethamine **52**, 231; **65**, 115
Azidoprofen **89**, 51, 65; **90**, 129
5(1-Aziridinyl)-3-hydroxymethyl-2(3-hydroxy-1-propenyl)-1-methyl-4,7 **100**, 181
Azithromycin **133**, 237
Azlocillin **26**, 133; **58**, 103; **135**, 111
Azo compound **106**, 255; **121**, 211
Azo dye **106**, 255
Azo reductase **106**, 255; **121**, 211; **139**, 187
Azobenzene derivative **87**, 37; **99**, R5
Azobenzene **106**, 255
Azone derivative **116**, 201
Aztreonam **67**, R5; **91**, 217; **110**, 249

B

Bacampicillin **9**, 91; **13**, 89
Bacillus cereus **21**, 265
Bacillus megaterium **30**, 237
Bacillus pumilis **7**, 279; **21**, 265
Bacillus subtilis **17**, 291; **25**, 105; **31**, 231; **34**, 51; **35**, 121; **37**, 113; **65**, 127; **119**, 247; **126**, 261
Bacitracin **12**, 67; **116**, 77
Baclofen **118**, 181; **132**, 63; **137**, 125, 137
Bacterial cell wall **25**, 239; **131**, 83
Bacterial colonization **58**, 165
Bacterial count **60**, 147
Bacterial growth **2**, 325; **19**, 239; **25**, 105; **32**, 55; **58**, 155, 165; **73**, 97; **101**, R7–R20
Bacterial resistance **12**, 295; **17**, 291; **19**, 239; **34**, 115; **75**, 81; **87**, 195; **101**, R7–R10

Bactericidal agent **4**, 57
Bacterium **1**, 165; **16**, 127; **25**, 329; **61**, 57; **99**, 293
Bacterium adherence **57**, 1; **58**, 165; **113**, 257
Bacterium contamination **1**, 165; **72**, 111
Bacterium mutant **34**, 115
Bacterium spore **16**, 127; **34**, 51; **126**, 261
Badimol **93**, 21
Balsalazide **97**, 1
Bambuterol **124**, 149
Bamifylline **35**, 129; **43**, 59
Barbital **3**, 341; **6**, 19; **7**, 129; **14**, 1; **20**, 73; **26**, 157; **42**, 181, 97; **60**, 235; **65**, 141; **72**, 157; **103**, 19
Barbital **42**, 97, 181; **60**, 235; **65**, 141; **72**, 157; **103**, 19
Barbituric acid derivative **17**, 1; **20**, 73; **32**, 187; **42**, 181; **62**, 37; **66**, 175; **83**, 25, 39; **84**, 233; **91**, 15; **103**, 19; **112**, 1; **123**, 1
Barium carbonate **3**, 39
Barium sulfate **3**, 39; **48**, 51; **69**, 233; **90**, R1; **114**, 1; **118**, 95; **119**, 203; **122**, 35; **145**, 157; **147**, 95; **147**, 79
Barquat **81**, 111
Base **29**, 19; **31**, 69
Bay-l-5240 **31**, 179
Beclometasone dipropionate **101**, 1; **112**, 199; **115**, 209; **116**, 11, 169; **138**, 45; **139**, 255; **142**, 129
Beclometasone **83**, 163
Beclovent **8**, 1; **10**, 1
Bee venom **37**, 163
Behavior **72**, 233
Behenic acid **148**, 47
Benazepril **136**, 117
Bendazac lysine **82**, 179
Bendazac **82**, 179
Bendroflumethiazide **4**, 67; **17**, 325; **94**, 231; **114**, 121
Benfluorex **91**, 151
Benoxaprofen **11**, 215, 227
Benperidol **34**, 23
Benserazide **62**, 207
Bentone-27 **132**, 131
Bentonite **3**, 13; **128**, 129
Benzaldehyde **13**, 253; **22**, 45; **35**, 13
Benzalkonium **72**, 51; **113**, 263; **120**, 179; **127**, 239; **145**, 175
Benzalkonium bromide **39**, 229
Benzalkonium chloride **1**, 165; **9**, 1, 49; **16**, 127; **25**, 73, 105, 329; **37**, 27; **46**, 141; **47**, 21; **49**, 205; **55**, 175; **75**, 81; **81**, 111; **87**, 195; **93**, 239; **126**, 281; **132**, 165; **135**, 137; **136**, 181; **139**, 63; **141**, 151; **142**, 103
Benzalkonium chloride derivative **12**, 87
Benzamide **2**, 155; **7**, 129; **22**, 45; **25**, 57; **30**, 161; **71**, 31; **123**, 95
Benzamide derivative **9**, 295; **45**, 47
Benzathine cloxacillin **20**, 25
Benzene **15**, 159; **16**, 127; **17**, 77; **32**, 187; **34**, 217; **129**, 283; **137**, 149; **140**, 237
Benzene derivative **47**, 185; **51**, 73; **77**, 151, 163; **85**, 199; **125**, 297; **137**, 149
Benzenesulfonamide derivative **29**, 177; **56**, 273
Benzethonium chloride **36**, 191; **129**, 113
Benzimidazole derivative **77**, 265; **98**, 1; **137**, 149
Benzimidazole **137**, 149
Benziodarone **17**, 1
Benzisothiazolone **66**, 201; **74**, 195
5(7-Benzo[b]furanyl)-8-chloro-2,-3,-4,-5-tetrahydro-3-methyl-1H-3-benzazepin-7-ol **79**, 205
Benzocaine **2**, 337; **6**, 179; **8**, 89; **8**, 81; **50**, 27, 53; **55**, 199; **59**, 165; **77**, 239; **79**, 61; **81**, 103; **96**, 167; **118**, 129; **127**, 27
Benzodiazepine **13**, 67; **16**, 327; **61**, 189; **80**, 135; **110**, 169
1,4-Benzodiazepine **3**, 205
Benzodiazepine derivative **3**, 205; **17**, 357
Benzodiazepine receptor **36**, 113
N[4(1,4-Benzodioxan-6-yl)-2-thiazolyl]oxamic acid ethanolamine **50**, 7
Benzoic acid **1**, 323; **7**, 99, 189; **8**, 143; **9**, 97; **11**, 119, 131; **18**, 277; **20**, 43; **22**, 127; **27**, 205; **30**, 63; **32**, 223; **42**, 181; **48**, 149; **57**, 205; **71**, 223; **72**, 51; **82**, 135; **83**, 53; **86**, 211; **92**, 15; **96**, 167; **113**, 97; **127**, 115; **133**, 155; **137**, 217; **140**, 237; **142**, 251
Benzoic acid derivative **50**, 117; **73**, 1, 221; **86**, 43
Benzoic acid ester derivative **39**, 75
Benzophenone **1**, 173
Benzoquinolizine derivative **79**, 39
Benzoquinone **14**, 133
Benzoxazole derivative **98**, 1
1-Benzoyl-7-chloro-1,2-dihydro-3-methylaminoquinoxaline **3**, 205
5-Benzoyl-1,2-dihydro-3h-pyrrolo[1,2a]pyrrole-1-carboxylic acid **5**, 139
Benzoyl peroxide **133**, 215
Benzoylarginine ethyl ester **138**, 13
1a Benzoylcarbonylmitomycin C **15**, 61
O-Benzoyldextran derivative **27**, 205, 219
N-Benzoylglycylvaline-*n*-ethylamide **50**, 117
1a Benzoylmitomycin C **15**, 49, 61
1a Benzoyloxymethylmitomycin C **15**, 49, 61
Benzydamine **83**, 73; **118**, 129; **133**, 1
N-(Benzylaminomethyl)benzamide **9**, 7
Benzyl alcohol **2**, 295; **9**, 1; **30**, 63; **35**, 73; **35**, 13; **80**, 213; **103**, 187; **105**, 227; **124**, 183; **132**, 23; **140**, 155; **141**, 71
Benzyl benzoate **113**, 57; **133**, 107; **145**, 241
Benzyl nicotinate **61**, 205
Benzyl paraben **125**, 141
Benzylamine **88**, 111
1a Benzylcarbonylmitomycin C **15**, 49, 61; **25**, 95
6-*O*-Benzylguanine **90**, 239; **125**, 55
N-Benzylidene-tert-butylamine-*n*-oxide derivative **20**, 209
1a Benzylmitomycin C **15**, 49, 61; **25**, 95
1a Benzyloxycarbonylmitomycin C **15**, 49
Benzyltriphenylphosphonium salt **30**, 101
Berberine **29**, 177
Bergapten **133**, 245
Bestatin **30**, 73; **76**, 247; **113**, 1
Betahistine **102**, 141
Betaine derivative **91**, 95; **131**, 257
Betamethasone **94**, 211; **148**, 1
Betamethasone dipropionate **86**, 201

Betamethasone valerate **41**, 63; **70**, 271; **96**, 105; **116**, 19; **140**, 51
Betatrop **108**, 11
Betaxolol **58**, 115; **84**, 283; **133**, 171; **142**, 103
Bevantolol **99**, 303
Bicarbonate **19**, 169; **20**, 99; **21**, 341; **28**, 249; **29**, 253; **34**, 105; **35**, 157; **45**, 19; **54**, 131; **64**, 27; **68**, 11; **80**, R7; **105**, 97; **143**, 233; **148**, 117
Bichromate potassium **12**, 251
Bidisomide **136**, 117
Biguanide derivative **117**, 123
Bile **5**, 117; **43**, 41; **49**, 173, 179
Bile acid **37**, 127; **74**, 127; **80**, 63; **89**, 81, 245; **91**, 105; **101**, 209; **107**, 159; **109**, 197; **115**, 249; **121**, 37; **134**, 223
Bile salt **2**, 101; **8**, 303; **12**, 219; **28**, 33; **31**, 193; **39**, 87; **95**, 101; **103**, 27; **109**, 147; **111**, 147, 213; **114**, 65; **125**, 327; **142**, 1, 103; **143**, 1; **144**, 233
Binary code **38**, 71
Binding capacity **40**, 51
Binding kinetics **58**, 209; **76**, 257
Binding site **124**, 165
Bioavailability **1**, 71, 151, 213, 323; **2**, 101, 113, 173, 215; **3**, 55, 187; **6**, 43, 301; **7**, 83, 245, 301; **8**, 175; **9**, 17, 91, 233, 239; **10**, 181; **11**, 81; **13**, 1, 287; **14**, 59, 251; **15**, 235, 261; **16**, 23, 47, 171, 241; **17**, 47, 225, 273; **18**, 79; **19**, 233; **20**, 99, 187, 273; **21**, 3, 51, 187, 251; **22**, 31, 75, 137, 257, 311; **23**, 117; **23**, 25, 35, 277, 289; **24**, 173; **25**, 73, 113; **26**, 99, 145, 157, 203, 243, 317; **27**, 71, 81, 117, 145, 157, 245; **28**, 85, 95, 151, 167, 193, 201, 239; **29**, 113, 169, 191, 223; **30**, 73, 91, 111, 225, 247; **31**, 15, 83, 169, 193, 231, 239, 261; **32**, 103, 199, 219, 225, 235; **34**, 35, 131, 189, 207, 259; **35**, 47, 157, 181, 211, 243; **36**, 41, 81, 105, 127, 185; **37**, 11, 19, 97, 145, 227; **42**, 155; **43**, 1, 17, 31, 53, 83, 93, 101, 139, 261; **44**, 9, 15, 25, 57, 87, 97, 107, 197; **45**, 1, 59; **46**, 1, 9, 133; **47**, 37, 89, 209; **48**, 1, 51, 195; **49**, 23, 63, 109, 121, 231, 241; **50**, 15, 21, 79, 83, 253; **51**, 103, 233; **53**, 85; **54**, 113, 199; **55**, 17, 91, 105, 229; **56**, 43, 97; **57**, 125, 255, 163; **59**, 105, R1; **60**, 151; **62**, 119, 133, 187; **63**, 77, 143, 201; **64**, 61, 75; **65**, 159, 183, 195; **69**, 29; **72**, 131, 175; **73**, 157; **76**, 71; **76**, 71, 177; **78**, 199, 217; **79**, 205; **86**, 123, 147, 201; **94**, 69; **99**, 135, 189, 311, 337; **100**, 133, 249; **102**, 71, 213, 257; **104**, 87, 145, 175; **106**, 161; **111**, 171, 181; **113**, 209; **114**, 137, 171, 227; **115**, 95, 209; **117**, 31, 95, 113, 119, 129, 147, 197; **118**, 23; **123**, 1; **143**, 265; **145**, 137, 157; **147**, 71
Biocompatibility **58**, 79; **148**, 231
Biodegradation **58**, 79; **70**, 77; **84**, 141; **89**, 25, 251; **96**, 23; **100**, 55; **115**, 61; **116**, 1; **117**, 209; **143**, 187, 195; **144**, 115
Bioequivalence **16**, 47; **28**, 167; **54**, 33; **74**, 1; **86**, 201
Biogenic amine **12**, 97
Biological activity **36**, 233
Biopharmaceutics **5**, 127
Biostim **37**, 33; **52**, 27; **105**, 235
Biosynthesis **96**, 1
Biotin **81**, 1; **138**, 175
Biotransformation **2**, 127; **69**, 83; **75**, 37; **107**, 9; **102**, 241; **116**, 147
Biphenyl **18**, 47; **31**, 225
Biphenyl derivative **116**, 231
2,5-Bis(acyloxymethyl)allopurinol **24**, 307
1,3-Bis(acyloxymethyl)-5-fluorouracil **24**, 43
2,7-Bis(carboxyethyl)-6-carboxyfluorescein **68**, 231
Bis(cyclopentylcarbonyl)adrenalone **1**, 189
3,17-β-Bis[[(1,4-Dihydro-1-methyl-3-pyridyl)-carbonyl]oxy]-estra-1,3,5(10) **35**, 47
Bis(2,2 dimethylbutyryl)adrenalone **1**, 189
N,*n*-Bis(2-hydroxyethyl)l-inoleamide **88**, 89
Bis(2-hydroxyethyl)oleylamine **32**, 257
1[2,4-Bis(hydroxyiminomethyl)pyridinio]-1′-(4-carbamoylpyridinio)dimethyl-ether diiodide **109**, 35
1,4-Bis(methacryloylamino)azobenzene **87**, 37
Bis(2-methylvaleryl)adrenalone **1**, 189
21[4[2,6-Bis(1-pyrrolidinyl)-4-pyrimidinyl]-1-piperazinyl]-16α **66**, 63
Bis(tert butylacetyl)adrenalone **1**, 189
α-Bisabolol **70**, 87
Bisacodyl **118**, 103; **143**, 13
Bisanhydrodaunomycinone **34**, 247
Biskaol **28**, 239
Bismuth carbonate **7**, 83; **19**, 307; **28**, 239
Bismuth citrate **19**, 307
Bismuth gallate **19**, 307
Bismuth nitrate **34**, 207; **36**, 185
Bismuth salicylate **19**, 307; **30**, 225; **47**, 209
Bismuth **87**, 203
Bisoprolol fumarate **130**, 137
Bisphosphonic acid derivative **123**, 13, 71; **133**, 171
Bispilocarpic acid monoester derivative **79**, 233, 243
Bisulfate sodium **135**, 31
Bisulfite salts **13**, 273
Bitolterol **124**, 149
Blanaose **141**, 109
Blanching reaction **19**, 245
Blanose **84**, 69; **106**, 103; **110**, 285; **117**, 41
Bleaching **140**, 51
Bleogin **108**, 69
Bleomycin **1**, 41; **2**, 245; **8**, 291; **13**, 321; **35**, 177; **55**, 31; **65**, 49; **68**, 151; **108**, 69; **123**, 1; **145**, 9; **147**, 119
Bleomycin A2 **124**, 205
Bleomycin in 111 **1**, 41
Bleomycin PT 195M **1**, 41
Blood **1**, 41; **12**, 331; **17**, 91; **57**, 133
Blood brain barrier **32**, 79; **35**, 47; **45**, 249; **57**, 77; **84**, 39; **104**, 157; **108**, 225; **140**, 77; **145**, 9
Blood clotting **7**, 293
Blood clotting factor 10 **37**, 65
Blood clotting time **9**, 321
Blood flow **51**, 137; **88**, 327
Blood level **46**, 1, 69, 89; **48**, 247; **49**, 135; **50**, 35, 155, 175; **54**, 89; **57**, 125, 177
Blood rheology **75**, 155; **91**, 59, 123, 133; **95**, 201; **113**, 89
Blood sampling **10**, 177; **11**, 11
Blood toxicity **59**, 231
Blue dextran **79**, 11
BMY-43748 **140**, 1

BMS-180431 **105**, 209
Body distribution **34**, 145; **121**, 233
Body posture **21**, 341
Body surface **47**, 185
Bone marrow **68**, 121; **83**, 273
Borate sodium **9**, 337; **10**, 99; **14**, 299, 313; **28**, 113; **134**, 147
Boric acid **42**, 47
Boroleucine **105**, 7
Boron derivative **80**, 9
Boronic acid derivative **117**, 181
Bovine insulin **126**, 89; **138**, 75
Bovine serum albumin **3**, 65; **33**, 137, 147; **43**, 257; **47**, 79; **50**, 181; **59**, 263; **76**, 161; **78**, 103; **90**, 255; **106**, 221; **125**, 1; **126**, 179; **129**, 51; **134**, 15, 247; **143**, 1; **144**, 107, 241; **145**, 231; **147**, 173
Bowman birk inhibitor **78**, 1, 9
Brain **45**, 249
Brain injury **140**, 33
Brain ischemia **134**, 89
Brain perfusion **57**, 77
Brassicasterol **107**, 1; **114**, 33
Breast **53**, 43
Breast cancer **109**, 73
Breast carcinoma **80**, 271
Brij-36t **87**, 261, 265
Brilliant blue **61**, 109; **147**, 119
Bromazepam **16**, 327; **37**, 167; **61**, 189; **72**, 207
Bromelain **76**, 199
Bromerguride **31**, 69
Bromhexine **9**, 305; **43**, 77; **100**, 227
3-Bromo-4,4-dimethyl-2-oxazolidinone **3**, 151
3-Bromo-*n*-[(1-ethyl-2-pyrrolidinyl)methyl]-2,6-dimethoxybenzamide **15**, 87
1-Bromobutane **28**, 75
Bromocresotamide **15**, 273
Bromodiphenhydramine **7**, 343; **102**, 109
Bromoketamine **25**, 147
4-Bromophenol **10**, 89
Bromopride **123**, 285
1-Bromopropane **28**, 75
N-Bromosuccinimide **3**, 151
5(2-Bromovinyl)uracil arabinoside **91**, 217
Brompheniramine **91**, 75
Bromphenol blue **4**, 281
Bronchodilatation **81**, 199; **91**, 111
Bronchodilating agent **35**, 81; **124**, 149; **130**, 1; **137**, 261
Bropirimine **41**, 41; **75**, 211; **77**, 247; **83**, 241; **96**, 5; **123**, 65
Broxuridine **123**, 65
BRIJ-36 **49**, 39; **57**, 87
BRIJ-96 **57**, 87
BRIJ-721 **91**, 59
Buccal absorption **40**, 119
Bucillamine **105**, 11
Bucolome **7**, 317
Bucrilate **58**, 115; **84**, 1, 13; **100**, 55; **138**, 237; **142**, 75
Budesonide **86**, 9, 137; **101**, 1; **116**, 169; **138**, 45
Bufexamac **83**, 73
Buffer **10**, 143; **29**, 103; **33**, 125; **37**, 19; **44**, 97; **73**, 97; **78**, 25; **80**, R7; **87**, 11, 161; **90**, 239; **98**, 189; **109**, 27; **112**, 97; **113**, 215, 257; **114**, 197; **115**, 175; **117**, 189; **120**, 179; **124**, 219; **132**, 53; **133**, 191; **134**, 1; **139**, 249; **142**, 85; **144**, 91; **146**, 115
Buflomedil **72**, 79; **140**, 131
Bumadizone **34**, 189
Bunitrolol **48**, 1
α-Bungarotoxin **123**, 137
Bupivacaine **84**, 33; **92**, 1; **107**, 41; **131**, 219; **136**, 165
Bupranolol **36**, 165; **46**, 95; **49**, 141; **55**, 265; **76**, 25
Buprenorphine **21**, 233; **132**, 81
Buprenorphine H 3 **21**, 233
Burn **75**, 81; **142**, 235
Buserelin acetate **123**, 103; **141**, 39
Busulfan **108**, 69
Butacaine **12**, 77
Butadiene styrene copolymer **34**, 137
Butanediol **15**, 185
1,3-Butanediol **121**, 19; **144**, 127
Butanol **4**, 205; **16**, 127; **29**, 177; **42**, 19; **50**, 213; **68**, 285; **76**, 91; **78**, 17; **80**, 17; **84**, R5; **92**, 81; **95**, 23; **106**, 51; **125**, 107; **126**, 219; **132**, 23; **138**, 149; **148**, 41
1-Butanol **1**, 15
2-Butanone **126**, 219
3-Butanoyloxymethyl-5,5-diphenylhydantoin **126**, 11
Butaperazine **31**, 69
1-Butenyl acetate **34**, 137
Butethal **41**, 139; **42**, 97; **60**, 235; **72**, 157; **103**, 19
Buthionine sulfoximine **44**, 265
Butibufen **62**, 21
O-(Butyryloxymethyl)salicylamide **30**, 111
5-Butyl-2-hydrazinopyridine **46**, 203
Butyl paraben **30**, 35; **32**, 223; **33**, 105; **34**, 115; **35**, 227; **69**, 103; **83**, 211; **90**, 161, 203; **109**, 155; **125**, 141; **127**, 27; **148**, 1
2-Butyl-1[2′(1H-tetrazol-5-*yl*)biphenyl-4-yl]methyl-1H-benzimidazole-7-carboxylic acid **103**, 1
4-Butylaniline **37**, 49; **57**, 189; **100**, 1
Butylated hydroxyanisole **144**, 153; **147**, 31
Butylcaine **77**, 239; **127**, 27
1-Butylcarbamoyl-5-fluorouracil **36**, 147
Butylcresol **29**, 73; **105**, 227; **140**, 169; **144**, 153; **147**, 31
Butylparaben **27**, 163; **30**, 35
Butyric acid **32**, 79; **33**, 235; **42**, 181; **86**, 43; **126**, 219; **132**, 23
γ-Butyrolactone **7**, 169
Butyrylcholinesterase **26**, 15
7-(Butyryloxymethyl)theophylline **12**, 299
1-Butyryloxymethylallopurinol **24**, 307
1-Butyryloxymethylfluorouracil **12**, 153
α-Butyryloxystyrene **9**, 29
O-Butyryltimolol **33**, 15

C

Cadaver **22**, 291
Cadmium **25**, 57; **54**, 143; **79**, 213

Caffeine **7**, 99, 343; **14**, 1, 123; **16**, 271; **18**, 127, 259, 299; **19**, 149; **22**, 291; **26**, 329; **31**, 201; **32**, 79, 187, 257; **48**, 51, 103; **50**, 205; **51**, 1, 263; **53**, 47; **70**, 191; **79**, 257; **80**, 267; **81**, 143; **82**, 117; **90**, 213; **95**, 93; **96**, 197, 205, 217; **99**, R5; **100**, 41; **103**, 131; **106**, 33; **109**, 9, 209, 221; **111**, 127; **112**, 37; **115**, 217; **120**, 63, 83; **121**, 141; **125**, 157; **127**, 185; **129**, 185, 233; **138**, 199; **140**, 237; **141**, 53; **148**, 1
Calcein **114**, 33; **124**, 19; **133**, 171
Calcification **52**, 139; **59**, 95; **98**, 165
Calcitonin **33**, 89; **63**, 135; **75**, 97; **89**, 169; **101**, 115; **115**, 9; **126**, 245; **127**, 115; **130**, 33; **134**, 47; **137**, 103; **144**, 19
Calcitonin[1,7-aminosuberic acid] **33**, 89
Calcitonin derivative **127**, 31, 43; **134**, 47
Calcium **13**, 9; **52**, 79; **64**, 217; **77**, 57; **79**, 159; **81**, 1; **87**, 73, 203; **90**, 229; **93**, 121; **105**, 209; **106**, 229; **107**, 121; **111**, 271; **114**, 221; **123**, 13, 187; **126**, 199; **127**, 135; **139**, 105
Calcium acetate **72**, 43; **90**, 203
Calcium alginate **25**, 121; **95**, 77; **97**, 183; **103**, 267; **111**, 271; **112**, 241
Calcium antagonist **64**, 117; **79**, 199; **98**, 113; **140**, 145; **148**, 123
Calcium carbonate **19**, 307; **28**, 239, 249; **35**, 157; **47**, 223; **49**, 1; **64**, 55; **66**, 273, 283; **68**, 11, 157; **70**, 241; **72**, 43; **83**, 147; **90**, 203; **105**, 97; **144**, 187; **147**, 95
Calcium channel blocking agent **131**, 153, 183
Calcium chloride **1**, 141; **19**, 177; **27**, 17; **28**, 249; **79**, 11; **87**, 239; **93**, 131; **115**, 45; **137**, 133, 187
Calcium fluoride **16**, 171
Calcium fluoride CA 45 **15**, 177
Calcium formate **72**, 43; **90**, 203
Calcium gluceptate **24**, 1; **30**, 161
Calcium ion **113**, 257
Calcium lactate **72**, 43; **90**, 203
Calcium nitrate **7**, 83
Calcium phosphate **22**, 127; **25**, 207; **40**, 223; **49**, 1; **64**, 55; **68**, 157; **72**, 43, 117; **73**, 75; **88**, 285; **90**, 203; **92**, 55, 123, 137, 157; **110**, 37
Calcium phosphate dibasic **7**, 269; **45**, 79; **67**, 79; **71**, 201; **75**, 181; **80**, 259; **82**, 71; **88**, 285; **89**, 223, R17; **90**, R1; **92**, 55, 123, 137, 157; **94**, 181; **95**, 29, 117; **97**, 119; **99**, 7; **103**, 9, 55; **104**, 271; **106**, 149; **107**, 29, 219, 247; **108**, 213; **109**, 1; **110**, 37; **112**, 225; **114**, 23, 85; **115**, 35; **118**, 151; **121**, 65; **123**, 143; **124**, 69; **126**, 57; **128**, 277; **131**, 209; **133**, 29; **134**, 105; **143**, 233; **149**, 131
Calcium pyrophosphate **2**, 145; **44**, 177; **51**, 77
Calcium sulfate **26**, 165; **90**, R1
Calcium sulfate dihydrate **38**, 255
Caloric restriction **4**, 175
Camostat mesilate **81**, 119; **113**, 1
Campesterol **107**, 1; **114**, 33
Camphor **124**, 279
Camphorquinone **86**, 221
Cancer **13**, 303; **35**, 219
Cancer cell **17**, 65; **63**, 121; **98**, 19
Cancer chemotherapy **2**, 245; **13**, 303; **15**, 49, 61; **21**, 219; **22**, 75, 137; **36**, 147; **37**, 135
Cancer diagnosis **80**, 271
Cancer model **110**, 117
Cancer patient **130**, 257
Candida albicans **2**, 325; **46**, 141; **54**, 51; **56**, 169; **73**, 43; **84**, 77; **86**, 193; **93**, 201; **105**, 39, 71
Candidiasis **81**, R5; **138**, 67
Cannulation **24**, 185
Capderm **126**, 119; **131**, 143
Capillary permeability **145**, 9
Capmul-MCM90 **106**, 15
Capping phenomenon **72**, 117
Caprolactam **140**, 169
Caprolactam derivative **52**, 47; **76**, 37; **79**, 141
Caproylated calcitonin **134**, 47
Capsaicin derivative **145**, 175
Capsaicin **124**, 279; **126**, 119; **128**, 169; **130**, 121; **131**, 143
Capsule **2**, 91; **4**, 291; **6**, 301; **7**, 245; **8**, 35; **12**, 251, 341; **21**, 187; **22**, 311
Captopril **11**, 329; **41**, 245, 255, 263; **75**, 117; **86**, 99; **107**, 77; **112**, 133; **128**, 229; **134**, 147; **136**, 117; **143**, 93, 101, 119; **145**, 215; **148**, 41
Carazolol **64**, 171; **105**, 189
Carbamazepine **10**, 181; **20**, 307; **24**, 297; **30**, 91; **50**, 97, 111; **53**, 25; **54**, 1; **73**, 17; **77**, 287; **82**, 67, 117; **85**, 175; **86**, 259; **93**, 7; **94**, 15; **98**, 239; **110**, 169; **126**, 65; **129**, 79; **148**, 1
Carbamazepine-10,11-oxide **30**, 91
Carbamic acid derivative **71**, 31; **79**, 205; **81**, 253; **91**, 39
Carbamic acid phenyl ester derivative **81**, 253
1-Carbamoyl-5-fluorouracil derivative **23**, 209
2-Carbamoyloxymethyl-4(2,3-dichlorophenyl)-1,4-dihydro-6-methyl-3,5-pyridinedicarboxylic-acid-3-isopropyl-5-methyl ester **124**, 27
1(4-Carbamoylpyridinio)-1′-(2-hydroxyiminomethylpyridinio) dimethyl ether **40**, 193; **46**, 247; **109**, 35; **114**, 125; **137**, 167
Carbanilide **49**, 205; **56**, 43
Carbazepine **45**, 13
Carbazole **101**, 199
Carbazole derivative **104**, 125; **140**, 237
Carbenicillin **1**, 95; **26**, 133; **30**, 181; **37**, 113; **46**, 45; **82**, 107; **148**, 191
Carbenoxolone **119**, 133
Carbidopa **6**, 225; **72**, R5; **108**, 95
Carbocisteine **52**, 155; **77**, 41
Carbohydrate **2**, 325; **3**, 1; **16**, 319; **28**, 151; **107**, 99; **136**, 107
Carbohydrate derivative **79**, 159; **87**, 105
Carbomer **2**, 27; **12**, 173; **15**, 13; **48**, 247; **51**, 203; **53**, 79; **61**, 109; **67**, 113; **73**, 69; **83**, 221; **84**, 69; **100**, 207; **102**, 19; **103**, 97; **104**, 107; **109**, 115; **110**, 241; **117**, 75; **124**, 173, 183; **126**, 119; **129**, 137; **133**, 265; **135**, 115; **136**, 155; **137**, 233; **138**, 13, 103; **140**, 251; **141**, 39, 109; **142**, 169; **145**, 231
Carbomer 940 **110**, 241; **111**, 261
Carbon 14 **65**, 7, 149, 235
Carbon black **53**, 47
Carbon dioxide **148**, 123
Carbon tetrachloride **17**, 77; **20**, 203; **34**, 217
Carbonate dehydratase **51**, 27; **47**, 79; **90**, 95
Carbonate dehydratase inhibitor **56**, 273
Carbonic acid **34**, 105

Carboplatin **69**, 197; **101**, 257; **108**, 69; **124**, 205; **129**, 275; **138**, 125; **146**, 263; **147**, 153
Carbopol **40**, 111
Carbopol 934 **3**, 73; **31**, 15; **73**, 131; **138**, 103; **141**, 39; **142**, 169; **147**, 119
Carbopol 940 **12**, 173; **61**, 15; **109**, 115; **130**, 121; **133**, 1; **138**, 103; **142**, 169; **145**, 175, 231
Carbopol 974P **111**, 251; **131**, 47
Carboprost **124**, 205
Carbowax 1540 **33**, 249
Carbowax 1540 **44**, 261
Carbowax 4000 **28**, 113
3(2-Carboxyethylthio)-2-hydroxy-3[2(8-phenyloctyl)phenyl] propionic acid **120**, 235
Carboxyfluorescein **35**, 263; **57**, 241; **60**, 219; **77**, 31; **79**, 159; **91**, 69; **95**, 51; **101**, 81; **105**, 1; **108**, 117; **111**, 15; **117**, 225; **119**, 231; **141**, 27
6-Carboxyfluorescein **17**, 203; **31**, 231, 239; **36**, 147; **48**, 125; **105**, 259
Carboxylate **49**, 15
Carboxylesterase **112**, 181
Carboxylic acid **18**, 235; **27**, 39; **81**, 133; **90**, 203; **101**, 89; **108**, 225
Carboxylic acid derivative **18**, 235; **42**, 181; **103**, 253; **108**, 141; **131**, 257
Carboxymethyl-β-cyclodextrin **124**, 205; **126**, 73
Carboxymethyl starch **136**, 107
Carboxymethylcellulose **6**, 285; **7**, 285; **9**, 59; **11**, 309; **12**, 341; **19**, 107; **20**, 187; **24**, 61; **27**, 157; **31**, 169; **34**, 183; **39**, 39; **39**, 47; **41**, 159; **44**, 125, 151, 177; **45**, 39; **46**, 9; **47**, 133; **48**, 51, 247; **53**, 79; **55**, 115; **75**, 155; **77**, 261; **78**, 43; **79**, 97; **81**, 49; **84**, 69; **85**, 141; **89**, 223; **92**, 55; **94**, 15; **106**, 103; **110**, 169, 285; **117**, 41; **118**, 207; **119**, 149; **130**, 41; **133**, 71; **135**, 115; **138**, 75, 103; **140**, 85; **141**, 109; **142**, 251; **144**, 71; **145**, 175; **146**, 1
Carboxymethylethylcellulose **4**, 291
Carboxymethylstarch sodium **12**, 341; **62**, 27; **146**, 71
N[*n*(3-Carboxyoxirane-2-carbonyl)-leucyl]agmatine **107**, 133; **112**, 133
Carboxypeptidase **48**, 189; **107**, 133; **117**, 95; **136**, 61
Carboxypeptidase a **26**, 15; **107**, 133; **113**, 25
Carcinoma VX2 **2**, 245
Cardamom **126**, 111; **131**, 137; **134**, 183
Cardanol **66**, 107
Cardiac glycoside **14**, 123; **29**, 73
Cardiovascular agent **81**, 1; **92**, 1
Cardiwell plus **108**, 11
Carindacillin **4**, 299
Carmethizole **71**, 157
Carmofur **24**, 79; **36**, 147; **38**, 191, 239; **49**, 163
Carmustine **68**, 151; **108**, 69; **124**, 205; **145**, 9
Carnauba wax **112**, 37; **123**, 85; **138**, 215
β-Carotene **88**, 211; **94**, 1; **117**, 85
Carrageenan **19**, 107; **86**, 159; **109**, 115; **115**, 103; **121**, 11; **139**, 53
Carrier protein **146**, 205
Carsalam **78**, 199
Carteolol **95**, 117; **109**, 229; **114**, 13; **118**, 151; **128**, 81
Carveol **74**, 157
Casein **37**, 103; **92**, 97; **94**, 223; **106**, 221; **117**, 23
Castor oil **78**, 95; **108**, 207; **115**, 217; **132**, 33
Catechin **65**, 261
Catechol **140**, 155
Catechol derivative **141**, 239
Cathepsin B **81**, 119; **107**, 133
Cathepsin **113**, 1
Cation **77**, 107; **106**, 229; **139**, 105
Cedrox **119**, 213
Cefacetrile **7**, 317
Cefaclor **9**, 305; **12**, 209; **50**, 233; **111**, 31; **115**, 175; **134**, 213; **145**, 115
Cefadroxil **8**, 25; **9**, 263; **12**, 209; **16**, 23; **17**, 29; **82**, 165; **104**, 187; **119**, 213; **145**, 115
Cefalexin **30**, 181; **32**, 99; **34**, 75, 241; **43**, 187; **47**, 37; **82**, 165; **98**, 83; **100**, 213; **104**, 187; **106**, 33; **115**, 79; **118**, 1; **122**, 9; **125**, 195; **140**, 175; **145**, 115; **148**, 1
Cefaloglycin **3**, 151; **12**, 209; **34**, 241; **43**, 187
Cefaloridine **3**, 151; **14**, 1; **16**, 319; **28**, 103; **30**, 181; **34**, 241; **82**, 53
Cefalotin **3**, 151; **16**, 319; **23**, 117; **25**, 105; **30**, 161, 181; **32**, 151; **34**, 241; **44**, 205; **50**, 233; **51**, 55; **62**, 65, 165; **75**, 131, 161
Cefamandole nafate **4**, 263; **30**, 161; **44**, 205
Cefamandole **17**, 237; **23**, 117; **30**, 161; **50**, 233
Cefapirin **23**, 117; **44**, 205
Cefazaflur **34**, 241
Cefazolin **3**, 151; **4**, 281; **7**, 317; **16**, 319; **17**, 29; **23**, 117; **30**, 57, 161, 181; **31**, 231; **34**, 241; **40**, 175; **50**, 233; **67**, 259; **137**, 1; **144**, 177
Cefixime **43**, 53; **56**, 125; **59**, 217; **106**, 33
Cefmenoxime **23**, 117
Cefmetazole **10**, 291; **21**, 239; **33**, 89; **123**, 1
Cefodizime **77**, 47
Cefonicid **44**, 205; **105**, 189
Cefoperazone **131**, 67
Ceforanide **17**, 29
Cefotaxime **17**, 29; **23**, 117; **34**, 1; **44**, 205; **46**, 45; **137**, 1
Cefotiam **19**, 345; **23**, 117; **38**, 179
Cefotiam derivative **38**, 179
Cefotiam pivaloyloxymethyl ester **23**, 117
Cefoxitin **5**, 117; **6**, 225; **7**, 279; **21**, 239; **32**, 151; **33**, 99; **123**, 1; **143**, 93
Cefradine **12**, 209; **30**, 181; **34**, 241; **43**, 187; **47**, 1; **85**, 181; **104**, 187; **115**, 79
Cefroxadine **12**, 209; **17**, 29
Cefsulodin **111**, 31
Ceftazidime **36**, 55; **50**, 233; **51**, 55; **147**, 127
Ceftezole **23**, 117
Ceftibuten **63**, 101
Ceftizoxime **23**, 117; **30**, 181
Ceftriaxone **19**, 199; **92**, 47; **123**, 1; **137**, 1
Cefuroxime **75**, 131; **77**, 13; **137**, 1; **147**, 127
Celiprolol **107**, 243
Cell adhesion **53**, 253; **80**, 33; **84**, 77; **99**, 93, 293

Cell culture **32**, 55, 79; **36**, 253; **40**, 215; **42**, 117; **45**, 249; **46**, 45, 69, 141, 149; **47**, 129; **102**, 55; **104**, 157, 187; **108**, 225; **109**, 73; **110**, 29; **111**, 25; **114**, 141, 151; **115**, 79; **116**, 147; **123**, 13, 65, 187; **125**, 1, 165; **129**, 103, 223; **133**, 47; **145**, 1
Cell growth **70**, 159; **75**, 1; **102**, 55; **107**, 189; **125**, 165; **129**, 105
Cell membrane **1**, 127; **10**, 153; **16**, 271; **24**, 297; **53**, 61
Cell membrane permeability **5**, 229
Cell membrane transport **1**, 127
Cell proliferation **123**, 65
Cell receptor **128**, 65
Cell strain HL 60 **62**, 133
Cell transport **79**, 39; **83**, 131
Cellaburate **48**, 217; **92**, 35
Cellacefate **18**, 1; **28**, 157; **31**, 175; **34**, 93, 259; **42**, 105; **47**, 21; **48**, 51; **87**, 239; **109**, 9; **114**, 205; **115**, 29; **120**, 205; **124**, 87; **126**, 1; **145**, 247
Cellactose **95**, 201
Cellulose **2**, 145; **6**, 11; **7**, 269; **11**, 323; **12**, 135, 341; **15**, 251; **16**, 349; **17**, 313; **22**, 127; **30**, 161; **31**, 137; **35**, 91, 103, 157, 235; **36**, 29, 51, 205; **37**, 15, 155; **38**, 227; **40**, 267; **41**, 159; **42**, 15, 77; **44**, 43, 151, 177; **45**, 79, 111, 259; **46**, 35; **50**, 79; **48**, 51; **52**, 213; **53**, 47; **55**, 143; **57**, 139; **59**, 155; **61**, 145; **64**, 35, 207, 227; **65**, 1; **66**, 207, 273, 283; **67**, 39, 231; **70**, 261; **71**, 81, 127, 201; **72**, 43, 111, 243; **73**, 37, 75; **76**, 239; **77**, 269; **79**, 47; **80**, 179; **81**, 205, 225; **83**, 11, 147, 187; **89**, 1, 223, R17; **90**, 203; **91**, 123, 133, 143, 173, 183, 241, 247; **92**, 55; **93**, 249; **94**, 49, 181; **95**, 57, 135; **96**, 59, 197, 205, 217, 225; **97**, 15, 79; **99**, 7, 157, 197; **100**, 33, 49, 71; **101**, 169; **102**, 81; **105**, 177; **106**, 149; **107**, 29, 219; **108**, 31, 207; **109**, 209, 221; **111**, 89; **112**, 153, 225; **114**, 1; **116**, 65, 179, 191; **118**, 213; **119**, 71; **120**, 95, 101; **121**, 65, 73, 81; **122**, 49; **124**, 9; **125**, 257; **128**, 123; **130**, 41, 147; **131**, 47, 201, 209; **132**, 207; **133**, 133; **134**, 129; **136**, 71; **137**, 41; **139**, 25, 87; **141**, 53, 101; **142**, 175; **144**, 187; **145**, 215; **148**, 1
Cellulose acetate **86**, 35; **88**, 405; **92**, 197; **120**, 119; **125**, 215; **129**, 279
Cellulose acetate trimellitate **109**, 9; **115**, 29
Cellulose derivative **27**, 267; **48**, 1; **62**, 97; **95**, 219; **109**, 9; **122**, 49; **124**, 107; **126**, 147; **128**, 9, 23; **135**, 147; **139**, 15
Cellulose-ethyl-hydroxyethyl-ether **83**, 1; **95**, 219; **124**, 107; **125**, 205; **128**, 23, 9
Cellulose phthalate **33**, 71
Cellulose propionate **34**, 137
Centella asiatica **146**, 63
Centperazine **31**, 169
Central nervous system **35**, 47; **104**, 157; **141**, 239
Cephalexin **3**, 151; **12**, 209
Cephalosporin **16**, 319; **17**, 29; **19**, 345; **23**, 117; **26**, 317; **34**, 241; **73**, 259; **147**, 127
Cephalosporin C **30**, 181
Cephalosporin derivative **32**, 151; **75**, 131; **77**, 13; **81**, 1; **82**, 165; **104**, 187; **137**, 1; **140**, 175; **145**, 115
Cephalosporin pivaloyloxymethyl ester **23**, 117
Ceramide **70**, 173; **98**, 203; **126**, 49; **128**, 203; **132**, 263; **141**, 17
Ceranapril **140**, 175
Ceraphyl **105**, 227
Cesapectin **133**, 1
Cetirizine **136**, 181
Cetomacrogol **65**, 225; **122**, 27; **130**, 203; **143**, 75, 83; **147**, 31
Cetomacrogol 1000 **49**, 29; **82**, 223
Cetomacrogol A14 **82**, 223
Cetostearylic alcohol **25**, 179, 237; **27**, 311; **31**, 281; **63**, 219; **68**, 205; **83**, 95; **91**, 59; **125**, 315; **137**, 243
Cetrimide **19**, 97, 239; **25**, 179, 237; **27**, 311; **30**, 101; **31**, 281; **38**, 211; **49**, 129; **63**, 219; **71**, 213; **78**, 157; **84**, 77; **86**, 43, 193; **87**, 261, 265; **107**, 239; **111**, 73; **113**, 263; **124**, 107; **126**, 281; **128**, 29; **139**, 69; **144**, 99; **145**, 175
Cetrimonium salt **9**, 49
1-Cetyl-2-piperidone **68**, 239
Cetylmannoside **70**, 225
Cetylpyridinium salt **34**, 51, 115; **36**, 191; **44**, 243; **47**, 13; **58**, 229; **63**, 183; **84**, 77; **86**, 193; **113**, 263; **126**, 281; **127**, 239; **131**, 33; **143**, 119; **145**, 175
Charcoal **19**, 307; **80**, 271
Cheek mucosa **47**, 13; **48**, 189; **49**, 141; **56**, 135; **76**, 25; **82**, 165; **84**, 77; **85**, 129; **86**, 193; **101**, 15, 121; **121**, 249; **125**, 165; **133**, 1; **141**, 217
Cheek pouch **81**, 39
Chelating agent **88**, 345; **90**, 221; **123**, 13
Chelation **52**, 79; **69**, R1; **123**, 187; **124**, 129; **127**, 135
Chelerythrine **46**, 255
Chelidonine **46**, 255
Chemical analysis **62**, 97; **63**, 219; **100**, 181, 207, 213, 227; **111**, 83; **114**, 247; **115**, 113; **115**, 249; **118**, 121; **141**, 27
Chemical injury **34**, 75
Chemical kinetics **1**, 95; **2**, 275, 295, 337; **4**, 171; **7**, 119, 197; **8**, 45; **10**, 1, 281; **11**, 71; **12**, 251; **13**, 273; **15**, 37; **23**, 163
Chemical modification **134**, 147; **136**, 61; **140**, 185, 195; **141**, 257
Chemical reaction **108**, 95
Chemical reaction kinetics **98**, 209
Chenodeoxycholic acid **74**, 127; **114**, 65; **149**, 1
Chewing gum **121**, 117
Chibro pilocarpine **95**, 229
Chitosan **61**, 35; **67**, 11; **78**, 43; **80**, 33; **88**, 9; **89**, 1, 19, 177; **92**, 211; **93**, 233; **96**, 33, 139; **98**, 121, 181; **99**, 13; **100**, 257; **101**, 227; **111**, 217; **116**, 39; **118**, 257; **120**, 115; **121**, 113; **122**, 69; **131**, 101; **134**, 239; **135**, 63, 115, 183; **139**, 25; **145**, 231; **146**, 193; **147**, 71; **148**, 231
Chitosan derivative **98**, 121; **148**, 231
Chitosan malate **98**, 181
Chitosome **146**, 193
Chitosonium malate **100**, 257
Chloral hydrate **6**, 205
Chlorambucil **8**, 217; **53**, 195; **57**, 63; **66**, 289; **71**, 73; **87**, 141; **108**, 69; **124**, 205
Chlorambucil derivative **28**, 1
Chloramphenicol **1**, 165; **6**, 29, 205; **7**, 89; **12**, 67; **14**, 1, 123; **17**, 1; **20**, 265; **25**, 105; **28**, 67; **32**, 91; **37**, 11; **38**, 247; **41**, 75; **46**, 169; **70**, 137; **75**, 161; **88**, 313; **110**, 109; **112**, 241
Chloramphenicol derivative **28**, 1
Chloramphenicol hemisuccinate **15**, 37

Chloramphenicol palmitate **14**, 1; **30**, 161
Chloramphenicol 3 succinate **15**, 37
Chlorbutol **39**, 229; **123**, 47; **135**, 137
Chlorcyclizine **63**, R1; **102**, 109
Chlordiazepoxide **3**, 205; **5**, 85; **14**, 1; **16**, 327; **42**, 235; **61**, 189; **75**, 161
Chlorhexidine **1**, 127, 165; **2**, 343; **9**, 49; **19**, 239; **33**, 201; **34**, 51; **36**, 191; **49**, 205; **55**, 209; **75**, 81; **84**, 85; **114**, 257; **143**, 25
Chlorhexidine acetate **84**, 85; **131**, 33; **143**, 25
Chlorhexidine diacetate **25**, 329; **34**, 51, 115; **36**, 191
Chlorhexidine digluconate **9**, 1
Chlorhexidine gluconate **39**, 229; **49**, 129; **65**, 127; **84**, 77; **86**, 193; **101**, 121; **119**, 247; **131**, 33; **147**, 243
Chloride **11**, 131; **16**, 127; **69**, 197; **130**, 179
Chloride ion **63**, 167
2-Chloro-10-cyclopropylmethyl-9-acridinone **3**, 109
7-Chloro-1-cyclopropylmethyl-5-(2-fluorophenyl)-1,3-dihydro-2H-1,4 **3**, 109
5-Chloro-2-cyclopropylmethylamino-2′-fluorobenzophenone **3**, 109
2-Chloro-2′,3′-dideoxyadenosine **60**, 53
8-Chloro-5(2,3-dihydro-7-benzo[b]furanyl)-2,3,4,5-tetrahydro-3-methyl-1H-3 **79**, 205
6-Chloro-2,3-dimethoxy-1,4-naphthoquinone **35**, 61
6-Chloro-2,3-dimethoxy-1,4-naphthohydroquinone-1-acetate **35**, 61
7-Chloro-4,5-epoxy-4,5-dihydro-2-methylamino-5-phenyl-3H-1,4 **3**, 205
4-Chloro-5′-fluoro-2′-hydroxybenzophenone **46**, 231
2-Chloro-5a,6,7,8,9,9a-hexahydro-*n*(3-quinuclidinyl)dibenzofuran-4-carboxamide **121**, 95
5-Chloro-2-methyl-4-isothiazolin-3-one **66**, 201; **74**, 195
5-Chloro-2-(methylamino)benzophenone **31**, 9
9-Chloro-5-methylamino-2-phenyl-4H-benzo[g][1,3,6]oxadiazocine **3**, 205
1-Chloro-4-nitrobenzene **18**, 247
3[[3[2(7-Chloro-2-quinolyl)vinyl]phenyl]2(*n*,*n*)] **59**, 281
4-Chloro-5-sulfamoylanthranilic acid **11**, 237
Chloroaniline **6**, 261
3-Chloroaniline **6**, 261
4-Chloroaniline **6**, 261
4-Chlorobenzoic acid **28**, 47
(3-Chlorobenzyl)triphenylphosphonium chloride **30**, 101
Chlorocresol **2**, 295; **9**, 1; **25**, 105; **29**, 29; **34**, 115; **35**, 227; **100**, 189
Chlorodantrolene **75**, 193
1(2-Chloroethyl)-3-(2,6-dioxo-3-piperidyl)-1-nitrosourea **145**, 9
7-(2-Chloroethyl)theophylline **18**, 259
Chlorofluorocarbon **95**, 111; **123**, 291
Chloroform **2**, 81; **5**, 287; **16**, 127; **17**, 77; **21**, 119; **31**, 209; **34**, 217; **35**, 173; **99**, 93; **126**, 219; **129**, 283; **131**, 171; **140**, 237
4-Chloronitrobenzene **18**, 247
Chlorooxazone **7**, 129
α-Chloropenicillin G **17**, 47
Chloropheniramine maleate **6**, 243; **11**, 131
Chlorophenol **140**, 155
2-Chlorophenol **126**, 219
3-Chlorophenol **4**, 1
4-Chlorophenol **10**, 89; **100**, 189; **126**, 219
2-Chlorophenothiazine derivative **16**, 153
Chloroquine **12**, 265; **25**, 217; **26**, 267; **31**, 107; **34**, 137; **35**, 173; **43**, 225; **44**, 31; **51**, 161, 263; **52**, 179; **56**, 213; **62**, 81; **65**, 133; **70**, 241; **72**, 11; **92**, 15, 183; **98**, 181; **105**, 189; **133**, 155
6-Chloroquinoxaline derivative **3**, 205
8-Chlorotheophylline **18**, 259
Chlorothiazide **7**, 129; **17**, 1; **80**, 129
Chloroxylenol **2**, 295; **9**, 49; **19**, 239; **32**, 55, 91; **118**, 41; **138**, 149
Chlorphenesin carbamate **100**, 133; **117**, 31
Chlorpheniramine **10**, 17; **16**, 47; **41**, 83; **53**, 91; **91**, 75; **131**, 47; **138**, 57; **143**, 107
Chlorpheniramine maleate **44**, 125; **70**, 59; **88**, 159; **91**, 75; **101**, 23; **147**, 135
Chlorphenotane **7**, 99; **24**, 185; **25**, 57; **30**, 161; **33**, 165; **73**, 177; **80**, 51
Chlorproethazine **35**, 129; **94**, 135
Chlorpromazine **3**, 231; **5**, 25, 273; **12**, 219; **13**, 227; **14**, 159; **16**, 153, 271; **17**, 183; **18**, 225; **35**, 129; **88**, 261; **89**, 177; **94**, 135
Chlorpropamide **146**, 93
Chlorprothixene **17**, 1; **35**, 129
Chlortalidone **29**, 73; **65**, 177
Chlortetracycline **5**, 273; **17**, 1
3[(3-Cholamidopropyl)-dimethylammonio]-1-propanesulfonic acid **90**, 187
Cholecystokinin octapeptide **105**, 113
Cholecystokinin receptor blocking agent **92**, 63
Cholera toxin **119**, 251
Cholera toxin B subunit **119**, 251
Cholera vaccine **119**, 251
Cholestasis **128**, 223
Cholesterol **6**, 179; **8**, 101; **11**, 27; **13**, 263; **16**, 305; **20**, 119, 139; **23**, 1; **28**, 113; **32**, 39; **37**, 75, 127; **42**, 257; **47**, 21; **69**, 13; **70**, 173; **75**, 45; **76**, 161; **80**, 93; **84**, 205; **85**, 97; **87**, 183; **88**, 201; **89**, 139; **90**, 59, 73, 255; **94**, 1; **96**, 1; **98**, 19, 149, 203; **102**, 151; **103**, 81, 237; **105**, 1, 189; **107**, 1; **113**, 141; **114**, 33, 221; **118**, 207; **121**, 195; **124**, 19, 75; **125**, 1; **126**, 41, 49; **127**, 273; **131**, 57, 103; **133**, 171; **140**, 119; **145**, 95
Cholesterol acetate **84**, 107; **88**, 53
Cholesterol derivative **58**, 41; **88**, 53; **124**, 53
Cholesterol ester **84**, 107; **94**, 89
Cholesterol esterase **62**, 259
Cholesterol hydrogen succinate **132**, 9
Cholesterol oleate **114**, 43
Cholesterol sulfate **98**, 203; **126**, 49
Cholesterol synthesis **57**, 223; **96**, 1
Cholesteryl oleyl carbonate **124**, 53
Cholestyramine **5**, 167; **16**, 23
Cholic acid **6**, 137; **8**, 303; **9**, 173; **12**, 219; **27**, 117; **31**, 157, 193; **33**, 219; **37**, 127; **43**, 41; **67**, 207; **69**, 29; **74**, 95, 127; **89**, 81; **95**, 101; **101**, 15, R7; **105**, 137; **109**, 147; **114**, 65; **121**, 37; **125**, 327

Choline **39**, 263; **88**, 293
Choline acetyltransferase **63**, 149
Choline theophyllinate **25**, 113; **26**, 99; **36**, 73; **66**, 111
Cholinergic receptor **15**, 223; **123**, 137
Cholinergic receptor blocking agent **98**, 113; **114**, 111
Cholinergic receptor stimulating agent **15**, 223
Chondroitin **36**, 253
Chondroitin sulfate **19**, 107; **84**, 141
Chondroitin sulfate iron **61**, 199
Chromatography **17**, 1; **22**, 327; **24**, 27; **26**, 259; **137**, 111
Chromocarb **23**, 43
Chronic obstructive lung disease **8**, 1
Chronic pain **139**, 237
Chylomicron **34**, 175; **73**, 177
Chymostatin **78**, 1, 9; **146**, 247
Chymotrypsin **12**, 295; **26**, 15; **95**, 171; **97**, 171; **117**, 95; **146**, 55, 247
Chymotrypsin A **26**, 15; **76**, 99; **77**, 65; **78**, 1
CI 977 **83**, 65
CI 987 **104**, 169
Ciclopiroxolamine **105**, 77
Cigarette smoking **33**, 55; **100**, 253
Ciglitazone **15**, 261
Ciliary body **11**, 21; **13**, 183; **95**, 101
Ciliary body epithelium **86**, 59
Ciliary motility **12**, 57, 67, 77; **107**, 77; **123**, 47; **128**, 9; **135**, 137; **136**, 181
Ciliated epithelium **12**, 77; **31**, 193
Cimetidine **5**, 167; **17**, 1; **31**, 33; **35**, 181; **43**, 59; **54**, 89; **81**, 1; **127**, 185; **140**, 195
Cinchocaine **9**, 1; **24**, 231
Cineole **74**, 157; **105**, 47; **111**, 235; **124**, 279; **127**, 9; **130**, 141; **145**, 37, 49; **146**, 41
Cinnamaldehyde **148**, 131
Cinnamon oil **111**, 235
Cinnarizine **29**, 113; **48**, 1; **126**, 287
Cinoxacin **111**, 83
Ciprocinonide **11**, 257
Ciprofloxacin **63**, 237; **77**, 193; **83**, 267; **88**, 379; **89**, 125; **93**, 121; **95**, 51; **101**, R7; **102**, 249; **106**, 229; **122**, 9; **123**, 173; **127**, 279; **131**, 131; **132**, 53; **136**, 117; **138**, 113; **143**, 143; **146**, 225
Ciprofloxacin derivative **102**, 249
Circular dichroism **34**, 247; **108**, 49
Circulation **90**, 73
Cisplatin **10**, 77; **24**, 91; **41**, 67; **57**, 241; **68**, 151; **85**, 39; **89**, 229; **99**, 343; **106**, 249; **124**, 165; **138**, 121; **146**, 263
Citicoline **134**, 89
Citrate calcium **3**, 127; **7**, 83; **28**, 23
Citrate potassium **36**, 233; **68**, 11
Citrate sodium **1**, 141; **63**, 183; **130**, 245
Citric acid **44**, 225; **45**, 13; **49**, 57; **52**, 79; **56**, 29; **84**, 167; **101**, 89; **110**, 47; **132**, 53; **147**, 31
Citric acid tributyl ester **103**, 293; **113**, 17; **127**, 203
Citric acid triethyl ester **103**, 293; **113**, 17; **127**, 203; **133**, 161; **138**, 57
CL 487 **91**, 23
CL 490 **91**, 23
CL 940 **139**, 79
Clarithromycin **109**, 45; **114**, 65; **118**, 47
Clay **22**, 357
Clebopride **123**, 285
Clenbuterol **71**, 237, 245
Clindamycin **34**, 75; **81**, 1; **85**, 97; **134**, 71
Clindamycin phosphate **74**, 215; **134**, 71
Clinifeed **28**, 249
Clioquinol **131**, 243
Clobazam **16**, 327; **61**, 189
Clobetasol propionate **26**, 145; **41**, 63; **94**, 211
Clobetasone butyrate **41**, 63
Clodronic acid **107**, 189
Clofazimine **54**, 155; **58**, 107; **80**, 277; **105**, 137; **109**, 147
Clofazimine derivative **58**, 107
Clofibrate **135**, 63, 161
Clofibride **76**, 225
Clomethiazole **35**, 129; **132**, 231
Clomipramine **34**, 75; **35**, 129; **94**, 135
Clonazepam **13**, 67; **16**, 327; **38**, 247; **41**, 75; **61**, 189
Clonidine **12**, 277; **24**, 267; **35**, 21; **43**, 201; **52**, 37; **65**, 159; **118**, 113; **130**, 83
Clopamide **99**, 327
Clopenthixol **6**, 185
Cloprostenol **4**, 1
Cloprothiazole **35**, 129
Clorazepate **61**, 189
Clorofene **9**, 49
Clotrimazole **10**, 119; **35**, 129; **88**, 9; **93**, 233; **105**, 77; **110**, 109; **138**, 67
Cloxacillin **3**, 151; **9**, 273; **26**, 133
CNV 8912 **138**, 113
Coagulation **10**, 133
Coated tablet **42**, 161
Coating **10**, 43, 57; **13**, 159; **39**, 1
Cobalt **40**, 43; **68**, 105
Cocaine **12**, 77; **17**, 1
Cod liver oil **9**, 17
Codeine **7**, 343; **9**, 245; **17**, 1; **32**, 7; **44**, 257; **45**, 1; **85**, 45; **87**, 31; **94**, 1; **102**, 71; **138**, 149
Codeine phosphate **80**, 263
Colchicine **68**, 135; **110**, 127; **118**, 55
Colecalciferol **29**, 73; **84**, 151; **118**, 55
Colestipol **109**, 107
Colestyramine **64**, 155; **109**, 107; **128**, 129
Colipase **144**, 115
Colistin **36**, 55; **82**, 157; **90**, 59
Colitis **126**, 139
Collagen **47**, 79; **88**, 171; **91**, 209; **120**, 115; **138**, 121
Collagen derivative **47**, 95
Collagen gel **123**, 173
Collagenase **144**, 27
Colloid **62**, 1; **63**, 177; **75**, 45; **94**, 115; **96**, 129; **113**, 57; **135**, 63; **148**, 165
Colloidal gold **90**, 119

Colon **2**, 101; **4**, 281; **30**, 95; **33**, 89; **69**, 221; **87**, 215; **95**, 193; **101**, 115; **106**, 255
Colon absorption **31**, 239; **79**, 135; **126**, 245; **130**, 33; **143**, 101
Colon adenocarcinoma **83**, 197; **104**, 187; **128**, 155; **129**, 223; **140**, 175
Colon carcinoma **95**, 209; **126**, 41
Color **83**, 73
Colorimetry **10**, 199; **17**, 161, 325; **57**, 235; **61**, 199, 261; **81**, 153; **113**, 263
Coloring agent **72**, 259; **86**, 25
Compaction **22**, 337; **31**, 201; **40**, 267
Compartment model **17**, 23
Compensation **40**, 1
Complement receptor **32**, 165
Complex formation **42**, 193; **43**, 225; **49**, 91, 163, 249; **50**, 45; **51**, 213; **54**, 175; **56**, 249; **57**, 63, 107; **58**, 19, 41, 221; **59**, 35, 45, 225; **61**, 199; **63**, 95, 259, 267; **67**, 11; **68**, 105; **69**, 211; **70**, 35; **72**, 123; **73**, 43, 81, 9; **74**, 127; **75**, 193; **77**, 247, 303; **80**, 9, 243, 253; **82**, 229; **85**, 175, 189; **87**, 73, 79, 203; **88**, 1, 189; **89**, R5; **90**, 35, 181; **91**, 15; **96**, 91, 139; **99**, 119, 275; **106**, 179; **108**, 69, 173; **109**, 99; **110**, 169; **112**, 249; **114**, 13, 95; **115**, 255; **117**, 247; **118**, 77; **127**, 135, 239, 245; **128**, 81; **129**, 63; **131**, 219; **133**, 237; **134**, 213; **135**, 63; **136**, 165; **139**, 105; **141**, 1, 117; **142**, 183; **143**, 113; **148**, 219; **149**, 1
Complexation **28**, 95
Component analysis **19**, 27
Compressed tablet formulation **40**, 129
Compression **10**, 209; **27**, 127; **49**, 1, 157, 209; **57**, R1; **61**, 251; **63**, 23, 275; **64**, 27, 35, 223; **66**, 273, 283; **67**, 113, 217, 231; **68**, 51, 157; **70**, 103, 241; **71**, 81; **72**, 111, 117, 233; **73**, 37, 239; **82**, 71; **84**, 191; **85**, 121; **88**, 9, 261, 285; **89**, R17; **93**, 249; **94**, 181; **97**, 195; **99**, 99; **103**, 155; **112**, 173; **114**, 23; **124**, 69, 231; **128**, 1; **129**, 1, 21; **132**, 183; **142**, 97; **146**, 101
Compression speed **40**, 15
Compril **144**, 147
Computer **17**, 167; **61**, 225; **87**, 89
Computer program **55**, 193; **73**, 147; **79**, 1; **98**, 209; **129**, 243
Computer simulation **87**, 175; **104**, 253; **110**, 101
Concentration **37**, 19; **90**, R9
Conductance **55**, 77; **61**, 57; **76**, 207; **137**, 243
Conductivity **13**, 67
Conformation **73**, 185; **79**, 199
Conjugated estrogen **67**, 211
Conjugation **57**, 263
Conjunctiva **11**, 21; **16**, 163; **19**, 107; **30**, 73; **53**, 219; **106**, 161
Conjunctiva epithelium **84**, 217
Contact lens wetting **23**, 357
Container **54**, 181; **58**, 25; **59**, 245; **68**, 281; **77**, 47; **78**, 115; **86**, 89; **88**, 351; **94**, 223; **108**, 1; **130**, 251; **140**, 169
Contamination **28**, 211; **48**, 255; **51**, 55; **58**, 155; **74**, 175
Contramid **136**, 43
Contrast medium **92**, 1; **117**, 1; **132**, 9; **140**, 13
Controlled drug release **40**, 33; **146**, 55
Controlled release formulation **144**, 37; **148**, 85, 219
Convulsion **1**, 315; **48**, 91; **93**, 7
Coordination compound **133**, 237; **148**, 191
Copolymer **38**, 231; **48**, 1; **65**, 201; **70**, 77, 147; **80**, 277; **81**, 31; **84**, 23; **87**, 37; **91**, 29; **95**, 105; **100**, 9; **103**, 37, 293; **104**, 227; **112**, 79; **113**, 223; **120**, 33; **123**, 33; **124**, 149; **126**, 129; **128**, 99; **129**, 113; **131**, 229; **132**, 195; **136**, 31; **137**, 67, 199; **146**, 55, 71; **148**, 139; **149**, 93
Copper **68**, 105; **99**, 119; **115**, 175; **121**, 107; **133**, 237
Copper acetate **33**, 105
Copper complex **148**, 191
Copper ion **22**, 283; **90**, 181; **134**, 213
Copper sulfate **104**, 247
Copper zinc superoxide dismutase **132**, 263
Coralyne **10**, 239
Cordycepin **59**, 45
Corn oil **106**, 15; **130**, 203; **142**, 1; **147**, 31
Corn starch **1**, 65
Cornea **2**, 239, 337; **11**, 21; **29**, 43, 259; **33**, 15; **34**, 163; **37**, 19; **46**, 95; **55**, 237; **61**, 15, 163; **63**, 1; **66**, 23, 131; **68**, 135, 231; **74**, 221; **83**, 177; **88**, 111, 125; **93**, 49; **103**, 283; **119**, 149; **145**, 231
Cornea epithelium **123**, 173
Cornea membrane **2**, 239
Cornea permeability **2**, 239, 337; **16**, 203; **39**, 229; **63**, 49; **110**, 211; **118**, 243; **127**, 85; **132**, 165; **133**, 171; **137**, 209; **142**, 103
Corpus striatum **72**, R5
Corticosteroid **2**, 265; **6**, 195; **19**, 245; **22**, 137; **55**, 1; **74**, 1; **80**, 253; **94**, 211; **95**, 105; **103**, 19; **110**, 169; **123**, 1, 159
Corticosteroid derivative **138**, 95
Corticosterone **71**, 55; **132**, 23; **138**, 95, 149
Corticosterone **4**, 249
Corticotropin **75**, 97
Corticotropin[1–10] **50**, 261
Cortisone **7**, 99; **10**, 1; **14**, 1, 191; **16**, 305; **17**, 1; **20**, 347; **21**, 155; **27**, 287; **125**, 195; **132**, 23
Cortisone 21 acetate **10**, 1; **17**, 91; **27**, 287
Cortisone acetate **21**, 155
Cortisone behenate **17**, 91
Cortisone butyrate **17**, 91; **27**, 287
Cortisone butyrate **21**, 155
Cortisone derivative **17**, 91
Cortisone ester **27**, 287
Cortisone hexanoate **17**, 91; **21**, 155; **27**, 287
Cortisone octanoate **16**, 305; **17**, 91; **20**, 347; **21**, 155; **27**, 287
Cortisone palmitate **16**, 305; **17**, 91; **20**, 347
Cortodoxone **50**, 253; **132**, 23; **138**, 149
Corvic **60**, 197
Cosurfactant **148**, 47
Cotinine **80**, 227
Cotrimoxazole **59**, 255; **63**, 173; **72**, 141; **85**, 209; **90**, 51
Cotton seed oil **119**, 173; **121**, 149; **130**, 57; **132**, 33
Cough syrup **32**, 7
Coulter counter **122**, 57
Coumachlor **59**, 137
Coumarin **80**, 75; **94**, 1
Coumarin derivative **59**, 137
Coumetarol **12**, 231; **59**, 137

Cream **41**, 49; **66**, 53; **68**, 205; **70**, 167; **77**, 199; **86**, 9, 147, 201; **137**, 243
Creatine kinase **120**, 197
Creatinine **125**, 205; **145**, 9
Creatinine blood level **34**, 231
Creatinine clearance **34**, 231
Cremophor **10**, 109; **13**, 303; **35**, 1; **39**, 87; **54**, 171; **108**, 207; **127**, 65; **130**, 203; **139**, 113; **147**, 31
Cresol **25**, 105; **34**, 115; **125**, 205
Cresotamide **15**, 273; **20**, 129
Cresotamide derivative **15**, 273; **20**, 129
Critical micelle concentration **13**, 47; **24**, 297; **49**, 129
Crohn disease **68**, 199
Cromoglycate disodium **12**, 77, 87; **16**, 163; **23**, 333; **35**, 139; **37**, 239; **39**, 101, 107, 189; **42**, 211; **53**, 219; **57**, 205; **58**, 49, 57; **64**, 1; **73**, 131; **74**, 203; **75**, 73; **80**, R1; **83**, 233; **87**, 255; **101**, 1; **102**, 127; **103**, 27; **106**, 69; **116**, 11; **124**, 149; **134**, 137; **135**, 53; **138**, 45
Cromoglycate disodium-tc-99m **37**, 239
Cromoglycic acid derivative **55**, 1
Cromoglycic acid **16**, 163
Cromolyn **6**, 261
Croscarmellose sodium **38**, 255; **39**, 201; **45**, 111; **47**, 89; **54**, 161; **57**, 139; **62**, 125; **83**, 65; **85**, 141; **92**, 55; **97**, 119; **109**, 173; **110**, 257; **130**, 41; **144**, 195; **147**, 11
Crospovidone **33**, 115; **39**, 201; **44**, 177; **50**, 147; **97**, 119; **101**, 45, 127; **106**, 149; **110**, 257; **126**, 65; **128**, 129
Cross linking **57**, 205; **59**, 73; **96**, 33
Cross linking reagent **128**, 197
Cryoprotection **107**, 99; **131**, 257
Cryoprotective agent **107**, 99; **131**, 257
Crystal **8**, 11; **14**, 243; **17**, 237; **19**, 35; **20**, 3, 307; **24**, 239; **42**, 123; **69**, 255; **72**, 233
Crystal structure **41**, 29; **59**, 281; **63**, 35; **92**, 183; **95**, 93; **101**, 127, 169; **109**, 83; **125**, 257; **127**, 109; **133**, 117; **137**, 255; **146**, 93
Crystal violet **32**, 91
Crystalline substance **1**, 33; **24**, 1
Crystallization **14**, 325; **25**, 41; **34**, 153; **41**, 29; **42**, 123; **50**, 111; **51**, 147; **56**, 125; **58**, 185; **60**, 11, 243; **61**, 27; **62**, 65; **65**, 141; **66**, 47, 143; **67**, 59, 131; **68**, 25, 35, 179; **73**, 9; **74**, 33; **75**, 219; **76**, 193; **79**, 171; **82**, 185, 195; **85**, 87, 149; **87**, 1; **88**, 345; **98**, 189; **101**, 237; **104**, 169; **105**, 125; **108**, 195; **110**, 9; **112**, 17, 55, 65, 79, 215; **115**, 139, 151, 245; **116**, 113; **118**, 1; **119**, 25, 139; **121**, 19, 73, 81; **123**, 265; **126**, 21; **126**, 11; **127**, 53; **128**, 1; **129**, 79, 147, 159, 289; **130**, 195; **137**, 21; **139**, 33; **141**, 257; **142**, 115, 175; **143**, 247; **144**, 195
Crystallography **2**, 81; **3**, 261; **14**, 325; **24**, 239; **43**, 241
Curcuma longa **87**, 79
Curcumin **38**, 247; **41**, 75; **50**, 67, 91, 179, 259; **57**, 173; **58**, 237; **76**, 257; **87**, 79; **90**, 221; **100**, 93; **110**, 161; **131**, 1; **132**, 251
Curcumin derivative **51**, 179, 259
Curcuminoid **131**, 1
Cyanamide **142**, 227
Cyanide **33**, 1
Cyanoacrylate derivative **80**, 1
Cyanoacrylate **16**, 105; **84**, 1, 13; **101**, 165; **139**, 1
2-Cyanoacrylic acid ethyl ester **84**, 1, 13
2-Cyanoacrylic acid isohexyl ester **80**, 1; **84**, 1; **84**, 13
2-Cyanoacrylic acid methyl ester **84**, 1; **84**, 13
Cyanocobalamin **38**, 239; **81**, 1; **145**, 1
Cyanonaphthalene **121**, 27
4-Cyanophenol **121**, 27
Cyclacillin **9**, 273; **12**, 185; **17**, 47; **30**, 181; **85**, 181
Cyclic amp **58**, 129
Cyclizine **17**, 1
Cycloalkane derivative **83**, 39
Cycloalkanone **77**, 21
Cyclobarbital **20**, 73; **26**, 157; **79**, 21
Cyclodextrin **13**, 253; **16**, 327; **20**, 155; **31**, 231; **63**, 259; **65**, 101; **67**, R5; **68**, 271; **70**, 35; **71**, 19; **72**, 123; **73**, 9, 43; **74**, 85, 127; **75**, 37, R5; **78**, 25; **79**, 107, 273, 289; **84**, 129; **88**, 189; **89**, R5; **95**, 1; **96**, 91; **98**, 225, 239; **104**, 145, 239; **110**, 169; **115**, 255; **117**, 85; **124**, 149; **133**, 191; **135**, 137, 161, 183; **138**, 37; **142**, 153
α-Cyclodextrin **10**, 1; **13**, 253; **20**, 155; **31**, 231; **36**, 223; **43**, 41; **48**, 119; **61**, 27; **78**, 25; **101**, 97; **109**, 99; **110**, 93; **127**, 239; **128**, 239; **137**, 103; **138**, 37; **142**, 153; **148**, 33
β-Cyclodextrin **10**, 1; **13**, 253; **20**, 155; **21**, 51; **23**, 35; **24**, 79; **26**, 77; **28**, 47, 95; **30**, 35; **31**, 231; **33**, 105; **35**, 193; **36**, 223; **38**, 191; **43**, 93; **46**, 49; **48**, 83, 119; **49**, 91, 163; **50**, 45; **51**, 213; **52**, 115; **54**, 175; **56**, 249; **57**, 107; **58**, 19, 221; **59**, 45; **62**, 153; **63**, 95, 259, 267; **64**, 195; **66**, 289; **69**, 211; **75**, 147, 193; **77**, 247, 297, 303; **78**, 25, 183; **79**, 149; **85**, 1, 175; **86**, 263; **88**, 1; **90**, 35; **90**, 247; **93**, 201; **94**, 69; **96**, 5; **99**, 51; **100**, 25; **101**, 71, 97, 127; **104**, 181; **106**, 63, 135, 179; **109**, 99; **110**, 93; **111**, 111; **112**, 249; **114**, 95; **117**, 247; **120**, 1, 189, 197; **123**, 53; **124**, 231; **126**, 275; **127**, 239; **128**, 81; **129**, 63, 113; **131**, 219; **132**, 271; **136**, 43, 165; **137**, 103; **138**, 37; **141**, 117; **142**, 121, 153, 183; **148**, 211
γ-Cyclodextrin **10**, 1; **20**, 155; **13**, 253; **29**, 161; **48**, 119; **59**, 121; **78**, 25; **79**, 289; **82**, 29; **101**, 97; **105**, 249; **110**, 93; **124**, 213; **127**, 239; **133**, 191; **137**, 103; **138**, 37; **142**, 153; **143**, 113; **148**, 33
Cyclodextrin derivative **35**, 193; **62**, 243; **90**, 187; **101**, 97; **120**, 189, 197; **125**, 17, 31, 91; **126**, 73; **127**, 239; **129**, 63; **137**, 103, 209; **141**, 151; **144**, 225; **148**, 33
β-Cyclodextrin derivative **26**, 77; **46**, 217; **49**, 163; **91**, 15; **99**, 85; **124**, 205; **125**, 55; **129**, 113; **132**, 189; **138**, 37; **141**, 117; **145**, 241; **148**, 33; **149**, 1
γ-Cyclodextrin derivative **133**, 191
α-Cyclodextrin epichlorohydrin polymer **23**, 35
β-Cyclodextrin epichlorohydrin polymer **23**, 35
γ-Cyclodextrin epichlorohydrin polymer **23**, 35
Cyclodisone **66**, 253
Cycloheptane derivative **144**, 81
Cyclohexane **13**, 75; **14**, 263; **27**, 39; **29**, 177; **82**, 145; **113**, 97; **122**, 49; **129**, 51; **138**, 149; **140**, 237
Cyclohexanol **13**, 75; **17**, 115; **29**, 177; **140**, 155
Cyclohexanone **14**, 263
N-Cyclohexyl-*n*-methyl-4(1,2,3,5-tetrahydro-2-oxoimidazo [2,1b]quinazolin-7 **45**, 129
Cyclohexylethylmethylidenemalonate **92**, 71

Cyclopentane **138**, 149
Cyclopenthiazide **69**, 265; **73**, 137
Cyclopentolate **120**, 169; **138**, 37
Cyclophosphamide **7**, 1; **68**, 151; **108**, 69; **124**, 205
Cyclo-s,s[acetylcysteinyl(*n*-α-methylarginyl)glycylaspartylpenicillamine **106**, 95
Cycloserine **22**, 197
Cyclosporin **57**, 125, 133; **75**, 117; **103**, 277; **120**, 127; **121**, 181; **136**, 117
Cyclosporin A **44**, 107; **72**, 97; **81**, 21; **82**, 79, 85; **92**, 197, 203; **97**, 205; **99**, 263; **110**, 155; **114**, 221; **115**, 87; **120**, 127; **121**, 181; **126**, 169; **130**, 75; **141**, 81, 227; **142**, 1; **147**, 131
Cyoctol **65**, 77
Cysteine **28**, 119; **64**, 161; **72**, 149
Cystine **7**, 19; **28**, 119; **33**, 1; **64**, 161
Cytarabine **31**, 185; **68**, 151; **81**, 1; **108**, 69; **117**, 151; **124**, 61, 149, 205
Cytidine **29**, 177
Cytochalasin B **68**, 135; **88**, 125
Cytochrome *c* **65**, 261
Cytolysis **110**, 161
Cytosine **145**, 9
Cytoskeleton **68**, 135
Cytosol aminopeptidase **62**, 259; **111**, 147; **113**, 25; **135**, 191
Cytotoxic agent **21**, 219
Cytotoxicity **15**, 49; **29**, 169; **36**, 147; **37**, 135; **49**, 205; **59**, 27; **65**, 57, 219, 249; **84**, 13; **87**, 141; **89**, 139; **93**, 1; **98**, 19; **99**, 275; **101**, 97, 121; **102**, 55; **111**, 25; **122**, 107; **123**, 71; **131**, 1, 57; **138**, 85, 237; **139**, 69; **148**, 65, 231

D

DA 125 **103**, 147; **109**, 181
Dabicycline **124**, 271
Dacarbazine **108**, 69; **124**, 205
Dactinomycin **25**, 105
Daichitosan **78**, 43
Danazol **125**, 91, 99; **128**, 45; **145**, 137
Dantrolene **17**, 357; **64**, 161; **75**, 193
Dantron **55**, 221
Dapsone **32**, 21; **39**, 267; **47**, 1; **52**, 249; **121**, 157
Dapsone derivative **121**, 157
Daren **67**, 259
Data analysis **26**, 339
Daunomycinone **34**, 247; **35**, 219
Daunorubicin **22**, 137; **31**, 75; **34**, 247; **35**, 219; **56**, 217; **67**, 169; **72**, 123; **82**, 29; **111**, 25; **122**, 107; **135**, 167
Daunorubicin derivative **34**, 247; **122**, 107
Daunorubicinol **35**, 219
Deacetyldiltiazem **76**, 71
Deacetyllanatoside A **2**, 127
Deacetyllanatoside B **2**, 127
Dealkylflurazepam **49**, 83
Deaminase **21**, 265
3-Deazauridine **19**, 17
Decaethoxyoleyl ether **2**, 343
Decane **34**, 217; **123**, 71
Decanoic acid **33**, 235; **95**, 161, 181; **106**, 15; **109**, 45; **126**, 267; **136**, 147; **141**, 71; **145**, 215
Decanol **15**, 185; **29**, 177; **148**, 41
Decanoyl-*n*-methylglucamide **105**, 219
Decarboxylation **63**, 213
Decolorization **56**, 125
Decomposition **64**, 45; **86**, 231
Decompression **132**, 183
Decongestive agent **12**, 57
Decyl-glucoside **79**, 159; **105**, 219
Decylmaltoside **105**, 219; **132**, 107
Decyl-methyl-sulfoxide **27**, 89; **33**, 225; **43**, 1, 17; **49**, 39; **57**, 29; **87**, 261
3-Decyldimethylsilyl-*n*[2(4-methylphenyl)-1-phenylethyl]-propionamide **96**, 1
Deferoxamine **22**, 137
8-Defluorolomefloxacin **79**, 89
Deformity **67**, 59; **95**, 29; **106**, 187
Deglycyrrhizinated liquorice **119**, 133
Degradation kinetics **40**, 157
Degradation **30**, 133; **43**, 131; **50**, 91; **64**, 99; **67**, 155; **73**, 191; **80**, R11; **99**, 73; **109**, 191, 261; **134**, 27
Dehydration **59**, 217; **104**, 271; **106**, 33; **120**, 83; **134**, 27; **147**, 41
Dehydrocholic acid **6**, 137; **69**, 21; **74**, 127
Dehydropenicillin F **17**, 47
Dehymuls **99**, 29
Deiten **99**, 351
Demeclocycline **5**, 273
4-Demethoxydaunorubicinone **82**, 29
4′-Demethylpodophyllotoxin-4,6-*O*-benzylidene-β-dextro-glucopyranoside **122**, 57
Demoxepam **5**, 85
Dental caries **58**, 203
Denzimol **50**, 97
Deoxyacyclovir **99**, 79
7-Deoxydaunorubicinone **34**, 247
Deoxyadenosine **59**, 45
5′-Deoxyadenosine **59**, 45
Deoxycholate sodium **14**, 159; **65**, 243; **81**, R5; **128**, 29; **134**, 47; **142**, 103; **149**, 123
Deoxycholic acid **6**, 137; **8**, 303; **12**, 219; **14**, 159; **28**, 33; **38**, 211; **56**, 181; **66**, 131; **67**, 207; **69**, 29; **74**, 127; **75**, 45; **85**, 129; **89**, 81, 169; **92**, 197; **93**, 91; **101**, R7; **103**, 27; **105**, 137; **111**, 147; **114**, 65; **121**, 37; **125**, 327; **138**, 139
Deoxycorticosterone **4**, 249; **6**, 185; **14**, 191; **42**, 161; **132**, 23; **138**, 95, 149
Deoxyribonuclease **144**, 47
Deoxyribonucleic acid **16**, 153
Depolymerization **51**, 259; **142**, 175
Depten **108**, 11
Deptropine **34**, 75
Dequalinium **84**, 77; **86**, 193; **92**, 151; **104**, 247; **115**, 241; **121**, 107; **124**, 279; **137**, 143
Derivative UV spectroscopy **40**, 85
Dermatitis **49**, 29
Dermatological agent **148**, 131

Dermatophagoides pteronyssinus **119**, 239
Dermis **100**, 1; **102**, 241
Desciclovir **57**, 229
Desintegrating agent **15**, 115, 167
Desipramine **35**, 129; **92**, 15; **94**, 135
Deslanoside **2**, 127
Desmopressin **59**, 263; **64**, 181; **75**, 117; **81**, 119; **114**, 141; **127**, 115; **135**, 137; **136**, 117
Desorption **31**, 137; **127**, 43; **136**, 141
Desquamation **44**, 261
Detergent **34**, 183; **85**, 19
Detoxification **129**, 295
Deuteration **19**, 271
Deuterium ion **19**, 271
Deuterium oxide **113**, 189
Development **69**, R1
Dexamethasone **4**, 249; **10**, 1; **12**, 77; **17**, 1; **19**, 245; **39**, 229; **47**, 21; **59**, 225; **62**, 1; **80**, 253; **91**, 235; **92**, 105; **98**, 239; **104**, 181; **126**, 139
Dexamethasone derivative **89**, 157; **126**, 139
Dexamethasone sodium phosphate **59**, 225; **130**, 49
Dexamethasone-21-palmitate **89**, 157
Dexamphetamine **21**, 75
Dextran **8**, 131; **19**, 107; **22**, 137; **27**, 205; **37**, 135, 145; **44**, 125, 265; **49**, 23; **51**, 233; **52**, 55; **54**, 95; **57**, 263; **73**, 81; **77**, 141; **79**, 11, 159, 191; **83**, 1, 233; **87**, 255; **90**, 229; **92**, 71, 105; **100**, 199; **110**, 21; **119**, 149; **120**, 73; **124**, 37; **128**, 9, 23, 189; **129**, 175; **138**, 1; **143**, 133
Dextran-20 **120**, 73
Dextran-40 **16**, 319
Dextran-70 **16**, 319; **30**, 17; **65**, 7; **78**, 103
Dextran derivative **49**, 23; **113**, 149
Dextran sulfate **13**, 321; **19**, 107; **42**, 193; **46**, 103; **105**, 19; **118**, 121; **137**, 85
Dextran T 20 **27**, 219
Dextran T 40 **27**, 219
Dextran T 70 **27**, 219; **78**, 43
Dextran T 110 **27**, 219
Dextrin **98**, 219; **106**, 25; **120**, 21; **134**, 27
Dextromethorphan **14**, 173; **53**, 91; **73**, 51
Dextromorphan **44**, 257
Dextropropoxyphene **34**, 75; **102**, 213
Dextrorphan **14**, 173
Di-*o*-methyl-*β*-cyclodextrin **30**, 35; **31**, 25; **38**, 265
Di-tab **110**, 37
Diabetes mellitus **1**, 121; **12**, 107; **122**, 91
Diabetic ketoacidosis **1**, 121
Diacetolol **31**, 179
N,*o*-Diacetylsalicylamide **30**, 111
Diacylglycerol **115**, 45
Diagnosis **1**, 41; **76**, 183
Dialdehyde starch **30**, 133
1,2-Dialkyl-3-hydroxy-4-pyridone derivative **114**, 185
Dialkylmethylidenemalonate **92**, 71
Dialysis **4**, 171; **11**, 11; **13**, 67; **16**, 153; **18**, 225; **23**, 109; **27**, 45; **30**, 123, 189; **55**, 165; **79**, 83
Diamide **86**, 231
2,4-Diamino-6-hydroxymethylpteridine **16**, 141
2,7-Diamino-1-hydroxymitosene **24**, 219
Diamorphine **13**, 219
Dianhydrogalactitol **145**, 9
Diapam **110**, 197
Diazepam **1**, 141, 173; **3**, 195; **5**, 127, 161; **6**, 137, 195, 205; **7**, 111; **10**, 339; **11**, 11; **13**, 67, 287; **14**, 1; **16**, 327; **17**, 1; **23**, 195; **26**, 77; **33**, 37; **34**, 17; **39**, 213; **40**, 223; **47**, 1, 111; **49**, 91, 163; **51**, 213, 263; **54**, 1, 171; **60**, 35; **61**, 189; **62**, 207; **63**, 95; **64**, 195; **66**, 29, 263; **67**, 201; **71**, 1; **80**, 81; **81**, 1; **83**, 241; **89**, 139, 207; **91**, 227; **98**, 219; **110**, 169, 197; **129**, 253; **130**, 213; **136**, 13; **146**, 11; **148**, 47
Diazepine **1**, 173
Diaziquone **47**, 215
Diaziquone derivative **47**, 215
Dibehenoylphosphatidylcholine **128**, 163
Dibekacin **14**, 279; **30**, 181; **42**, 193
4,7-Dibenzyl-2,3,4,5,6,7-hexahydro-5,6-dihydroxy-1,3-bis[4-(hydroxymethyl)benzyl]-2H-1,3-diazepin-2-one **144**, 99
Dibenzylamine **67**, 259
Dibrompropamidine **105**, 71
Dibrompropamidine-isetionate **34**, 115; **36**, 191; **75**, 81; **82**, 107; **87**, 195; **92**, 237
Dibutyl ether **13**, 75
α(Dibutylaminomethyl)-6,8-dichloro-2-(3,4-Dichlorophenyl)-4 **6**, 195
Dibutyryladrenalone **1**, 189
Dicalcium-hydrogen-phosphate **12**, 341; **13**, 287; **17**, 39, 313; **24**, 355; **28**, 23; **31**, 15, 201; **35**, 29; **37**, 15, 119; **44**, 151; **45**, 79; **46**, 35; **48**, 173; **51**, 63, 77; **54**, 131; **55**, 135; **56**, 51; **57**, 139; **59**, 145; **60**, 197; **62**, 125; **67**, 247; **69**, 43; **71**, 81, 201; **73**, 75, 249; **75**, 181; **86**, 187
Dicarboxylic acid derivative **111**, 153
Dicetyl-phosphate **105**, 1; **121**, 195; **140**, 119
Dichlorodifluoromethane **124**, 195
Dichloroethane **17**, 77; **35**, 173
Dichloromethane **21**, 119; **28**, 75; **35**, 173; **76**, 91; **98**, 157; **99**, 93; **113**, 257; **126**, 129; **129**, 51; **131**, 171; **136**, 165; **142**, 67
2′,4′-Dichloropenicillin G **17**, 47
5-(3,4-Dichlorophenyl)-2,3-dihydro-6,7-bis(hydroxymethyl)-1H-pyrrolizine **13**, 303
Diclofenac **24**, 115; **27**, 245; **33**, 181; **42**, 251; **46**, 1; **48**, 1; **53**, 7; **66**, 223; **70**, 69; **73**, 239; **74**, 85, 105; **79**, 21; **79**, 263; **83**, 73; **85**, 57; **100**, 241; **89**, 191; **90**, 171; **92**, 211; **99**, 173; **108**, 109, 187; **113**, 57, 165; **120**, 119, 157; **121**, 19, 239; **126**, 95, 111; **130**, 141; **133**, 149; **135**, 177; **136**, 37, 101, 117; **137**, 11; **138**, 215; **139**, 53; **140**, 131; **147**, 41, 219
Diclofenac-diethylamine **125**, 231
Diclofenac-*n*(2-hydroxyethyl)-pyrrolidime **136**, 101
Dicoumarol **19**, 307; **29**, 73; **37**, 103; **59**, 137
Didecanoyl-α-phosphatidylcholine **89**, 147
Didecanoylphosphatidylcholine **128**, 239
2′,3′-Didehydro-2′,3′-dideoxythymidine **58**, R1
2′,3′-Didehydro-2′,3′-dideoxythymidine derivative **58**, R1
2′,3′-Dideoxyadenosine **45**, 27; **59**, 35, 45
2′,3′-Dideoxycytidine **95**, 1
2′,3′-Dideoxyguanosine **73**, 105

2′,3′-Dideoxyinosine **45**, 27; **95**, 1; **98**, 149; **109**, 125
2′,3′-Dideoxyinosine-triphosphate **98**, 149
2′,3′-Dideoxynucleoside derivative **95**, 1
Didodecyldimethylammonium bromide **139**, 69
1,3-Didodecylurea **56**, 43
Dielectric constant **46**, 267; **47**, 261; **61**, 251; **84**, 283; **88**, 437
Diet **6**, 301; **52**, 155
Diet restriction **59**, 105
Dietary fiber **76**, 123
Diethanolamine **15**, 185; **137**, 143
Diethazine **94**, 135
7-Diethylamino-4-methylcoumarin **12**, 355
N-(Diethylaminomethyl)benzamide **9**, 7
Diethylaminoethyldextran **76**, 61; **78**, 43; **117**, 139
Diethylcarbamazine **19**, 229; **24**, 355; **27**, 157
Diethyldithiocarbamic acid **11**, 45; **124**, 165
Diethyldithiocarbamic acid methyl ester **11**, 45
Diethylene glycol **15**, 185
Diethylene-glycol-monoethyl-ether **92**, 219; **105**, 227; **111**, 191; **118**, 221; **121**, 89; **131**, 183; **140**, 45; **147**, 165
1-(*N*,*n*-Diethylglycyloxymethyl)allopurinol **27**, 71
Diethylmalonuric acid-*β*-*n*,*n*-dimethylaminoethyl ester **6**, 19
Diethylstilbestrol **11**, 27; **32**, 187
Diethyltoluamide **49**, 199; **141**, 17
Difenoxin **4**, 39
Differential pulse polarography **48**, 167
Differential scanning calorimetry **2**, 203; **20**, 217; **27**, 311; **28**, 95; **40**, 129; **62**, 217; **72**, 181; **89**, 119; **106**, 115; **135**, 13; **138**, 191
Diffraction **88**, 141; **127**, 219; **141**, 257; **143**, 113
Diffusion **2**, 41; **18**, 235, 247; **47**, 79; **50**, 197; **56**, 1, 197, 261; **71**, 237; **77**, 1; **80**, 145; **100**, 155; **101**, 35, 45; **122**, 49, 79; **126**, 219; **132**, 23
Diffusion cell **37**, 251
Diffusion coefficient **3**, 143; **9**, 285; **14**, 223; **57**, 197; **61**, 173; **113**, 165; **120**, 229; **121**, 27; **145**, 49
Difloxacin **63**, 237; **83**, 267; **88**, 379; **93**, 121
Diflunisal **51**, 225; **54**, 57; **57**, 197
α-(2,4-Difluorophenyl)-α[1[2(2-pyridyl)phenyl]-ethenyl]-1H-1,2,4; **94**, 11
Digestive system injury **106**, 63
Digitalis glycoside **2**, 127
Digitonin **66**, 131; **88**, 125; **142**, 103
Digitoxin **2**, 127; **26**, 77; **54**, 1
Diglycine **71**, 117
Digoxin **1**, 3; **2**, 127, 173; **5**, 167; **6**, 205; **12**, 1; **13**, 353; **14**, 1, 123; **26**, 267; **27**, 361; **47**, 223; **49**, 241; **53**, 181; **54**, 1; **72**, R1; **81**, 1; **82**, 185, 195; **94**, 1; **97**, 1; **118**, 129; **133**, 39; **136**, 117
Digoxin blood level **13**, 353
Digoxin-hemihydrate **82**, 185, 195
Dihexanoyladrenalone **1**, 189
Dihydralazine **61**, 261
1-(2,3-Dihydro-1-methyl-2-oxo-5-phenyl-1H-1,4-benzodiazepin-3-yl)-3(3-methylphenyl)urea **96**, 147; **130**, 195
17-*β*[[(1,4-Dihydro-1-methyl-3-pyridyl)-arbonyl]oxy]-3-methoxyestra-1,3,5(10)-triene **35**, 47
3[[(1,4-Dihydro-1-methyl-3-pyridyl)-carbonyl]oxy]-estra-1,3,5(10)-trien-17-one **35**, 47
17-*β*[[(1,4-Dihydro-1-methyl-3-pyridyl)-carbonyl]oxy]-estra 1,3,5(10)-trien **35**, 47
3,4-Dihydro-3-(2-nitratoethyl)-2H-1,3-benzoxazin-4-one **128**, 277
3,4-Dihydro-2H-pyran derivative **123**, 95
Dihydroartemisinine **69**, 83
Dihydroergocornine **4**, 111; **6**, 87, 183
Dihydroergocristine **4**, 111; **6**, 87, 183; **7**, 19
Dihydroergocryptine **4**, 111; **6**, 87, 183
Dihydroergotamine **4**, 111; **6**, 183; **7**, 19; **56**, 181; **67**, 97, 169; **130**, 225
Dihydroergotamine-mesilate **117**, 147; **135**, 137
Dihydroergotoxine **6**, 87
Dihydrofolate-reductase **116**, 217
Dihydropyridine derivative **79**, 273; **140**, 33
Dihydrostreptomycin **19**, 263; **105**, 189
9[(1,3-Dihydroxy-2-propoxy)-methyl]-guanine **35**, 73
Dihydroxyaluminum sodium carbonate **19**, 307; **34**, 189
3[3[2-(3,4-Dihydroxyphenyl)-ethylamino]-butyl]-benzamide **123**, 181
1-(3,4-Dihydroxyphenyl)-2-propylamine derivative **99**, 135
Diisopentylamine **15**, 185
Diisopropanolamine **8**, 193
Diisopropyl ether **29**, 177
3,5-Diisopropylsalicylic acid **121**, 107
Diisovaleryladrenalone **1**, 189
Dilazep **46**, 173
Diloxanide furoate **13**, 145, 159, 171; **134**, 231
Diltiazem **17**, 183; **30**, 29; **50**, 71; **53**, 7; **76**, 71, 133; **79**, 183; **88**, 99; **95**, 161; **104**, 95; **109**, 271; **118**, 257; **131**, 153; **137**, 159; **138**, 225; **140**, 131; **147**, 135
Dilution **115**, 225
Dimerization **137**, 11
1[4[3[*N*(3,4-Dimethoxyphenethyl)-*n*-methylamino]propoxy] benzenesulfonyl]-2-isopropylindolizine **123**, 273
3,11b-Dimethyl-1,4-benzodiazepino[5,4b]-oxazole **81**, 179
Dimethyl-*β*-cyclodextrin **64**, 61; **132**, 189
2,6-*O*-Dimethyl-*β*-cyclodextrin **90**, 187; **141**, 151
N,*n*-Dimethyl-1,4-phenylenediamine **93**, 13
3,4-Dimethyl-5-phenyloxazolidine **30**, 151
3,4-Dimethyl-5-phenyloxazolidine derivative **30**, 151
1,5-Dimethyl-2-pyrrolidone **95**, 43
Dimethyl-sulfoxide **4**, 309; **7**, 99; **14**, 263; **15**, 185; **26**, 275; **36**, 211; **48**, 141; **49**, 199; **53**, 13; **55**, 129; **59**, 211; **69**, 193; **71**, 193; **76**, 217; **105**, 47; **125**, 17, 31, 265; **129**, 283; **137**, 125
2,2-Dimethyl-*n*-(2,4,6-trimethoxyphenyl)-dodecanamide **108**, 85; **128**, 29
N,*n*-Dimethylacetamide **13**, 303; **14**, 263; **17**, 77; **49**, 199; **129**, 283
6-*N*,*n*-Dimethyladenosine **59**, 45
Dimethylamino-curcuminoid **131**, 1
1-Dimethylamino-2-propanol-dodecanoate **118**, 113
2-(*N*,*n*-Dimethylamino)propionic acid dodecyl ester **127**, 245
Dimethylaminoacetic-acid-dodecyl-ester **99**, 253
6-*N*-(3,3-Dimethylallyl)adenosine **65**, 57
4-Dimethylaminoazobenzene **7**, 99
4-Dimethylaminobenzoic acid **50**, 27

4-Dimethylaminobenzoic-acid-2-ethylhexyl-ester **50**, 27
N,*n*-Dimethyllauramide **76**, 151
N,*n*-Dimethylcapramide **76**, 151
N,*n*-Dimethylformamide **14**, 263; **17**, 77; **22**, 291; **36**, 211; **48**, 195; **49**, 199; **56**, 181; **82**, 215; **108**, 149; **129**, 283
7-(*N*,*n*-Dimethylglycyloxymethyl)theophylline **12**, 299
Dimethylglyoxime **29**, 177
5,5-Dimethylhydantoin **5**, 67
3,4-Dimethylphenol **18**, 247
N-(2,6-Dimethylphenyl)acetamide derivative **22**, 345
N-(2,6-Dimethylphenyl)-2-dimethylsulfonylacetamide **22**, 345
N-(2,6-Dimethylphenyl)-2-ethylthioacetamide **22**, 345
N-(2,6-Dimethylphenyl)-2-methylthioacetamide **22**, 345
2-(2,3-Dimethylphenyl)-2,3,5,6-tetrahydroimidazo[2,1b] thiazole **33**, 201
7-(*N*,*n*-Dimethylsuccinamoyloxymethyl)theophylline **12**, 299
1,3-Dimethyluric acid **33**, 55
Dimeticone **15**, 73; **19**, 169; **21**, 341; **22**, 257; **28**, 249; **35**, 21, 91, 103; **45**, 1; **50**, 181; **94**, 81, 153; **119**, 81; **123**, 33; **125**, 297; **131**, 229; **137**, 149; **139**, 161
Dimyristoyllecithin **13**, 227; **14**, 191; **20**, 119, 139; **21**, 155
Dimyristoylphosphatidylcholine **50**, 261; **69**, 13; **80**, 93; **85**, 189; **89**, 183; **91**, 1; **98**, 149; **103**, 217; **118**, 65; **128**, 163; **130**, 65; **132**, 231; **146**, 123
Dimyristoylphosphatidylethanolamine **85**, 189
Dimyristoylphosphatidylglycerol **146**, 123
2,4-Dinitrophenol **1**, 127; **18**, 259; **103**, 27
Dioctadecyldimethylammonium bromide **139**, 69
Dioctanoyladrenalone **1**, 189
Dioctyl sulfosuccinate sodium **9**, 315
Dioleoyllecithin **4**, 281
Dioleoylphosphatidylcholine **103**, 81
Dioxane **17**, 77; **32**, 187; **38**, 71; **123**, 71; **129**, 283; **140**, 237
1,4 Dioxane **4**, 309
1,3-Dioxolane **15**, 223; **129**, 51
1,3 Dioxolane derivative **15**, 223
Dipalmitoyllecithin **13**, 227; **14**, 191; **16**, 305; **20**, 119, 139, 347; **23**, 299
Dipalmitoyllecithin I^{125} **20**, 347
Dipalmitoylphosphatidylcholine **42**, 97; **43**, 207; **61**, 189; **65**, 211; **76**, 217; **80**, 93; **89**, 183; **90**, 73; **91**, 1; **98**, 149; **103**, 237; **107**, 1; **113**, 141; **114**, 33; **118**, 65; **124**, 1, 19; **126**, 209; **128**, 163; **130**, 65; **132**, 89, 231, 263; **134**, 119; **138**, 139, 191; **140**, 119; **146**, 31, 81, 123
Dipalmitoylphosphatidylpolyglycerol **111**, 103
Dipeptide **20**, 273; **71**, 45; **100**, 271; **115**, 79; **140**, 175; **146**, 205
Dipeptidyl-carboxypeptidase **112**, 133; **134**, 147
Dipeptidyl-carboxypeptidase inhibitor **107**, 77; **128**, 229; **140**, 175
Dipeptidyl-peptidase **107**, 133
Diphenadione **19**, 307
Diphenhydramine **1**, 257; **6**, 205; **7**, 343; **8**, 277; **12**, 77; **30**, 209; **31**, 43; **35**, 259; **37**, 211; **50**, 75; **53**, 91; **60**, 109; **72**, 163; **81**, 75; **102**, 109
1,6-Diphenyl-1,3,5-hexatriene **98**, 203; **126**, 49; **138**, 139; **147**, 153
5[5-(4,6-Diphenyl-2-pyridyloxy)-1,1-dimethylpentyl]-tetrazole **118**, 221
5,5-Diphenyl-3-sulfonyloxymethylhydantoin **14**, 113
Diphenylpyraline **102**, 109
Diphtheria toxoid **76**, R5; **85**, R5
Dipicolinic acid **16**, 127
Dipivalyladrenalin **1**, 189; **17**, 299
Dipivalyladrenalone **1**, 189
Dipivefrine **17**, 299; **133**, 171
9[(1,3-Dipropionyloxy-2-propoxy)-methyl]-guanine **35**, 73
Diprophylline **48**, 1; **50**, 205; **61**, 127; **91**, 167
Dipropyl ether derivative **13**, 75
Dipropylene glycol **15**, 185
Diprotin A **113**, 1
Dipyridamole **81**, 49; **85**, 141; **88**, 31; **95**, 1; **103**, 243; **108**, 11
Dipyrone **18**, 9; **31**, 201
Disinfectant agent **17**, 291; **19**, 239; **25**, 245; **34**, 115
Disinfection **9**, 49; **65**, 35; **73**, 191
Disintegrating agent **147**, 95
Disintegration **50**, 147; **60**, 61
Disodium–hydrogen–phosphate **19**, 177; **78**, 237; **92**, 177; **124**, 219
Disopyramide **28**, 145; **54**, 1; **92**, 1
Disopyramide phosphate **28**, 145
Dispersion **28**, 11; **31**, 157; **35**, 7; **45**, 245; **56**, 197; **57**, 197; **58**, 1, 221, 229; **59**, 79; **60**, 171; **67**, 139, 201; **69**, 53; **71**, 229; **73**, 117; **75**, 25, 45, 155; **76**, 17, 207; **77**, 119; **78**, 175; **83**, 47, 115, 163, 211; **84**, 161, 197; **88**, 369, 417; **93**, 221; **96**, 129; **97**, 101; **103**, 47, 137; **104**, 125, 169; **108**, 101; **110**, 179, 267; **111**, 15; **115**, 225; **118**, 55; **123**, 25, 53; **139**, 223; **143**, 59
Dispersion coefficient **32**, 13
Dissociation **3**, 231; **18**, 207; **84**, R1
Dissociation constant **31**, 69; **42**, 181; **61**, 173, 265; **67**, R1; **74**, 105, 215; **88**, 379; **92**, 15
Dissolution **9**, 337; **11**, 81; **14**, 299; **26**, 165; **27**, 117; **28**, 23, 33, 95, 167; **29**, 113; **30**, 133; **31**, 157; **32**, 243; **36**, 73; **41**, 55, 159; **44**, 43, 51, 67, 223, 249, 261, 263; **49**, 91, 121, 179, 241, 261; **50**, 205, 222; **51**, 129, 213, 245; **53**, 7, 35; **54**, 123, 161; **55**, 25, 77, 193, 217; **56**, 151, 249; **57**, 107, 117, 197; **58**, 1, 19; **59**, 115, 255; **60**, 151; **61**, 173; **62**, 105, 143; **63**, 49, 95, R9; **65**, 169, R5; **69**, 13, 53, 255, 265; **73**, 17, 51, 221, 239; **74**, 43, 49, 169; **75**, 37, 63, 141, R1; **76**, 17, 49; **77**, 89, 183, 193, 255; **78**, 175, 183, 203; **79**, 67, 83, 289; **80**, 129, 151, 191, 243, 253, 277; **81**, 75; **83**, 65; **84**, 151, 161, 167, 223; **85**, 1, 57, 103, 113, 141, 149, R1; **86**, 9, 79; **87**, 161; **88**, 23, 31, 75, 141, 151, 159, 211, 261, 423; **89**, 111, 119, 207, R13; **90**, 51, 105; **91**, 23; **92**, 97, 115, 211; **93**, 111, 209, 215, 233; **94**, 59, 125; **95**, 67, 117, 143; **96**, 5, 91, 119, 139, 167, 175, 231; **97**, 93, 119; **98**, 29; **99**, 321; **100**, 25, 33, 115, 143, 155, 175, 207, 241; **102**, 47, 71, 91, 231; **102**, 213; **102**, 185; **103**, 97, 137; **104**, 11, 107, 125, 169; **106**, 25, 141, 149; **106**, 109; **108**, 101; **109**, 271, 291; **110**, 223, 249, 257; **111**, 117, 159, 171, 181, 217; **112**, 215, 233, 241; **113**, 97, 103; **114**, 95, 185, 205, 257; **115**, 1, 29, 123, 133, 201; **117**, 31, 41; **118**, 103; **121**, 29, 37, 129; **122**, 35; **123**, 53, 273; **125**, 99; **126**, 57, 65, 155, 161, 199,

287; **127**, 261, 283; **128**, 29; **129**, 73; **129**, 253; **130**, 41, 65, 213; **131**, 41, 273; **133**, 17, 139; **134**, 223; **135**, 73; **136**, 13, 43, 101, 175; **138**, 13, 57, 185; **139**, 169, 223, 231; **140**, 25, 69, 131, 247; **141**, 117, 179; **142**, 53; **143**, 59, 113, 179, 203, 211, 223, 247; **146**, 1; **148**, 73
Dissolution in vivo **40**, 165
Distearoyllecithin **13**, 227; **14**, 191; **20**, 119, 139; **23**, 299; **37**, 75; **126**, 41; **132**, 9, 263; **140**, 119; **143**, 1; **144**, 11; **146**, 123
Distribution coefficient **18**, 353; **31**, 209; **56**, 135
Disulfide bond **85**, 39
Disulfiram **11**, 45; **35**, 129
Di-tab **106**, 149; **109**, 1; **110**, 37
Ditekiren **73**, 231
1,2-Dithiole-3-thione derivative **129**, 295
Dithiothreitol **16**, 127; **89**, 51; **106**, 249; **134**, 147
Dithranol **49**, 63, 103; **55**, 221; **75**, 89
Dithranol derivative **55**, 1
Diuretic agent **6**, 205; **29**, 73
Divinylbenzene **88**, 303
DMV 171780 **90**, 213
DNA **72**, 251; **105**, 181; **139**, 69
DNA binding **49**, 103
DNA degradation **139**, 69
Dobutamine **92**, 1
Docusate-sodium **37**, 179; **45**, 91; **50**, 213; **124**, 97; **128**, 29; **146**, 271
Dodecyl-*n*,*n*-dimethylamino-acetate **92**, 89; **101**, 57; **103**, 77
Dodecyl-glucoside **105**, 219
Dodecyl-maltoside **105**, 219; **132**, 107, 115
1-Dodecyl-2-piperidone **68**, 239; **125**, 63
1-Dodecyl-2-pyrrolidinone **60**, 177; **125**, 63
Dodecyl sulfate **8**, 303; **9**, 173; **14**, 159; **16**, 127; **24**, 165, 355; **26**, 191; **27**, 117
Dodecyl–sulfate–lithium **110**, 267
Dodecyl–sulfate–magnesium **30**, 237
Dodecyl–sulfate–sodium **28**, 113; **38**, 211; **39**, 263; **47**, 13; **48**, 159; **49**, 147, 189; **51**, 9; **57**, 205; **60**, 61; **63**, 183; **68**, 205; **69**, 53; **71**, 213; **75**, 155; **79**, 123; **79**, 135, 159; **81**, 39, R1; **83**, 47, 53; **85**, 129; **87**, 261, 265; **90**, 105, 187, 195; **91**, 23, 105; **99**, 285; **104**, 145; **107**, 239; **110**, 147, 223, 267; **111**, 73; **120**, 179; **122**, 1; **123**, 71; **125**, 99; **128**, 29; **129**, 113, 123; **130**, 121; **131**, 41; **132**, 131; **133**, 71; **134**, 247; **135**, 73; **136**, 147; **137**, 125, 243; **138**, 185; **142**, 189; **143**, 119; **144**, 99; **145**, 175; **148**, 201
4-Dodecyl-3-thiomorpholinone **76**, 217
Dodecylsulfate–potassium **110**, 267
Dodecyl-sulfate **135**, 13
N-Dodecyldiethanolamine **125**, 63
Dodecyltrimethylammonium **120**, 179; **124**, 107
Dodecyltrimethylammonium-bromide **48**, 159; **57**, 189; **79**, 123
1-Dodecylurea **56**, 43
Dolasetron-mesilate **130**, 159
Dolichol **105**, 259
Dolomite **36**, 205
N-(4-Doluidinomethyl)succinimide **9**, 7
Dolviran **7**, 343
Domiphen-bromide **36**, 191
Domperidone **28**, 133; **103**, 179; **123**, 285
Dopa **116**, 51
Dopamine **72**, R5; **79**, 21; **127**, 115; **136**, 31
Dopamine derivative **28**, 1
Dopamine-1 receptor-blocking agent **79**, 205
Dopamine receptor-stimulating agent **80**, 39; **87**, 67
Dopamine-2 receptor-stimulating agent **88**, 165
Dose kidney function relation **5**, 117; **30**, 57
Dose response **1**, 95, 281; **3**, 13; **12**, 295; **13**, 23, 67, 75; **14**, 113; **24**, 297, 327, 339; **25**, 41, 199, 217; **27**, 45, 89; **28**, 47, 85, 119; **29**, 29, 37, 169; **31**, 1, 55, 231, 275; **32**, 133, 223; **33**, 45; **35**, 73, 227; **97**, 213
Dose time effect relation **12**, 277; **26**, 275; **27**, 45; **30**, 95; **33**, 55; **34**, 137, 145, 213; **37**, 179
Dosimetry **32**, 251
Dosulepin **52**, 123
Down regulation **54**, 259
Doxazosin **111**, 181
Doxepin **94**, 135
Doxorubicin **14**, 73; **16**, 79; **17**, 135; **22**, 299; **23**, 1; **30**, 17; **32**, 123; **33**, 137, 147; **38**, 257; **42**, 145; **43**, 161, 167; **48**, 1; **51**, 253; **53**, 67; **54**, 113; **56**, 217; **59**, 57; **68**, 151, 271; **72**, 123; **79**, 289, R1; **89**, R9; **96**, 157; **102**, 55; **103**, 87; **107**, 149; **108**, 69; **110**, 117; **111**, 25; **120**, 51; **121**, 1; **124**, 205; **126**, 41; **127**, 73; **129**, 113; **131**, 91; **135**, 167; **138**, 237; **144**, 11; **145**, 9; **146**, 31, 81; **148**, 139
Doxorubicin derivative **103**, 147; **109**, 181
Doxorubicinol **32**, 123; **121**, 1; **131**, 91
Doxorubicinone **23**, 1
Doxycycline **96**, 13
Doxylamine **53**, 91; **63**, 35
5-Doxylstearic acid **104**, 61
Droperidol **17**, 1
Droplet size **14**, 349; **17**, 203; **19**, 43; **27**, 99; **41**, 147; **46**, 177; **50**, 141; **58**, 13; **83**, 227; **88**, 417; **118**, 213; **123**, 247, 257
Drug **25**, 255; **82**, 1; **86**, 123; **105**, 57; **137**, 111; **144**, 141; **148**, 1
Drug absorption **1**, 323; **2**, 101, 215; **3**, 247, 299; **4**, 27, 99, 205, 271, 281, 291; **5**, 25; **6**, 43, 243, 285; **7**, 19, 83, 99, 137; **8**, 193, 253, 263; **9**, 17, 107, 165, 173; **10**, 143, 339; **11**, 119; **12**, 107; **13**, 99, 103; **14**, 251, 279; **15**, 125, 167, 235; **16**, 23, 163, 181, 191, 241, 255, 285, 327, 339, 353; **17**, 23, 47, 183, 273, 299, 331; **18**, 139, 157, 259; **19**, 75, 297; **20**, 43, 99, 119, 139, 163; **21**, 3, 17, 233, 239; **22**, 89, 177, 311, 345; **23**, 35, 43, 247; **24**, 61, 115, 173, 209, 259, 267; **25**, 27, 85, 95, 255; **26**, 157, 203, 317; **27**, 29, 89; **28**, 47, 95, 133, 151, 201, 219; **29**, 43, 113, 169; **30**, 35, 63, 73, 91, 95, 151, 225, 247; **31**, 33, 83, 107, 179, 185, 193, 231, 239; **32**, 13, 257; **33**, 37, 45, 89, 99, 181, 235, 249, 253; **34**, 35, 131, 137, 179; **35**, 91, 243; **36**, 41, 61, 67, 91, 105, 121, 185; **37**, 19, 27, 49, 97, 127, 245; **38**, 33; **41**, 139; **43**, 41, 59, 119, 225, 257; **44**, 65, 71, 187; **45**, 123, 161, 169, 197; **46**, 1, 19, 95, 193, 199; **47**, 113, 43, 185; **48**, 23, 91, 133, 159, 195, 141, 183, 213, 231; **50**, 53, 79, 83; **51**, 9, 47, 137, 183, 213; **52**, 191; **53**, 157, 227; **54**, 171; **55**, 1, 165, 265; **56**, 1, 29, 111, 181; **57**, 49, 73, 177, 189; **59**, 1, 255, 263; **60**, 1, 41, 125, 191; **61**, 1, 43, 51, 135, 241; **62**, 259; **63**,

101, 207; **64**, 61, 75, 127, 139, 161, 171, 181, 195, 207; **65**, 19, 49, 63, 77, 85, 115, R5; **66**, 53, 157, 193, 243; **67**, 1, 11, 259; **68**, 11, 121, 167, 191, 231, 239; **69**, 29, 103, 221; **70**, 19, 43, 87, 271; **71**, 25, 55, 105, 193, R9; **72**, 175, 193, R1; **73**, 89, 221, 231; **74**, 1, 9, 95, 115, 137, 147, 157, 189, 203, 229; **75**, 25, 73, 89, 97, 117, R1; **76**, 25, 37, 61, 151, R1; **77**, 71, 75, 141, 151, 177, 297, 303, 309; **81**, 119, 127, 193; **82**, 165; **84**, 117, 129, 141, 161, 217, 223, 245, 261; **85**, 175, 181; **86**, 239; **87**, 105, 125, 215, 229, 261, 265; **88**, 63, 89, 111, 125, 151, 211, 221, 237, 243, 251, 319, 423, 433; **89**, 41, 75, 147, 169, 199; **90**, 129; **99**, 165, 173, 209, 311; **102**, 223; **103**, 259; **104**, 145; **105**, 137, 189, 219; **106**, 15, 95; **107**, 159; **110**, 65; **113**, 1, 83; **117**, 129, 139; **118**, 55, 221, 251, 257; **125**, 205; **126**, 89; **127**, 115; **135**, 191; **137**, 33, 85, 103; **143**, 179; **144**, 171

Drug absorption(gastrointestinal) **40**, 165

Drug accumulation **15**, 185; **18**, 1; **22**, 137; **23**, 299; **27**, 145, 287, 361; **32**, 79; **37**, 33, 145, 245; **43**, 267; **65**, 249

Drug activity **54**, 241, 251; **58**, 129

Drug additive **72**, 243; **76**, 1, 49; **77**, 75; **78**, 35; **79**, 141, 257; **90**, 187, 213; **91**, 23; **92**, 197, 203; **95**, 117, 161; **101**, 71, 89; **103**, 9, 97; **105**, 219; **105**, 227, 255; **109**, 1; **110**, 155, 223; **112**, 17; **118**, 151; **119**, 65, 71; **121**, 81; **124**, 213; **126**, 147; **132**, 107, 115; **134**, 247; **140**, 169

Drug adhesion **45**, 91; **49**, 231

Drug administration **30**, 95; **34**, 157; **48**, 15; **55**, 1; **75**, 1, 97, 117

Drug administration route **17**, 273; **21**, 233; **24**, 79, 115; **27**, 81; **28**, 1, 47; **30**, 95, 215, 247; **31**, 185, 193; **32**, 103; **33**, 99, 235, 249; **34**, 35, 75, 131; **35**, 139; **36**, 81, 127; **37**, 145, 227, 239, 245

Drug adsorption **13**, 145; **16**, 181; **17**, 183; **19**, 307; **22**, 357; **27**, 71; **30**, 17; **34**, 189; **42**, 41; **44**, 1; **47**, 209; **48**, 125; **53**, 175; **55**, 209; **64**, 89; **67**, 89; **70**, 129; **84**, 261, 283; **86**, 89; **88**, 171, 327; **89**, 33; **108**, 31

Drug analysis **1**, 15, 49, 71, 95, 249, 275, 315; **2**, 81, 155, 203, 257, 317; **3**, 109, 205, 261, 275; **4**, 121, 39, 47, 111; **5**, 85; **6**, 87, 183; **7**, 189; **8**, 143, 211; **9**, 245, 195; **10**, 281; **11**, 131, 237; **12**, 163, 173, 185, 209, 265, 271; **23**, 13; **38**, 9, 39, 59, 65, 191; **40**, 193; **41**, 49, 63, 241; **41**, 147; **43**, 131; **43**, 207; **48**, 167; **49**, 129; **51**, 115; **52**, 155, 225; **75**, 249; **81**, 169; **85**, 163; **87**, 11; **126**, 281

Drug antagonism **23**, 357; **32**, 13; **34**, 51; **37**, 127

Drug antibody **120**, 247

Drug antigenicity **120**, 247

Drug bile level **27**, 45

Drug binding **1**, 61; **3**, 341; **15**, 97; **16**, 153; **17**, 65; **18**, 277; **22**, 75; **25**, 217; **30**, 181; **32**, 269; **33**, 45; **34**, 189; **35**, 121, 219; **36**, 113, 199; **37**, 135, 145; **44**, 213; **49**, 189; **54**, 113; **59**, 87; **60**, 1, 35; **61**, 189; **62**, 153; **67**, 123; **76**, 145; **81**, 143; **85**, 39, 45; **87**, 31, 37, 105, 203; **99**, 275, 285; **107**, 223; **144**, 71; **147**, 135

Drug binding protein **35**, 211

Drug bioavailability **41**, 255, 263; **45**, 197; **46**, 261; **52**, 37, 159; **53**, 157; **54**, 33, 65; **60**, 27, 191; **61**, 15, 43, 135; **62**, 53; **65**, 85, 133, R1; **66**, 23; **67**, 185; **68**, 11, 35, 167; **69**, 123, 169; **70**, 137; **73**, 89, 231; **74**, 1; **75**, 37, 55; **77**, 141, 199, 297; **81**, 133, 193; **84**, 223, 273; **85**, 45, 175, 209; **86**, 1, 35; **87**, 31, 59; **88**, 63, 165, 243, 409, 423; **89**, 9, 75, 245; **104**, 1; **106**, 95; **107**, 57; **113**, 83; **120**, 111; **137**, 233; **144**, 217; **148**, 155

Drug blood level **2**, 173; **7**, 99; **11**, 323; **12**, 277, 299; **13**, 1, 99, 103, 239, 353; **14**, 113, 173, 251; **16**, 23, 285; **17**, 23, 273; **18**, 1, 353; **19**, 229, 297, 345; **20**, 53, 139; **21**, 3, 187; **22**, 345; **23**, 35, 117, 277, 289; **24**, 61, 79, 101, 115, 173; **25**, 27, 347, 113; **26**, 99, 157, 317; **27**, 45, 71, 81, 245; **28**, 85, 95, 133, 151, 193; **29**, 53, 113, 191, 223; **30**, 57, 91, 95, 111, 143, 225, 247; **31**, 15, 33, 83, 165, 179, 185, 231; **32**, 13, 103, 133, 159; **33**, 1, 55, 89, 99, 219, 235, 249; **34**, 35, 207, 225, 259; **35**, 47, 173, 181; **36**, 41, 121, 147, 253; **37**, 97, 145, 163, 219, 245; **41**, 255, 263; **52**, 159; **53**, 157, 181; **60**, 35; **61**, 51, 119, 135; **64**, 109, 155, 187, 195; **67**, 67, 185, 259; **68**, 61, 97, 167; **69**, 5, 169; **71**, 25; **73**, 177, 231; **74**, 9, 147; **77**, 177; **82**, 21; **87**, 223, 229

Drug brain level **10**, 239; **72**, R5

Drug capsule **61**, 145; **63**, 275; **69**, 123; **71**, 127, R5; **73**, 89; **84**, 49; **95**, 1; **98**, 29, 219; **103**, 259, 283; **110**, 249, 257, 291; **112**, 207; **121**, 37, 117, 239; **122**, 79; **124**, 219; **125**, 99; **127**, 203; **128**, 123, 197; **130**, 203; **133**, 139; **134**, 223; **135**, 53; **136**, 147, 155; **138**, 13; **139**, 53, 187; **142**, 97

Drug carrier **14**, 43; **17**, 65; **22**, 137; **26**, 113; **28**, 125; **30**, 17, 133; **33**, 137, 147; **36**, 121, 253; **39**, 51; **40**, 201, 207; **41**, 197; **46**, 31, 149; **47**, 51, 95; **53**, 67; **57**, 55; **58**, 1, 49, 57, 115; **61**, 109; **63**, 183; **67**, 11, 29, 103, 139; **68**, 69, 111, 121; **69**, 129; **70**, 1, 129, 159; **74**, 9; **77**, 297; **81**, 103, 199, 235; **84**, 197, 283; **85**, 1, 29, 51, 75; **86**, 25, 137; **89**, 19, 25, 119, 139; **90**, 105, 119, 195; **91**, 157, 209, 227; **93**, 189, 215; **94**, 115, 143; **99**, 1, 13, 135; **100**, 49; **101**, 1, 165; **106**, 109; **108**, 21; **110**, 21; **112**, 215; **116**, 1; **126**, 139, 253; **128**, 261; **129**, 113, 159; **130**, 25, 213; **131**, 191; **132**, 195; **133**, 107; **134**, 119; **138**, 1, 75, 85; **139**, 25, 87; **140**, 247; **142**, 121, 227; **143**, 247; **145**, 241; **146**, 123; **147**, 153; **149**, 93

Drug cerebrospinal fluid level **8**, 253

Drug clearance **2**, 337; **16**, 31, 191; **19**, 75; **20**, 347; **22**, 137; **23**, 25; **26**, 303; **27**, 45; **28**, 85; **30**, 143; **32**, 103, 133; **36**, 127; **37**, 145, 163, 219; **43**, 221; **46**, 203, 241; **54**, 41, 113; **56**, 269; **64**, 109, 155; **65**, R1; **66**, 169; **73**, 231; **85**, 75; **135**, 115

Drug coating **17**, 347; **22**, 17, 265, 273; **27**, 267; **31**, 55; **43**, 155; **45**, 245; **50**, 163; **51**, 19; **57**, 183; **58**, 73, 89, 197; **60**, 109, 171; **63**, 43; **67**, 21, 29, 79; **68**, 157; **72**, 163; **73**, 197; **77**, 89; **78**, 49; **86**, 49, 113, 247; **87**, 37, 133; **88**, 75, 409; **89**, 25; **100**, 107, 115; **104**, 203; **106**, 149; **109**, 9, 271; **113**, 113; **119**, 81, 193; **125**, 243, 289; **132**, 267; **133**, 139, 253; **138**, 57; **139**, 187; **141**, 179; **144**, 47; **148**, 73

Drug comparison **1**, 49, 249, 275; **3**, 275; **4**, 1, 13; **9**, 295; **12**, 315, 355; **13**, 9, 67, 75; **14**, 113; **15**, 61, 235; **17**, 357; **18**, 235, 247, 325; **19**, 233, 239; **20**, 99, 273; **21**, 35, 51, 349; **21**, 3, 155, 251; **22**, 75, 131, 137, 283, 345; **23**, 13, 25, 117, 163, 209, 223; **24**, 43, 91, 165, 173, 231, 297, 307, 355; **25**, 27, 73, 95, 217, 265; **26**, 45, 77, 113, 157, 203, 227, 243, 317, 329; **27**, 1, 17, 71, 89, 219, 335; **28**, 1, 11, 41, 67, 75, 119, 145, 157, 167, 201, 219, 239, 249; **29**, 9, 37, 43, 53, 67, 73, 137, 169, 177, 237, 253; **30**, 17, 91, 111, 123, 151, 209; **31**, 55, 137, 165, 175, 193, 231, 261; **32**, 123, 199, 207, 223; **33**, 15, 45, 89, 105, 225, 253; **34**, 93, 105, 137, 145, 189, 207, 217, 241; **35**, 73, 91, 111, 129, 173, 193, 227, 243; **36**, 7, 41, 61, 67, 127, 147, 165, 191, 253; **37**, 27, 49, 87, 135, 145, 185, 251, 257

Drug conjugate **37**, 135, 145
Drug conjugation **52**, 55; **67**, 177, 207; **68**, 105; **68**, 61, 77, 191; **77**, 13; **83**, 233; **87**, 59, 255; **89**, 91; **128**, 261; **136**, 31; **137**, 33; **138**, 167
Drug contamination **24**, 239; **25**, 57; **29**, 233; **30**, 161, 237; **31**, 9, 65; **32**, 21; **51**, 129; **54**, 143; **55**, 175
Drug control **1**, 257, 315; **10**, 119; **11**, 237
Drug cytotoxicity **15**, 49; **22**, 75; **23**, 299; **29**, 169; **36**, 147; **37**, 135
Drug decomposition **1**, 173; **7**, 1, 119, 197; **8**, 45, 111, 121, 323; **11**, 271; **18**, 287; **22**, 327; **52**, 179; **57**, 139; **60**, 185; **62**, 65; **63**, 17, 155, 213; **75**, 211; **83**, 87; **84**, 279; **89**, 213; **113**, 199; **114**, 121; **120**, 235; **128**, 277; **130**, 251; **142**, 251
Drug degradation **1**, 95; **10**, 109, 165; **12**, 331; **19**, 207; **30**, 199; **31**, 1, 75, 125; **35**, 165; **41**, 41, 67, 105, 169, 227; **42**, 263; **45**, 181, 189, 207; **46**, 223, 231, 267; **51**, 27; **52**, 19, 71, 123, 191, 239; **56**, 103; **57**, 41, 217; **58**, 103, 143; **59**, 217; **60**, 99, 157; **62**, 27; **63**, R1, R5; **65**, 137; **66**, 253; **66**, 63, 71, 97, 289; **67**, R5; **68**, 87, 105; **69**, 197; **70**, 29, 95, 195; **71**, 65, 157; **72**, 89, 123; **73**, 23, 43, 81, 105, 147, 163; **74**, 67, 85, 203; **75**, 231, R5; **76**, 99, 113, 171, 261; **77**, 169; **78**, 1, 25, 59; **79**, 183, 243; **80**, 203; **87**, 79; **88**, 89, 177, 189, 229, 275, 397; **89**, 65, 169, 213, R5; **91**, 39; **92**, 63; **98**, 225; **99**, 165, 263, 351; **102**, 193; **104**, 247; **105**, 249; **112**, 97; **120**, 169; **121**, 95; **123**, 209, 217, 229; **124**, 129; **128**, 99; **129**, 95, 185; **135**, 81; **138**, 75; **139**, 113, 249; **140**, 219, 251; **141**, 39; **142**, 61; **143**, 163; **144**, 91, 123; **146**, 263; **147**, 31, 127
Drug delivery system **1**, 219; **2**, 41, 245; **6**, 19; **7**, 119, 129, 169; **8**, 291; **10**, 181; **13**, 23, 321; **14**, 123, 343; **15**, 177; **16**, 47, 191, 339; **17**, 203; **18**, 67, 79, 325; **19**, 107, 263; **20**, 119, 139, 187, 235; **22**, 137; **24**, 79, 267; **25**, 1, 265; **26**, 303; **27**, 145, 255; **28**, 67, 125; **29**, 147; **30**, 1, 17, 47, 73, 133, 151, 215; **31**, 15, 43, 65; **32**, 223; **33**, 71, 225; **34**, 57, 75, 125, 163; **35**, 21, 47, 121, 145, 243; **36**, 55, 81, 147, 253; **37**, 41, 119, 135, 145, 175, 239; **41**, 213; **42**, 19, 117, 243; **45**, 1, 217, 229; **47**, 129; **48**, 159; **49**, 15; **50**, 133; **51**, 1, 203; **53**, 195; **54**, 241; **56**, 217, 227; **57**, 17, 87, 133, 149, 155, 205; **58**, 49, 57, 115, 215; **59**, 1, 173, 197; **60**, 219; **61**, 35, 109, 127, 235, 241; **62**, 119; **63**, 43, 83, 121, 177, 207; **64**, 1, 187; **65**, 1, 49, 57, 159; **66**, 39, 79, 175, 223; **67**, 1, 21, 67, 97, 259, 265; **68**, 43, 87, 215; **69**, 63, 143, 155, 169; **70**, 53, 235; **71**, 105, 237; **72**, 19, 211; **73**, 111, 125, 171; **74**, 1; **75**, 45, 97, 117, 241; **76**, 77, 193, R5; **77**, 31, 81, 141, 151, 177, 221, 231; **78**, 165, 237; **79**, 11, 29, 191, 213, 223; **80**, 19, 63, 93, 145, 151, 161, R15; **81**, 31, 75, 103, 119; **82**, R1; **83**, 155, 177, 251; **84**, 23, 69, 141, 253; **85**, 129, 203, R1; **86**, 1, 35, 137; **87**, 21, 37, 67, 89, 141, 183; **88**, 1, 63, 201, 211, 251, 313, 319, 327, 335, 639, 391, 409, 423; **89**, 251, R1, R9, R13; **90**, 73, 95, 247, 255; **91**, 1, 29, 95, 209, 235; **92**, 71, 177, 225; **93**, 27, 49; **94**, 1, 115, 125; **95**, 1, 51, 77, 85, 105, 111, 153, 193, 219, 229; **96**, 147; **97**, 1, 133; **98**, 9, 19, 45, 113, 121, 149, 157, 235; **100**, 55, 199; **101**, 1, 23, 81, 191; **102**, 25, 47, 127, 141, 159; **103**, 37, 119, 131, 217, 237, 253, 283; **104**, 157; **105**, 65, 189; **106**, 15, 63, 69, 221, 255; **107**, 99, 149, 189, 205, 223; **108**, 225, 241; **109**, 35, 73, 135; **110**, 1, 93, 117, 127, 211, 285, 291; **111**, 31, 73, 103, 271, 295; **112**, 37, 47, 199, 207, 215, 241, 249; **113**, 25, 141, 175; **114**, 141, 151, 227; **115**, 9, 87; **116**, 1, 51, 131, 223; **117**, 243; **118**, 23, 113, 129, 199, 251; **119**, 81, 91, 103, 127, 203, 239, 251; **120**, 127, 247; **121**, 37, 45; **122**, 121; **123**, 33, 159, 173, 181; **124**, 37, 119, 149, 195, 261, 271; **125**, 17, 31, 73, 107, 283; **126**, 41, 119, 139, 235, 253; **127**, 1, 135, 165, 191; **128**, 9, 23, 65, 139, 145; **129**, 73, 175, 191; **130**, 1, 83, 169, 231; **131**, 57, 229; **132**, 9, 71, 81, 153, 175, 243, 267; **133**, 223; **134**, 89, 223, 239; **136**, 23, 43, 147; **137**, 67, 75, 95, 103, 261; **138**, 13, 45, 75, 121, 139, 175, 237; **139**, 1; **140**, 33; **141**, 9, 39, 109, 125, 227; **142**, 9, 25, 75, 115, 129, 227; **144**, 19, 177; **145**, 77; **146**, 71; **147**, 1, 173; **149**, 93
Drug derivative **3**, 1
Drug design **6**, 225; **14**, 123; **22**, 45; **52**, 231; **89**, 251; **125**, 117
Drug determination **1**, 185, 257; **2**, 317; **3**, 1, 93, 151; **4**, 111, 337; **5**, 85, 287; **6**, 183, 261; **7**, 55, 177, 343; **8**, 81, 121, 203; **9**, 245; **10**, 119, 177, 199; **11**, 257, 329; **13**, 89, 205, 219, 333; **15**, 37, 115, 149; **16**, 65; **17**, 1, 161, 325; **18**, 277; **19**, 199; **20**, 65, 285, 335; **21**, 265; **22**, 105, 345; **23**, 13, 277, 289; **24**, 27, 61, 101, 165, 173, 193, 219, 327; **25**, 85, 95, 113; **26**, 203, 227, 259; **27**, 45, 185, 205; **28**, 11, 23, 41, 47, 95, 113, 133, 193; **29**, 1, 161, 233; **30**, 29, 57; **31**, 9, 15, 25, 33, 43, 65, 75, 125, 157, 165, 231; **32**, 7, 21, 133, 171, 199, 243; **33**, 1, 55, 89, 99, 225, 235, 249; **34**, 153, 189, 213, 225, 259; **35**, 1, 21, 47, 157, 173, 181, 219; **36**, 41, 147; **37**, 11, 65, 87, 97, 135, 145, 163, 239, 257; **41**, 75, 245; **43**, 111; **45**, 245; **46**, 89, 199, 255; **49**, 79, 83, 135; **50**, 75, 175; **51**, 161, 169, 263; **52**, 11, 55, 129, 225; **53**, 91, 257; **54**, 89; **57**, 235; **58**, 103; **60**, 1, 11, 27, 41, 53, 185, 235; **61**, 43, 67, 85, 199, 261, R1; **63**, 173; **64**, 139, 235; **65**, 115, 137, 177, R1; **66**, 23, 53, 63, 71, 223, 289; **67**, 131, 185, 211, 259, R5; **69**, 1, 109, 155, 169, 181; **70**, 9, 95, 187, 195, 201; **71**, 1, 73, 187; **72**, 207; **73**, 1, 23, 33, 231; **74**, 33, 43, 67, 77, 127, 183, 215, 221; **75**, 161; **75**, 37, 55, 73, R5; **76**, 171; **77**, 41, 151, 163, 193, 211, 279, 303; **78**, 77, 85, 89; **80**, 263, 267; **81**, 143, 169, 193, 253; **82**, 179, 215; **84**, 279, R1; **85**, 163, 199; **86**, 99; **87**, 1, 223; **88**, 15, 141, 151, 177, 379; **89**, 51; **99**, 327; **106**, 167; **107**, 243; **108**, 11; **114**, 215; **119**, 109, 115; **121**, 123; **123**, 229; **134**, 231
Drug detoxification **22**, 327
Drug development **75**, 1; **82**, 9, 21, 47
Drug diffusion **17**, 331; **18**, 247; **20**, 107; **22**, 89; **35**, 91; **56**, 135; **57**, 253; **58**, 197; **60**, 83, 139; **63**, 281; **64**, 117; **65**, 133; **67**, 265; **68**, 43; **69**, 143; **71**, 95, 245; **72**, 1; **73**, 97, 249; **74**, 25; **75**, 63; **77**, 81, 163, 221, 255, 261; **78**, 123, 137; **80**, 191; **85**, 181; **86**, 9, 69; **88**, 99, 111, 125, 237, 293, 351; **89**, 111; **90**, 95, 161
Drug disposition **11**, 21; **16**, 203; **23**, 175; **26**, 243; **31**, 179; **53**, 219; **65**, 77; **66**, 169; **68**, 97; **73**, 131; **86**, 239; **108**, 241; **114**, 33; **119**, 91, 103; **142**, 129; **149**, 63
Drug dissociation **46**, 231
Drug dissolution **1**, 33; **2**, 113, 343; **3**, 127, 261, 175; **4**, 13, 39, 67, 159, 171; **5**, 149, 161; **6**, 137; **7**, 29, 129, 245, 327; **8**, 153; **9**, 97; **10**, 17, 89, 143; **11**, 181, 209; **12**, 1
Drug distribution **1**, 41, 213; **3**, 247; **6**, 179; **10**, 99; **16**, 339; **19**, 75; **22**, 137; **23**, 25; **24**, 79, 173; **25**, 191; **25**, 347; **26**, 113, 157, 243; **27**, 145; **28**, 1, 85; **29**, 53; **30**, 143; **31**, 179; **32**, 79, 103, 133; **34**, 145; **36**, 67, 127, 147, 253; **37**, 33, 145, 163, 219, 239; **43**, 267; **46**, 111, 179; **47**, 21, 111, 157, 265; **49**, 147, 249; **50**, 15, 97, 197; **51**, 85; **52**, 27, 87; **53**, 13; **54**, 41, 149; **55**, 39;

56, 269; **57**, 77; **61**, 85, 119; **62**, 1; **63**, 227; **64**, 109, 155, 171; **65**, 1; **66**, 53, 201, 263; **68**, 1, 61; **69**, 77; **70**, 19, 87, 271; **71**, 7; **72**, R5; **73**, 177; **74**, 59, 105, 203, 209; **77**, 297; **81**, 127; **84**, 273; **85**, 75; **88**, 159, 165, 335, 369; **89**, 91, 103; **90**, 119, 141, 171; **99**, 343; **108**, 85; **123**, 291; **124**, 19

Drug distribution volume **18**, 345

Drug DNA binding **72**, 251

Drug-dosage form **22**, 219; **27**, 185; **60**, R1; **63**, 77, 83, 113; **67**, 51, 79; **71**, 229, R5; **72**, 35, 141, 163; **73**, 177; **76**, 49, 55; **77**, 65, 71; **78**, 1, 9, 69, 217; **79**, 67, 83; **80**, 63, 145, 151, 227; **83**, 65; **84**, 59; **91**, 23, 75; **92**, 55, 167; **95**, 1, 29, 67; **98**, 37, 113; **101**, 63; **103**, 97, 243, 259; **104**, 253; **106**, 25; **109**, 115, 125, 135, 173; **110**, 189; **111**, 217; **113**, 231; **114**, 1, 13, 103, 125; **118**, 251; **119**, 1, 47, 71, 127, 165; **120**, 21, 33, 119, 189, 197; **121**, 37, 117, 129, 169; **122**, 9; **124**, 61, 97, 173; **125**, 17, 31, 99, 205, 215; **126**, 83, 147; **127**, 73, 115, 177; **128**, 73, 123; **129**, 13, 137; **129**, 73; **130**, 257; **131**, 137; **132**, 131, 189, 207; **133**, 29, 59, 107, 155, 365; **134**, 71, 129; **136**, 81, 117, 155; **136**, 81; **137**, 49; **138**, 13, 25, 67, 185; **139**, 45, 95, 223; **140**, 111, 131, 195; **143**, 93, 101; **148**, 1

Drug dose **1**, 197, 233; **3**, 335; **10**, 99; **13**, 239; **17**, 23; **20**, 99; **23**, 345; **31**, 83, 107, 179; **32**, 133; **34**, 35, 213; **37**, 19, 27, 127; **38**, 117

Drug efficacy **1**, 315; **13**, 183; **15**, 13, 235; **16**, 241, 271; **17**, 291, 299; **18**, 39, 299; **19**, 245; **20**, 73, 187; **21**, 51, 75, 341; **22**, 75, 137, 147, 291, 345; **23**, 87, 117, 209, 223, 247, 299; **24**, 61, 165, 307, 355; **25**, 73, 105, 121, 155, 199, 329; **26**, 45, 113, 133, 243, 267, 275; **27**, 29, 81, 117, 335; **28**, 1, 47, 201, 249; **29**, 223; **30**, 17, 73; **31**, 137, 165; **32**, 13, 39, 47, 133, 143, 165, 223; **33**, 37, 45, 253; **33**, 181; **34**, 35, 51, 111, 115, 241; **35**, 121, 139, 173, 181; **36**, 41, 121, 147, 191, 233; **37**, 19, 27, 33, 127, 135, 145, 163, 219, 245, 251; **46**, 141; **48**, 149; **49**, 205; **50**, 87; **51**, 175, 203; **61**, R1; **65**, 101; **68**, 111; **74**, 209; **75**, 45

Drug elimination **1**, 323; **2**, 173; **4**, 291; **5**, 117; **8**, 277, 323; **14**, 113; **15**, 261; **16**, 31, 285, 353; **18**, 79; **19**, 345; **23**, 25; **24**, 355; **25**, 155; **26**, 157; **27**, 117, 157; **28**, 133; **28**, 85; **30**, 143; **31**, 83, 169; **32**, 103, 133; **34**, 213; **35**, 157; **36**, 105, 127, 253; **37**, 145; **47**, 43; **52**, 159; **61**, 119; **64**, 109, 155; **68**, 11; **69**, 103; **81**, 127; **85**, 75, 175; **88**, 433

Drug encapsulation **22**, 137

Drug erythrocyte binding **42**, 225

Drug excretion **1**, 3; **6**, 205; **31**, 83; **47**, 89; **49**, 173, 249; **50**, 35; **52**, 27; **56**, 111; **60**, 27; **65**, 77

Drug-excipient interaction **40**, 129

Drug extravasation **144**, 11

Drug formulation **1**, 71, 95, 151, 165, 249, 315; **2**, 113; **3**, 81, 239, 247; **4**, 13, 39, 67; **5**, 67; **6**, 185, 195, 277;**7**, 1, 245; **8**, 89, 183, 217; **9**, 245; **11**, 277, 323, 329; **12**, 315, 341; **13**, 1, 9, 23, 89, 109, 243, 287, 313; **14**, 1, 59, 95, 123, 191, 209, 223, 251, 299, 313, 333, 319; **15**, 13, 73, 115, 167, 185, 235; **16**, 47, 65, 115, 163, 171, 349; **17**, 39, 225; **18**, 1, 25, 67, 127, 157, 207, 225, 269; **19**, 1, 27, 35, 63, 89, 139, 229, 263, 297; **20**, 139, 155, 187, 265, 307; **21**, 3, 35, 51, 187, 219, 239; **22**, 45, 105, 127, 131, 137, 299, 311; **23**, 35, 277, 289, 299, 333; **24**, 27, 61, 79, 91, 115, 165, 173, 275, 327, 339, 355; **25**, 57, 73, 85, 113, 121, 191, 199, 225; **26**, 45, 77, 99, 113, 165, 191, 243, 289, 303; **27**, 29, 81, 89, 117, 145, 157, 185, 233, 287, 311, 335; **28**, 11, 23, 47, 85, 113, 167, 193, 201, 239, 249; **29**, 1, 37, 53, 73, 83, 95, 113, 169, 195, 223, 233, 243, 267; **30**, 17, 95, 161; **31**, 9, 15, 33, 43, 65, 157, 169, 231, 239, 261, 271, 281; **32**, 7, 21, 71, 85, 99, 207, 223, 229, 237, 265; **33**, 1, 37, 45, 65, 71, 89, 99, 115, 137, 147, 201, 219, 225, 235, 249; **34**, 17, 105, 225, 259; **35**, 1, 7, 21, 61, 73, 81, 91, 121, 139, 145, 157, 187, 201, 243, 263; **36**, 55, 73, 99, 105, 147; **37**, 11, 15, 19, 97, 135, 145, 155, 175, 211; **38**, 255; **41**, 55, 67, 83, 91, 115, 139, 159, 197, 205, 213, 227, 241, 245, 255, 263; **43**, 179; **44**, 131; **45**, 229; **46**, 183, 241; **47**, 13, 51, 67, 89, 111, 133, 171, 179, 233, 263; **48**, 15, 119, 133, 217; **49**, 1, 29, 63, 69, 115, 147, 179; **51**, 19, 39; **52**, 1, 11, 37, 55, 71, 83, 91, 109, 149, 203, 249, 265; **53**, 99, 129, 145; **54**, 71, 103; **55**, 17, 105, 247, R1; **56**, 249, 261; **57**, 17, 155; **58**, 63; **59**, 69; **60**, 27, 35, 41, 89, 125, 197, 203, 229; **61**, 1, 27, 35, 51, 225, R1; **62**, 15, 27, 75, 81, 87, 207, R1; **63**, 65; **64**, 89, 99, 117, 147, 207, 223; **65**, 1, 19, 43, 57, 63, 69, 85, 101, 235; **66**, 29, 47, 53, 79, 111, 207, 213, 223, 263, 283, 289; **67**, 67, 97, 103, 113, 139, 163, 169, 185, 195, 201, 207, 211, 247, 265, 275, R5; **68**, 19, 25, 69, 87, 97, 111, 167, 299, 205, 255, 265, 271, R1; **69**, 5, 83, 93, 123, 129, 175, 181; **70**, 1, 9, 19, 35, 43, 53, 59, 69, 103, 119, 129, 153, 159, 241; **71**, 7, 81, 95, 127, 137, 147, 213; **72**, 79, 131; **73**, 17, 23, 51, 69, 75, 89, 197, 221, 239, 249, 259; **74**, 25, 49, 85, 95, 115, 127, 147, 169, 183, 203; **75**, 1, 25, 37, 45, 55, 63, 73, 89, R1, R5; **77**, 81, 89, 107, 119, 127, 141, 163, 169, 177, 183, 193, 199, 211, 221, 231, 239, 247, 255, 287, 315; **80**, R1; **81**, 103, 111, 187, 199, 205, 225, 235, 243, 253; **82**, 129; **84**, 93, 107, 117, 141, 151, 161, 167, 175, 191, 197, 217, 261, R5; **85**, 19, 29, 51, 57, 65, 97, 103, 113, 121, 129, 141, 175, 189, 203, 209, R1, R5; **86**, 9, 25, 35, 49, 79; **87**, 21, 31, 37, 89, 117, 133, 183, 229, 239, 247, 255; **88**, 1, 9, 23, 31, 39, 53, 75, 99, 159, 189, 261, 275, 313, 319, 327, 335, 345, 369, 407, 417, 423; **89**, 1, 19, 25, 33, 111, 119, 133, 139, 157, 177, 183, 207, 213, 223, 229, 235, 245, 251, R5, R9, R17; **90**, 89, 95, 105, 119, 129, 141, 151; **101**, 63, 191; **104**, 97, 107, 181; **105**, 1, 65, 181; **107**, 141, 189, 239, 247; **108**, 57, 117, 155; **109**, 135, 173; **111**, 31; **113**, 17, 57, 65, 93, 83, 89, 103, 123; **115**, 9, 17, 45, 53, 103, 193, 217, 225, 259;**116**, 31, 39, 45, 65, 87, 169, 253; **117**, 1, 23, 49, 197, 243; **118**, 47, 221; **120**, 127, 189, 197; **121**, 149; **122**, 91; **123**, 25, 199, 237; **124**, 27, 61, 183, 195, 261, 271; **127**, 115, 165, 229; **128**, 73, 163; **132**, 131, 267; **133**, 29, 155; **135**, 111, 137, 147, 151, 183; **137**, 133, 199, 249; **140**, 45, 119; **141**, 71, 227; **142**, 53; **143**, 133, 151, 163, 171, 179, 187, 195, 255; **146**, 271; **147**, 233; **148**, 117

Drug granulation **64**, 207; **69**, 43, 233, 239; **71**, 201; **72**, 141, 163, 243; **75**, 181; **76**, 239; **92**, 143; **95**, 135; **98**, 181; **110**, 101; **119**, 65; **130**, 147; **131**, 47; **132**, 221; **133**, 127, 133; **134**, 105, 243; **139**, 125, 139, 149; **140**, 207; **141**, 109, 161; **142**, 97; **145**, 203; **148**, 103

Drug granule **99**, 197, 337; **109**, 173; **110**, 101; **113**, 9; **119**, 65; **120**, 101; **129**, 1; **130**, 149; **133**, 149; **138**, 215; **141**, 161

Drug half life **12**, 331; **14**, 113; **15**, 61, 261; **18**, 79; **22**, 345; **23**, 25, 209; **25**, 27; **28**, 85, 133; **30**, 111, 143, 199; **32**, 103, 133; **36**, 127; **37**, 97, 163, 219; **46**, 203; **48**, 223; **50**, 35; **52**, 159; **54**, 57, 65; **57**, 125; **61**, 119; **64**, 109, 155; **71**, 45, 73, 117; **77**, 297; **88**, 391; **99**, 343

Drug hydrolysis **37**, 257; **42**, 51, 217, 263; **45**, 207; **52**, 55, 71, 239, 255; **53**, 189; **54**, 57, 65; **61**, 43, 67; **63**, 29, 129; **64**, R1; **66**, 39, 97, 149, 253; **68**, 77; **70**, 137; **71**, 31, 45, 105, 117; **73**, 33, 105, 221; **74**, 67; **75**, 211, 231, 259; **75**, 231, 259; **77**, 65; **79**, 183; **82**, 205; **84**, 175; **86**, 43; **87**, 59; **88**, 221, 229, 237, 397; **89**, 65; **90**, 239; **104**, 29; **107**, 67; **126**, 209; **135**, 177
Drug identification **3**, 109, 205; **11**, 237, 329; **12**, 355; **13**, 47, 89, 109; **14**, 113; **15**, 149; **18**, 53; **20**, 155, 273; **21**, 99, 251, 349; **22**, 45, 75, 197, 345; **23**, 117, 163, 209, 223, 265; **24**, 193, 219, 275, 307; **25**, 27, 293, 313; **26**, 227, 317; **27**, 175, 219; **28**, 1, 95, 119, 157; **29**, 9, 19, 67, 73, 95, 233; **31**, 125; **32**, 21, 123, 151; **33**, 105, 115; **34**, 81, 247; **35**, 39, 61, 73, 165, 243, 259; **36**, 7, 147; **37**, 65, 135, 145, 185; **46**, 25, 179; **49**, 83; **63**, 29; **84**, 279
Drug impurity **53**, 257; **127**, 65; **140**, 169
Drug incompatibility **18**, 287; **118**, 181; **119**, 71; **127**, 293; **128**, 283; **130**, 251; **138**, 125; **139**, 243
Drug induced disease **1**, 183
Drug industry **1**, 249; **39**, 1; **44**, 169; **103**, 9
Drug interaction **1**, 281; **2**, 195, 257; **4**, 263; **5**, 167; **8**, 277; **11**, 309; **13**, 9, 67, 353; **14**, 243; **15**, 347; **16**, 23, 215, 271, 305, 345; **17**, 161, 291; **18**, 9, 235, 247, 287; **19**, 17, 233, 307; **20**, 99, 155; **21**, 75, 99, 277; **22**, 257, 283, 291, 345; **23**, 163; **24**, 91, 231, 239, 275, 297, 339; **25**, 41, 57, 225, 359; **26**, 77, 133, 267; **27**, 1, 17, 39, 117, 311; **28**, 95, 119, 133, 201, 239; **29**, 29, 103, 137, 147, 161, 169, 177, 237; **30**, 17, 29, 35, 101, 143, 225; **31**, 25, 33, 55, 107, 137, 157, 185, 193; **32**, 39, 207; **33**, 1, 55, 105, 125, 253; **34**, 93, 137, 189, 207, 219; **35**, 73, 111, 129, 193; **36**, 121, 185, 233; **37**, 113, 167, 175, 227; **47**, 21, 209; **48**, 113; **50**, 35, 45, 261; **56**, 269; **57**, 173; **59**, R1; **64**, 155; **65**, 63; **69**, 221; **72**, 175; **76**, 187; **79**, 61; **87**, 73; **88**, 31, 433; **89**, 75; **98**, 51, 231; **106**, 229; **109**, 107; **124**, 205; **131**, 67, 153; **146**, 181
Drug isolation **15**, 149; **18**, 53; **34**, 225; **37**, 65; **63**, 29; **76**, 199; **79**, 61
Drug labeling **80**, R1
Drug lymph level **8**, 253; **24**, 79, 173; **28**, 193; **33**, 99; **36**, 147; **37**, 145; **73**, 177; **80**, 51; **119**, 173
Drug manufacture **144**, 131; **147**, 23, 79, 95
Drug mechanism **1**, 15, 127; **8**, 323; **16**, 127; **19**, 169; **24**, 297; **28**, 249; **29**, 29, 43, 169; **31**, 131, 231; **33**, 253; **34**, 35, 51; **36**, 253; **37**, 127, 251; **40**, 235; **41**, 95, 105; **45**, 153, 169; **46**, 203; **49**, 199; **50**, 27, 91; **51**, 241, 259; **68**, 1; **85**, 181; **87**, 79; **88**, 391; **104**, 29
Drug metabolism **1**, 95; **2**, 127, 215; **3**, 195, 341; **5**, 67; **7**, 179; **12**, 277, 331; **13**, 273; **14**, 113, 133, 173; **15**, 235, 309; **17**, 299; **18**, 185; **19**, 1; **20**, 43, 73, 265; **22**, 105, 345; **23**, 117, 209; **24**, 43, 61, 101, 307; **25**, 155; **26**, 15, 227, 317; **26**, 227, 317; **27**, 71, 325; **28**, 1, 47, 133; **29**, 9, 137; **30**, 111, 119, 247; **31**, 165; **32**, 199; **33**, 15, 55; **35**, 47, 73, 219; **36**, 41, 147; **37**, 87, 163, 219; **41**, 121; **46**, 111; **47**, 43; **52**, 27, 155; **53**, 249; **54**, 41; **55**, 1; **62**, 9; **64**, 7, R1; **65**, 77, 109, 265; **66**, 183; **71**, 175; **74**, 67; **76**, 177; **77**, 265; **81**, 253; **87**, 59; **88**, 391, 397, 423; **105**, 7; **116**, 231; **117**, 95; **127**, 213; **128**, 229
Drug metabolite **1**, 173; **3**, 109, 205; **8**, 121; **12**, 331; **14**, 173; **35**, 219; **46**, 111; **49**, 79, 83; **55**, 67; **71**, 175; **76**, 71, 171; **77**, 265; **82**, 1; **84**, 253; **87**, 59; **88**, 177; **89**, 65; **91**, R1; **95**, 127; **97**, 1; **98**, 231; **103**, 147; **104**, 175; **109**, 181; **112**, 199; **119**, 109; **121**, 1, 55; **124**, 119; **131**, 263; **134**, 157; **145**, 145; **146**, 181
Drug mixture **14**, 231; **17**, 39; **21**, 51, 265; **22**, 105; **27**, 39, 127, 245, 311, 335; **29**, 253; **30**, 225; **31**, 25, 179, 193, 239; **32**, 7, 187, 207; **34**, 207; **37**, 113; **49**, 129; **50**, 75, 213; **51**, 169, 263; **53**, 47, 75; **55**, 135; **63**, 173; **64**, 235; **65**, 177; **66**, 47; **70**, 187; **71**, 187; **80**, 171; **86**, 187, 211, 221; **103**, 225; **139**, 149
Drug monitoring **18**, 353; **19**, 169; **21**, 341; **24**, 61, 79, 297; **25**, 191; **26**, 113, 203, 259, 303; **27**, 45, 71, 81, 145; **28**, 95, 193, 201, 219, 249; **29**, 43, 53, 169; **30**, 29, 57, 123; **31**, 15; **33**, 89, 249; **34**, 29; **35**, 91, 157; **37**, 145; **43**, 145; **47**, 265; **48**, 79, 223, 247; **50**, 53, 155; **51**, 183; **54**, 95; **55**, 105
Drug particle **37**, 239; **59**, 79; **63**, 177
Drug partitioning **17**, 183
Drug penetration **27**, 361; **33**, 225; **44**, 15, 87, 269; **52**, 47; **53**, 13, 43, 61, 119; **57**, 29, 77; **58**, 93; **60**, 139, 177; **61**, 205; **63**, 49; **68**, 127, 151, 285; **74**, 1, 59, 95, 89, 157, 221, 229; **75**, 89; **76**, 37, 77; **77**, 1, 221, 231; **84**, 93, 117, 129, 253; **85**, 7, 129, 181; **86**, 69; **87**, 67, 125, 175, 261; **88**, 89, 111, 125, 165, 237, 293; **89**, 41, 81, 147, 191, 199, R1, R13; **90**, 161, 171; **107**, 231; **124**, 53; **135**, 91, 127; **141**, 171; **145**, 187
Drug polymer interaction **2**, 295
Drug potentiation **25**, 245; **28**, 151, 201; **29**, 29; **31**, 239; **33**, 45, 89, 99, 225; **35**, 227
Drug prescription **32**, 47
Drug production **47**, 171, 179; **49**, 1; **55**, 135, 143, 199; **63**, 57; **99**, 99, 239; **106**, 77; **112**, 165; **125**, 315; **134**, 105; **134**, 247
Drug protein binding **2**, 283; **3**, 65; **6**, 205; **7**, 111; **11**, 11, 45; **15**, 273; **16**, 215; **23**, 109, 209; **25**, 147; **27**, 45; **30**, 123, 189; **35**, 211; **36**, 113; **37**, 103; **42**, 225; **47**, 163; **49**, 103; **50**, 97; **51**, 253; **54**, 1, 123; **55**, 221; **56**, 235; **59**, 137; **62**, 21; **65**, 109; **65**, 149; **67**, 177; **68**, 61, 191; **75**, 45; **87**, 141; **108**, 241; **114**, 177; **118**, 85; **119**, 57; **128**, 223; **134**, 15
Drug purification **27**, 17; **29**, 73; **30**, 237; **34**, 183; **76**, 199
Drug purity **20**, 217
Drug quality **53**, 53; **54**, 143; **74**, 77
Drug receptor **2**, 127
Drug receptor binding **15**, 223; **54**, 259; **63**, 135; **85**, 75
Drug release **1**, 151, 197; **2**, 91, 113, 195; **6**, 29, 185, 195; **10**, 25; **11**, 323; **12**, 251; **13**, 89, 145, 159, 171; **14**, 95, 191; **15**, 115, 167, 177; **16**, 79; **17**, 91, 99, 225, 273; **18**, 127; **19**, 177; **20**, 335, 347; **21**, 119; **22**, 75, 131; **24**, 43, 275, 327, 339, 355; **25**, 1, 121, 199; **26**, 45, 57, 203; **27**, 29, 145, 157, 205, 233; **28**, 67, 113, 239; **29**, 1, 37, 83, 147, 195, 243, 267; **30**, 1, 9, 17; **31**, 15, 43, 169; **32**, 21, 71, 85, 99, 143, 223, 243; **33**, 65, 71, 137, 147; **34**, 57, 93, 137, 169, 259; **35**, 1, 7, 21, 39, 121, 145, 201; **36**, 147; **37**, 11, 41, 87, 119, 135, 167, 179, 211, 245; **39**, 267; **40**, 111, 223; **41**, 115, 245; **42**, 15, 161; **43**, 167; **44**, 131, 159, 225; **45**, 111, 145, 217, 229; **46**, 49, 69, 133; **47**, 37, 95, 133, 163, 233, 249; **48**, 1, 15, 51, 125, 217; **49**, 29, 63, 109, 183, 231; **50**, 21, 39, 133, 155, 181, 205; **51**, 1, 103, 213, 245, 253; **52**, 37, 83, 109, 139, 191, 203, 249; **53**, 53; **54**, 15, 71, 161; **55**, 25, 31, 47, 53, 143, 147, 157, 199; **56**, 65, 71, 197, 243, 261; **57**, 107, 139, 149, 155, 163, 169, 197, 241, 263; **58**, 1, 19, 31, 49, 63, 79, 89, 197, 215; **59**, 1, 9, 79, 165; **60**,

83, 109, 151, 171, 219, 229, R1; **61**, 1, 127, 235; **62**, 9, 15, 53, 81, 105, 113, 119, 125, 157, 189, 249; **63**, 43, 83, 143, R9; **64**, 171; **65**, 19, 57, 69, 85, 133; **66**, 29, 111, 213; **67**, 21, 67, 79, 177, 265; **68**, 87, 97, 111, 215, 255, 265, R1; **69**, 5, 63, 69, 123; **70**, 59, 69, 129, 153; **71**, 7, 95, 201, 213, 245; **72**, 19, 35, 141, 163, 199; **73**, 17, 51, 89, 125, 221, 239; **74**, 25, 49, 169, 183; **75**, 25, 181, 219; **76**, 55, 77, 161, 193, R5; **77**, 87, 177, 297; **78**, 1, 237; **79**, 123, 223, R1; **80**, 161, R15; **81**, 103, 193, 235, 263; **82**, 53, 61, 59, 95; **84**, 33, 85, 141, 151, 167, 175, 217; **85**, 29, 51, 57, 65, 103, 113, 141, 189, 203, R1, R5; **86**, 9, 35, 79, 113, 131, 137, 147, 159; **87**, 21, 31, 89, 117, 141, 183, 215, 229, 255; **88**, 23, 31, 39, 75, 99, 165, 201, 391, 405, 409; **89**, 9, 19, 91, 111, 133, 177, 229, 235, 245, 251, R13; **90**, 1, 43, 89, 151; **91**, 95; **92**, 177; **95**, 219; **98**, 219, 235; **99**, 135, 145, 173, 229, 247, 263, R5; **103**, 81, 217, 243, 267; **104**, 107, 181; **105**, 241; **106**, 1, 15; **107**, 41, 179; **108**, 77; **109**, 271; **110**, 285, 291; **111**, 251, 271, 283; **112**, 37, 47, 105, 117; **113**, 39; **113**, 231; **114**, 221, 257; **116**, 51, 125; **117**, 147, 151; **118**, 103, 151, 237, 251, 257; **119**, 11, 165, 203, 231; **120**, 63, 119; **121**, 113, 117, 141, 149, 239; **122**, 9; **123**, 85, 119; **124**, 9, 107, 149, 219, 271; **125**, 7, 151, 215, 223, 231, 243, 289, 297; **126**, 79, 83, 147, 169; **127**, 177; **128**, 261; **129**, 13, 175, 233, 243, 263, 279; **130**, 25, 49, 83, 115; **131**, 9, 47, 73, 97, 101, 183, 191, 247; **132**, 1, 153, 243, 259; **133**, 1, 59, 65, 89, 97, 149, 215, 265; **134**, 37, 71, 203, 239; **135**, 73; **136**, 23, 31, 89, 155; **137**, 49, 57, 95, 159, 225; **138**, 45, 75, 121, 215, 225; **139**, 1, 45, 53, 79, 87, 187;**140**, 85, 145; **141**, 9, 27, 53, 109; **142**, 153, 163; **143**, 13, 25, 107, 143, 255; **144**, 37, 159, 209; **145**, 37, 49, 61, 77; **146**, 123, 175, 193; **147**, 1, 11, 23, 71, 135; **148**, 23, 55, 85, 177

Drug research **87**, 11

Drug residue **131**, 171

Drug resistance **12**, 295; **17**, 291; **19**, 239; **34**, 115

Drug safety **29**, 233; **31**, 193; **37**, 167; **75**, R1

Drug saliva level **15**, 167; **31**, 15; **34**, 207; **54**, 199; **68**, 11; **82**, 233

Drug screening **1**, 141; **3**, 13; **9**, 7; **14**, 113; **22**, 345; **23**, 223; **26**, 317; **27**, 29; **28**, 1, 75, 249; **29**, 253; **31**, 137; **32**, 187; **33**, 71; **34**, 225; **36**, 29, 61, 147, 243; **37**, 135

Drug sensitivity **25**, 105; **27**, 163; **32**, 55, 91; **34**, 115; **35**, 227; **36**, 191; **46**, 45; **49**, 29; **55**, 209; **141**, 217

Drug solubility **11**, 27, 81; **12**, 1, 251, 341; **13**, 1, 23, 67, 145, 159, 171; **14**, 59, 79, 113, 251, 263, 291, 299, 313, 333; **15**, 25, 73, 87, 97, 159, 235, 251; **16**, 65, 97, 171, 295, 327; **17**, 1, 77, 115, 193; **18**, 25, 47, 89, 127, 149, 207, 213, 225, 247, 311; **19**, 17, 35, 89, 149, 297, 307; **20**, 25, 99, 273; **21**, 61, 123, 155, 187, 251, 349; **22**, 283; **23**, 35, 97, 117, 195, 209, 223, 277, 289; **24**, 1, 43, 61, 79, 165, 173, 231, 275, 307; **25**, 27, 85, 95, 135, 199, 265, 359; **26**, 77, 123, 165, 203, 215, 243, 317; **27**, 1, 39, 71, 89, 117, 219; **28**, 1, 11, 23, 33, 41, 67, 75, 95, 157, 183, 201, 219; **29**, 19, 73, 95, 103, 113, 161, 177, 223; **30**, 9, 101, 111; **31**, 25, 45, 69, 157, 175, 225; **32**, 21, 187, 223, 229, 265; **33**, 15, 115, 125, 219, 225, 235; **34**, 125, 189, 217, 241; **35**, 7, 73, 91, 243; **36**, 7, 41, 61, 67, 73, 105, 211; **37**, 103, 179, 251; **38**, 257; **40**, 223; **41**, 197, 205; **42**, 1, 35, 135; **43**, 67, 119; **45**, 1, 27, 129, 139, 153, 157, 259; **46**, 77, 95, 133, 217, 247; **47**, 103, 223; **48**, 83, 207; **49**, 163, 189, 223; **50**, 7, 103, 111, 117, 222; **51**, 9, 39, 195, 213, 225; **52**, 11, 19, 63, 109, 129; **53**, 7, 99; **54**, 175; **55**, 77, 123; **56**, 151; **57**, 63, 149, 229; **58**, 177, 221, R1; **59**, 271, 281; **60**, 1, 11, 41, 53, 61, 163, 191, 226, 243; **61**, 1, 43, 173; **62**, 15, 37; **63**, 167, 237; **64**, 147; **65**, 109, 115, 141; **66**, 29, 39, 71, 111, 149, 175, 193, 213, 233, 289; **67**, 11, 79, 131, 139, 201, 207, 211, 247; **68**, 1, 19, 25, 35, 43, 167, 179, 255, 271, R1; **69**, 93, 129, 175, 265; **70**, 9, 43, 59, 69, 137, 153, 191, 219, 261; **71**, 19, 105, 117, 201; **72**, 29; **73**, 1, 9, 17, 23, 33; **74**, 77, 85, 105, 169; **75**, 63, 141, 147, 193, 201, 259, R13; **77**, 89, 211, 239, 247, 255, 261; **78**, 17, 189; **79**, 273; **81**, 263; **82**, R7; **84**, 107, 167, 197, 233; **85**, 1, 45, 57, 149, 175, 203; **86**, 79; **87**, 47, 95, 161, 203; **88**, 31, 39, 53, 151, 261, 345, 437; **89**, 41, 119, 207, 229; **90**, 141; **99**, 157; **105**, 137; **108**, 149; **112**, 1; **113**, 103; **121**, 157; **122**, 57; **125**, 55; **126**, 73, 95, 199; **127**, 27, 293; **128**, 29, 45; **129**, 13, 63, 283; **130**, 75, 179; **131**, 41; **133**, 17, 191; **134**, 59; **135**, 161; **136**, 165; **137**, 167, 209; **139**, 33, 215; **140**, 25, 131, 247; **141**, 53, 171, 197; **142**, 183, 189; **144**, 127, 153, 213; **146**, 213; **147**, 131; **148**, 1, 123, 211; **149**, 1

Drug solution **3**, 319; **7**, 197; **8**, 45; **35**, 1; **80**, 267; **87**, 229; **107**, 85; **114**, 125; **119**, 139, 183; **122**, 27; **124**, 195; **147**, 135

Drug solvent **148**, 1

Drug stability **3**, 109; **4**, 1, 21, 75; **5**, 67, 161, 345; **7**, 197; **9**, 1, 29, 59; **10**, 29; **11**, 71, 257, 277, 329; **12**, 331, 355; **13**, 9, 145, 159, 171, 219, 243, 253, 303, 313, 333; **14**, 73, 113, 209; **15**, 61, 73, 149, 251, 293; **16**, 93, 115, 141, 319; **17**, 29, 91, 313; **18**, 79, 89, 139, 213, 287, 325; **19**, 1, 199, 207; **20**, 25, 155, 265, 273, 307; **21**, 35, 201, 211, 251, 349; **22**, 45, 105, 127, 197, 299; **23**, 1, 117, 209, 223, 299; **24**, 27, 91, 173, 193, 219, 355; **25**, 225, 265, 293, 313; **26**, 15, 77, 123, 227, 243, 259, 289, 303, 317; **27**, 1, 17, 81, 157, 175, 185, 205, 219, 311; **28**, 1, 47, 75, 119; **29**, 9, 19, 67, 103, 137, 147, 161; **30**, 29, 111, 161, 199, 229; **31**, 1, 9, 25, 43, 65, 75, 125, 169, 239, 281; **32**, 21, 111, 123, 151, 207; **33**, 1, 15, 71, 201; **34**, 17, 153, 225, 247; **35**, 7, 21, 39, 47, 61, 73, 139, 165, 193, 243, 359; **36**, 7, 29, 81, 99, 205, 147; **37**, 49, 87, 167, 179, 185, 245, 257; **38**, 147, 161; **41**, 41, 75, 95, 105; **42**, 27, 41; **43**, 23, 167, 191, 215; **43**, 161; **44**, 31, 117, 151, 205, 225, 235; **45**, 27, 47, 79, 129, 165, 181, 237; **46**, 49, 57, 77, 159, 223; **47**, 103, 163, 195, 215, 239, 261; **48**, 119, 159, 179, 195; **49**, 79, 83, 91, 115, 157, 163; **50**, 1, 71, 87, 189, 219, 233; **51**, 157, 189; **52**, 11, 71, 101, 123, 191, 239; **53**, 25, 181, 237; **54**, 65, 181, 211; **55**, 47, 59, 67, 91, 123, 143, 185, R1; **56**, 37, 103, 125, 159, 175; **57**, 63, 87, 217; **58**, 57, 103, 143, R1; **59**, 69, 121, 131, 217, 271; **60**, 1, 11, 27, 41, 53, 61, 99, 157, 185, 253; **61**, 9, 67; **62**, 65, 75, 81, 87, 125, 165, 193, 207, 243, R1; **63**, 17, 35, 65, 113, 213, R1; **64**, 67, 139, 195, R1; **65**, 49, 137, 169, 225, R9; **66**, 1, 71, 253; **67**, 103, 169, 177, 195, R5; **68**, 105, 121, 271, 277, 281; **69**, 1, 197; **70**, 29, 95, 137, 251; **71**, 19, 73, 117, 137, 157; **72**, 11, 57, 123; **73**, 23, 33, 51, 69, 81, 105, 147, 209; **74**, 67, 85, 137; **75**, 45, 97, 147, 171, 211, 231, 259, R5; **76**, 99, 113, 225; **77**, 21, 57, 65, 119, 169, 247, 269, 279, 309, 315; **78**, 199; **79**, 107, 273, 289; **80**, 93, 151, 227, R7; **81**, 153, 179, 253, 263; **82**, 29, 79, 91; **83**, 263; **84**, 175, 233, 279; **85**, 87; **86**, 25, 231; **87**, 133, 167, 203; **88**, 1, 177, 189, 201, 251, 275, 335, 397, 423; **89**, 125, 139, 207, R5; **90**, 19; **92**, 47; **98**, 149; **99**, 85, 229, 333; **101**, 165, 257; **102**, 201; **105**, 83, 89;

106, 7, 135; **107**, 1, 111, 141, 199, 243, 247; **110**, 241; **112**, 93, 133, 143, 181; **113**, 25, 57, 199, 215; **115**, 161, 167; **121**, 95, 157, 217; **122**, 17; **123**, 95, 143; **125**, 55, 315; **126**, 29, 103; **127**, 255, 273; **128**, 89, 283; **129**, 147, 191; **130**, 137; **131**, 19, 247; **132**, 33, 85, 107, 115, 191, 257; **135**, 81, 103; **136**, 81; **137**, 209; **138**, 1, 13, 37, 125, 167; **139**, 219, 243; **141**, 125; **142**, 85, 125; **143**, 1, 93; **144**, 153, 225; **145**, 95; **146**, 213; **147**, 153; **148**, 47, 165

Drug sterility **24**, 307; **142**, 125; **148**, 47; **149**, 85

Drug storage **2**, 265, 295; **5**, 345; **30**, 229; **39**, 51; **42**, 41; **67**, 169; **68**, 281; **68**, 277; **70**, 119; **71**, 19, 137; **72**, 57; **73**, 51, 209, 249; **74**, 175; **79**, 61; **86**, 89; **88**, 351; **89**, 125, 139; **90**, 105; **127**, 65; **131**, 247, 257; **132**, 1; **135**, 53

Drug structure **1**, 281; **2**, 81; **3**, 1; **4**, 39; **8**, 211; **13**, 145, 349; **14**, 159, 325; **15**, 49; **16**, 93; **17**, 77, 161; **19**, 149; **21**, 277; **23**, 13, 55, 333; **24**, 1, 239; **25**, 41, 57, 121, 225; **26**, 25, 123, 215; **27**, 335; **28**, 11, 125, 183; **29**, 95; **30**, 161; **31**, 25, 43, 99, 131, 157; **33**, 115, 219; **34**, 23, 75, 105, 111, 137, 183; **35**, 81, 111, 139, 145, 181; **36**, 165; **37**, 175, 219, 251; **43**, 241, 249; **47**, 79, 95, 141, 171; **49**, 261; **50**, 103; **52**, 173; **53**, 25; **55**, 257; **59**, 217; **61**, 157; **65**, 225; **73**, 1; **87**, 1, 79; **88**, 379; **99**, 125, 303; **110**, 75; **118**, 11; **119**, 25; **124**, 107; **141**, 257

Drug synthesis **1**, 183, 189; **3**, 73, 81; **8**, 35; **10**, 29, 307; **12**, 113; **14**, 113, 123; **15**, 49, 61, 235; **16**, 65; **17**, 313; **21**, 51, 251, 277, 349; **22**, 75, 345; **23**, 13, 117, 209, 223, 333; **24**, 307; **25**, 27; **26**, 45, 227, 317; **27**, 17; **27**, 205; **28**, 1, 125; **29**, 9, 19, 73; **30**, 209, 215, 237; **32**, 251; **33**, 15, 71; **34**, 183; **35**, 47, 243; **36**, 7, 81, 147; **37**, 87, 135, 145, 185; **38**, 179, 199; **45**, 229; **47**, 103, 195; **48**, 91, 195; **51**, 27; **52**, 47, 255; **53**, 195; **54**, 57; **57**, 41, 253; **60**, 163; **61**, 43; **62**, 193; **68**, 77, 87, 191, 215; **70**, 137; **73**, 259; **74**, 67; **75**, 131, 249; **77**, 13, 21, 265; **79**, 233; **81**, 253, 263, 267; **84**, 175; **85**, 189; **89**, 51, 157; **92**, 105; **101**, 145; **105**, 57; **121**, 211; **123**, 65; **141**, 125; **146**, 247

Drug targeting **26**, 113; **30**, 95; **31**, 185; **40**, 201, 207; **42**, 69, 145; **47**, 129; **56**, 217; **57**, 241; **59**, 27, 57; **63**, 121, 177; **72**, 211; **90**, 119; **105**, 19; **110**, 277; **128**, 65; **131**, 73; **138**, 85; **139**, 1; **145**, 157

Drug tissue level **11**, 323; **16**, 105; **17**, 299; **25**, 347; **26**, 113, 203; **27**, 45, 89; **28**, 47; **29**, 43, 53; **30**, 57; **34**, 145; **35**, 47, 219; **36**, 147; **37**, 11, 219; **59**, 57; **61**, 119; **63**, 227; **67**, 67; **68**, 61, 127; **70**, 271; **89**, 91

Drug tolerability **137**, 249

Drug tolerance **64**, 187

Drug toxicity **12**, 57, 67; **14**, 113; **15**, 49, 235; **16**, 127; **22**, 75, 137; **28**, 1; **30**, 215; **31**, 275; **32**, 251; **34**, 17; **58**, 41

Drug transformation **118**, 161; **120**, 235

Drug transport **12**, 87, 97; **18**, 259, 311; **25**, 85; **29**, 177; **30**, 63; **33**, 165; **34**, 175; **37**, 103,251; **43**, 233; **48**, 141, 231; **49**, 15, 39, 57, 199; **50**, 27, 197; **52**, 27, 79; **53**, 13; **55**, 265; **56**, 75, 87; **59**, 263; **62**, 53; **63**, 281; **65**, 115; **66**, 193, 243; **68**, 43, 231, 285; **69**, 103, 109, 143, 193; **72**, 219; **74**, 9, 189; **76**, 1, 25, R1; **77**, 107; **80**, 39; **85**, 129; **87**, 215; **88**, 111, 125, 313; **89**, 147; **90**, 119, 129, 161, 171; **108**, 85; **114**, 151; **119**, 173; **120**, 229; **125**, 1; **126**, 179; **127**, 103; **139**, 25; **144**, 61; **145**, 115, 145, 175; **147**, 61

Drug uptake **28**, 201; **53**, 67; **61**, 85; **62**, 133; **65**, 7; **66**, 201; **74**, 221; **82**, 21, 99; **84**, 245; **85**, 75; **110**, 29

Drug urine level **6**, 301; **13**, 99; **14**, 173; **15**, 309; **18**, 1; **19**, 233, 297; **22**, 345; **24**, 101, 355; **27**, 45, 117, 157; **28**, 167; **31**, 83, 169; **32**, 133; **33**, 55; **34**, 29, 189; **35**, 157; **36**, 105; **37**, 145, 219, 227; **53**, 181; **60**, 27; **68**, 61; **70**, 191; **75**, 37

Drug vehicle **1**, 71; **2**, 145; **3**, 39; **9**, 107; **10**, 165; **16**, 115, 163; **22**, 131; **26**, 165; **31**, 261; **33**, 165; **35**, 121; **37**, 167; **41**, 49; **54**, 223; **57**, 29; **58**, 197, 209; **60**, 139; **63**, 201, 275, 275; **69**, 43; **72**, 131; **79**, 257; **80**, 119, 179, 271; **81**, 59; **83**, 1, 241; **86**, 187; **90**, 213; **92**, 219, 243; **95**, 117; **105**, 189, 227; **109**, 135; **110**, 137; **111**, 235; **113**, 189; **119**, 173; **120**, 95, 101; **121**, 27, 89; **122**, 49; **124**, 149; **125**, 231; **126**, 287; **130**, 57; **131**, 137; **132**, 71; **136**, 53; **139**, 161; **141**, 71; **142**, 1; **147**, 165; **148**, 1

Dry deposition **135**, 53

Dry powder **149**, 51

Dry skin **105**, 103; **113**, 247

Drying system **51**, 19

DT0 **35**, 47

Duodenum **21**, 233; **102**, 177

Durene **15**, 347

Dye **48**, 63, 113; **83**, 263; **99**, 173; **103**, 97, 283; **119**, 81

Dyflos **107**, 133; **112**, 133

Dymel-11 **110**, 9

Dynacel **111**, 43

Dynasan-112 **129**, 159

Dynasan-114 **129**, 159

Dynasan-116 **129**, 159

Dynasan-118 **129**, 159

Dynaspirin **108**, 11

E

Eastman cap **109**, 9

Eastman cat **109**, 9

Ebifuramin **52**, 71

Echinococcosis **100**, 271

Echogum T **134**, 173

Econazole **90**, 247; **105**, 77; **110**, 169

Econazole nitrate **10**, 119

Edema **28**, 47

Edetate disodium **33**, 89

Edetate sodium **33**, 89; **121**, 217; **139**, 63

Edetic acid **9**, 49; **19**, 239; **23**, 357; **28**, 249; **35**, 227; **36**, 191; **56**, 169; **66**, 131; **68**, 135; **87**, 195; **90**, 229; **93**, 91; **121**, 95; **142**, 103

Efrotomycin **94**, 1

Egtazic acid **79**, 159; **123**, 13

Ehrlich ascites tumor **1**, 41; **83**, 131

Ehrlich ascites tumor cell **1**, 41; **22**, 75

Elaidic acid **98**, 203

Elastase **95**, 171; **97**, 171; **104**, 151; **117**, 95; **125**, 117

Elastase inhibitor **125**, 117

Elasticity **57**, R1; **73**, 75; **137**, 243; **141**, 101

Elastin **125**, 117

Elastolysis **125**, 117

Elastomer **87**, 117

Elcema **91**, 183; **97**, 15

Electric current **96**, 51; **105**, 47

Electric field **60**, 133; **125**, 123

Electric potential **64**, 67; **87**, 167
Electric resistance **85**, 129
Electricity **65**, 7; **137**, 261
Electrochemical detection **35**, 219
Electrochemistry **55**, 237
Electrolyte **1**, 141; **26**, 25; **27**, 17; **58**, 13, 229; **64**, 67; **67**, 155; **70**, 251; **81**, 1; **85**, 57; **87**, 167; **100**, 189; **101**, 175; **107**, 121; **109**, 191
Electron microscopy **1**, 315; **13**, 23; **23**, 333; **34**, 173
Electron spin resonance **104**, 61; **142**, 247
Electrophoresis **7**, 285; **13**, 9; **71**, 223; **92**, 225
Electrophysiology **63**, 1; **68**, 135
Embonic acid **24**, 355
Emdex **30**, 237; **48**, 149; **112**, 55; **114**, 23
Emepronium bromide **34**, 75
Emulsifying agent **23**, 69; **25**, 237; **84**, 107, R5; **88**, 53; **106**, 15; **114**, 103; **125**, 73; **146**, 239
Emulsion **9**, 17; **11**, 277; **17**, 203; **25**, 179; **30**, 47; **33**, 235; **38**, 133; **42**, 19; **44**, 169; **45**, 169; **46**, 25; **50**, 141; **51**, 47; **52**, 165; **57**, 87; **59**, 69; **61**, 51; **62**, 81, 249; **63**, 65, 219; **64**, 217; **65**, 133, 225; **66**, 1, 29, 79; **67**, 195; **68**, 19, 205, 277, 281; **69**, 193; **70**, 251; **72**, 65; **73**, 23, 177; **75**, 171; **76**, 207, 225; **77**, 169, 211; **78**, 217; **79**, 113; **81**, 111; **82**, 99; **84**, 93, 107, R5; **85**, 19; **86**, 25, 187; **87**, 167, 239, 247; **88**, 53, 171, 417; **89**, 139; **90**, 43; **91**, 157; **93**, 61, 161; **94**, 59, 89, 115; **95**, 77; **96**, 147; **98**, 173; **99**, 1; **100**, 55; **101**, 175; **104**, 49; **106**, 51; **108**, 57, 117; **109**, 45, 251; **110**, 189; **111**, 63, 137, 213, 235; **114**, 43, 103; **115**, 1, 45, 129, 225; **116**, 253; **117**, 119, 197; **118**, 47; **119**, 139; **120**, 127; **121**, 45, 169; **124**, 61; **125**, 107, 223; **126**, 119; **129**, 159; **130**, 203; **132**, 33, 259; **134**, 119; **136**, 147; **137**, 57, 177, 243; **140**, 97; **141**, 27, 205, 227; **142**, 1, 75; **143**, 67; **145**, 17; **146**, 239
Enalapril **75**, 117
Enalapril maleate **28**, 183; **40**, 129; **93**, 245; **107**, 77
Enalkiren **65**, 195; **120**, 127
Enamel **49**, 147
Enantiomer **68**, 97; **87**, 95; **92**, 1; **94**, 171; **99**, 303; **127**, 53; **137**, 21
Enbucrilate **80**, R11; **110**, 21
Encapsin **75**, R5
Encapsulation **56**, 103; **57**, 241; **59**, 9; **63**, 149; **66**, 207, 273, 283; **68**, 19; **69**, 69; **70**, 261; **71**, 127; **72**, 141; **74**, 183; **77**, 119, 177; **78**, 227; **79**, 11; **81**, 235; **87**, 183; **88**, 211, 313; **89**, 133; **91**, 69; **96**, 67, 157, 231; **100**, 71; **101**, 191; **111**, 271; **117**, 225, 237; **120**, 21; **123**, 33; **129**, 113, 289; **131**, 25, 101, 171, 191; **132**, 9, 263; **134**, 89; **135**, 63; **137**, 133, 187; **138**, 113, 191; **141**, 81; **142**, 135; **143**, 1, 171; **145**, 241; **147**, 153; **148**, 139
Encompress **35**, 157; **62**, 125; **64**, 35; **89**, R17; **90**, 213
Endocytosis **37**, 33; **85**, 75
Endolipide **99**, 1
α-Endorphin **50**, 261
Endothelium cell **68**, 231; **78**, 103; **110**, 29
Endotoxin **27**, 17; **40**, 173
Endotoxin cr 51 **27**, 17
Enema **25**, 191
Energy **72**, 117, 157; **97**, 101; **120**, 13
Enkephalin **12**, 271
Enkephalin derivative **12**, 271; **70**, 111; **75**, 117; **76**, 113
Enoxacin **63**, 237; **83**, 267; **88**, 379; **93**, 121
Enprofylline **56**, 235; **58**, 129
Enrofloxacin **95**, 51
Enteric coated tablet **4**, 291; **34**, 259; **42**, 105; **80**, 63; **112**, 199, 207; **113**, 17; **125**, 151; **133**, 139, 149; **141**, 9; **145**, 247
Enteritis **122**, 121
Enterobacter aerogenes **2**, 325; **34**, 241
Enterobacter cloacae **82**, 107; **87**, 195
Enterohepatic circulation **83**, 257
Entex lA **103**, 97
Enthalpy **40**, 1; **42**, 19; **50**, 213; **86**, 211
Entropy **28**, 103; **40**, 1; **78**, 157
Enzyme **75**, 97, 117; **101**, 165
Enzyme activity **50**, 67; **62**, 259; **74**, 195; **81**, 119, 267; **86**, 17; **88**, 221, 229, 293; **89**, 65; **97**, 171; **111**, 147; **114**, 197; **117**, 181
Enzyme immobilization **34**, 225
Enzyme immunoassay **31**, 193; **33**, 89; **46**, 9
Enzyme inducing agent **129**, 295
Enzyme inhibition **58**, 129; **74**, 195; **81**, 119, 267; **86**, 17; **144**, 27
Enzyme inhibitor **74**, 195; **75**, 97, 117; **95**, 101; **99**, 51; **105**, 227; **106**, 85; **107**, 133; **113**, 1; **121**, 217; **125**, 117; **128**, 229
Enzyme localization **64**, 7
Enzyme stability **90**, 187
Enzymuria **46**, 103
Eosin **113**, 263; **126**, 281
Ephedrine **4**, 135; **6**, 195; **9**, 285; **10**, 165; **12**, 57; **30**, 151; **34**, 75; **41**, 55, 205; **51**, 161; **58**, 89; **86**, 211; **94**, 171; **100**, 257; **119**, 37; **137**, 21
Ephedrine naphthalenesulfonate **94**, 171
Epiallomuscarine **15**, 223
Epianhydrotetracycline **1**, 185
Epichlorohydrin **23**, 35
4-Epichlortetracycline **96**, 13
Epicillin **4**, 299; **9**, 273
7-Epidaunomycinone **34**, 247
Epidermis **2**, 265; **26**, 89; **62**, 9; **65**, 211; **74**, 59; **78**, 209; **98**, 141; **102**, 241; **105**, 113; **114**, 237
Epidermal growth factor **46**, 9; **49**, 173; **54**, 259; **75**, 117; **138**, 75
Epidermis horny layer **2**, 265; **8**, 193; **16**, 255; **22**, 31, 31; **24**, 267; **26**, 275; **27**, 287; **31**, 247
Epikuron **84**, 107, R5; **116**, 253
Epikuron-200 **111**, 63
Epilepsy **16**, 285; **23**, 25; **86**, 259
Epimerization **1**, 281
Epimuscarine **15**, 223
Epipodophyllotoxin **145**, 9
Epirubicin **32**, 123; **41**, 213; **65**, 49; **108**, 69; **124**, 205
Epitetracycline **26**, 259
Epithelial cell **10**, 153; **64**, 181; **71**, 55
Epithelium **53**, 253; **75**, 73
Epsilon caprolactone **113**, 223
Equilibrium constant **80**, 135

Equipment **88**, 285
Erbium **32**, 251
Erbium 170 **32**, 251
Erbium 171 **32**, 251
Ergocalciferol **81**, 1
Ergot alkaloid **75**, 97
Ergotamine **7**, 19
Erosion **117**, 41
Erythema **83**, 73; **93**, 1; **113**, 159; **131**, 143
Erythrocyte **8**, 323; **22**, 137; **33**, 1; **53**, 61; **110**, 161
Erythrocyte membrane **94**, 161
Erythroleukemia cell **139**, 69
Erythromycin **1**, 165; **5**, 345; **14**, 1; **17**, 225; **30**, 199; **34**, 75; **38**, 33; **55**, 59, 67; **56**, 159; **62**, R1; **67**, 195; **72**, 57; **80**, R7; **106**, 33; **110**, 109; **113**, 215; **115**, 123
Erythromycin acetate **56**, 159
Erythromycin benzoate **56**, 159
Erythromycin estolate **63**, 113; **115**, 123
Erythromycin ethylsuccinate **56**, 159; **115**, 123
Erythromycin glutarate **56**, 159
Erythromycin methylcarbonate **56**, 159
Erythromycin propionate **56**, 159; **115**, 123
Erythromycin salicylate **56**, 159
Erythromycin stearate **17**, 225
Erythropoietin **123**, 1
Erythrosine **136**, 155
Escherichia coli **2**, 325; **9**, 49; **12**, 295; **15**, 49; **21**, 265; **23**, 357; **25**, 329; **27**, 163; **34**, 241; **35**, 227; **36**, 191; **45**, 153; **46**, 141; **54**, 51; **55**, 209; **56**, 169; **57**, 1; **59**, 205; **99**, 93; **113**, 257
Escherichia coli endotoxin **10**, 133
Escin **142**, 103
Esophagus **22**, 219; **34**, 75; **61**, 109
Esophagus cancer **147**, 119
Esorubicin **32**, 123
Ester **76**, 99, 177; **77**, 21; **79**, 233; **82**, 205; **99**, 135; **101**, 145; **103**, 1, 293; **107**, 9, 91; **112**, 181; **126**, 139
Ester derivative **144**, 107
Esterase **13**, 183; **17**, 299; **27**, 325; **35**, 73, 121; **40**, 249; **44**, 125; **48**, 189; **62**, 9; **70**, 271; **82**, 205; **88**, 221, 229; **89**, 65; **101**, 165
Esterase liver level **27**, 325
Esterolysis **19**, 1
Estradiol **2**, 113, 215; **15**, 185; **29**, 73; **32**, 159; **35**, 47; **42**, 235; **57**, 29; **61**, 135; **64**, 7, 61; **74**, 157; **76**, 177; **77**, 75, 231; **78**, 123, 137; **79**, 21; **80**, 17; **86**, 69; **89**, 251, R13; **90**, 171; **93**, 233; **110**, 169; **114**, 237; **116**, 1, 101; **117**, 101; **127**, 155; **131**, 117; **132**, 23; **135**, 137; **138**, 149; **145**, 37, 49; **146**, 11, 41; **148**, 55
Estradiol acetate **76**, 177
Estradiol-3-acetylsalicylate **76**, 177
Estradiol-3-anthranilate **76**, 177
Estradiol benzoate **76**, 177
Estradiol cipionate **76**, 177
Estradiol derivative **127**, 155
Estradiol-3-methyl-ether **35**, 47
Estradiol undecylenate **6**, 185
Estradiol valerate **76**, 177
Estramustine **63**, 121; **68**, 87; **79**, 107; **81**, 31; **108**, 69; **124**, 205
Estramustine phosphate **108**, 69
Estriol **132**, 23; **138**, 149
Estrogen **6**, 195; **28**, 1; **35**, 47; **71**, 25
Estrone **2**, 215; **35**, 47; **42**, 235; **78**, 137; **92**, 1; **132**, 23; **138**, 149
Estrus **24**, 209
Etacrynic acid **51**, 55
Etarotene **101**, 249
Ethallobarbital **3**, 341
Ethanol **4**, 205; **5**, 167; **7**, 189; **16**, 127; **17**, 77; **18**, 149; **19**, 149; **25**, 359; **26**, 275; **27**, 1; **28**, 193; **32**, 187; **33**, 37; **34**, 217; **39**, 183
Ethanol derivative **3**, 1
Ethanolamine derivative **137**, 143
Ethenzamide **108**, 167
Ether **13**, 75; **17**, 77; **29**, 177; **34**, 217; **35**, 173; **126**, 219; **140**, 237
Ethinylestradiol **15**, 149; **137**, 95
2[2[(1-Ethoxycarbonyl-3-phenylpropyl)amino]propionyl] × 1,2,3,4-tetrahydro **60**, 99
7-(Ethoxycarbonyloxymethyl)theophylline **12**, 299
Ethoxydiglycol **131**, 183
Ethoxylated stearyl alcohol **25**, 199
7-(Ethoxymethyl)theophylline **12**, 299
Ethoxymethylenemalonic acid diethyl ester **33**, 89
4-Ethoxyphenol **10**, 89
Ethoxzolamide **51**, 27
Ethyl biscoumacetate **7**, 293; **12**, 231; **14**, 123; **18**, 9; **59**, 137
Ethyl cellulose **2**, 91; **3**, 247; **12**, 251; **16**, 65; **21**, 119; **24**, 355; **29**, 1, 37; **31**, 55, 175, 271; **37**, 119; **43**, 155; **44**, 125, 249; **47**, 21; **48**, 51; **49**, 213; **52**, 1; **58**, 31, 73; **61**, 101; **64**, 99; **67**, 21; **68**, 199; **70**, 59; **76**, 91, 193; **77**, 89; **79**, 223; **80**, 171; **81**, 49; **82**, 129; **85**, 29; **86**, 247; **88**, 39, 405; **91**, 167; **92**, 29, 35, 55; **98**, 37; **103**, 47; **104**, 95, 203; **111**, 117, 181; **112**, 37, 117, 173; **113**, 9; **114**, 257; **115**, 217; **117**, 13; **118**, 199; **123**, 85, 119; **124**, 219; **125**, 289; **126**, 1, 83; **130**, 187; **131**, 25; **133**, 97, 161; **140**, 85; **141**, 137
1-Ethyl-1,4-dihydro-7-methoxy-4-oxo-3-quinolinecarboxylic acid **22**, 283
2-(1-Ethyl-1,4-dihydro-7-methoxy-4-oxo-3-quinolyl)acetic acid **22**, 283
4-Ethyl-2-hydroxyimino-5-nitro-3-hexenamide **110**, 1
2-Ethyl-2-hydroxymethylpropane-1,3-diol **130**, 83
Ethyl paraben **2**, 337; **19**, 1; **27**, 1, 163; **29**, 103; **32**, 223; **33**, 105; **34**, 115; **80**, 203; **83**, 211; **90**, 203; **109**, 155; **112**, 65, 79; **125**, 141; **127**, 27; **133**, 17; **140**, 237
Ethylaniline **37**, 49; **57**, 189; **100**, 1
Ethylbenzene **103**, 163; **126**, 219
Ethylene glycol **15**, 185; **30**, 1; **36**, 211; **57**, 87; **82**, 145
Ethylene glycol dimethacrylate **7**, 189
Ethylene maleic anhydride copolymer **31**, 261
Ethylene oxide **9**, 359; **16**, 127; **79**, 251; **113**, 223; **116**, 95

Ethylene vinyl acetate copolymer **18**, 247; **45**, 1; **48**, 1, 247; **137**, 95
Ethylenediamine **86**, 231
3-Ethylgentisic acid **5**, 149
4-Ethylgentisic acid **5**, 149
(2-Ethylhexyl)phthalate **29**, 233
O-Ethylhydroxylamine **32**, 257
Ethylmethylglycinexylidide **8**, 81
4-Ethylphenol **10**, 89
Ethylurea **7**, 189
Eticol-90 **94**, 223
Eticol-372 **94**, 223
Etidocaine **136**, 165
Etidronic acid **52**, 139; **59**, 95
Etilefrine **26**, 243
Etodolac **49**, 121
Etofylline **30**, 143; **50**, 205
Etoposide **21**, 219; **41**, 169; **68**, 151; **84**, 223; **109**, 73; **113**, 103; **118**, 161
Etorphine **138**, 149
Etretin **115**, 255
Etretinate **101**, 249; **130**, 57
Eudraflex **133**, 161
Eudragit **27**, 233; **31**, 15, 43, 271; **32**, 143; **37**, 119, 211; **45**, 229; **47**, 21, 37, 263; **48**, 51; **49**, 213; **50**, 133; **55**, 47, 157, 247; **57**, 183; **58**, 89; **62**, 97, 105; **67**, 163; **70**, 153, 261; **72**, 1; **74**, 169; **85**, R1; **86**, 35, 113, 159; **87**, 247; **88**, 31; **89**, 9; **91**, 29; **94**, 125; **95**, 117, 193; **96**, 129, 175; **99**, 181; **101**, 23; **102**, 101, 185; **103**, 243, 293; **106**, 149; **111**, 89; **113**, 17; **114**, 13; **116**, 31; **118**, 103, 151; **119**, 165; **123**, 145; **126**, 1, 79; **127**, 203; **130**, 187; **131**, 9, 183, 247; **133**, 65; **137**, 159; **138**, 215; **139**, 237; **140**, 145; **142**, 143; **144**, 195; **145**, 157; **147**, 1; **148**, 219
Eudragit l 30 D **34**, 259; **37**, 203
Eudragit RS **79**, 97; **81**, 49; **86**, 159; **98**, 63, 75; **101**, 23; **106**, 213; **113**, 39; **118**, 151; **126**, 79; **129**, 185; **130**, 115; **131**, 9, 183, 247; **133**, 65; **137**, 159; **139**, 177; **140**, 145; **144**, 37, 195; **146**, 175; **148**, 85
Eumulgin **62**, 217; **92**, 191
Evaporation **68**, 19, 205; **84**, 151
Excipient **64**, 227; **68**, 51; **84**, 167; **89**, 1; **91**, 173; **97**, 161, 195; **103**, 9, 155; **109**, 237; **112**, 225; **115**, 35, 201; **116**, 125; **132**, 183; **135**, 53, 147; **139**, 53; **143**, 211; **144**, 147; **145**, 65; **146**, 1, 71; **148**, 1, 155
Experimental infection **79**, 29; **83**, 123, 251
Expidet **46**, 241
Explosion **128**, 277
Extraction **35**, 173; **76**, 151; **84**, 151
Eye **1**, 165; **11**, 21, 155; **13**, 263; **16**, 163, 339; **17**, 299; **20**, 187, 285; **23**, 175; **26**, 203, 243; **29**, 43; **31**, 261; **34**, 163; **47**, 21; **54**, 95, 149; **55**, 105; **57**, 163; **59**, 1; **61**, 241; **81**, 59; **137**, 233; **138**, 175
Eye albinism **13**, 183
Eye drops **1**, 165; **20**, 265; **37**, 19; **42**, 47; **64**, 235; **75**, R1; **82**, 179; **84**, 217; **123**, 173; **137**, 249; **147**, 127
Eye inflammation **110**, 211
Eye irritation **100**, 219; **127**, 85; **133**, 171

F

Famotidine **54**, 65; **58**, 19; **61**, 173; **66**, 71; **81**, 1; **102**, 271; **108**, 31
Farmarubicin **124**, 205
Farmitrexate **54**, 181
Fast dissolving dosage form **40**, 119
Fat **28**, 151; **29**, 191; **64**, 217; **70**, 251; **81**, 1; **85**, 103; **99**, 1
Fat emulsion **13**, 9, 303; **29**, 233
Fatigue **12**, 277
Fatty acid **6**, 185, 195; **31**, 239; **33**, 225; **70**, 173; **85**, 103; **88**, 201; **99**, 109; **102**, 25; **104**, 1, 87; **106**, 1, 15; **109**, 147; **111**, 213; **126**, 49, 267; **131**, 103; **132**, 33, 89; **144**, 115; **145**, 187
Fatty acid derivative **88**, 89
Fatty acid ester **95**, 181; **137**, 117
Fatty alcohol **33**, 225; **106**, 1; **112**, 165; **148**, 41
Fatty aldehyde **9**, 49
Felbinac **52**, 115; **57**, 55; **59**, 19; **83**, 73
Felbinac ethyl ester **57**, 107
Felodipine **47**, 67; **60**, 151; **62**, 143; **68**, 25; **81**, R1; **87**, 1; **104**, 253; **106**, 141; **122**, 57; **131**, 183
Femoxetine **21**, 51
Fenbufen **40**, 181, 187; **44**, 261; **126**, 95
Fenbufen derivative **40**, 187
Fenclofenac **126**, 95
Fenfluramine **21**, 75
Fenofibrate **103**, 137
Fenoprofen **2**, 283; **60**, 243; **69**, 175; **75**, 63; **96**, 79
Fenoterol **101**, 1; **116**, 169; **142**, 125; **149**, 85
Fentanyl **38**, 261; **44**, 257; **101**, 105; **120**, 221; **138**, 149
Fenthion **114**, 171
Ferric chloride **90**, 221
Ferric gluconate **61**, 199
Ferric ion **57**, 173
Ferric nitrate **127**, 279
Ferrite **147**, 119
Ferritin **61**, 199; **90**, 119
Ferrous sulfate **34**, 75; **40**, 201; **68**, 11; **90**, 221; **104**, 247
Fertility **69**, 93
Fertirelin **108**, 49
Ferulic acid **80**, 75
Feruloylmethane **110**, 161
Fetindomide **45**, 207
Fibrin **90**, 95
Fibrinogen **63**, 177; **90**, 95; **91**, 235
Fibrinogen I^{125} **8**, 35
Fibrinogen receptor antagonist **146**, 147
Fibrinopeptide A **50**, 261
Fibroblast **64**, 139; **70**, 159; **110**, 277
Fibronectin **119**, 149
Fibrosarcoma **142**, 25; **146**, 81
Ficoll **19**, 107
Film **45**, 39; **61**, 101; **62**, 97; **71**, 147; **73**, 185; **78**, 49; **104**, 203
Film coating **145**, 197
Filter **1**, 133; **103**, 87; **133**, 9
Filtration **117**, 225
Flavine mononucleotide **125**, 151
Flavine nucleotide **35**, 157

Flavonoid **86**, 17; **124**, 1; **145**, 87
Flavonol derivative **86**, 17
Flavoring agent **72**, 97
Flecainide **54**, 1
Fleroxacin **63**, 237; **83**, 267; **88**, 379; **93**, 121, 131
Flocculation **9**, 315; **77**, 57; **118**, 229
Flow **130**, 103
Flow injection analysis **30**, 123
Flow kinetics **103**, 155; **146**, 159
Fluanisone **38**, 261
Flucloxacillin **15**, 309; **32**, 151; **54**, 1; **143**, 93
Flucytosine **97**, 1
Flucytosine derivative **35**, 243
Fludarabine **71**, 73
Fludiazepam **6**, 147; **16**, 327
Fludrocortisone **1**, 165
Flufenamic acid **2**, 283; **6**, 195, 205; **21**, 187; **32**, 13; **67**, 1; **68**, 1
Fluidized bed **86**, 247; **93**, 101
Flumazenil **98**, 51
Flunisolide **1**, 71
Flunitrazepam **6**, 147; **16**, 327; **29**, 67; **36**, 113; **61**, 189
Flunitrazepam-H-3 **36**, 113
Fluocinonide **11**, 257; **26**, 145
Fluocinolone acetonide **10**, 1; **19**, 245; **40**, 105; **118**, 129
Fluocortolone **14**, 1
Fluorescein **1**, 71; **6**, 205; **13**, 321; **38**, 231; **42**, 267; **59**, 9; **68**, 231; **80**, 213; **90**, 229; **98**, 19
Fluorescein isothiocyanate **8**, 217; **73**, 81; **77**, 141; **79**, 191; **93**, 91; **103**, 27
Fluorescein isothiocyanate albumin **62**, 133
Fluorescein isothiocyanate dextran **38**, 231
Fluorescein sodium **139**, 197
Fluorescence **31**, 75; **46**, 25; **67**, R1
Fluorescence microscopy **144**, 11
Fluorescence polarization **33**, 253
Fluorescent dye **141**, 27
Fluorescent probe **40**, 93
Fluoride **15**, 177; **58**, 203; **123**, 159
Fluoride ion **131**, 33
Fluoride sodium **49**, 147; **110**, 127
Fluorine-18 **76**, 183
5-Fluoro-1,3-bis(morpholinomethyl)uracil **21**, 251
5-Fluoro-1-octadecylcarbamoyluracil **36**, 147
5-Fluoro-1-octylcarbamoyluracil **36**, 147
Fluorocarbon **8**, 1
Fluorometry **1**, 71; **25**, 217; **31**, 231; **35**, 157, 173
8-Fluoronorfloxacin **79**, 89
8-Fluoropefloxacin **79**, 89
4-Fluorophenol **10**, 89
Fluorophotometry **54**, 95
2-Fluorothymine-arabinoside **84**, 39
Fluorouracil derivative **13**, 89; **21**, 251, 349; **24**, 43; **44**, 87; **60**, 1; **93**, 27, 37; **111**, 223; **129**, 203, 223; **132**, 243
Fluorouracil **8**, 217; **12**, 153; **13**, 321; **23**, 209; **24**, 43; **25**, 347; **27**, 89; **36**, 41, 147; **38**, 239; **41**, 91; **43**, 1, 17; **44**, 87; **48**, 1; **50**, 15; **54**, 181; **55**, 31, 185; **56**, 43; **57**, 29; **60**, 1, 41; **61**, 51; **65**, 1; **68**, 61, 151; **70**, 87; **74**, 157; **79**, 21; **81**, 1, 39; **86**, 69; **92**, 89; **93**, 27, 37; **94**, 189; **99**, 145; **106**, 161; **108**, 241; **111**, 223; **116**, 1, 201, 237; **117**, 101; **126**, 169; **129**, 33, 203, 223; **131**, 73, 117; **133**, 97; **138**, 139; **139**, 205; **143**, 255; **145**, 9, 37, 49, 157; **146**, 41
Fluphenazine **3**, 231; **6**, 185; **7**, 1; **12**, 219; **13**, 227; **16**, 153, 271; **35**, 129; **36**, 185; **94**, 135
Fluphenazine enanthate **6**, 185
Fluprednisolone **14**, 1
Fluprostenol **4**, 1
Flurandrenolide **18**, 53
Flurazepam **6**, 147; **16**, 327; **42**, 235
Flurbiprofen **17**, 273; **45**, 165; **53**, 7; **62**, 21; **70**, 153; **74**, 169; **79**, 21; **85**, R1; **95**, 51; **115**, 119; **120**, 21; **126**, 267
FMC-2843 **90**, 213
Foam **25**, 191
Folic acid **38**, 239; **65**, 63; **81**, 1
Folinic acid **79**, 71; **87**, 195; **92**, 237; **116**, 217
Food **25**, 113; **28**, 151; **29**, 191; **34**, 213; **35**, 187; **37**, 227, 245; **38**, 221
Food drug interaction **6**, 301; **17**, 225; **25**, 113; **28**, 151, 249; **29**, 191; **34**, 213; **37**, 227, 245; **44**, 9; **50**, 79, 83, 155; **54**, 123; **55**, 25; **56**, 151; **60**, 125; **108**, 125; **112**, 87; **136**, 147; **139**, 95; **144**, 233
Food dye **115**, 241
Food intake **4**, 175; **14**, 123; **109**, 91; **112**, 207; **123**, 145
Food processing **72**, 97
Formaldehyde **9**, 49; **12**, 185; **16**, 127; **19**, 239; **27**, 175; **38**, 59; **72**, 51; **73**, 97, 191; **107**, 141; **110**, 257; **114**, 121; **133**, 139; **148**, 73
Formamide **36**, 211
Formic acid **11**, 131; **67**, 59; **114**, 185
Formic acid methyl ester **4**, 309
Formoterol **101**,
Foscarnet **63**, 213; **104**, 157; **108**, 225
Fourier transformation **63**, 219; **85**, 163
Fracture **66**, 143
Fragmentation **23**, 79
Free energy **17**, 39; **40**, 1; **42**, 181
Free fatty acid blood level **11**, 11
Free radical **104**, 49; **127**, 73; **131**, 1; **144**, 123
Freeze drying **60**, 203; **61**, 27; **62**, 165; **71**, 137; **72**, 97; **77**, 119; **88**, 319; **111**, 205; **113**, 133; **114**, 159; **124**, 241; **140**, 13
Freeze fracture **35**, 263
Freezing **12**, 173; **103**, 93
Friction **16**, 349; **26**, 191
Fructose **16**, 319
Fruit **76**, 199
Fucose **124**, 173
Fumagillol chloroacetylcarbamate **123**, 237
Fumaric acid **111**, 89
Fungus growth **80**, 33
2,5-Furandicarboxylic acid **12**, 31
Furfural curcuminoid **131**, 1
Furosemide **4**, 13; **11**, 81, 237; **22**, 105; **31**, 65; **38**, 247; **41**, 75; **42**, 217; **45**, 65; **47**, 141; **50**, 222; **53**, 99; **60**, 11, 163; **67**, 113, 169; **76**, 193; **80**, 129; **88**, 275; **89**, 19; **99**, 333; **111**, 217; **136**, 117, 175

Furosemide derivative **60**, 163
Furosemide S 35 **4**, 27
Fusafungine **123**, 291
Fusidate sodium **72**, 57; **119**, 57; **128**, 223
Fusidic acid **72**, 57; **119**, 57; **128**, 223
Fusogenic fatty acid **31**, 239

G

Gabapentin **78**, 25
Gabexate mesilate **29**, 137
Gadolinium pentetate **117**, 1; **132**, 9
Galactitol **145**, 9
Galactomannan **136**, 43
Galactosamine **87**, 105
Galactose **128**, 65; **147**, 143
Galactose binding protein **42**, 69
β-Galactosidase **22**, 137; **68**, 265; **71**, 137; **90**, 187
Galactosylceramide **103**, 81
Gallic acid propyl ester **121**, 95
Gallium **107**, 189
Gallium-67 **88**, 327
Gallopamil **64**, 117
Ganglioside **44**, 257; **46**, 31; **90**, 59
Ganglioside-GM1 **109**, 99; **113**, 141
Ganglioside-GM3 **90**, 73
Gantrez copolymer **78**, 237; **80**, 277
Gantrez copolymer derivative **26**, 45
Gas **106**, 77
Gas chromatography **14**, 113; **25**, 147; **29**, 113; **142**, 67
Gas liquid chromatography **27**, 45
Gas sterilization **9**, 359
Gastrin **75**, 117; **75**, 97
Gastrocote **28**, 249
Gastroesophageal reflux **21**, 341; **34**, 105
Gastrointestinal absorption **16**, 241; **17**, 47; **56**, 97; **83**, 155, 257; **91**, 29; **92**, 197; **98**, 165; **102**, 133; **136**, 117; **137**, 85
Gastrointestinal clearance **22**, 1; **32**, 85
Gastrointestinal hemorrhage **10**, 143; **91**, 75
Gastrointestinal motility **49**, 183; **59**, 105; **78**, 69; **92**, 167
Gastrointestinal mucosa **104**, 227; **128**, 99
Gastrointestinal passage **4**, 127
Gastrointestinal secretion **37**, 219
Gastrointestinal symptom **70**, 35
Gastrointestinal toxicity **27**, 245; **32**, 13; **33**, 219, 235; **106**, 63; **126**, 275
Gastrointestinal tract **7**, 19, 137; **17**, 47; **21**, 167; **34**, 35; **69**, 181; **84**, 245; **87**, 37, 105; **95**, 171; **96**, 183; **105**, 137, 157; **108**, 39, 125; **111**, 99
Gastrointestinal transit **34**, 213; **42**, 105; **49**, 183; **53**, 79, 145; **60**, 89; **72**, 79; **76**, 123, 133, 183; **78**, 69; **83**, 155; **84**, 59; **92**, 167; **94**, 235; **97**, 61; **100**, 81; **101**, 263; **112**, 199, 207; **114**, 1; **119**, 1, 47, 223; **125**, 185; **137**, 225; **139**, 95; **140**, 111, 229
Gaucher disease **22**, 137
Gaviscon in 113m **28**, 249
Gaviscon **19**, 169; **21**, 341; **105**, 97
Gel **21**, 277; **25**, 179, 237; **14**, 223; **58**, 203; **61**, 225; **62**, 217; **70**, 147; **71**, 193; **74**, 25; **78**, 209; **80**, 119, 271; **83**, 221; **88**, 1; **90**, 43; **95**, 153, 219; **98**, 37, 57, 113, 131; **99**, 13; **100**, 143, 165, 199; **102**, 19; **105**, 31; **106**, 103, 125; **107**, 85; **109**, 115; **110**, 241; **111**, 191, 261, 271; **121**, 11; **124**, 107, 173; **125**, 7, 205, 231; **127**, 73, 115; **128**, 9; **129**, 13; **130**, 121; **131**, 101, 137; **136**, 155; **137**, 117, 233; **140**, 251; **142**, 9; **145**, 129
Gel chromatography **13**, 109
Gelatin **2**, 91; **4**, 175; **6**, 277; **7**, 285; **8**, 131; **9**, 59; **13**, 23; **17**, 59; **19**, 107; **20**, 87; **22**, 137; **25**, 121; **26**, 203; **31**, 99; **32**, 207; **34**, 125; **35**, 157, 177; **38**, 103; **42**, 77; **44**, 43, 151; **47**, 21; **49**, 157; **50**, 205; **51**, 161; **55**, 31, 53; **65**, 69, 85; **71**, 147, 187; **73**, 69, 185; **79**, 67; **83**, 139; **84**, 49; **85**, 1; **95**, 85; **99**, 321; **103**, 259; **105**, 181; **110**, 291; **112**, 17; **113**, 17; **115**, 201; **117**, 151; **118**, 199; **121**, 37, 73, 117, 149; **122**, 79; **126**, 161; **127**, 203; **128**, 261; **130**, 203; **133**, 139; **134**, 223; **135**, 53, 63; **139**, 53; **142**, 25, 97; **143**, 133, 255; **144**, 209; **145**, 137; **146**, 271; **147**, 31; **148**, 23, 73
Gelatin microsphere **2**, 245
Gelatin solution **27**, 299
Gelita collagel **99**, 321
Gellan gum **73**, 117; **95**, 219, 229
Gelrite **57**, 163; **73**, 117; **95**, 219; **95**, 229
Gelucire **50**, 133; **58**, 215; **72**, 35; **78**, 35; **80**, 145, 151; **110**, 75; **111**, 51; **125**, 123; **126**, 287; **132**, 1; **141**, 9
Generol **99**, 29
Genetic polymorphism **59**, 281; **63**, 35; **69**, 265; **73**, 137; **92**, 183
Genital system **54**, 231
Gentamicin **2**, 101; **18**, 353; **21**, 239; **26**, 133; **30**, 181; **31**, 193; **35**, 121; **37**, 113; **42**, 193; **46**, 103, 261; **53**, 85; **56**, 29; **78**, 165; **81**, 1; **90**, 59; **111**, 197; **116**, 1; **123**, 173; **127**, 115; **133**, 265; **137**, 75; **143**, 187, 195
Gentisic acid **5**, 149; **35**, 73
Gentisic acid ethanolamide **35**, 73
Geometry **108**, 187
Gepirone **136**, 117
Geraniol **130**, 141
1-Geranylazacycloheptan-2-one **135**, 91; **145**, 103
Germination **34**, 51
Ghost cell **22**, 137
Gill permeability **47**, 185
Gingivitis **51**, 103
Gitoxin acetate **2**, 127
GLA-60 **111**, 15
Glafenine **4**, 195
Glass **16**, 181; **25**, 217; **35**, 111; **53**, 75; **99**, 59; **101**, 81; **113**, 133; **120**, 179; **121**, 81; **127**, 65; **130**, 251; **146**, 213
Glass bead **28**, 23
Glaucoma **1**, 189; **17**, 299; **33**, 45; **53**, 189; **58**, 115; **106**, 161
Gliadin **121**, 117; **131**, 191
Glibenclamide **38**, 123; **50**, 103; **67**, 131; **122**, 35, 57; **126**, 155; **136**, 117; **140**, 69
Gliclazide **148**, 211
Glipizide **140**, 69
Glucagon **132**, 189
Glucagon-like-peptide-1 **136**, 53
Glucagon-like-peptide-1[9–36]-amide **136**, 53
β-1,3-Glucan **46**, 149

Glucocerebrosidase **22**, 137
Glucocorticoid **92**, 105
Gluconate calcium **51**, 55; **72**, 43; **90**, 203
Glucosamine **87**, 105; **149**, 15
Glucose **1**, 121; **2**, 325; **7**, 189; **11**, 131; **12**, 31; **13**, 9; **14**, 223; **16**, 319; **27**, 17; **29**, 43; **36**, 61, 67; **43**, 225; **49**, 1; **51**, 55; **57**, 217; **58**, 177; **64**, 217; **66**, 149; **67**, 79; **71**, 201; **72**, 97; **75**, 181; **80**, 9; **81**, 1; **87**, 167; **88**, 141; **91**, 29; **92**, 47; **101**, 15, 191; **107**, 99, 199; **119**, 71, 127; **121**, 169; **123**, 281; **127**, 95; **128**, 113; **129**, 1, 275; **130**, 149, 251; **134**, 193; **135**, 53; **136**, 1; **146**, 263
Glucose blood level **1**, 315; **5**, 223; **23**, 87; **30**, 95; **34**, 157; **38**, 83; **54**, 223; **62**, 187; **78**, 9; **83**, 1; **119**, 127; **132**, 189
Glucose oxidase **101**, 191
β-Glucosidase **22**, 137
Glucoside **105**, 219; **107**, 1; **117**, 129
β-Glucuronidase **68**, 265
Glutamic acid **72**, 149; **88**, 111; **117**, 129
Glutamic acid-4-methoxy-2-naphthylamide **30**, 73
Glutaminase **34**, 225
Glutamine **34**, 225; **38**, 239; **72**, 149
Glutaraldehyde **4**, 57; **16**, 127; **25**, 105; **42**, 61; **57**, 205; **58**, 63; **90**, 95; **106**, 221; **111**, 217; **121**, 233; **126**, 103; **127**, 165; **128**, 251; **131**, 191; **134**, 247; **135**, 183; **144**, 241
O-Glutarylsalicylamide **13**, 273
Glutathione **28**, 119; **33**, 1; **44**, 265; **57**, 263; **63**, 227; **76**, 257; **88**, 201; **133**, 85; **134**, 147
Glutathione dextran conjugate **57**, 263
Glutathione transferase **76**, 257
Glycerogelatin **122**, 79
Glycerol **15**, 185; **16**, 319; **17**, 39; **30**, 1; **31**, 185; **32**, 79; **33**, 71; **36**, 61, 67; **43**, 155, 225; **55**, 185; **57**, 87; **82**, 145; **88**, 293; **104**, 107; **105**, 189; **109**, 45; **112**, 117; **119**, 183, 193; **122**, 79; **123**, 71; **125**, 205; **131**, 183; **137**, 243; **142**, 115, 251; **144**, 99; **145**, 9; **146**, 271; **147**, 51
Glycerol behenate **86**, 159; **105**, 227
Glycerol derivative **135**, 1
Glycerol-1,2-dinitrate **22**, 327; **71**, 175
Glycerol-1,3-dinitrate **22**, 327; **71**, 175
Glycerol dinitrate **12**, 331
Glycerol dioleate **4**, 271, 281
Glycerol laurate **11**, 27; **135**, 1
Glycerol myristate **11**, 27
Glycerol oleate **4**, 219, 271, 281; **24**, 173; **31**, 185; **79**, 113; **86**, 137; **98**, 173; **101**, 115; **110**, 137; **125**, 7; **137**, 85; **142**, 9; **147**, 135
Glycerol stearate **6**, 11; **11**, 27; **37**, 127; **50**, 141; **118**, 95; **119**, 203; **137**, 243; **147**, 31, 79, 95
Glyceryl trinitrate **4**, 75; **5**, 335; **8**, 323; **10**, 339; **12**, 331; **22**, 327; **26**, 89; **29**, 73; **32**, 243; **39**, 183; **43**, 201; **53**, 119; **55**, 147; **56**, 87; **64**, 187; **65**, 159, 265; **71**, 175; **73**, 125; **82**, R1; **89**, R1; **110**, 197; **131**, 229; **135**, 1
Glycine **1**, 173; **3**, 109; **28**, 119; **38**, 239; **71**, 117; **72**, 97, 149; **104**, 187; **114**, 159; **115**, 139; **121**, 157
Glycine derivative **93**, 1
Glycine-4-methoxyphenyl ester **75**, 211
Glycocholate sodium **12**, 1; **69**, 21; **95**, 209; **107**, 159; **111**, 213; **113**, 83; **134**, 47; **141**, 151
Glycocholic acid **6**, 137; **8**, 303; **9**, 173; **11**, 27; **12**, 219; **31**, 193; **69**, 29; **84**, 107; **88**, 53; **89**, 81; **93**, 91; **94**, 89; **101**, 209; **103**, 27; **107**, 231; **111**, 147
Glycodeoxycholic acid **31**, 193; **74**, 95; **111**, 147; **114**, 65; **127**, 239
Glycofurol **33**, 249; **95**, 209; **113**, 83; **119**, 37; **128**, 287
Glycofurol-75 **119**, 37
Glycol **140**, 237
Glycol methacrylate **7**, 189
Glycolamide derivative **43**, 101
Glycolic acid **100**, 9; **115**, 61; **142**, 85
Glycolipid **107**, 99; **115**, 183; **128**, 65
Glycopeptide **144**, 71
Glycophorin **90**, 73
Glycoprotein **28**, 33; **35**, 211; **37**, 33; **94**, 195; **107**, 223; **131**, 159
Glycosamine derivative **14**, 123
Glycosaminoglycan **87**, 21
Glycosylated protein **85**, 75
Glycosylation **61**, 219
Glycylproline **104**, 187
Glycyrrhetinic acid **84**, 279; **122**, 91
18α-Glycyrrhetinic acid **84**, 279
Glycyrrhiza **119**, 133
Glycyrrhizic acid derivative **94**, 161
Glyloid **133**, 1
Glyloid-3S **134**, 173
Gonadorelin agonist **123**, 103
Gonadorelin derivative **16**, 181; **48**, 15; **105**, 7; **110**, 137; **121**, 45; **138**, 167
Goserelin **59**, 173
Gossypol **14**, 123
Gram negative bacterium **34**, 241
Gram positive bacterium **34**, 241
Gramicidin **1**, 165
Gramicidin S **80**, 253
Granulac-200 **146**, 1
Granulac-220 **135**, 53
Granulation **1**, 65; **12**, 163; **31**, 277; **32**, 177; **40**, 73
Granule **7**, 261; **10**, 99; **15**, 97; **31**, 131
Granulocyte **99**, 1; **138**, 85
Granulocyte colony stimulating factor **81**, 133; **114**, 177; **127**, 191
Granulocyte-macrophage-colony stimulating factor **97**, 213
Granuloma **46**, 149
Green S **83**, 263
Griseofulvin **2**, 81; **6**, 179, 195; **13**, 35; **14**, 1; **17**, 77; **25**, 57; **28**, 23; **30**, 161; **33**, 115, 235; **40**, 257; **45**, 111; **47**, 51, 223; **54**, 161; **62**, 143; **63**, 49; **67**, 139, 247; **69**, 13, 53; **70**, 241; **79**, 123; **80**, 191; **82**, 47; **83**, 47, 147; **89**, 119; **90**, 1, 105; **93**, 215; **94**, 1, 31; **98**, 239; **110**, 267; **111**, 159; **122**, 35, 57; **133**, 117; **135**, 13; **136**, 117; **142**, 189; **147**, 135; **148**, 1, 63
Growth **32**, 55
Growth hormone **23**, 13; **34**, 35; **45**, 197; **59**, 173; **74**, 147; **75**, 97; **90**, 81; **128**, 239

Growth hormone releasing factor **106**, 95
Guaifenesin **54**, 15; **103**, 97; **114**, 171
Guaifylline **65**, R5
2-Guanidinothiazole derivative **50**, 117
Guanethidine **20**, 65; **47**, 209
Guanfacine **7**, 19
Guanine derivative **90**, 239; **125**, 55; **139**, 231
Guar gum **19**, 107
Gum arabic **119**, 71
Gum tragacanth **3**, 13; **73**, 163; **95**, 23

H

Halcinonide **70**, 167
Halofenate **6**, 225
Haloperidol **17**, 1; **67**, 169
Haloprogin **7**, 99; **110**, 109
Halothane **149**, 123
Hardness **7**, 327; **44**, 43; **114**, 85
Hartmann solution **22**, 105
HCO 60 **31**, 231, 239
Health care personnel **58**, 25
Heart arrhythmia **22**, 345
Heart muscle contractility **123**, 181
Heart rate **57**, 9
Heart rhythm **22**, 345
Heart transplantation **115**, 87
Heart valve **52**, 139
Heart valve bioprosthesis **59**, 95
Heat **83**, 11; **86**, 263
Heat of solution **40**, 1
Heat sterilization **59**, 131
Heating **118**, 161; **126**, 29
Hematoporphyrin derivative **70**, 129
Hemoglobin **33**, 1
Hemolysis **4**, 75; **7**, 111; **16**, 271; **49**, 163; **50**, 45; **53**, 61; **58**, 41; **110**, 161; **132**, 251; **133**, 171
Heparin **4**, 219; **5**, 167; **9**, 321; **11**, 11; **13**, 9; **18**, 39; **19**, 107; **37**, 65; **48**, 15; **53**, 237; **54**, 251; **59**, 173; **79**, 191; **81**, 1; **82**, 171; **93**, 233; **102**, 19; **107**, 111; **110**, 117; **111**, 191; **120**, 51; **121**, 245
Heparin fraction **18**, 39
Heparin lithium **9**, 321
Heparin lyase **88**, 293
Heparinoid **37**, 65
Heparitinase **88**, 293
Hepatitis B virus **58**, 25
Hepatitis B vaccine **106**, 89
Hepatitis B surface antigen **106**, 89; **115**, 193; **147**, 143
Hepatomegaly **46**, 149
Hepatosplenomegaly **57**, 23
Heptakis(2,6-*o*-dimethyl)-β-cyclodextrin **20**, 155; **35**, 193; **38**, 265; **50**, 45; **57**, 107; **79**, 107; **125**, 17; **125**, 31; **131**, 219; **136**, 165; **137**, 209; **138**, 37; **149**, 1
Heptakis(2,6-*o*-dimethyl)-γ-cyclodextrin **59**, 121
Heptakis(2,3,6-*o*-trimethyl)-β-cyclodextrin **35**, 193
Heptakis(2,3,6-*o*-trimethyl)-γ-cyclodextrin **59**, 121
Heptane **15**, 159; **17**, 77; **27**, 39; **34**, 217; **35**, 173; **123**, 71
Heptanoic acid derivative **126**, 219
Heptanol **1**, 15; **68**, 285; **132**, 23
Herpes labialis **123**, 159
Herpes simplex **123**, 159
Herpes simplex virus **48**, 141; **106**, 85
Herpes simplex virus 1 **65**, 183; **93**, 139
Herpes virus **49**, 205
Herpes zoster **123**, 159
Hesperetin **124**, 1; **145**, 87
Hetastarch **51**, 233; **62**, 27; **78**, 43; **106**, 41; **139**, 25
Heterocyclic compound **137**, 149
Heweten **97**, 15
Hexachlorobenzene **80**, 51
Hexachlorophene derivative **28**, 1
Hexadecanol **3**, 247; **27**, 311; **28**, 47, 113; **31**, 15; **125**, 315; **126**, 119
Hexadecyl-β-D-galactopyranoside **107**, 99
Hexadecylmaltoside **132**, 107
Hexadecylmorpholine **8**, 193
Hexadimethrine bromide **19**, 107
Hexamethylbenzene **15**, 347
Hexane **13**, 75; **15**, 159; **17**, 77; **27**, 39; **29**, 177; **34**, 217; **78**, 17; **82**, 223; **92**, 81; **101**, 63; **113**, 97; **140**, 237
Hexanetriol **15**, 185
Hexanoic acid **20**, 3; **33**, 235; **49**, 15; **86**, 43; **109**, 45; **126**, 219; **129**, 283; **132**, 23; **141**, 71
Hexanoic acid methyl ester **33**, 235; **40**, 257
Hexanol **1**, 15; **13**, 75; **29**, 177; **68**, 285; **126**, 219; **132**, 23; **148**, 41
7-(Hexanoyloxymethyl)theophylline **12**, 299
Hexapeptide **115**, 225; **135**, 191
Hexobarbital **3**, 341; **14**, 123; **18**, 185; **20**, 73; **26**, 157; **92**, 1
Hexobendine **46**, 173
1-Hexyl-2-pyrrolidinone **60**, 177
4-Hexylaniline **100**, 1
Hexylglucoside **105**, 219
Hexylparaben **125**, 141
Hexylresorcinol **18**, 247
High density lipoprotein **96**, 1
High performance liquid chromatography **34**, 231; **35**, 13, 39, 47, 61, 181, 219; **36**, 7, 41, 147; **37**, 97, 257; **46**, 89; **54**, 89; **58**, 103; **63**, 113; **86**, 211; **91**, R1; **93**, 183, 239, 245; **94**, 231; **102**, 271; **103**, 187; **105**, 235; **106**, 167
High pressure liquid chromatography **3**, 93; **4**, 21; **9**, 295; **10**, 119; **11**, 237, 329; **13**, 1, 333; **14**, 73; **15**, 49, 61, 115, 261; **16**, 115, 141, 203; **18**, 53; **19**, 17; **20**, 155, 273, 285; **21**, 35, 211, 239; **22**, 105, 345; **23**, 163, 209, 223, 277, 289, 315; **24**, 91, 173, 193, 219, 239, 307; **25**, 27, 85, 95, 293; **26**, 123, 227, 317; **27**, 1, 185, 205, 219; **28**, 47, 95, 119; **29**, 9, 19, 67, 161, 233; **30**, 29, 57, 151, 199; **31**, 9, 15, 33, 65, 75, 83, 125, 157, 165; **32**, 7, 21, 123, 151, 199; **33**, 89, 225, 235, 249; **34**, 189, 259
Hippuric acid **1**, 323
Hippuric acid derivative **37**, 185
Hisetal **101**, 57; **103**, 77; **107**, 23
Histamine **68**, 191
Histamine H_2 receptor antagonist **81**, 1

Histidine **72**, 149; **85**, 7; **99**, 43; **123**, 41
Histidyl-dextro-tryptophylalanyltryptophyl-dextro-phenylalanyllysinamide **115**, 225; **144**, 91
Histopathology **68**, 239
Histrelin **108**, 49
Hoechst-605331 **90**, 213
Homatropine **1**, 165
Homatropine methyl bromide **9**, 245
Hormone **75**, 117; **75**, 97
Hormone transport **32**, 39
Horseradish peroxidase **101**, 115; **138**, 75; **148**, 231
Hospital infection **1**, 133
HPC H **134**, 173
HPC-101 **123**, 103
Human growth hormone **23**, 13; **63**, 207; **113**, 89; **127**, 115; **128**, 239; **140**, 155
Human immunodeficiency virus **58**, 25
Human immunodeficiency virus infection **93**, 139; **95**, 1
Human insulin **36**, 199; **38**, 83; **95**, 209; **111**, 213; **122**, 91; **123**, 199; **125**, 205; **140**, 77
Human interferon **42**, 261
Human serum albumin **2**, 283; **69**, 5, R1; **103**, 253; **114**, 65; **128**, 197; **132**, 143
Human skin (in vivo) **40**, 105
Humidifier **123**, 47
Humidity **30**, 229; **54**, 131; **62**, 65; **63**, 23; **67**, 163; **73**, 249; **81**, 153; **83**, 59; **94**, 49; **114**, 23; **120**, 83; **135**, 53; **145**, 221
Hyalastin **72**, 131
Hyalectin **72**, 131; **86**, 159
Hyalection **72**, 131
Hyaluronic acid **19**, 107; **50**, 91; **51**, 203, 259; **72**, 131; **80**, 161, 213; **82**, 205; **83**, 1; **85**, 51; **86**, 159; **100**, 199; **106**, 161; **107**, 91; **111**, 205, 295; **118**, 243; **119**, 149; **127**, 115; **129**, 137
Hyaluronic acid benzyl ester **111**, 295
Hyaluronic acid derivative **80**, 161, 213; **87**, 21; **107**, 91; **111**, 295; **129**, 137; **144**, 19
Hyaluronidase **69**, 29; **82**, 205; **88**, 293
Hycanthone **10**, 199
Hydralazine **17**, 161; **47**, 209
Hydration **14**, 29; **20**, 307; **51**, 73; **55**, 129; **56**, 87; **73**, 209; **77**, 269; **80**, 213; **83**, 11; **85**, 149; **91**, 51; **105**, 103; **112**, 29; **129**, 253; **132**, 1
Hydrazone derivative **65**, 109
Hydrocarbon **77**, 47; **86**, 43; **99**, 93; **113**, 257
Hydrochloric acid **29**, 253
Hydrochlorothiazide **5**, 167; **7**, 129, 327; **12**, 341; **17**, 161; **20**, 65; **51**, 169; **56**, 97; **57**, 9; **76**, 49, 187; **86**, 99; **91**, 217; **93**, 111, 245; **100**, 207; **107**, 141; **108**, 207; **110**, 203, 249, 257; **135**, 147; **142**, 61
Hydrochlorothiazide plus propranolol **55**, 25
Hydrochlorothiazide plus triamterene **57**, 9
Hydrocinnamic acid **79**, 135
Hydrocolloid **87**, 229; **91**, 51; **99**, 13; **112**, 29; **117**, 49
Hydrocortisone **1**, 165; **4**, 249; **5**, 167, 229; **7**, 99; **10**, 1, 307; **11**, 119; **13**, 205; **14**, 191; **16**, 255; **17**, 1, 23; **19**, 245; **20**, 43, 139; **21**, 155; **25**, 199; **26**, 77; **27**, 287, 325; **32**, 187; **36**, 67; **42**, 97, 161; **50**, 21, 253; **53**, 43, 53; **56**, 75; **60**, R1; **66**, 79; **71**, 55; **76**, 77; **77**, 231; **78**, 137; **80**, 17, 213; **81**, 89; **82**, 205; **93**, 233; **95**, 161, 181; **103**, 19, 93, R1; **106**, 7; **110**, 169, 211; **116**, 19, 201; **118**, 113, 129; **126**, 73; **127**, 293; **132**, 23; **138**, 95, 149; **139**, 63; **148**, 1
Hydrocortisone acetate **10**, 1, 339; **27**, 287, 325; **47**, 223; **50**, 39; **76**, 1; **91**, 85; **94**, 203; **110**, 231; **138**, 95; **146**, 255
Hydrocortisone 21 acetate **10**, 307; **27**, 325
Hydrocortisone butyrate **27**, 325; **96**, 91; **116**, 19
Hydrocortisone 21 butyrate **27**, 325
Hydrocortisone 17 butyrate 21 propionate **27**, 29
Hydrocortisone caproate **1**, 165
Hydrocortisone derivative **15**, 235
Hydrocortisone 21 diethylsuccinamate **15**, 235
Hydrocortisone 21 diethylsuccinamate 17 valerate **15**, 235
Hydrocortisone ester **27**, 287
Hydrocortisone 21 ester **36**, 67
Hydrocortisone 21 succinamate derivative **15**, 235
Hydrocortisone hexanoate **21**, 155; **27**, 287
Hydrocortisone hyaluronate ester **82**, 205
Hydrocortisone octanoate **21**, 155; **27**, 287
Hydrocortisone-21-octanoate **51**, 39
Hydrocortisone propionate **21**, 155; **27**, 287
Hydrocortisone sodium phosphate **106**, 7
Hydrocortisone sodium succinate **80**, 213; **106**, 7
Hydrocortisone valerate **21**, 155; **27**, 287
Hydroflumethiazide **4**, 67; **17**, 1; **114**, 121
Hydrogel **48**, 1; **58**, 135; **59**, 173; **61**, 79; **70**, 53; **76**, R5; **78**, 43, 77; **80**, 9; **92**, 203; **93**, 233; **100**, 25, 65; **116**, 19; **118**, 207, 237; **127**, 165; **128**, 251; **130**, 83; **133**, 265; **138**, 103, 121; **142**, 169
Hydrogen **99**, R5; **104**, 107
Hydrogen bond **14**, 263; **81**, 187; **113**, 97
Hydrogen peroxide **16**, 127; **49**, 205; **57**, 217; **65**, 261; **72**, 89; **92**, 151; **104**, 151, 247; **121**, 107; **124**, 279; **137**, 143
Hydrogenated castor oil **115**, 217; **130**, 203; **146**, 63
Hydrogenated phospholipid **79**, 263
α-Hydroxyhippuric acid **37**, 185
Hydrolase **113**, 25
Hydrolysis **1**, 49; **4**, 89; **6**, 147; **9**, 7, 29; **10**, 29; **12**, 355; **13**, 89, 219; **17**, 91, 357; **21**, 201, 349; **22**, 197; **24**, 43, 193; **25**, 1, 27; **26**, 15, 227, 339; **27**, 175; **32**, 199; **35**, 39; **46**, 57, 77, 159; **47**, 103; **48**, 91, 159; **50**, 1; **51**, 233; **56**, 159; **58**, 143; **59**, 35; **60**, 99, 163; **62**, 193; **69**, 1; **74**, 85, 221; **75**, R5; **76**, 99, 113; **78**, 157; **80**, R11; **81**, 253; **93**, 75; **99**, 21; **102**, 109; **105**, 169; **123**, 95
Hydromorphone **56**, 1; **138**, 149
α-Hydroxypenicillin G **17**, 47
Hydroperoxide **142**, 215
Hydrophilic compound **15**, 13
Hydrophilicity **62**, 75; **64**, 171; **67**, 113; **68**, 43, R1; **70**, 69; **74**, 157; **81**, 111; **85**, 113, 141; **88**, 99, 125; **94**, 15, 31, 143; **99**, 29; **100**, 263; **103**, R1; **106**, 125; **116**, 95
Hydrophobicity **16**, 271; **31**, 193; **33**, 201; **34**, 131; **58**, 129; **67**, 29; **68**, R1; **70**, 69, 111, 153, 219; **74**, 169; **77**, 151; **87**, 47; **89**, 25, 33; **94**, 103; **103**, R1; **120**, 215; **131**, 83
Hydrotalcite **13**, 313; **52**, 173

Hydroxamic acid **69**, R1
Hydroxide **15**, 213
3-Hydroxy-1,2-dimethyl-4-pyridone **88**, 345
2-Hydroxy-3-(2-hydroxyethylamino)-5,8-dimethoxytetralin **38**, 99
5-Hydroxy-1-methyl-4-nitroimidazole **35**, 165
9-(2 Hydroxy 3 nonyl)adenine **19**, 17
5-Hydroxy-2[*n*-propyl-*n*[2(2-thienyl)-ethyl]-amino]-tetralin **88**, 165
Hydroxyapatite **79**, R1; **86**, 137; **112**, 215
Hydroxybenzoic acid **38**, 257; **81**, 31
3-Hydroxybenzoic acid **45**, 153
4-Hydroxybenzoic acid **11**, 131; **27**, 1; **29**, 237; **32**, 223; **33**, 105; **34**, 125; **109**, 155
Hydroxybenzoic acid derivative **72**, 29; **90**, 95; **109**, 155
4-Hydroxybenzoic acid derivative **35**, 227
4-Hydroxybenzoic acid ester **19**, 1; **29**, 237; **33**, 105; **69**, 103 **72**, 29, 51; **74**, 189; **84**, 233; **90**, 203; **93**, 201; **99**, 21; **109**, 155; **119**, 183; **125**, 141
4-Hydroxybenzoic acid ester derivative **49**, 223
4-Hydroxybenzoic acid heptyl ester **125**, 141
N-(α Hydroxybenzyl)benzamide **22**, 45
4-Hydroxybutyric acid **7**, 169
Hydroxychloroquine **43**, 215
25-Hydroxycholesterol **96**, 1
Hydroxycoumarin **12**, 355
4-Hydroxycoumarin **12**, 231, 355; **59**, 137
Hydroxyethylcellulose **11**, 309; **15**, 251; **45**, 39; **49**, 231; **54**, 95; **57**, 163; **84**, 69; **95**, 229; **101**, 127; **119**, 71, 149; **138**, 225; **139**, 113
Hydroxyethyl-β-cyclodextrin **104**, 239; **109**, 99
N-(2-Hydroxyethyl)-lactamide **61**, 205
2-Hydroxyethyl-methacrylate **39**, 59; **87**, 37; **111**, 127; **120**, 33
1[4(2-Hydroxyethyl)-phenoxy]-3-isopropylamino-2-propanol **66**, 23
N-(2-Hydroxyethyl)-2-pyrrolidone **95**, 181
N-(1 Hydroxyethyl)thiobenzamide **22**, 45
Hydroxyethylmethylcellulose **11**, 131
Hydroxyl radical **57**, 173; **58**, 237; **87**, 79; **92**, 151; **104**, 247; **107**, 199; **115**, 241; **124**, 279; **131**, 1; **137**, 143
16-Hydroxyhexadecanoic acid **121**, 211
4-Hydroxyisophthalic acid **40**, 85
12-Hydroxylauric acid **121**, 211
3-Hydroxymethyl-5,5-diphenylhydantoin-*n*,*n*-dimethylglycine-ester-methanesulfonate **75**, 201; **86**, 175
3-Hydroxymethyl-5,5-diphenylhydantoin acetate **75**, 219
5-Hydroxymethyl-2-furoic acid **12**, 31
7-(Hydroxymethyl)theophylline **12**, 299
1-Hydroxymethylallopurinol aminoacid ester **25**, 27
N-Hydroxymethylbenzamide **22**, 45; **37**, 185
2-Hydroxymethylbenzamide **29**, 9
2-Hydroxymethylbenzamide derivative **29**, 9
Hydroxymethylcellulose **99**, 37; **100**, 241; **145**, 175
Hydroxymethylglutaryl-coenzyme-A-reductase inhibitor **105**, 209; **108**, 195; **125**, 141
3-Hydroxymethylphenytoin sulfate potassium **14**, 113
N-(Hydroxymethyl)thiobenzamide **22**, 45
7-*N*-(4-Hydroxyphenyl)mitomycin C **45**, 189; **105**, 249
Hydroxyphenylacetic acid **81**, 31
5′-Hydroxypiroxicam **24**, 61, 101
Hydroxyprogesterone **50**, 253; **132**, 23; **138**, 95, 149
11-α-Hydroxyprogesterone **42**, 161
17β-(2-Hydroxypropionyl)-17-α-methylestra-4,9(10)-dien-3-one **72**, 1
Hydroxypropyl-cyclodextrin **80**, 243, 253
Hydroxypropyl-α-cyclodextrin **101**, 97
Hydroxypropyl-β-cyclodextrin **91**, 15; **93**, 201; **110**, 137, 169; **136**, 43
2-Hydroxypropyl-β-cyclodextrin **29**, 73; **57**, 63, 107; **59**, 35, 45, 225, 231; **69**, 211; **77**, 303; **78**, 25; **79**, 273, 289; **80**, 243, 253; **85**, 175; **86**, 263; **90**, 187, 247; **98**, 225, 239; **101**, 97; **104**, 181, 239; **106**, 63, 135; **108**, 69; **118**, 77; **121**, 217; **123**, 53, 103; **125**, 17, 31, 91; **127**, 239; **128**, 45; **129**, 63; **131**, 219; **132**, 271; **133**, 191; **136**, 165, 43; **138**, 37; **139**, 141, 215; **141**, 1, 171; **142**, 153; **143**, 203; **145**, 137; **148**, 33
Hydroxypropyl-γ-cyclodextrin **101**, 97
2-Hydroxypropyl-γ-cyclodextrin **79**, 289; **80**, 253; **98**, 239
Hydroxypropyl starch **136**, 107; **37**, 211; **45**, 39; **47**, 21; **49**, 231; **61**, 109; **77**, 71; **78**, 43; **84**, 69; **85**, 29; **86**, 79; **90**, 151; **96**, 119; **99**, 229; **100**, 33; **109**, 209, 221; **111**, 181; **113**, 9; **114**, 137; **119**, 71, 149; **126**, 79, 147; **134**, 173; **136**, 107; **138**, 215; **141**, 109; **143**, 151; **146**, 1, 71; **147**, 119
Hydroxypropylcellulose **11**, 309; **15**, 251; **17**, 347; **20**, 187; **25**, 73; **31**, 43, 175; **34**, 93
N-(2-Hydroxypropyl)-methacrylamide **128**, 99
N-(2-Hydroxypropyl)-methacrylamide copolymer **44**, 213; **87**, 105
Hydroxypropylmethyl cellulose **7**, 285; **11**, 309; **15**, 251; **22**, 17, 219, 265, 273; **24**, 327, 339; **29**, 37; **31**, 55, 169, 175; **34**, 93; **35**, 157; **38**, 65; **39**, 39, 51; **40**, 223; **41**, 83; **43**, 155;, 221; **44**, 125; **45**, 245; **47**, 133, 205; **48**, 51, 63, 113; **49**, 157, 213; **50**, 163; **51**, 19; **52**, 149; **53**, 47, 75; **54**, 95; **55**, 123; **59**, 155; **61**, 101, 127; **66**, 233; **71**, 95, 213; **72**, 163, 243; **73**, 69, 111, 197; **75**, 181; **77**, 89, 177, 183; **78**, 49; **79**, 97, 223; **80**, 63, 179; **81**, 49, 243; **85**, 57; **86**, 159; **88**, 405; **91**, 151, 167; **93**, 21; **94**, 15; **95**, 57; **98**, 57, 157; **99**, 37, 173; **100**, 33, 143, 155, 165, 175, 241, 263; **102**, 207; **103**, 205, 259; **104**, 115, 181; **107**, 91; **109**, 9, 91, 115; **111**, 251; **112**, 17, 37, 191; **113**, 65; **114**, 85; **115**, 201; **116**, 31, 65, 159; **119**, 149, 193; **121**, 11, 129, 239, 249; **123**, 85; **124**, 87, 97; **125**, 185, 289; **126**, 147, 189; **127**, 293; **129**, 21, 233; **133**, 17; **135**, 115; **136**, 43; **138**, 57, 103; **139**, 223; **140**, 85; **141**, 109; **142**, 53; **145**, 157, 175, 197, 231; **146**, 1, 239
Hydroxypropylmethylcellulose acetate succinate **126**, 1
Hydroxypropylmethylcellulose phthalate **28**, 157; **31**, 175; **34**, 93; **35**, 111; **42**, 105; **79**, 97; **121**, 239; **124**, 97; **125**, 151; **132**, 131; **146**, 233
3-Hydroxypyridine derivative **114**, 185
4-Hydroxytamoxifen **99**, 59
Hygroscopicity **62**, 65
Hymecromone **12**, 355
Hyoscyamine **28**, 249
Hyperlipidemia **100**, 279
Hypermer A60 **99**, 29

Hypertension **131**, 229
Hyperthermia **57**, 241; **135**, 167
Hypertonic solution **123**, 41
Hypnorm **38**, 261; **149**, 123
Hypocalcemia **134**, 47; **149**, 93
Hypochlorite sodium **17**, 291; **50**, 87
Hypoglycemia **1**, 121; **5**, 223; **111**, 213; **114**, 137
Hypoprothrombinemia **131**, 131
Hypothermia **9**, 97
Hypoxanthine **42**, 263; **145**, 145

I

Ibufenac **62**, 21
Ibuprofen **2**, 283; **11**, 11; **14**, 59; **15**, 251; **21**, 99; **28**, 95; **32**, 229; **37**, 175; **45**, 139; **49**, 1; **50**, 155; **59**, 115; **61**, 1; **62**, 21; **66**, 193, 243; **68**, 97; **70**, 69; **72**, 219; **73**, 111, 157; **75**, 25; **79**, 21; **83**, 73, 95; **84**, 167; **87**, 95; **89**, 9; **91**, 173; **92**, 1; **92**, 29, 97, 151, 219; **93**, 111; **94**, 59; **99**, 125; **102**, 185; **106**, 25; **107**, 179; **108**, 109; **112**, 37, 233; **113**, 17, 175; **116**, 125, 159; **118**, 129, 191; **119**, 133; **120**, 13; **121**, 107, 205; **123**, 85; **124**, 97; **125**, 157, 243; **126**, 95; **127**, 203; **130**, 25; **133**, 155; **139**, 79, 187; **143**, 211, 223; **147**, 23; **148**, 1
Ibuprofen octyl ester **113**, 175
Ibuproxam **90**, 35
Ibuterol **124**, 149
Idarubicin **32**, 123; **89**, R9
Idoxuridine **1**, 165; **43**, 1; **48**, 141; **123**, 159
Idrorammosan **49**, 147
Ifosfamide **139**, 249
Ileum **46**, 111; **80**, 1
Ileum contraction **142**, 163
Image analysis **144**, 141; **148**, 149
Imidazole **125**, 141; **137**, 149
Imidazole derivative **90**, 247; **125**, 141
4-Imidazolidinone derivative **20**, 273; **23**, 163; **27**, 175; **71**, 45; **76**, 113
Imidazoline derivative **71**, 45
Imipenem **94**, 1
Imipramine **18**, 235; **35**, 129; **54**, 1; **94**, 135; **99**, 67
Immune response **57**, 133; **85**, R5; **88**, 251, 335; **103**, 303; **128**, 23
Immune system **37**, 33
Immunity **86**, 1
Immunization **88**, 335; **90**, 255; **108**, 133
Immunogenicity **85**, R5; **119**, 251; **122**, 121; **147**, 143
Immunoglobulin **75**, 97, 117; **77**, 31; **106**, 249; **119**, 251; **120**, 247; **124**, 165
Immunoglobulin A **108**, 133; **128**, 23
Immunoglobulin Fc fragment **124**, 165
Immunoglobulin G **17**, 65; **98**, 9; **108**, 133; **128**, 23
Immunological adjuvant **115**, 193; **147**, 143
Immunology **1**, 315
Immunomodulating agent **75**, 97, 117; **105**, 235
Immunomodulation **51**, 137
Immunoreactivity **54**, 251; **59**, 27
Immunosuppressive treatment **57**, 23
Implant **54**, 223; **76**, 161; **78**, 165
Imwitor-742 **114**, 103
In vitro diffusion **40**, 93
In vitro skin penetration **40**, 101
In vitro study **19**, 307
Indacrinone **92**, 1
Index **30**, 209
Indicine-*n*-oxide **8**, 45
Indium **25**, 57
Indium-111 **19**, 97; **37**, 245; **68**, 199; **79**, 77; **85**, 75; **87**, 215; **100**, 81; **101**, 263; **108**, 125; **118**, 243; **119**, 223
Indium 113m **19**, 169
Indium hydroxide in 111 **19**, 169
Indole **137**, 149
Indole-3-curcuminoid **131**, 1
Indometacin **4**, 89; **7**, 137; **8**, 203, 263, 303; **10**, 125; **13**, 35, 313; **14**, 1; **16**, 191; **17**, 273; **19**, 35; **24**, 165, 275; **26**, 77; **28**, 11, 47; **29**, 73; **30**, 35; **31**, 157; **32**, 13, 207; **33**, 219; **34**, 75; **39**, 235; **40**, 223; **42**, 257; **46**, 19, 149; **48**, 1; **49**, 79; **52**, 55, 101, 191; **55**, 17, 53, 123, 157, 217, R1; **56**, 249; **60**, 27, 191, 229; **61**, 225; **62**, 105, 157; **63**, 77; **64**, 109, 155; **65**, R1; **66**, 223; **67**, 11, 79, 185, 207; **69**, 211; **70**, 69; **71**, 201; **74**, 85; **75**, 181; **77**, 21; **79**, 21, 263; **80**, 63, 259; **83**, 163, 187; **84**, 141; **85**, 103, 113; **88**, 39; **90**, 161; **91**, 75; **92**, 55, 191; **93**, 21; **94**, 115; **95**, 51; **98**, 235; **100**, 165, 257; **104**, 1, 11, 87; **106**, 25, 63, 115; **112**, 87; **113**, 209; **116**, 1, 201; **118**, 95, 113, 129, 237; **119**, 203; **120**, 63, 145; **123**, 127; **124**, 45, 219; **126**, 95, 111, 147, 161, 275; **127**, 245, 283; **128**, 251; **129**, 103, 113, 233, 279; **131**, 9, 159, 247, 273; **133**, 107; **137**, 117; **139**, 169; **141**, 53; **145**, 241; **146**, 1; **147**, 79; **148**, 1, 201
Indometacin derivative **43**, 101; **77**, 21; **123**, 127; **124**, 45
Indometacin zinc **60**, 27
Indomethacin zinc **120**, 145
Indoprofen **126**, 95
Infection **110**, 189
Infection prevention **1**, 133; **77**, 13
Inflammation **15**, 235; **54**, 149; **58**, 79; **61**, 1; **70**, 35; **104**, 151; **106**, 63
Infrared spectrometry **15**, 235; **18**, 247; **21**, 251; **22**, 345; **23**, 117; **26**, 123, 317; **28**, 11, 95; **35**, 139; **74**, 77
Infrared spectrophotometry **35**, 243
Infrared spectroscopy **63**, 219; **81**, R11; **148**, 191
Infusion **36**, 199; **72**, 57
Infusion bag **10**, 339; **17**, 183; **28**, 211
Infusion fluid **33**, 261; **65**, 49; **70**, 119; **80**, R7
Infusion pump **19**, 75; **144**, 177
Infusion system **17**, 145; **77**, 279; **110**, 197
Inhalation **81**, 199; **106**, 69; **130**, 13, 103; **149**, 51
Injection **77**, 47; **79**, 301; **81**, 193; **109**, 35; **114**, 125
Injection fluid **7**, 111
Injection pain **109**, 45; **114**, 65; **146**, 63
Injection site **47**, 1
Inorganic compound **14**, 123; **135**, 31
Inorganic salt **130**, 179; **134**, 193; **137**, 167; **145**, 129
Inosine **145**, 145
Inspiratory capacity **138**, 45
Instrument sterilization **50**, 87; **133**, 9
Instrumentation **107**, 219

Insulin **1**, 121, 315; **7**, 317; **9**, 165, 173; **12**, 107; **13**, 263; **15**, 13; **23**, 87; **30**, 95, 247; **34**, 157; **36**, 199; **38**, 83; **44**, 197; **47**, 233; **48**, 15; **53**, 227; **54**, 223, 241, 251; **57**, 49; **59**, 173; **61**, 163; **62**, 53, 187; **64**, 89; **65**, 243; **69**, 29; **74**, 95; **75**, 97, 117; **76**, 61; **78**, 1, 9; **80**, 9; **81**, 1; **83**, 1; **84**, 129; **86**, 1; **87**, 125, 255; **88**, 63, 319; **89**, 147; **91**, 29; **95**, 1, 101, 171; **101**, 15, 191; **103**, R1; **105**, 113, 219; **106**, 255; **111**, 147; **114**, 137; **117**, 129; **118**, 129; **119**, 127; **124**, 37, 149; **125**, 205; **126**, 89; **127**, 115; **128**, 215, 287; **131**, 19; **132**, 107, 115; **141**, 39; **142**, 135; **143**, 163; **149**, 123
Insulin blood level **30**, 247
Insulin I^{125} **1**, 315; **30**, 95
Insulin pump **119**, 127
Insulin zinc suspension **1**, 315; **114**, 137
Interaction **19**, 307
Interdroplet potential **64**, 67
Interferon **18**, 117; **34**, 111; **42**, 261; **48**, 15; **75**, 97, 117
α-Interferon **32**, 103; **34**, 111; **46**, 199; **51**, 241; **72**, 97; **104**, 145; **115**, 235; **128**, 89
β-Interferon **18**, 117; **32**, 103; **34**, 111; **64**, 139; **127**, 115
γ-Interferon **123**, 33; **142**, 85
α-2-Interferon **32**, 103
α-2A-Interferon **134**, 99
Interleukin-2 **59**, 173; **96**, 41
Intermedin **101**, 57; **102**, 193; **103**, 77
Intermedin derivative **107**, 23; **135**, 81
Internalization **65**, 219
Interstitial fluid **81**, 143
Intestine **2**, 101; **4**, 271, 281; **5**, 167; **7**, 83, 145; **9**, 107; **23**, 247; **30**, 181; **31**, 231, 239; **34**, 175; **47**, 185; **51**, 137; **64**, 161, 181; **70**, 235; **71**, 55; **91**, 105; **101**, 209; **102**, 177, 261
Intestine absorption **2**, 101; **3**, 13; **4**, 271; **5**, 25; **7**, 83, 145; **14**, 279; **16**, 47, 191, 241; **18**, 259; **19**, 75; **23**, 247; **25**, 255; **28**, 151; **29**, 191; **30**, 95, 181, 225; **31**, 179, 231, 239; **33**, 181; **34**, 111; **36**, 121; **37**, 33, 127; **59**, 263; **76**, R1; **79**, 159; **98**, 83; **101**, 115; **109**, 125, 147; **110**, 47127; **115**, 79; **120**, 41, 127; **122**, 1; **123**, 13, 71; **128**, 129, 155; **132**, 63, 115, 123; **133**, 47; **134**, 15, 47; **140**, 175; **142**, 1; **143**, 101; **145**, 1; **146**, 205
Intestine brush border membrane **141**, 39
Intestine cell **99**, 209
Intestine epithelium **33**, 253
Intestine flora **97**, 133
Intestine fluid **143**, 1
Intestine juice **3**, 299
Intestine lymph **33**, 165; **130**, 57
Intestine mucosa **4**, 219, 249; **20**, 107; **30**, 73; **38**, 211; **48**, 189; **78**, 43; **107**, 133; **113**, 223; **141**, 251; **146**, 247
Intestine peristalsis **7**, 145
Intestine secretion **19**, 75
Intestine transit time **20**, 315; **21**, 167; **37**, 227; **48**, 79
Intestine ulcer **126**, 275; **32**, 13
Intoxication **79**, 213
Intralipid **13**, 9; **24**, 173; **29**, 233; **39**, 33; **44**, 169; **48**, 133; **58**, 13; **64**, 217; **66**, 1; **70**, 251; **77**, 57; **87**, 167; **99**, 1; **107**, 121; **134**, 235; **137**, 249
Intranasal drug administration **31**, 193; **39**, 87; **57**, 49; **74**, 147; **93**, 91
Intraperitoneal drug administration **98**, 165
Intratracheal drug administration **88**, 63
Intravenous drug administration **7**, 111; **144**, 177
Intravenous infusion **1**, 105, 133;
Inulin **13**, 263; **29**, 43; **38**, 9; **44**, 97; **78**, 103; **118**, 129; **129**, 185
Inulin clearance **34**, 231
Iodide **69**, 197
Iodinated poppyseed oil **36**, 121; **123**, 237
Iodine **15**, 159; **35**, 129; **46**, 179; **103**, 199; **131**, 243; **140**, 237
Iodine-125 **65**, 235; **84**, 273; **99**, 275
4-Iodophenol **10**, 89
2-Iodosobenzoic acid **78**, 157; **86**, 43
Ion exchange **85**, 45
Ion exchange resin **40**, 119; **53**, 79; **59**, 9; **87**, 31, 215
Ion transport **12**, 87; **32**, 39; **55**, 237, 265; **123**, 41
Ion **124**, 107; **137**, 243
Ionic strength **15**, 37; **30**, 101; **70**, 53; **75**, R9; **80**, R21; **90**, 151; **96**, 51; **102**, 193
Ionization **12**, 231; **140**, 1; **141**, 63; **54**, 155; **61**, 265; **63**, 167, 237; **83**, 267; **84**, R1; **94**, 11; **96**, 13; **99**, 79
Ionization constant **8**, 25
Ionizing radiation **40**, 173
Ionophore X 537 A **12**, 97
Iontophoresis **101**, 105, R5; **30**, 63; **44**, 197; **49**, 15, 57; **56**, 13; **60**, 133; **61**, 179; **69**, 109; **77**, 107; **92**, 23; **106**, 7; **111**, 73; **117**, 165; **120**, 221; **123**, 159, 173, 181; **124**, 137; **129**, 41; **131**, 19; **141**, 63; **145**, 175
Iopromide **117**, 1; **132**, 9; **140**, 13
Ipecac **7**, 343
Ipratropium bromide **114**, 111
Iproniazid **29**, 73
Ipsalazide **97**, 1
Iris **13**, 183; **66**, 23
Iron **64**, 27; **87**, 79, 203; **100**, 93; **104**, 247; **106**, 229; **132**, 153, 251
Iron chelation **88**, 345
Iron oxide **47**, 205; **48**, 63, 113; **50**, 163; **53**, 47; **65**, 249; **103**, 69; **128**, 269; **147**, 95
Iron protein succinylate **61**, 199
Iron saccharate **61**, 199
Iron salt **127**, 279
Irradiation **29**, 67; **57**, 217; **98**, 63
γ-Irradiation **63**, 155; **65**, 201; **70**, 147; **99**, 37; **106**, 103; **139**, 219; **142**, 125
Irritable bowel **40**, 151
Irritant agent **113**, 159
Isethionic acid derivative **34**, 51
Isoascorbic acid **22**, 147
Isobutanol **95**, 23; **106**, 51; **129**, 51
Isocyclosporin A **82**, 79; **82**, 85
Isoelectric point **71**, 147
Isoleucine **72**, 149
Isomer **81**, 179
Isoniazid **2**, 257; **29**, 195; **30**, 133; **43**, 119; **57**, 235; **70**, 201; **126**, 179; **134**, 15; **148**, 73
Isonicotinic acid **34**, 81

Isooctane **1**, 15; **3**, 275; **15**, 159; **27**, 39
Isopentyl alcohol **141**, 71
Isophane insulin **1**, 315
Isopilocarpic acid **20**, 285; **76**, 171; **88**, 397
Isopilocarpine **10**, 281; **20**, 285; **76**, 171
Isoprenaline **3**, 239; **79**, 21; **83**, 163; **93**, 221; **124**, 149; **130**, 179; **144**, 123; **149**, 85
Isopropamide iodide **8**, 143; **45**, 59
Isopropyl acetate **119**, 139
N-Isopropylacrylamide **118**, 237
Isoquinoline **42**, 97; **137**, 149
Isoquinoline derivative **137**, 149
Isosorbide **1**, 197; **5**, 335
Isosorbide dinitrate **34**, 75; **39**, 183; **65**, 159; **79**, 21; **93**, 1; **97**, 1; **107**, 111; **113**, 189; **124**, 9; **127**, 213; **129**, 211; **131**, 229
Isosorbide-2-nitrate **127**, 213
Isosorbide-5-nitrate **127**, 213
α-Isobutyryloxystyrene **9**, 29
Isosulfan blue **72**, 259
Isotherm **34**, 207
Isothipendyl **35**, 129
Isotonic salt solution **25**, 73
Isotope labeling **69**, 189; **76**, 55; **79**, 77; **98**, 63, 75; **102**, 101
Isotope tracing **1**, 41
Isotretinoin **101**, 249; **119**, 173; **130**, 57; **134**, 157
Isovaleric acid **42**, 181
Isovaleryl-phenyl-alanyl-nor-leucyl-statinyl-alanyl-statine **115**, 45
Isoxsuprine **130**, 179
Ispagula **19**, 107; **148**, 117
Itazigrel **102**, 133
Ivermectin **103**, 93

J

Jejunum **18**, 259; **20**, 119; **37**, 127
Jet injection **137**, 75
Josamycin **88**, 151

K

Kabimix **134**, 235
Kanamycin **81**, 1; **90**, 59; **99**, 189; **121**, 55
Kaolin **5**, 223; **7**, 83; **18**, 157; **19**, 233; **27**, 17; **28**, 239; **30**, 225; **31**, 43; **34**, 207; **36**, 185; **37**, 211; **89**, 75; **132**, 131
Kaolin pectin **47**, 209; **89**, 75
Karaya gum **139**, 53
KE-298 **124**, 129
Keratin **4**, 309; **24**, 209; **125**, 265
Keratinization **47**, 13
Keratinocyte **68**, 285; **99**, 67
Keratitis **123**, 173
Ketamine **25**, 147; **92**, 1
Ketanserin **87**, 161
Ketastarch **62**, 27
Ketoacidosis **1**, 121
Ketobemidone **84**, 253; **88**, 221, 229, 237, 243
Ketoconazole **64**, 161; **96**, 1; **142**, 183; **143**, 203
Ketoprofen **13**, 243; **14**, 251; **17**, 273; **34**, 189; **44**, 9; **49**, 213; **50**, 97; **65**, 85; **70**, 191; **74**, 115; **77**, 177; **79**, 21; **86**, 113; **88**, 1, 433; **99**, 109; **105**, 31; **106**, 25; **108**, 77, 101; **109**, 107; **113**, 65; **119**, 71; **121**, 149; **124**, 279; **126**, 95; **135**, 177; **144**, 195; **145**, 29; **147**, 71
Ketoprofen derivative **135**, 177
Ketorolac trometamol **41**, 105, 95; **96**, 231
Ketotifen fumarate **141**, 71
Khellin **57**, 223
Kidney **16**, 191; **80**, R15
Kidney cancer **134**, 99
Kidney clearance **120**, 111
Kidney failure **9**, 263; **23**, 345
Kidney injury **5**, 117
Kinetics **1**, 95; **2**, 275, 295, 337; **4**, 171; **7**, 119, 197; **8**, 45; **10**, 1, 281; **11**, 71; **12**, 251; **13**, 273; **14**, 73, 291; **15**, 37; **16**, 141; **17**, 237; **18**, 247; **19**, 1, 35; **21**, 107; **60**, 83; **63**, R5; **72**, 19, 89; **76**, 145; **115**, 175, 183, 241
Kinetin **68**, 135
Klebsiella edwardsii **26**, 133
Klebsiella pneumoniae **9**, 49
Kleptose **69**, 211; **101**, 71; **106**, 63, 179; **123**, 53; **124**, 231; **131**, 219; **136**, 43, 165; **141**, 117
Kupffer cell **70**, 225
Kyotorphin **57**, 77

L

L-44-O **84**, 93
L-679336 **125**, 141
L-683453 **136**, 147
Labetalol **63**, 73; **130**, 179; **130**, 179
Labrafac-CM-6 **106**, 15
Labrafac-CM-8 **106**, 15
Labrafac-CM-10 **106**, 15
Labrafac Hydro **106**, 15
Labrafac hydrophile **121**, 89
Labrafil **49**, 199
Labrafil-M-1944-CSD **106**, 15
Labrafil-M-2125-CS **96**, 147; **106**, 15
Labrafil-NA-10 **106**, 15
Labrasol **105**, 227; **106**, 15; **118**, 221
Lachesine **1**, 165
Lacrimal fluid **60**, 219; **80**, 161
β-Lactam antibiotic **82**, 157; **83**, 123; **89**, 103; **111**, 31
β-Lactamase **82**, 53
Lactase **79**, 257; **107**, 219
Lactate dehydrogenase **104**, 73; **109**, 17; **114**, 197
Lactate sodium **61**, 205; **113**, 247
Lactic acid **33**, 235; **81**, 31; **100**, 9; **108**, 225; **115**, 61; **141**, 71
Lactic acid ethyl ester **33**, 235
Lactone **113**, 223
Lactone derivative **138**, 1
Lactosaminated albumin **80**, 101
Lactose **1**, 65, 183, 249; **2**, 145; **3**, 127; **7**, 269; **10**, 43, 57, 99, 209; **11**, 131; **14**, 1, 231; **15**, 73, 293; **16**, 345, 349; **17**, 313; **20**, 171; **22**, 127; **24**, 355; **25**, 225; **26**, 165; **28**, 23; **30**, 237; **31**, 15, 99, 277; **33**, 249; **35**, 29, 157, 235; **36**, 29, 205; **37**, 15;

38, 231; **39**, 201, 207; **40**, 15, 223; **43**, 225; **44**, 125, 151, 177; **45**, 79, 111; **46**, 35; **47**, 89; **48**, 51; **49**, 1, 209; **51**, 9, 161, 213; **52**, 91; **53**, 47; **54**, 131; **55**, 135, 175; **57**, 205, R1; **59**, 145; **62**, 229; **63**, 23; **64**, 35; **66**, 47, 207, 273, 283; **67**, 79, 231; **68**, 157; **69**, 239; **70**, 241; **71**, 81, 127, 201; **72**, 111, 163, 233; **73**, 209; **75**, 181, 211; **76**, 9, 49; **77**, 269; **79**, 47; **81**, 75, 153, 205, 225; **83**, 11, 187; **84**, 191; **85**, 121; **86**, 187; **87**, 149; **88**, 141, 159; **89**, 1, R17; **90**, 213, R1; **92**, 81, 137, 143; **93**, 249; **94**, 49, 181; **95**, 29, 117, 201; **96**, 59; **97**, 29, 79, 101, 119, 161, 195; **99**, 181, 197; **101**, 45, 127; **102**, 1, 117; **103**, 55; **104**, 135; **105**, 125, 255; **106**, 149, 157; **107**, 29, 141, 219; **109**, 155, 173; **110**, 203, 249, 257; **112**, 225; **114**, 23, 55, 85; **116**, 179, 191; **117**, 253; **118**, 95, 151, 213, 229; **119**, 25, 65, 71, 203; **120**, 101; **120**, 95; **121**, 81, 187; **123**, 265; **124**, 27; **125**, 157; **127**, 95; **129**, 1, 253; **130**, 149, 213; **131**, 201; **132**, 131, 221; **133**, 1, 29, 127, 139; **134**, 105, 243; **135**, 1; **136**, 13, 71, 141; **137**, 41; **139**, 125, 139, 149; **140**, 207; **141**, 161; **142**, 53, 61, 97; **143**, 233; **144**, 47, 153; **145**, 65, 197, 221; **146**, 1, 101, 271; **148**, 149; **149**, 51

α-Lactose **35**, 139; **131**, 209; **135**, 53; **136**, 13; **141**, 93

β-Lactose **136**, 13

Lactulose **87**, 215

Lafarquim **26**, 317

Lafarquim derivative **26**, 317

Lafrabac hydrophile **132**, 71

Lameform **99**, 29

Lamotrigine **144**, 171

Lanatoside A **2**, 127

Lanatoside B **2**, 127

Lanatoside C **2**, 127

Lanolin **16**, 163; **26**, 243; **28**, 113; **35**, 91, 103

Lanolin alcohol **3**, 247

Lanthanum **118**, 129

Large intestine **13**, 321

Lasalocid **12**, 97; **54**, 41

Laser **66**, 131; **115**, 69; **125**, 45

Lasma **52**, 63

Latamoxef **44**, 205; **50**, 233; **60**, 203; **62**, 165

Latex **60**, 109; **63**, 177; **73**, 185; **86**, 239; **87**, 117; **90**, R9; **94**, 223; **103**, 47; **117**, 209; **125**, 243; **130**, 187; **133**, 203; **141**, 179

Lattice disruption **40**, 1

Lauric acid **4**, 271, 281; **11**, 27; **33**, 99, 225; **37**, 251; **48**, 103; **49**, 15; **67**, 97; **84**, 117; **99**, 109; **106**, 1; **110**, 137; **111**, 213; **123**, 1; **126**, 119, 267; **130**, 121; **145**, 215; **148**, 1

Laurocapram **27**, 89; **28**, 201; **31**, 231, 239; **33**, 37; **43**, 1, 207; **46**, 19; **47**, 43; **49**, 47, 199; **51**, 47; **53**, 61; **55**, 17; **56**, 1, 181; **57**, 29; **65**, 183; **66**, 53, 79; **68**, 239; **70**, 173; **71**, R9; **72**, 219; **76**, 37, 151, 217; **80**, 81; **81**, 39; **82**, 171; **85**, 7; **88**, 165; **90**, 129; **91**, 9, 85; **92**, 89; **98**, 113, 173; **99**, 253; **101**, 57; **103**, 77; **104**, 61; **110**, 137, 231; **116**, 201; **118**, 113; **125**, 63; **128**, 251; **129**, 33; **130**, 225; **137**, 125; **141**, 17; **143**, 37; **144**, 81

Laurocapram derivative **91**, 85

Lauromacrogol **10**, 109; **11**, 277; **24**, 79; **28**, 47; **33**, 45; **45**, 91; **50**, 27; **79**, 123; **82**, 223

N-Lauroylsarcosine **93**, 1; **113**, 159

Lauryl alcohol **72**, 219; **92**, 89; **99**, 253; **100**, 9; **125**, 7; **141**, 71; **148**, 41

N-Lauryl-β-D maltoside **134**, 47

Lauryl glucoside **79**, 159

Lauryl lactate **105**, 227

Lauryl maltoside **79**, 159

Lavage fluid **33**, 261

Lavender oil **111**, 235

Laxative **123**, 1

Leaching **78**, 115; **113**, 241

Lead **54**, 143

Lecithin **6**, 137; **8**, 101; **11**, 27; **13**, 263; **19**, 177; **23**, 1, 299; **25**, 57; **27**, 245; **29**, 53; **30**, 161; **32**, 39

Lecithin I'obnbop|125 **20**, 347

Lectin **107**, 223; **110**, 127; **114**, 227; **120**, 247; **128**, 65; **138**, 175

Lectin binding site **138**, 175

Lectin receptor **138**, 175

Leishmania donovani **40**, 215; **57**, 23

Leishmaniasis **53**, 129; **68**, 111; **126**, 253

Leprostatic agent **80**, 277

Lethargy **12**, 277

Leucine **72**, 149; **115**, 139; **121**, 157; **132**, 63

Leucine enkephalin **76**, 113; **117**, 181

Leucine-4-methoxy-2-naphthylamide **30**, 73

Leucine-4-nitroanilide **48**, 189

Leucyl-2-naphthylamide **13**, 183

Leukemia **21**, 219; **111**, 25; **128**, 139

Leukemia cell **65**, 57; **128**, 139

Leukemia l 1210 **15**, 49; **21**, 219; **22**, 75; **36**, 147; **37**, 135

Leukemia P 388 **15**, 49; **37**, 135; **65**, 219; **138**, 237

Leukemia P 388 cell **98**, 121; **101**, 97

Leukocyte count **101**, 89; **81**, 133

Leukotriene-receptor blocking agent **118**, 221; **120**, 235; **134**, 59

Leukotriene synthesis inhibitor **121**, 73

Leupeptin **89**, 169; **113**, 1

Leuprorelin **54**, 231; **61**, 135; **67**, 67; **75**, 97, 117; **90**, 141; **105**, 113; **107**, 57; **108**, 49; **120**, 41

Levallorphan **19**, 75

Levamisole **35**, 129; **39**, 255; **51**, 137; **91**, 95; **132**, 153; **145**, 77

Levobunolol **82**, 39; **100**, 219; **142**, 103

Levodopa **6**, 225; **62**, 207; **72**, R5; **79**, 21; **92**, 1; **97**, 1; **99**, 135; **104**, 157; **116**, 39; **136**, 117

Levodopa derivative **99**, 135

Levodopa plus benserazide **62**, 207

Levomepromazine **35**, 129; **94**, 135

Levonorgestrel **58**, 93

Liarozole **99**, 51

Librafil-M2125-MS **96**, 147

Lichen planus **95**, 105

Lidocaine **2**, 317; **5**, 167; **8**, 81, 89; **12**, 77; **14**, 223; **20**, 87; **29**, 147; **44**, 65; **47**, 1; **54**, 1; **62**, 249; **71**, 167; **79**, 21, 113; **86**, 137; **88**, 9; **93**, 233; **99**, 13; **101**,; **116**, 77; **123**, 47, 159; **131**, 219; **133**, 1; **136**, 165; **141**, 63

Ligand **67**, 155

Light **31**, 65; **43**, 215; **47**, 239; **50**, 219

Light exposure **107**, 199

Light scattering **105**, 235; **134**, 59

Limonene **61**, 225; **74**, 115, 157; **103**, 179; **105**, 31; **111**, 191; **127**, 9; **128**, 251; **130**, 141, 169; **143**, 37; **145**, 37, 49; **146**, 41
Limulus lysate test **10**, 133
Lincomycin **51**, 55
Lindane **7**, 99
Linear model **17**, 23
Linoleic acid **4**, 271, 281; **24**, 79; **37**, 127; **42**, 97; **56**, 1; **88**, 89; **98**, 203; **106**, 15; **109**, 147; **111**, 213; **117**, 219; **119**, 173; **126**, 267; **130**, 57
Linolenic acid **4**, 281; **126**, 267
Lipid **2**, 101; **8**, 193; **12**, 97; **23**, 69; **33**, 165; **37**, 127; **51**, 241; **68**, 1, 277, 281; **69**, 63; **76**, 145; **83**, 131; **84**, 205; **86**, 137; **90**, 59; **93**, 189; **95**, 43; **99**, 219; **106**, 85; **114**, 43; **115**, 129; **121**, 149
Lipid A **111**, 15
Lipid bilayer **57**, 29; **70**, 209; **71**, R1; **74**, 137; **76**, 217; **77**, 127; **79**, 141; **88**, 1; **98**, 173; **105**, 259; **126**, 49; **131**, 103
Lipid composition **106**, 89; **127**, 273; **131**, 103
Lipid emulsion **34**, 173; **39**, 33; **42**, 243; **46**, 177; **54**, 191; **58**, 13; **63**, 183; **64**, 67; **77**, 57; **107**, 121; **109**, 73; **113**, 175; **134**, 119, 235; **137**, 249
Lipid membrane **19**, 107; **21**, 75; **37**, 251; **38**, 99; **42**, 97; **52**, 115; **59**, 19; **66**, 131; **70**, 43, 111; **93**, 21; **103**, 217
Lipid monolayer **51**, 241
Lipid peroxidation **77**, 31; **90**, 221; **91**, 209; **100**, 93; **131**, 1; **132**, 251
Lipid solubility **15**, 347; **20**, 73, 247; **129**, 203
Lipofundin **83**, 227; **99**, 1; **101**, 175; **107**, 121; **134**, 235
Lipogenesis **54**, 251
Lipoid E80 **145**, 17
Lipolysis **11**, 11
Lipopeptide **137**, 33; **138**, 167
Lipophilicity **11**, 209; **12**, 231; **19**, 129; **23**, 209; **25**, 95; **32**, 79, 223; **37**, 49; **51**, 73, 95; **56**, 273; **57**, 229; **60**, 1; **64**, 127, 161, 171; **66**, 39, 183, 213; **68**, 215; **69**, 143; **70**, 173, 219; **71**, 105; **73**, 259; **74**, 59, 105, 157; **77**, 151; **79**, 89; **81**, 111, 235; **84**, 93, 175; **85**, 113, 189; **87**, 141; **88**, 211, 243; **89**, R9; **90**, 141; **91**, 157, 235; **99**, 21, 29; **101**, 209; **113**, 47; **116**, 95
Lipopolysaccharide **35**, 227; **111**, 15; **144**, 147
Lipoprotein **60**, 219; **110**, 277
Lipoprotein lipase **18**, 39
Lipoprotein metabolism **57**, 223
Lipoprotein receptor **96**, 1
Lipoprotein secretion **57**, 223
Liposome **8**, 101; **13**, 263; **14**, 191; **16**, 79, 305, 339; **17**, 91, 135, 215; **19**, 263; **20**, 119, 139, 347; **22**, 299; **23**, 1, 299; **26**, 303; **27**, 287; **32**, 237; **34**, 163, 179; **35**, 263; **36**, 147; **39**, 213;; **40**, 51, 81; **42**, 69; **43**, 257; **44**, 235; **46**, 25, 31; **47**, 129; **48**, 125; **49**, 39, 249; **51**, 39, 241; **56**, 23, 103; **57**, 125, 133, 211, 241; **58**, 49, 57; **59**, 19; **60**, 219; **62**, 1, 75, 133; **63**, 227; **64**, 89; **65**, 235; **67**, 123; **68**, 121; **69**, 77; **70**, 1, 43, 209, 225; **72**, 259; **74**, 183; **75**, 1, 97, 117; **76**, 217; **77**, 31, 119, 127; **78**, 227; **79**, 213; **80**, 93; **81**, 235; **82**, 53; **84**, 93; **85**, 97; **87**, 183; **88**, 1, 53, 201, 313, 327, 335, 409; **89**, 133, 157, 183; **90**, 59, 73, 221; **91**, 1, 69, 209; **95**, 1, 43, 51, 105; **96**, 51, 67, 157; **97**, 205; **98**, 9, 19, 131, 149, 203; **99**, 145, 219; **101**, 191, 227; **102**, 25, 35, 151, 159; **103**, 81, 131, 217, 237, 277; **105**, 189, 259; **106**, 115; **107**, 1, 99, 149, 189; **108**, 21; **110**, 161; **111**, 15, 103; **112**, 249; **113**, 141; **114**, 33, 43, 221; **115**, 183, 193, 209; **116**, 87; **117**, 1, 85, 159, 225, 237; **118**, 23, 65, 207; **119**, 231; **121**, 1, 195; **123**, 33; **124**, 1, 19, 149, 261; **125**, 31, 73, 231; **126**, 41, 49, 209, 235; **127**, 1, 273; **128**, 65, 89, 139, 163; **129**, 191; **130**, 1; **131**, 1, 57, 91, 257; **132**, 9, 263; **133**, 89, 171; **134**, 71, 89; **138**, 113, 139, 191; **139**, 69, 197; **140**, 13, 119; **141**, 17; **143**, 1, 171; **144**, 11, 131; **145**, 87, 95, 231; **146**, 31, 193; **147**, 109, 143, 153; **149**, 15
Liposome membrane **4**, 281
Liposyn **28**, 193
β-Lipotropin **50**, 261
Lipoxol-400 **139**, 87
Lipoxol-6000 **139**, 87
Lipoxygenase inhibitor **115**, 95; **123**, 217; **147**, 207
5-Lipoxygenase inhibitor **123**, 217
Lipsomal entrapment **40**, 143
Lipvenos **99**, 1
Liquid **57**, 17; **79**, 47; **82**, 145; **92**, 81; **118**, 213
Liquid chromatography **1**, 185, 281; **3**, 1; **19**, 271; **34**, 1; **63**, R1
Liquid membrane **24**, 297; **32**, 39; **38**, 239
Liquid paraffin **63**, 219; **91**, 59; **111**, 217; **126**, 119; **132**, 131; **137**, 243
Liquid petrolatum **22**, 131; **27**, 311; **31**, 281
Lisinopril **75**, 117; **107**, 77
Literature **21**, 135
Lithium **43**, 145
Lithium carbonate **3**, 81; **48**, 1
Lithium chloride **1**, 141; **105**, 103
Lithium sulfate **3**, 39, 179, 187
Liver **1**, 41; **8**, 131; **12**, 135; **20**, 73; **25**, 155; **29**, 53; **36**, 253; **44**, 265; **47**, 111; **65**, 7
Liver cell **42**, 69; **46**, 31; **53**, 67, 249; **54**, 259; **58**, 41; **67**, 123
Liver cell carcinoma **54**, 113; **146**, 31
Liver cell culture **57**, 223
Liver homogenate **41**, 121
Liver metabolism **114**, 43
Liver metastasis **54**, 27
Liver perfusion **80**, 101; **149**, 15
Liver protection **44**, 265
Liver scintiscanning **69**, 77
Liver toxicity **53**, 249
LL-D49194-α_1 **112**, 97
Local anesthesia **62**, 9; **94**, 75
Local anesthetic agent **12**, 77; **84**, 33; **131**, 219; **136**, 165
Lomefloxacin **63**, 237; **79**, 89; **83**, 267; **87**, 203; **88**, 379; **93**, 121, 131
Lomustine **62**, 243; **145**, 9
Lonapalene **35**, 61; **44**, 225
Loperamide **51**, 157; **119**, 1
Lorazepam **16**, 327; **21**, 211; **40**, 119; **54**, 171; **61**, 189
Lorcainide **54**, 1
Low density lipoprotein **21**, 219; **63**, 121; **99**, 275
Low-molecular-weight plasminogen activator **134**, 193
Low-molecular-weight protein **80**, R15

Lubricating agent **2**, 145; **42**, 89, 171; **46**, 211; **67**, 39; **70**, 103; **86**, 131; **117**, 13; **126**, 57
Lubrication **3**, 127; **26**, 191; **30**, 209; **31**, 131; **40**, 15; **48**, 149; **144**, 1
Lucanthone **10**, 199
Lucifer yellow **146**, 11
Ludipress **95**, 201; **103**, 155
Lung **1**, 41; **8**, 131; **12**, 135; **26**, 113, 303; **63**, 227; **95**, 111; **107**, 57; **115**, 95
Lung alveolus macrophage **78**, 227
Lung deposit **42**, 211; **64**, 1; **91**, 111; **97**, 47; **102**, 11, 127
Lung disease **124**, 149
Lung function **81**, 199; **87**, 183; **102**, 127
Lung injury **104**, 151
Lung lavage **88**, 201; **97**, 171
Lung parenchyma **128**, 55; **147**, 207
Lung resistance **87**, 183
Lung surfactant **103**, 119; **132**, 89
Lung toxicity **87**, 183
Lusopress **99**, 351
Lutrol **114**, 121
LY-231617 **140**, 33
Lymph **13**, 321; **69**, 169
Lymph circulation **24**, 185
Lymph flow **31**, 185
Lymph node **2**, 245; **72**, 259; **98**, 9
Lymph node metastasis **2**, 245
Lymphatic system **8**, 291; **24**, 79, 173; **31**, 185; **34**, 175; **110**, 155; **144**, 61
Lymphoblastoid cell **34**, 111
Lymphocyte **63**, 135; **75**, 1
Lymphoid tissue **75**, 1
Lysine **9**, 321; **72**, 149; **82**, 179; **88**, 111; **89**, 111; **99**, 43; **105**, 209; **121**, 157; **123**, 41
Lysine acetylsalicylate **16**, 241
Lysol **29**, 177
Lysophosphatidylcholine **56**, 29; **57**, 49; **63**, 207; **65**, 243; **74**, 147; **76**, 61; **101**, 15; **114**, 141, 151; **117**, 139; **132**, 89; **141**, 151; **143**, 67
Lysophosphatidylglycerol **77**, 75
Lysozyme **47**, 79; **75**, 97; **80**, R15; **90**, 95; **115**, 259; **133**, 257; **148**, 131
Lysosome enzyme **107**, 133; **133**, 257

M

Macrogol **7**, 145; **10**, 25; **15**, 13, 25, 185; **17**, 39, 347, 351; **19**, 107, 297; **24**, 61, 165; **25**, 359; **28**, 11; **28**, 201; **31**, 55, 157, 175, 271; **35**, 1, 7; **39**, 235; **40**, 181; **42**, 27; **47**, 51; **48**, 1, 207; **51**, 225; **57**, 197; **67**, 201; **69**, 53; **70**, 9, 261; **72**, R1; **73**, 69, 197; **74**, 33, 43; **76**, 17; **82**, 215; **83**, 47; **84**, 161; **85**, 103; **88**, 125, 327; **90**, 105; **94**, 103; **95**, 153; **99**, 337; **104**, 145; **105**, 189; **106**, 15, 109, 149; **111**, 51; **113**, 17, 65, 103, 209; **118**, 169; **119**, 203; **120**, 73; **121**, 89; **122**, 27; **123**, 1; **124**, 183; **125**, 73; **126**, 41; **127**, 191; **130**, 25, 83; **131**, 229; **132**, 9, 95, 195; **134**, 119, 129, 147; **135**, 115; **138**, 1; **139**, 187; **142**, 115; **143**, 247; **144**, 11; **146**, 31; **147**, 187; **148**, 1
Macrogol 200 **30**, 1; **31**, 55, 175; **55**, 247; **121**, 19; **131**, 183
Macrogol 300 **31**, 157
Macrogol 400 **19**, 149; **22**, 265; **28**, 113; **31**, 55, 175; **33**, 225; **78**, 95; **90**, 187; **103**, 259; **108**, 207; **112**, 191; **120**, 189, 197; **121**, 19, 27; **126**, 119; **127**, 203; **129**, 279; **130**, 203; **131**, 183; **137**, 95; **138**, 57; **139**, 87; **144**, 99, 217; **147**, 31, 207; **148**, 41
Macrogol 600 **31**, 55, 175; **77**, 193; **83**, 211; **131**, 183
Macrogol 1000 **109**, 17; **131**, 183
Macrogol 1500 **112**, 191
Macrogol 2000 **139**, 125, 139
Macrogol 3000 **28**, 201; **54**, 161; **110**, 267; **132**, 221; **134**, 105; **135**, 13; **139**, 125, 139, 149; **142**, 189; **145**, 197
Macrogol 3350 **118**, 221
Macrogol 3400 **78**, 175
Macrogol 4000 **84**, 197; **98**, 29; **101**, 115; **112**, 191; **113**, 223; **118**, 169; **123**, 187; **126**, 155; **130**, 25; **137**, 103; **142**, 163; **147**, 199; **148**, 123
Macrogol 6000 **28**, 11; **31**, 15, 55, 175; **33**, 219; **34**, 93; **37**, 119; **44**, 125, 249; **50**, 163; **78**, 175; **89**, 119; **93**, 215; **94**, 69; **95**, 117; **96**, 5; **99**, 321; **102**, 207; **103**, 137; **108**, 101; **110**, 223; **114**, 121; **118**, 151, 169; **119**, 71; **123**, 53, 273; **126**, 155; **127**, 261; **130**, 25; **132**, 153, 221, 271; **134**, 105; **138**, 67; **139**, 87, 125, 139; **142**, 189; **147**, 199
Macrogol 8000 **29**, 169; **31**, 43; **118**, 169; **139**, 125, 139; **147**, 1; **148**, 139
Macrogol 10000 **78**, 175; **90**, 213; **118**, 169; **139**, 125, 139
Macrogol 20000 **28**, 11; **78**, 175; **118**, 169; **139**, 125, 139
Macrogol derivative **79**, 251; **111**, 63; **128**, 287; **131**, 183; **132**, 221; **134**, 105; **139**, 125, 139; **142**, 189; **143**, 75, 83; **144**, 99; **146**, 81; **147**, 71, 199
Macrogol stearate **33**, 45; **50**, 141; **81**, 67, R5; **99**, 29; **132**, 1
Macrogol stearate 40 **7**, 145; **29**, 169
Macrolide **88**, 151; **110**, 223; **113**, 215; **114**, 65
Macromolecule **27**, 205, 219; **29**, 43; **35**, 39; **65**, 7; **105**, 113
Macrophage **23**, 69, 299; **29**, 53; **30**, 215; **32**, 165; **45**, 169; **46**, 149; **52**, 183; **65**, 249; **68**, 265
Macrophage activation **23**, 299
Mafenide acetate **80**, 213
Magaldrate **28**, 239; **52**, 173; **69**, 189
Magnesium **13**, 9; **22**, 283; **61**, 109; **79**, 159; **87**, 73, 203; **90**, 229; **93**, 121; **105**, 209; **106**, 229; **107**, 121; **114**, 221; **123**, 187; **126**, 199; **139**, 105
Magnesium carbonate **9**, 315; **19**, 307; **29**, 253; **34**, 207; **36**, 185; **49**, 1; **52**, 173; **60**, 197; **90**, 203
Magnesium chloride **1**, 141; **87**, 239; **93**, 131; **105**, 103; **134**, 193
Magnesium hydroxide **19**, 169; **22**, 257; **28**, 151, 239, 249; **34**, 207; **73**, 171
Magnesium oxide **19**, 307; **21**, 341; **28**, 239; **147**, 95
Magnesium palmitate **23**, 55
Magnesium stearate **1**, 249; **2**, 145; **3**, 127; **7**, 269, 327; **12**, 341; **13**, 35, 287; **14**, 1; **18**, 287; **21**, 99; **22**, 127; **23**, 55; **24**, 327, 339; **26**, 191; **31**, 15, 131; **35**, 29, 157; **36**, 51; **37**, 119; **39**, 201; **42**, 89, 171; **43**, 249; **44**, 43; **45**, 79, 111, 259; **46**, 183, 211; **47**, 89; **48**, 51; **51**, 9, 63; **53**, 47; **55**, 175; **57**, 139; **60**, 61; **62**, 125; **64**, 27; **67**, 39; **69**, 239; **70**, 103, 261; **72**, 43; **76**, 49; **78**, 203; **79**, 257; **86**, 131; **88**, 261; **90**, 203; **91**, 217, 241; **94**, 39; **96**, 231; **97**, 195; **99**, 173; **101**, 45; **102**, 81; **110**,

37, 249; **114**, 23; **116**, 65; **119**, 71; **121**, 73, 81; **123**, 119; **126**, 57, 83; **132**, 131; **139**, 53, 79; **144**, 1, 187; **146**, 271
Magnesium trisilicate **7**, 83; **18**, 157; **19**, 307; **20**, 99; **28**, 249; **29**, 253; **30**, 225; **34**, 207; **36**, 185; **37**, 211; **47**, 209; **49**, 1; **65**, 63; **67**, 89; **68**, 11; **100**, 249; **105**, 97; **128**, 129
Magnetic field **61**, 109
Magnetite **43**, 167; **44**, 49; **47**, 265; **52**, 87; **130**, 49; **138**, 85
Maize starch **128**, 1
Malaria **26**, 267; **34**, 137; **69**, 83
Malathion **7**, 99; **17**, 23
Maleimide derivative **135**, 13
Malonaldehyde **108**, 95
Malonic acid derivative **79**, 29; **148**, 165
Malta gran 10 **114**, 23
Maltodextrin **86**, 131; **107**, 199; **114**, 23, 55; **132**, 183; **133**, 155; **144**, 1
Maltodextrin maltrin M 510 **132**, 183
Maltodextrin maltrin qdM 500 **132**, 183
Maltose **49**, 1; **65**, 235; **132**, 107, 115
Maltosyl-β-cyclodextrin **125**, 17; **126**, 73
Maltrin **103**, 155
Maltrin M500 **114**, 23
Maltrin M510 **114**, 23
Mandelic acid **81**, 31; **86**, 221
Mandrax **6**, 205
Manganese pentetate stearate **49**, 249
Mannitol **16**, 319; **19**, 297; **20**, 163; **21**, 17; **22**, 127; **23**, 247; **26**, 329; **32**, 79; **35**, 1; **37**, 15; **38**, 83; **45**, 111; **49**, 1; **55**, 185; **60**, 203; **63**, 281; **66**, 149; **69**, 109; **71**, 55; **72**, 97; **88**, 111, 141; **90**, 1; **95**, 117, 209; **96**, 197, 205, 217; **97**, 161; **98**, 83, 141; **99**, 209; **101**, 63, 81; **107**, 199; **113**, 133; **116**, 147; **117**, 165, 173; **118**, 129, 151; **119**, 71; **123**, 25, 187; **125**, 165, 195; **127**, 95; **129**, 1; **130**, 149; **133**, 47; **138**, 67; **139**, 149; **141**, 205; **142**, 97, 115; **143**, 151; **145**, 9
Mannityl tetramesylate **108**, 69; **124**, 205
Mannosamine **87**, 105
Mannose **70**, 225
Manucol **107**, 179
Manugel **107**, 179
Mass **124**, 241
Mass spectrometry **29**, 73, 113; **34**, 247; **35**, 139, 259
Mast cell **68**, 191
Mathematic model **1**, 121; **17**, 193; **32**, 143, 159
Mathematical model **37**, 15, 41; **56**, 111; **73**, 147
Mathematics **79**, 1; **80**, R21**134**, 1
Matrix tablet **146**, 175
Maxepa **24**, 173
MDL 201040 **111**, 89
MDL 201346A **135**, 43
MDX 44210 **57**, 149
Mebendazole **102**, 231; **104**, 175; **142**, 251
Mebendazole derivative **104**, 175
Mebeverine **67**, 89; **72**, 19; **142**, 163
Mechanical strength **10**, 209
Mechanical stress **57**, 95
Mechanics **145**, 165
Mecillinam **4**, 299
Meclofenamate sodium **134**, 129
Medazepam **10**, 339; **13**, 67; **16**, 327; **49**, 91; **61**, 189; **63**, 155
Medical instrumentation **48**, 255
Medicinal plant **63**, 57
Medisorb **87**, 247; **143**, 133
Medisorb 100l **131**, 171
Medium chain fatty acid **145**, 17
Medium chain triacylglycerol **107**, 121; **123**, 237; **139**, 161
Medium chain triglyceride **2**, 245
Medroxyprogesterone **47**, 1; **54**, 33
Medroxyprogesterone acetate **54**, 33; **98**, 225; **108**, 69; **124**, 205
Mefenamic acid **8**, 303; **21**, 187; **34**, 75
Mefloquine **60**, 157; **70**, 95; **101**, 127; **110**, 189
Megestrol **98**, 225; **125**, 99
Megestrol acetate **6**, 195
Melanoma **122**, 107
Melanoma B16 **35**, 219
Melanotan 1 **135**, 81
Melatonin **69**, 93; **92**, 23; **111**, 283; **124**, 119; **144**, 37
Melphalan **7**, 1; **8**, 217; **24**, 193; **37**, 257; **57**, 63; **71**, 73; **108**, 69
Melting point **16**, 93; **45**, 13
Membrane **2**, 41; **8**, 193; **12**, 87, 97, 219; **16**, 271; **28**, 145; **47**, 79; **58**, 155; **60**, 1; **61**, 57; **63**, 281; **67**, 103; **69**, 103, 109; **70**, 209; **71**, R1; **75**, 89; **77**, 1; **105**, 113
Membrane binding **90**, 59; **124**, 1
Membrane damage **55**, 209
Membrane fluidity **125**, 1
Membrane metalloendopeptidase **97**, 171
Membrane permeability **2**, 41, 239; **12**, 97; **24**, 209, 297; **25**, 265; **37**, 49; **42**, 161; **55**, 237; **56**, 135; **72**, 1; **72**, R1; **80**, 213; **87**, 125; **90**, 43, 229; **96**, 33; **98**, 37; **105**, 137, 259; **113**, 165; **120**, 115, 119, 205; **121**, 27; **123**, 13, 71, 103; **124**, 53; **126**, 179; **131**, 97; **137**, 149
Membrane potential **49**, 57; **69**, 21
Membrane stabilization **90**, 221; **107**, 99
Membrane structure **55**, 209
Membrane transport **61**, 163; **63**, 1; **76**, 1; **80**, 39; **92**, 23; **111**, 1; **123**, 187; **139**, 15; **145**, 115
Menadione **39**, 137; **50**, 213; **129**, 159
Menadione sodium bisulfite **39**, 137
Menatetrenone **93**, 85
Menthol **84**, 117; **91**, 9; **111**, 235; **113**, 189; **125**, 7; **130**, 141; **143**, 37; **145**, 29
Menthone **111**, 235
Mephaquin **101**, 127
Mephenhydramine-8-chlorotheophyllinate **4**, 135
Mephenytoin **17**, 1; **65**, 141
Mepivacaine **136**, 165
Meprobamate **17**, 1
Mepyramine maleate **14**, 223; **18**, 287; **53**, 175
Mepyramine **18**, 287
Mequinol **10**, 89
Mequitazine **12**, 219; **13**, 227; **35**, 129
Mercaptoethanol **16**, 127
3-Mercaptopropionic acid **28**, 119

Mercaptopurine **5**, 167; **28**, 119; **35**, 165; **36**, 211; **42**, 263; **57**, 41, 253; **68**, 105; **95**, 51; **106**, 33; **117**, 173; **123**, 1
6-Mercaptopurine derivative **48**, 195
Mercuric chloride **34**, 115; **53**, 13
Mercuric oxycyanide **54**, 51
Mercury **49**, 205; **84**, 191; **123**, 71
Mergital **99**, 29
Mesalazine **88**, 177; **122**, 121; **123**, 1
Mesenchymal cell **40**, 23
Mesentery lymph duct **24**, 185
Mesentery lymph node **102**, 261
Mesna **104**, 151
Mestanolone **14**, 325, 333
Mestranol **11**, 181; **15**, 115, 149
Mesuximide **17**, 1
N-Mesylampicillin **17**, 47
Mesylmesylic-acid-2-chloroethyl-ester **48**, 179
Meta bisulfite sodium **12**, 185
Meta cresol **10**, 89; **140**, 155
Metabolism **2**, 41; **5**, 167; **29**, 29
Metabolite **24**, 101
Metandienone **14**, 325, 333
Metenkephalin **76**, 113; **121**, 217
Metenkephalin[2-dextro-alanine] **48**, 189
Metenolone acetate **13**, 333
Metenolone enanthate **13**, 333
Metenolone **13**, 333
Metformin **34**, 75; **128**, 155
Methacrylic acid **14**, 95; **34**, 137; **72**, 1; **95**, 105, 153; **110**, 291; **111**, 127
Methacrylic acid derivative **144**, 195
Methacrylic acid methyl ester **84**, 23; **87**, 37; **95**, 105; **111**, 127; **137**, 67; **146**, 71
Methadone **33**, 249; **45**, 1; **95**, 1; **134**, 203; **140**, 219
Methanol **16**, 127; **17**, 77, 291; **18**, 299; **21**, 119; **34**, 217; **68**, 285; **76**, 91; **78**, 17; **80**, 17; **92**, 81; **95**, 23; **113**, 97, 215; **115**, 123; **126**, 219; **129**, 51; **131**, 171; **132**, 23; **135**, 151; **136**, 71; **140**, 237; **141**, 71; **142**, 251
Methaqualone **6**, 205; **17**, 1
Methazolamide **51**, 27; **138**, 207
Methemoglobin **33**, 1
Methenamine **14**, 1; **36**, 233
Methionine **72**, 149; **87**, 59
Methionyl growth hormone **23**, 13; **45**, 197
Methocel K100 **129**, 21
Methocel K100LV **126**, 189
Methohexital **14**, 123
Methopterin **16**, 141
Methotrexate **8**, 217; **11**, 323; **16**, 141; **21**, 219; **22**, 137; **25**, 85; **36**, 7; **45**, 237; **47**, 129, 163, 239; **54**, 181; **59**, 27, 271; **61**, R1; **67**, 177; **68**, 151, 179, 265; **71**, R9; **79**, 71; **81**, 1; **85**, 189; **89**, 91; **92**, 1; **95**, 85; **98**, 9; **100**, 233; **102**, 35, 241; **105**, 83; **106**, 221; **108**, 21, 69; **111**, 25; **116**, 201; **124**, 205; **128**, 261, 283; **131**, 67; **132**, 175; **138**, 121; **142**, 25; **143**, 255
Methotrexate-α-benzyl-γ-cholesteryl ester **21**, 219
Methotrexate derivative **36**, 7; **102**, 241; **107**, 9; **132**, 175
Methotrexate dibutyl ester **59**, 271
Methotrexate diethyl ester **59**, 271
Methotrexate diisopropyl ester **59**, 271
Methotrexate dimethyl ester **59**, 271
Methotrexate dipropyl ester **59**, 271
Methotrexate ester derivative **36**, 7
Methoxide sodium **17**, 237
Methoxsalen **110**, 65; **133**, 245
5-Methoxy-2-methyl-3-indoleacetic acid **28**, 47; **49**, 79
7-Methoxy-4-methylcoumarin **12**, 355
6-Methoxy-2-naphthylacetic acid **118**, 31
9-Methoxyacridine **17**, 161
2-Methoxybenzoic acid **11**, 131
(4-Methoxybenzyl)triphenylphosphonium chloride **30**, 101
3-Methoxycoumarin **12**, 355
4-Methoxycoumarin **12**, 355
Methoxycoumarin derivative **12**, 355
4-Methoxycinnamic-acid-2-ethylhexyl-ester **108**, 57
3-Methoxymorphinan **14**, 173
4″-*O*-(4-Methoxyphenyl-acetyl)-tylosin **85**, 87
5-Methoxysalicylic acid **21**, 239; **31**, 185
Methyl acetate **55**, 17
Methyl benzoate **6**, 261
Methyl-β-cyclodextrin **104**, 239; **126**, 73
Methyl glucoside **35**, 103; **35**, 91
4-Methyl-2-hexylamine **15**, 185
2-Methyl-4-hydroxytamoxifen **99**, 59
2-Methyl-4-isothiazolin-3-one **74**, 195
N-Methyl-4-nitroacetanilide **29**, 67
2-Methyl-5-nitroimidazole **125**, 141
2-Methyl-3-nitropyridine-6-carboxaldehyde **2**, 257
Methyl orange **3**, 65; **8**, 193; **80**, 243
Methyl paraben **9**, 1; **19**, 1; **27**, 1, 163; **32**, 223; **33**, 105; **34**, 115; **35**, 227; **39**, 47, 229; **55**, 175; **80**, 203; **81**, 39; **83**, 211; **90**, 203; **92**, 15; **99**, 21; **109**, 155; **119**, 183; **125**, 141; **127**, 27; **133**, 17; **139**, 63; **140**, 97; **147**, 51
4-Methyl-2-pentanone **32**, 187
1-Methyl-3-piperidinomethylurea **9**, 7
1-Methyl-3-propylxanthine **56**, 235
1-Methyl-2-pyrrolidinone **21**, 17; **43**, 233; **49**, 39; **60**, 177; **95**, 43; **125**, 7
5-Methyl-2-pyrrolidone **95**, 43
N-Methyl-*n*[7(1-pyrrolidinyl)-1-oxaspiro[4,5]dec-8-*yl*]benzo[b]furan-4-acetamide **83**, 65
N-(Methylaminomethyl)benzamide **9**, 7
4-Methylaniline **37**, 49; **57**, 189; **77**, 1; **100**, 1
N-Methylbenzamide **71**, 31, **123**, 95
3-Methylbenzothiazolin-2-one-hydrazone **8**, 121
Methylcellulose **3**, 13; **10**, 209; **11**, 309; **12**, 163; **31**, 99; **45**, 39; **51**, 19; **52**, 149; **53**, 47, 75; **54**, 95; **55**, 115, 123; **56**, 207; **60**, 197; **71**, 95; **72**, 163; **77**, 177, 89; **79**, 257; **81**, 75, 243; **84**, 69; **86**, 159; **100**, 143; **91**, 151; **91**, 167; **94**, 39; **99**, 173; **100**, 155, 165, 175; **101**, 127; **102**, 19; **109**, 91, 115; **112**, 191; **114**, 85; **116**, 65; **118**, 221; **119**, 71; **121**, 11; **125**, 185; **126**, 147; **127**, 115; **135**, 115; **136**, 147; **140**, 97; **145**, 175, 197; **146**, 239; **147**, 233
β-Methyldigoxin **2**, 127
Methyldopa **3**, 239; **34**, 75

4′-*O* Methyldoxorubicin **32**, 123
Methylene blue **78**, 89; **146**, 175
Methylene iodide **15**, 73
6-Methyleneandrosta-1,4-diene-3,17-dione **71**, 19; **75**, 141, 147
N-Methylformamide **129**, 283
3-Methylgentisic acid **5**, 149
4-Methylgentisic acid **5**, 149
O-Methylglucose **35**, 103
Methylhydroxyethylcellulose **11**, 309
Methylidenemalonic acid diethyl ester **79**, 29
2-Methylimidazole **125**, 141
Methylimidazole derivative **19**, 107
N-Methylisothiazolone **66**, 201
N-Methylnicotinamide **123**, 95
2-Methylmuscarine **15**, 223
Methylpentynol trimethylsilyl ether **28**, 1
Methylprednisolone **72**, 57; **80**, 161, 253; **92**, 105; **100**, 257; **110**, 9; **120**, 189
6-α-Methylprednisolone-21-phosphate **120**, 189
Methylprednisolone sodium succinate **120**, 189; **123**, 159
15-Methylprostaglandin F2 α **1**, 281
Methylpyrrolidine **24**, 61
N-Methylpyrrolidone **82**, 171; **92**, 243
6-Methylselenomethyl-19-norcholest-5(10)-en-3β-ol-se-75 **16**, 105; **26**, 113
6-Methylselenomethylnorcholestenol se 75 **26**, 113
2-Methyltamoxifen **99**, 59
Methyltestosterone **14**, 325, 333; **25**, 265
Methylthiouracil **57**, 41
1-Methyluric acid **33**, 55
3-Methylxanthine **18**, 259; **33**, 55
Metiamide **31**, 33
Meticillin **30**, 181
Metkephamid **141**, 39
Metoclopramide **2**, 155; **5**, 167; **9**, 295; **24**, 91; **25**, 255; **28**, 41; **31**, 33; **43**, 59; **54**, 1; **81**, 1; **88**, 433; **100**, 25, 65; **104**, 107; **123**, 113, 285; **144**, 171
Metodiclorofen **145**, 9
Metolose 60-SH **138**, 57
Metolose **35**, 201
Metopimazine **123**, 285
Metoprolol **34**, 75, 131; **37**, 251; **36**, 165; **46**, 95; **47**, 133; **56**, 135; **63**, 73; **64**, 171; **68**, 167; **71**, 55; **79**, 223; **87**, 89; **88**, 391; **91**, 151; **92**, 1; **104**, 253; **105**, 189; **120**, 215; **123**, 149; **125**, 165; **130**, 137; **136**, 117; **141**, 217
Metoprolol benzoate **79**, 223
Metoprolol fumarate **79**, 223
Metoprolol sorbate **79**, 223
Metoprolol succinate **79**, 223
Metoprolol tartrate **39**, 39; **130**, 137; **140**, 229
Metronidazole **10**, 199; **13**, 243; **14**, 251; **18**, 67, 79; **21**, 201; **23**, 277; **23**, 289; **26**, 227; **29**, 1, 83; **31**, 99; **32**, 199; **35**, 39; **38**, 65; **41**, 121; **43**, 233; **44**, 31; **57**, 217; **60**, 253; **61**, 119; **75**, 161; **76**, 261; **92**, 233; **93**, 233; **97**, 1; **98**, 231; **109**, 251; **121**, 169; **125**, 141; **129**, 113; **134**, 231; **139**, 219; **140**, 169; **141**, 17; **145**, 9
Metronidazole-β-alaninate **18**, 67
Metronidazole-3-dimethylaminopropionate **18**, 67
Metronidazole-4-dimethylaminobutyrate **18**, 67
Metronidazole-4-methyl-1-piperazineacetate **18**, 67
Metronidazole acetate **32**, 199
Metronidazole benzoate **15**, 213
Metronidazole bromoacetate **18**, 67
Metronidazole butyrate **32**, 199
Metronidazole chloroacetate **18**, 67
Metronidazole derivative **26**, 227; **32**, 199; **58**, 143
Metronidazole ester **26**, 227; **58**, 143
Metronidazole glycinate **18**, 67
Metronidazole hexanoate **32**, 199
Metronidazole hydrogen succinate **21**, 201; **35**, 39
Metronidazole hydrogen succinate dextran **35**, 39
Metronidazole iodoacetate **18**, 67
Metronidazole morpholinoacetate **18**, 67
Metronidazole-*n*-propylglycinate **18**, 67
Metronidazole-*n*,*n*-dimethylglycinate **18**, 67, 79
Metronidazole propionate **32**, 199
Metronidazole valerate **26**, 227; **32**, 199
Mevalonic acid **96**, 1
Mevinolin **66**, 97; **94**, 1; **96**, 1; **125**, 141
Mexazolam **80**, 135
Mexazolam derivative **80**, 135
Mexiletine **5**, 167; **81**, 127
Mezlocillin **26**, 133; **77**, 47
Micelle **2**, 101; **4**, 271, 281; **8**, 303; **13**, 47; **24**, 79; **26**, 25; **39**, 263; **42**, 1; **64**, 239; **65**, 201; **78**, 157; **83**, 53, 115; **135**, 43, 73; **145**, 129; **146**, 63
Miconazole **48**, 207; **90**, 247; **94**, 115; **96**, 1; **110**, 169
Miconazole nitrate **10**, 119; **130**, 257
Microbial growth **58**, 155; **73**, 97
Microbiology **37**, 145
Microcalorimetry **29**, 237; **42**, 19; **72**, 181; **83**, 11; **86**, 263; **89**, 33; **104**, 135; **125**, 133
Microcapsule **6**, 277; **12**, 251; **14**, 95; **16**, 65; **26**, 289; **29**, 83, 195; **40**, 165; **58**, 31; **59**, 9, 173; **62**, 27; **68**, 19; **69**, R1; **78**, 165; **82**, 129; **92**, 29, 55; **103**, 253; **106**, 25; **126**, 161; **134**, 129; **137**, 187; **138**, 25, 185; **143**, 93; **144**, 153, 209; **148**, 73
Microcel **128**, 123
Microcel MC 101 **115**, 53
Microcomputer **15**, 37
Microcrystalline cellulose **34**, 217; **57**, R1; **38**, 227, 231; **40**, 129, 267; **55**, 143; **57**, 95, 139, 155; **61**, 145; **63**, 251; **64**, 35, 55, 207, 227; **66**, 143, 207, 273, 283; **67**, 79; **69**, 233; **70**, 261; **71**, 81; **72**, 117, 243; **73**, 37, 75, 181; **76**, 239; **78**, 183; **79**, 47, 257; **80**, 179; **81**, 205, 225; **82**, 71; **83**, 11, 65, 147, 187; **86**, 187; **89**, 223, R17; **90**, 213; **91**, 23, 123, 133, 143, 173, 183, 241, 247; **92**, 55, 137, 157; **94**, 49, 181; **95**, 29, 135; **96**, 59, 197, 205, 217, 225; **97**, 15, 79; **99**, 7, 157; **100**, 49, 71; **101**, 71, 169; **102**, 81; **103**, 55, 259; **105**, 255; **106**, 149; **107**, 141; **108**, 207; **109**, 209, 221; **110**, 203; **111**, 43, 89; **112**, 225; **113**, 97; **114**, 1, 55, 85, 137; **115**, 35, 53; **116**, 65, 179, 191; **117**, 23, 57; **118**, 95, 213; **119**, 71; **120**, 95, 101; **121**, 65, 73, 205; **122**, 49, 91; **125**, 45, 257; **128**, 1, 123; **130**, 147; **131**, 201, 209; **132**, 207; **133**, 65, 133; **134**, 79; **136**, 71; **137**, 41; **138**, 57; **139**,

15, 25; **140**, 97; **142**, 97; **143**, 151, 233; **144**, 187; **146**, 1, 21, 159; **147**, 79, 95; **148**, 131, 149, 201
Microcrystalline wax **147**, 23
Microdialysis **81**, 143
Microemulsion **29**, 267; **61**, 213; **147**, 131
Microencapsulation **2**, 91; **25**, 121; **29**, 1; **49**, 69; **58**, 123; **60**, 83; **61**, 219; **62**, 27; **73**, 157; **81**, 111; **86**, 147; **88**, 39; **96**, 85; **97**, 111; **103**, 37; **115**, 1; **118**, 199; **124**, 87, 97; **126**, 129; **129**, 51, 263; **130**, 187; **138**, 25; **147**, 173; **148**, 23
Microfilament **68**, 135
Microflora **97**, 1
Microorganism **26**, 89
Microparticle **106**, 77
Microparticulate ophthalmic delivery system **40**, 81
Microscopy **66**, 131
Microsphere **11**, 323; **12**, 135; **22**, 63; **29**, 95; **32**, 165; **33**, 137, 147; **34**, 225; **35**, 145; **39**, 189; **44**, 49; **46**, 69, 149; **51**, 1, 85, 253; **52**, 87, 183, 221; **54**, 231; **57**, 205; **58**, 79, 123; **59**, 27, 57; **63**, 207; **64**, 99; **68**, 121, 265; **69**, 5, 69; **70**, 235; **71**, 7; **72**, 199; **75**, 25, 73; **76**, 55, 193, R5; **78**, 1, 9; **79**, 191; **80**, 1; **83**, 1, 273; **84**, 151, 245; **85**, R5; **86**, 147, 239; **87**, 21; **89**, 91; **95**, 77, 85; **96**, 23; **98**, 157; **100**, 123; **105**, 181; **106**, 41, 213, 221; **107**, 29, 41, 205; **111**, 217; **118**, 237; **124**, 37, 75; **126**, 129; **128**, 261; **130**, 49, 187; **131**, 25, 171; **133**, 97; **134**, 203, 247; **136**, 89; **137**, 57, 159; **140**, 145, 219; **141**, 205; **142**, 135; **143**, 255; **144**, 19, 107, 241; **145**, 157; **146**, 147; **147**, 173; **148**, 23, 55, 231
Microvasculature **45**, 249
Microwave radiation **51**, 19
Micturition **45**, 65
Midazolam **33**, 37; **38**, 261; **49**, 83; **56**, 175; **61**, 189; **105**, 189; **123**, 229
Midazolam derivative **49**, 83
Midazolam maleate **33**, 37; **49**, 83
Miglyol **12**, 315; **27**, 335; **33**, 99; **95**, 161; **113**, 17, 57, 89; **125**, 107; **127**, 203; **130**, 57; **145**, 241
Migration **59**, 245
Mild silver protein **12**, 67
Milk **33**, 125; **36**, 73; **44**, 9
Milk fat **54**, 123; **118**, 55
Millisrol **65**, 159
Mineral oil **33**, 225; **57**, 87; **78**, 217; **132**, 267; **133**, 97; **139**, 161; **148**, 1
Minimum inhibitory concentration **27**, 163; **32**, 55, 91; **34**, 115, 241
Minocycline **25**, 105
Minoxidil **49**, 109; **50**, 21, 53; **55**, 229; **68**, 127; **72**, 227; **96**, 111
Miosis **10**, 219; **12**, 147; **37**, 19; **51**, 203; **72**, 131; **86**, 159; **95**, 229; **127**, 85; **142**, 153
Misonidazole **145**, 9
Mitindomide **45**, 207
Mitobronitol **108**, 69
Mitolactol **108**, 69; **124**, 205; **145**, 9
Mitomycin **15**, 49; **22**, 75
Mitomycin A **52**, 239
Mitomycin B **52**, 239
Mitomycin C **4**, 21; **8**, 131; **15**, 49, 61; **24**, 219; **25**, 95, 105; **32**, 111, 133; **37**, 135, 145; **45**, 189; **52**, 239; **54**, 27; **55**, 31; **58**, 79; **59**, 121; **65**, 7; **78**, 103; **89**, R5; **98**, 121; **105**, 249; **108**, 69; **124**, 205
Mitomycin C derivative **25**, 95; **55**, 1; **56**, 71
Mitomycin C dextran **37**, 145
Mitomycin C polyaspartic acid **22**, 75
Mitomycin C polyglutamic acid **22**, 75
Mitomycin C polylysine **22**, 75
Mitomycin derivative **15**, 61
Mitoxantrone **68**, 151; **70**, 1; **103**, 81, 87; **107**, 149; **116**, 87; **128**, 139; **139**, 1
Mixing **32**, 177
MK 0476 **134**, 59
Mobility **7**, 285
Model **1**, 3, 15, 127; **13**, 99, 103, 345; **31**, 107; **44**, 125; **49**, 39; **140**, 111
Modelling of process **40**, 33
Moisture **42**, 77; **62**, 87; **68**, 51, 179; **77**, 315; **81**, 187; **83**, 87; **84**, 261; **87**, 149; **90**, R1; **97**, 79; **103**, 103, 205; **112**, 191; **113**, 247; **114**, 55; **118**, 191; **120**, 13; **121**, 65; **132**, 1; **134**, 79; **146**, 101
Molecular interaction **79**, 149
Molecular model **137**, 149
Molecular stability **139**, 33; **140**, 97
Molecusol **59**, 35
Monensin **110**, 277
Monitoring **32**, 177
Monoacylglycerol **85**, 103; **111**, 283; **126**, 83; **142**, 9, 199, 215; **147**, 135
Monoamine oxidase inhibitor **5**, 167
Monoclonal antibody **17**, 65; **32**, 165; **47**, 129; **59**, 27; **97**, 213; **98**, 149
Monomuls **99**, 29
Monosialoganglioside **87**, 21
Montmorillonite **6**, 243; **128**, 129
Morphine derivative **141**, 217; **146**, 213
Morphine sulfate **114**, 117; **138**, 129; **139**, 237
Morphine **9**, 245; **32**, 7; **38**, 171; **44**, 269; **45**, 161; **48**, 1; **56**, 1; **59**, 1; **71**, 105; **79**, 21; **81**, 1; **91**, 9; **101**, 219; **114**, 117; **118**, 181; **119**, 223; **123**, 1; **134**, 37; **139**, 237; **141**, 217; **146**, 213
N-Morpholinomethyl-*n'*-acetylthiourea **9**, 7
N-Morpholinomethylbenzamide **7**, 119
N-(Morpholinomethyl)salicylamide **9**, 7; **31**, 165
N-(Morpholinomethyl)salicylamide derivative **29**, 19
N-(Morpholinomethyl)thiobenzamide **9**, 7
N-Morpholinomethyl-4-toluenesulfonamide **9**, 7
Morphology **25**, 121; **40**, 23; **108**, 187; **120**, 157
Motion **142**, 33
Motion picture **20**, 87
Mouth **30**, 73
Mouth cavity **66**, 169; **74**, 9, 189
Mouth disease **139**, 63
Mouth mucosa **74**, 9, 189; **88**, 237, 293; **89**, 41
Moxisylyte **99**, 85
Mucin **19**, 107; **30**, 181; **53**, 1; **117**, 75; **126**, 179; **131**, 159; **134**, 15; **142**, 227

Mucociliary clearance **31**, 275; **122**, 69; **135**, 115; **149**, 123
Mucociliary transport **12**, 57, 67; **65**, 243; **73**, 131; **95**, 57; **122**, 69; **129**, 137; **136**, 181
Mucosa **30**, 73; **49**, 47; **80**, 1; **113**, 223
Mucosa inflammation **27**, 245; **133**, 1
Mucus **28**, 33; **49**, 231; **70**, 235; **79**, 97; **83**, 221; **94**, 195; **99**, 209; **124**, 173; **126**, 179
Mucus secretion **131**, 159
Multidrug resistance **138**, 237
Multiparin **53**, 237
Multivitaminic preparation **40**, 43
Muramyl dipeptide **23**, 299
Muramyl dipeptide **75**, 97
Muramyl tripeptide **89**, 139
Muscarine **15**, 223
Muscarinic agent **15**, 223
Muscarone **15**, 223
Muscle contraction **70**, 111
Muscle homogenate **17**, 91
Mussel adhesive protein **141**, 251
Mycobacterium intracellulare avium **78**, 227
Mycotoxin **128**, 129
Mydriasis **1**, 189; **20**, 187; **25**, 73; **51**, 203; **55**, 105
Myeloperoxidase **104**, 151
Mygliol 812 **145**, 17
Mygliol 840 **132**, 71
Mylanta II **28**, 151
Myoglobin **128**, 23
Myricetin **86**, 17
Myristic acid **4**, 271, 281; **11**, 27; **56**, 111; **106**, 1; **126**, 119, 267; **137**, 117; **145**, 215
Myristic acid isopropyl ester **4**, 205; **8**, 81, 193; **11**, 277; **12**, 219; **21**, 155; **33**, 225; **34**, 125; **35**, 243; **36**, 211; **39**, 149; **42**, 19, 97; **43**, 31, 233; **44**, 87; **48**, 195; **50**, 213; **52**, 165; **76**, 151; **84**, R5; **85**, 7; **88**, 417; **91**, 9; **93**, 27, 37, 161; **106**, 51; **111**, 63; **114**, 237; **116**, 253; **125**, 63; **129**, 13, 33; **130**, 121; **132**, 33; **137**, 243; **139**, 161; **141**, 71; **143**, 37, 67
Myristyl alcohol **148**, 41
Myristylpicolinium chloride **1**, 281
Myvacet **126**, 83; **133**, 161

N

Nabumetone **118**, 31
Nadolol **36**, 165; **37**, 127; **46**, 95
Nafamstat **88**, 63
Nafarelin **31**, 125; **108**, 49; **110**, 137; **116**, 1; **117**, 165; **121**, 45
Nafarelin acetate **31**, 125
Nafcillin **4**, 299; **32**, 151; **137**, 1; **144**, 177
Naftidrofuryl **87**, 223; **87**, 229
Nalbuphine **38**, 199
Nalidixic acid **6**, 205; **22**, 283; **38**, 71; **41**, 197; **63**, 237; **79**, 89; **88**, 379; **93**, 121; **140**, 1
Nalidixic acid butyryloxymethyl ester **55**, 91
Nalidixic acid derivative **55**, 91
Nalidixic acid isobutyryloxymethyl ester **55**, 91
Nalorphine **9**, 245
Naloxone **21**, 233; **33**, 225; **36**, 127; **44**, 269; **76**, 145
Naltrexone **7**, 1; **36**, 127; **38**, 171; **45**, 1; **148**, 219
Nandrolone **14**, 325, 333; **25**, 265
Nandrolone decanoate **6**, 185
Nandrolone phenpropionate **6**, 185
Nanocapsule **28**, 125; **36**, 121; **72**, 211; **113**, 57; **145**, 241
Nanoparticle **13**, 109; **17**, 65; **27**, 145; **29**, 267; **30**, 17; **33**, 45; **39**, 213; **40**, 23, 81; **42**, 145; **61**, 85; **62**, 1; **110**, 21; **120**, 1; **121**, 45; **126**, 103; **128**, 189; **129**, 95, 113, 159; **131**, 191; **132**, 123; **137**, 67; **139**, 113; **141**, 81; **142**, 121, 227; **144**, 115; **148**, 47, 165
Naphazoline **12**, 57; **46**, 193; **48**, 103; **50**, 75
Naphthalene **3**, 39; **31**, 225; **87**, 47; **94**, 1; **140**, 237
Naphthalene derivative **106**, 15; **137**, 21, 149; **145**, 145
1-Naphthol **13**, 183
2-Naphthol **126**, 219
1,4-Naphthoquinone derivative **120**, 137
1,2-Naphthoquinone-4-sulfonate sodium **17**, 325
1-Naphthyl-acetate **78**, 157
Naphthyl group **86**, 43
α Naphthylacetate **13**, 183
2-Naphthylamine **13**, 183
1,8-Naphthyridine derivative **140**, 1
Naproxen **5**, 139; **16**, 215; **32**, 85; **37**, 245; **38**, 117; **48**, 83; **50**, 97; **51**, 233; **53**, 157; **57**, 55; **64**, 195; **66**, 243; **70**, 35, 43; **75**, 37, 55; **78**, 183; **79**, 149; **82**, 129; **86**, 131; **88**, 1; **92**, 29; **94**, 23; **96**, 167; **97**, 119; **98**, 37; **100**, 99, 257; **106**, 179; **108**, 109; **114**, 55; **115**, 29; **125**, 309; **126**, 95; **127**, 103; **129**, 13; **137**, 49; **138**, 149
Naproxen derivative **43**, 101; **51**, 233; **100**, 99
Naringenin **124**, 1; **145**, 87
Natamycin **1**, 165
Navelbine **77**, 279
Nebulization **36**, 55; **58**, 57; **104**, 73; **109**, 17; **115**, 69; **127**, 191; **129**, 123; **130**, 245; **133**, 203; **148**, 93
Nebulizer **67**, 275; **103**, 119; **130**, 1; **148**, 93; **149**, 63
Nedocromil **82**, 117
Neobee M5 **24**, 173; **106**, 15
Neogelco **28**, 239
Neomycin **1**, 165; **9**, 359; **12**, 67; **108**, 39; **116**, 77
Neostigmine **13**, 183
Nephrotoxicity **46**, 103; **57**, 125; **59**, 231
Nerolidol **127**, 9; **130**, 141
Nerve growth factor **87**, 21; **141**, 239
Netilmicin **147**, 109
Neuroleptic agent **5**, 167
Neurotensin derivative **117**, 95
Neurotensin **112**, 133
Neutral insulin **69**, 29; **95**, 101; **125**, 205; **128**, 9
Neutron activation analysis **32**, 251
Newborn **24**, 259
Nialamide **17**, 1
Nicardipine **80**, 109; **136**, 117; **137**, 75; **138**, 25; **140**, 145
Nickel **45**, 101; **68**, 105
Nickel sulfate **3**, 261; **49**, 29
Nicodisc **148**, 177
Nicorandil **43**, 31; **79**, 21

Nicotinamide **19**, 17; **21**, 265; **29**, 177; **34**, 17; **123**, 95; **130**, 83; **147**, 1
Nicotinamide amidohydrolase **21**, 265
Nicotine **33**, 55; **42**, 97; **68**, 43; **80**, 227; **99**, 67; **129**, 175; **148**, 177
Nicotine tartrate **129**, 175
Nicotine-*n*,*n*′-dioxide **80**, 227
Nicotine-*n*′-oxide **80**, 227
Nicotinic acid **14**, 299; **81**, 1; **106**, 109; **124**, 285
Nicotinic acid butyl ester **15**, 125; **124**, 285
Nicotinic acid derivative **63**, 129; **66**, 213; **124**, 285
Nicotinic acid ethyl ester **3**, 143; **118**, 129
Nicotinic acid hexyl ester **15**, 125; **36**, 91; **118**, 129; **124**, 285
Nicotinic acid methyl ester **3**, 143; **12**, 219; **15**, 125; **16**, 255; **19**, 129; **48**, 103; **83**, 73; **87**, 261; **90**, 43; **106**, 63; **118**, 129; **121**, 89; **124**, 279, 285; **139**, 161
Nicotinic acid octyl ester **124**, 285
Nicotinic receptor **15**, 223
Nicotrans **148**, 177
Nifedipine **27**, 81; **31**, 179; **38**, 247; **41**, 75, 227; **49**, 163; **54**, 211; **67**, 169; **70**, 195; **80**, 151; **84**, 59, 161; **85**, 1; **87**, 133; **91**, 23; **93**, 209; **96**, 33; **98**, 157; **99**, 337; **103**, 69, 179; **108**, 11; **111**, 235; **118**, 23; **127**, 115, 261; **130**, 65; **132**, 131; **134**, 239, 247; **135**, 73, 183; **139**, 223; **144**, 241; **148**, 123
Niflumic acid **52**, 129; **68**, 1; **113**, 47
Nifurtoinol **3**, 93
Nikethamide **92**, 15
Nikkol SS10 **27**, 29
Nikkol TS10 **27**, 29
Nile red **45**, 169
Nimetazepam **16**, 327
Nimodipine **49**, 163; **79**, 273; **118**, 11; **119**, 115; **129**, 147
Nimustine **65**, 49; **99**, 73
Niosome **148**, 139
Nipecotic acid derivative **48**, 91
Nisoldipine **49**, 163; **79**, 273
Nitrate **17**, 145
Nitrazepam **10**, 339; **13**, 67; **16**, 327; **61**, 189; **63**, 29; **69**, 1; **80**, 129; **113**, 47
Nitrendipine **79**, 273; **99**, 351; **125**, 7; **136**, 117
Nitrite **92**, 233
3-Nitroaniline **6**, 261
4-Nitrobenzaldehyde **20**, 265
Nitrobenzene **2**, 295; **100**, 189; **129**, 51
4-Nitrobenzoic acid **20**, 265
4-Nitrobenzoic acid ester **8**, 167
4-Nitrobenzoic acid ethyl ester **8**, 167
4-Nitrobenzoic acid methyl ester **8**, 167
2-Nitrodantrolene **75**, 193
Nitrofural **8**, 111; **67**, 169; **70**, 9; **82**, 215, 223, 229; **143**, 75, 83
Nitrofurantoin **3**, 93; **5**, 67; **8**, 111, 175; **16**, 65; **17**, 357; **18**, 213; **19**, 297; **28**, 167; **29**, 191; **30**, 229; **33**, 125; **34**, 29; **35**, 145; **36**, 105; **37**, 103; **55**, 257; **56**, 111; **57**, 177; **67**, 59; **71**, 223; **96**, 243
5-Nitro-2-furaldehyde **8**, 111
4-Nitroimidazole **125**, 141
Nitropercuten **131**, 229
Nitrophenol **126**, 219
4-Nitrophenyl acetate **26**, 339
5-(4-Nitrophenyl)furfural **17**, 357
4-Nitrophenylacetic acid **48**, 189
4-Nitro-*n*-(piperidinomethyl)benzamide **9**, 7
Nitroprusside sodium **24**, 27; **33**, 1
4-Nitrosobenzoic acid **20**, 265
3(2-Nitro-4-trifluoromethylphenyl)-thioureidocarboxylic-acid-methyl-ester **77**, 265
Nitrous oxide **63**, 155
Nitroxinil **8**, 217
NNC 0112 **79**, 205
NNC 0756 **79**, 205
Nonane **34**, 217; **92**, 243
Nonanoic acid **137**, 117
Nonionic surfactant **9**, 173; **103**, 19; **125**, 283, 315; **130**, 203; **141**, 197
Nonivamide **126**, 119; **128**, 169; **130**, 121; **131**, 143; **143**, 47
Nonivamide acetate **126**, 119; **128**, 169; **131**, 143
Nonoxinol **120**, 41; **125**, 81
Nonoxinol-9 **125**, 81
Nonsteroid antiinflammatory agent **55**, 1; **57**, 55; **79**, 263; **83**, 73; **88**, 1; **89**, 51, 65; **92**, 151; **99**, 173; **101**, 209; **103**, 217; **108**, 109; **113**, 57; **119**, 133; **120**, 145; **121**, 107; **123**, 1; **124**, 97, 149; **125**, 231; **126**, 95, 161
2-Nonyl-1,3-dioxolane **125**, 63
Nonylglucoside **105**, 219
Noradrenalin **3**, 239; **21**, 75
Noradrenalin bitartrate **21**, 75
Noradrenalin derivative **28**, 1
Nordazepam **61**, 189; **106**, 109
Norephedrine **99**, 67
2-Norepimuscarine **15**, 223
Norethisterone **2**, 203; **3**, 275; **11**, 181; **15**, 115, 149; **116**, 1
Norethisterone acetate **3**, 275; **63**, 143; **142**, 115
Norethisterone benzoate **3**, 275
Norethisterone-3-(4-butylcyclohexyl)propionate **3**, 275
Norethisterone-4-bicyclohexylcarboxylate **3**, 275
Norethisterone-4-biphenylcarboxylate **3**, 275
Norethisterone-6-(4-chlorophenyl)hexanoate **3**, 275
Norethisterone-4-cyclohexylbenzoate **3**, 275
Norethisterone derivative **2**, 203; **3**, 319; **28**, 1
Norethisterone dimethylpropionate **3**, 275
Norethisterone enanthate **3**, 275
Norethisterone-4β,5β-epoxide **15**, 149
Norethisterone-4-hexylcyclohexylcarboxylate **3**, 275
Norethisterone pentamethyldisiloxyl ether **3**, 275
Norethisterone-4-phenoxybenzoate **3**, 275
Norfloxacin **63**, 237; **68**, 11; **79**, 89; **83**, 267; **87**, 73; **88**, 379; **93**, 121; **111**, 83; **121**, 123; **123**, 53, 173; **132**, 271; **138**, 113
Norflunitrazepam **36**, 113
Norflurane **123**, 291
Norgestrel **7**, 1; **71**, 7; **97**, 39
Norgestrel plus ethinylestradiol **33**, 55
Norindometacin **8**, 203
Norlevorphanol **14**, 173

2-Normuscarine **15**, 223
Nornicotine **80**, 227
Norselenocholesterol se 75 **16**, 105
Nortriptyline **76**, 17; **78**, 175; **94**, 135
Noscapine **9**, 245; **32**, 7; **58**, 197
Nose **9**, 165, 173; **30**, 73; **95**, 101; **112**, 181; **127**, 115
Nose cavity **128**, 229; **149**, 123
Nose drops **70**, 19
Nose injury **119**, 37; **130**, 93
Nose irritation **141**, 151
Nose mucosa **7**, 317; **9**, 173; **69**, 21; **73**, 131; **74**, 95, 147; **75**, 73; **76**, 247; **76**, 61; **84**, 129; **87**, 125; **89**, 147; **103**, 27; **105**, 7; **114**, 197; **116**, 77; **119**, 37; **123**, 103; **124**, 37; **125**, 205; **128**, 9, 239; **129**, 175; **134**, 173; **139**, 25; **141**, 151; **146**, 11
Novaminsulfon **7**, 293; **18**, 9
Noveon **133**, 1; **141**, 39
Noxytiolin **38**, 59
NSC 201047 **52**, 71
Nuclear magnetic resonance **15**, 235; **21**, 251; **22**, 345; **23**, 117; **24**, 307; **26**, 317; **29**, 73; **34**, 247; **35**, 243; **36**, 7
Nuclear magnetic resonance imaging **49**, 249
Nucleation **58**, 185
Nucleoside derivative **141**, 1
5′-Nucleotidase **114**, 197
Numerical deconvolution **40**, 165
Nutrient **28**, 151
Nylon **25**, 121; **39**, 183; **59**, 9; **120**, 119
Nymcel ZSB 10 **55**, 115
Nymcel ZSB 16 **55**, 115
Nystatin **7**, 285; **73**, 43; **88**, 9; **101**, 121

O

Obidoxime **109**, 35
Occlusion **133**, 245
Octadecanol **21**, 99; **27**, 311; **37**, 119; **112**, 165; **119**, 71; **125**, 315; **126**, 119; **130**, 121
9-Octadeceneamine hydrofluoride **131**, 33
Octadecylamine **98**, 19; **108**, 57; **127**, 273; **140**, 119
Octadecylmaltoside **132**, 107
Octadecyltrimethylammonium **124**, 107
Octane **27**, 39; **34**, 217; **82**, 145
Octanoic acid **4**, 271, 281; **11**, 131; **20**, 3; **33**, 253; **93**, 91; **95**, 161; **100**, 219; **106**, 15; **109**, 147; **124**, 183; **125**, 107; **126**, 219; **132**, 23; **136**, 147; **137**, 117; **139**, 205
Octanoin **110**, 137
Octanol **1**, 15; **4**, 205; **13**, 75; **16**, 127; **18**, 299; **29**, 103, 177; **31**, 209; **36**, 211; **68**, 285; **88**, 379; **92**, 81; **129**, 33; **132**, 23; **136**, 165; **138**, 149; **148**, 41
7-(Octanoyloxymethyl)theophylline **12**, 299
Octenylsuccinate starch **128**, 73
Octoxinol **91**, 95; **116**, 95
Octreotide **136**, 117; **137**, 103; **139**, 25
Octyl glucoside **79**, 159; **105**, 219
2-Octylcyclohexanone **68**, 239
Octylmaltoside **132**, 107
Octylthioglucoside **105**, 219
Oenanthe **48**, 119
Ofloxacin **50**, 83; **63**, 237; **79**, 89; **83**, 267; **89**, 125; **93**, 121; **106**, 229; **118**, 65; **121**, 123
Oil **5**, 273; **11**, 277; **113**, 89; **119**, 173; **125**, 107
Oil blue dye **13**, 243
Ointment **6**, 349; **22**, 31; **24**, 61, 115; **27**, 29; **28**, 47; **30**, 47; **37**, 11; **41**, 49; **68**, 1; **86**, 9, 201; **95**, 105; **126**, 35, 119; **127**, 1; **128**, 169; **130**, 121; **131**, 143; **137**, 117; **141**, 171
Ointment base **8**, 263; **24**, 61; **26**, 243; **28**, 113; **93**, 233; **126**, 119; **130**, 121; **131**, 143
Oleate sodium **28**, 193; **107**, 159; **143**, 101
Oleic acid **4**, 219, 271, 281; **11**, 11; **12**, 315; **24**, 173; **25**, 41; **28**, 75; **31**, 239; **33**, 235; **36**, 211; **37**, 127, 251; **43**, 9, 17; **48**, 103; **49**, 39; **57**, 29, 73; **65**, 211; **67**, 97; **71**, 193, R1; **74**, 59, 137; **76**, 217; **80**, R1; **82**, 171; **85**, 7; **87**, 175; **88**, 89, 423; **91**, 105; **92**, 89; **93**, 221; **95**, 181; **98**, 203; **101**, 57; **103**, 77; **105**, 47; **106**, 15; **109**, 45; **109**, 147; **110**, 9; **116**, 237; **117**, 173; **117**, 219; **119**, 173; **121**, 37; **123**, 281; **125**, 7, 107, 133; **128**, 145; **132**, 71; **134**, 223; **137**, 125; **141**, 71; **142**, 1; **143**, 101; **145**, 187
Oleic acid ethyl ester **11**, 277; **15**, 185; **16**, 31; **62**, 217; **91**, 157; **92**, 191; **135**, 1; **147**, 131
Oleic acid methyl ester **4**, 271, 281
Oleyl alcohol **4**, 271, 281; **13**, 75; **132**, 33
Oligomer **125**, 81
Oligonucleotide **105**, 181; **107**, 169; **143**, 259
Oligopeptide **146**, 205
Olive oil **11**, 277; **28**, 193; **44**, 107; **142**, 1
Olsalazine **37**, 155; **69**, 239; **97**, 1
Omeprazole **143**, 113
Ondansetron **123**, 1
Opagel **62**, 27
Ophthalmic agent **29**, 243
Ophthalmology **16**, 203
Opiate receptor **44**, 257
Opiate **32**, 7; **70**, 111; **83**, 65; **92**, 1
Opsonin **32**, 165
Optical rotatory dispersion **108**, 95
Oral antidiabetic agent **126**, 155
Oral contraceptive agent **33**, 55
Oral drug administration **27**, 267
Oral drug delivery **40**, 143
Oral form **40**, 33
Orciprenaline **41**, 55; **80**, R1; **124**, 149; **130**, 1; **142**, 125; **149**, 85
Organ distribution **147**, 143
Organ growth **54**, 231
Organic compound **135**, 43
Organic nitrate **128**, 277
Organic solvent **13**, 75; **113**, 97; **129**, 51; **130**, 187; **132**, 23; **135**, 151; **139**, 33, 87; **142**, 67
Organophosphorus compound **114**, 125
Oropharynx **102**, 11
Oros **22**, 321
Orosomucoid **59**, 137
Orotic acid **61**, 43
Orthanilamide **88**, 437
Ortho cresol **10**, 89

Ortho novin **11**, 181
Osmium tetraoxide **128**, 251
Osmolality **61**, 15; **85**, 19; **93**, 61
Osmosis **61**, 57, 101; **77**, 107
Osmotic minipump **56**, 227, 261
Osteogenic sarcoma **40**, 23
Osteoporosis **147**, 233
Osteosarcoma **17**, 65
Otilonium bromide **71**, 1
Ouabain **88**, 111; **98**, 83; **103**, 27
Ovalbumin **16**, 65; **47**, 79; **90**, 95; **101**, 89; **103**, 37; **108**, 133; **115**, 259
Ovothin **84**, R5; **116**, 253
Ovulation **69**, 93
Oxacillin **3**, 151
Oxalate sodium **14**, 299
Oxalic acid **51**, 147
Oxathiin carboxanilide **73**, 23
Oxazepam **10**, 339; **13**, 67; **16**, 327; **40**, 119; **45**, 111; **46**, 241; **47**, 223; **57**, 17; **61**, 189; **69**, 129; **80**, 191; **90**, 1; **99**, 321; **110**, 169; **114**, 95; **122**, 35; **141**, 53; **143**, 247
Oxazolam **80**, 135
Oxazole derivative **81**, 179
Oxazolidine **18**, 325
Oxazolidine derivative **18**, 325; **30**, 151
Oxazolidinone derivative **136**, 61
5-Oxazolidinone derivative **46**, 159
Oxazolo[4,5-b]pyridine derivative **98**, 1
Oxetacaine **28**, 239
Oxfenicine **19**, 149
Oxidative stress **110**, 161
Oxidoreductase **146**, 11
Oxipurinol **27**, 71
Oxodipine **89**, 119; **93**, 215
Oxolinic acid **22**, 283; **79**, 89
5-Oxoprolyl-peptidase **52**, 255
Oxprenolol **22**, 321; **29**, 73; **36**, 165; **37**, 251; **46**, 95, 193; **47**, 133; **48**, 1; **49**, 141; **57**, 9; **63**, 73; **84**, 175; **112**, 93; **120**, 215; **123**, 149; **125**, 165; **130**, 179; **136**, 117; **141**, 125, 217
Oxprenolol derivative **141**, 125
Oxprenolol ester **84**, 175
Oxygen **146**, 11
Oxygen radical **58**, 237; **87**, 79; **113**, 159
Oxymetazoline **12**, 57; **135**, 137
Oxymorphone **56**, 1
Oxyphenbutazone **8**, 303; **32**, 171; **43**, 119
Oxytetracycline **5**, 273; **15**, 293; **20**, 171; **34**, 75; **43**, 77; **75**, 161; **124**, 271
Oxytocin derivative **114**, 151
Oxytocin **5**, 167; **75**, 117; **124**, 149
Ozone **31**, 231, 239; **87**, 67

P

P 388 **22**, 75; **35**, 219; **37**, 135
Packaging **67**, 169; **74**, 175; **77**, 47; **83**, 59; **105**, 83, 89; **113**, 241
Pain **61**, 1; **124**, 279
Pain assessment **109**, 45
Palate **122**, 69
Palmitate magnesium **43**, 249
Palmitic acid **4**, 271, 281; **11**, 11, 27; **16**, 215; **37**, 127; **98**, 203; **106**, 1; **111**, 213; **115**, 45; **119**, 71; **126**, 49; **137**, 117
Palmitic acid isopropyl ester **92**, 191
Palmitic acid methyl ester **16**, 305
Palmitoleic acid **37**, 127; **111**, 213; **126**, 267
Palmitooleic acid **4**, 271, 281
Palmitoyl timolol malonate **128**, 179
[2(2-Palmitoylamidoethyl)-5-acetamido-3,-5-dideoxy-glycero-β-D-galacto-2-nonulipyranoside]onate; **113**, 141
Palmitoylcarnitine **56**, 29
3-Palmitoylcatechin **27**, 117
Palmitoyllysolecithin **114**, 151
Pamidronic acid **123**, 187
Panadiplon **133**, 17
Pancreas enzyme **48**, 51
Pancreas insufficiency **42**, 105; **48**, 51
Pancreas islet **23**, 87
Pancreatin **143**, 1
Pantothenic acid **81**, 1
Papaverine **9**, 245; **32**, 7; **46**, 173; **68**, 255
Para anisidine **6**, 261
Para cresol **10**, 89
Paracetamol glucuronide **146**, 181
Paracetamol sulfate **14**, 113; **121**, 55; **146**, 181
Paracetamol sulfate potassium **14**, 113
Paracetamol **1**, 249; **5**, 25, 109, 167; **6**, 29; **7**, 269; **8**, 277; **9**, 107; **10**, 209; **12**, 341; **14**, 113; **17**, 313; **24**, 239; **25**, 57; **28**, 75, 133; **29**, 73, 195; **30**, 161; **31**, 271; **33**, 125; **34**, 75; **36**, 29; **37**, 15; **40**, 15; **41**, 29; **42**, 123, 155; **44**, 43, 265; **47**, 249; **49**, 1, 261; **50**, 197; **51**, 115, 129, 263; **53**, 249; **54**, 199; **56**, 249; **57**, 117; **58**, 123, 177, 209; **67**, 231; **70**, 103, 241; **72**, 19, 117, 219; **78**, 17, 203; **79**, 83; **81**, 187; **82**, 233; **83**, 257; **84**, 233; **89**, 1, R17; **91**, 173; **92**, 15, 123; **93**, 111; **96**, 119; **99**, 99, 229; **102**, 71; **104**, 215, 253; **105**, 255; **106**, 187; **109**, 209, 221; **112**, 17, 241; **114**, 55; **119**, 1, 47, 203; **121**, 55, 117; **123**, 209; **125**, 157; **127**, 185; **128**, 1, 73; **133**, 17, 223; **136**, 117; **137**, 225; **139**, 95; **140**, 237; **141**, 53, 101; **142**, 143; **143**, 107; **144**, 187; **146**, 181; **148**, 73, 155
Paraffin **6**, 29, 349; **11**, 277; **12**, 315; **15**, 73; **17**, 39; **18**, 127; **26**, 243, 275; **28**, 23, 113; **49**, 29; **92**, 203; **98**, 57
Para-menthan-3-one **74**, 157
Parameters of drug loading **40**, 63
Paramethasone **10**, 1
Parathion **7**, 99; **14**, 123
Parathyroid hormone **75**, 97
Parenteral feeding **13**, 9; **16**, 57
Parenteral nutrition **34**, 173; **39**, 33; **64**, 217; **66**, 1; **77**, 57; **81**, 1; **107**, 121
Parenteral solution **40**, 175; **59**, 245; **66**, 87; **101**, 175; **103**, 125; **134**, 235; **140**, 169
Paroxetine **42**, 135
Partial thromboplastin time **37**, 65
Particle size **1**, 71; **7**, 261; **10**, 209; **13**, 9; **15**, 177, 293; **16**, 171, 345; **17**, 167; **20**, 171; **31**, 137; **34**, 173; **40**, 111; **42**, 267; **44**, 49; **45**, 245; **47**, 171, 179, 205, 223, 249; **49**, 241; **51**, 9, 55,

85; **52**, 91, 165, 173, 213, 221; **58**, 229; **59**, 145; **60**, 89, 235; **61**, 145; **62**, 1, 143, 229; **63**, 143; **64**, 227; **65**, 1, 43, 69; **67**, 231, 247; **68**, 19, 69, 111; **73**, 75; **74**, 49; **77**, 183, 211, 261; **81**, 187, 199, 205, 225, 243; **82**, R7; **84**, 107; **85**, 121; **88**, 141, 303; **89**, R17; **91**, 247; **93**, 101; **94**, 59, 89; **104**, 115; **120**, 95, 101, 157, 253; **126**, 189; **127**, 219, 261, 283; **128**, 261, 269; **129**, 113, 123; **131**, 273; **132**, 221; **133**, 203; **134**, 137, 235; **136**, 1; **137**, 41, 261; **139**, 139, 169, 255; **142**, 33; **144**, 141, 187; **145**, 95; **148**, 33, 149
Particulate matter **48**, 255
Partition coefficient **3**, 231; **4**, 47; **9**, 295; **13**, 75; **14**, 29; **18**, 247; **20**, 209; **21**, 349; **22**, 283; **24**, 173; **27**, 287; **29**, 177; **30**, 101; **34**, 241; **36**, 61, 67; **38**, 47; **43**, 187; **45**, 1, 157; **54**, 155; **56**, 71; **57**, 77, 189, 229; **58**, 107, 129, R1; **59**, 245; **63**, 73; **66**, 87; **71**, 105; **73**, 1; **74**, 95, 105, 221; **75**, 193; **76**, 151; **79**, 89, 135, 243, 251; **83**, 25, 39, 379; **88**, 379; **90**, 141; **113**, 47; **120**, 215; **126**, 219; **129**, 33; **140**, 1; **145**, 9
Paste formation **40**, 73
Paw edema **52**, 101
Paxedin **7**, 343
PD 144872 **102**, 63
Pectin **30**, 225; **34**, 207; **36**, 185; **78**, 43; **89**, 75, 177; **126**, 161; **129**, 73; **133**, 1; **136**, 43; **137**, 133; **139**, 25; **144**, 233
Pefloxacin **79**, 89; **89**, 125; **118**, 65; **121**, 123; **139**, 105
Peglicol-5-oleate **105**, 227; **106**, 15
Pellet **21**, 167
Pellet extrusion **146**, 21
Penclomedine **78**, 217; **80**, 51
Penetrance **82**, 145
Penetration **33**, 37
Penetration enhancing agent **76**, 217; **90**, 229; **91**, 9, 85; **92**, 89, 197, 203, 243; **95**, 43, 161, 181, 209; **98**, 113, 173; **99**, 109, 253; **101**, 15, 57, 115; **103**, 27, 77; **105**, 219; **110**, 231; **111**, 235; **112**, 165; **113**, 189; **117**, 173, 219; **118**, 113; **120**, 41; **121**, 89; **123**, 103, 281; **125**, 7, 63, 265; **126**, 35, 111, 267; **127**, 9, 115, 239, 245; **128**, 239, 251; **129**, 33; **130**, 141, 169, 225; **131**, 137; **133**, 71; **134**, 183; **139**, 161, 205; **141**, 17, 71; **142**, 103; **143**, 37, 119; **144**, 81; **145**, 29, 37, 49, 103, 145; **146**, 41; **147**, 165; **148**, 41
Penicillamine **4**, 337; **57**, 41; **92**, 1
Penicillin derivative **1**, 95; **14**, 123; **32**, 151; **73**, 259; **77**, 13; **81**, 1; **105**, 189; **137**, 1; **148**, 191
Penicillin G **1**, 95; **3**, 151; **7**, 55; **9**, 273; **12**, 67; **17**, 47; **21**, 239; **25**, 105; **26**, 133; **30**, 181; **37**, 113; **44**, 125; **77**, 13; **81**, 1; **98**, 83; **104**, 187; **115**, 79; **123**, 1; **124**, 149; **148**, 191
Penicillin G potassium **30**, 161
Penicillin K **17**, 47
Penicillin V **1**, 95; **3**, 151; **7**, 55; **11**, 131; **17**, 47; **20**, 25; **34**, 75; **67**, R5; **148**, 191
Penicillium chrysogenum **10**, 193
Pentaerythritol **76**, 183
Pentamethylbenzene **15**, 347
Pentamidine **67**, 275; **124**, 149; **126**, 179; **134**, 15
Pentamidine isethionate **85**, 203; **98**, 45; **99**, R1
Pentane **17**, 77
Pentanediol **15**, 185
Pentanol **1**, 15; **68**, 285; **80**, 17; **106**, 51; **126**, 219; **132**, 23
Pentapeptide **115**, 45
Pentazocine **19**, 75; **34**, 75
Pentetate technetium TC 99M **10**, 17; **25**, 191; **37**, 245; **43**, 221; **95**, 193, 229; **102**, 159
Pentetic acid **79**, 213
Pentetic acid stearate **49**, 249
Penthanil in 111 **14**, 343
Pentizidone **22**, 197
Pentobarbital **10**, 339; **20**, 73; **26**, 157; **29**, 169; **38**, 261; **41**, 139; **72**, 157; **149**, 123
Pentostatin **65**, 57
Pentyl acetate **29**, 177
4-Pentylaniline **100**, 1
1A-Pentyloxycarbonylmitomycin C **25**, 95
Peppermint oil **10**, 193; **40**, 151
Pepsin A **78**, 1; **101**, 165; **117**, 95
Pepstatin **89**, 169; **138**, 75
Peptic ulcer **28**, 85
Peptidase **113**, 1, 25; **128**, 99
Peptide **30**, 73; **48**, 15; **75**, 117; **75**, 97; **76**, 113; **76**, 99; **79**, 39; **82**, 85; **83**, 131; **87**, 141; **104**, 187; **106**, 85; **111**, 137; **113**, 25; **117**, 181; **120**, 127; **120**, 179; **121**, 45; **121**, 217; **123**, 199; **127**, 31; **127**, 43; **130**, 1; **136**, 61
Peptide bond **20**, 273
Peptide-4-methoxy-2-naphthylamide **30**, 73
Peptide-*t*-amide[1-dextro-alanine] **77**, 65
Peracetic acid **72**, 89
Percutaneous absorption **40**, 101, 105, 111
Perfluoro compound **69**, 181; **88**, 171
Perfluorodecalin **59**, 69, 131; **63**, 65
Perfluoroperhydrofluoranthene **63**, 65
Perfusion **63**, 1
Periciazine **3**, 231; **94**, 135
Perilla ketone **42**, 243
Periodontitis **142**, 9
Peristalsis **48**, 231
Peritoneum macrophage **107**, 149; **120**, 73
Peritonitis **147**, 109
Perlite **27**, 17
Permeability **17**, 215; **28**, 219; **29**, 259; **32**, 199; **45**, 59; **47**, 79; **48**, 103; **49**, 23, 47, 141; **50**, 61; **55**, 229; **56**, 1, 29, 43, 87; **57**, 211; **58**, 31; **61**, 101; **62**, 229; **63**, 189; **64**, 117, 181; **71**, 55; **77**, 1, 31; **78**, 103; **84**, 191; **99**, 197; **125**, 165; **129**, 185
Permeability barrier **112**, 191
Permeation **33**, 37; **49**, 189, 199
Peroxovanadium compound **146**, 115
Perphenazine **12**, 219; **13**, 227; **35**, 129; **36**, 185; **94**, 135
Persulfate ammonium **7**, 189
Pertechnetic acid tc 99m **20**, 315; **21**, 167; **25**, 191; **34**, 213; **40**, 201; **83**, 177
Pethidine **9**, 285; **45**, 1; **138**, 149
Petrolatum **24**, 61; **27**, 29; **107**, 51; **126**, 35
PF 127 **129**, 13
pH **1**, 151; **3**, 299; **6**, 243, 285; **7**, 137; **10**, 181; **11**, 71, 81, 329; **15**, 61; **16**, 319; **17**, 331; **18**, 325; **21**, 201, 349; **22**, 45, 197; **23**, 223; **24**, 43, 61; **25**, 85, 293; **26**, 259; **27**, 89, 117, 205; **28**, 113, 157, 239, 249; **29**, 253; **30**, 17, 29, 151, 199; **31**, 15, 75,

125, 137; **32**, 123, 143, 207; **34**, 189; **35**, 193, 211; **45**, 181; **57**, 197, 217; **59**, 35, 271; **62**, 113; **63**, 167, 237; **67**, 155; **80**, R21; **86**, 167; **89**, 103, 261
pH measurement **31**, 107
Phagocytosis **23**, 69; **29**, 53; **32**, 165; **52**, 183; **99**, 1; **104**, 41; **120**, 73; **121**, 233
Phalloidin **68**, 135
Pharmaceutical formulation **40**, 85
Pharmaceutical processing **40**, 1
Pharmaceutics **40**, 129; **69**, 239; **82**, 85, 135; **102**, 267; **135**, 1, 13, 43
Pharmacoat **39**, 1
Pharmacoat 603 **80**, 179; **99**, 37
Pharmacodynamics **6**, 205; **54**, 1; **57**, 9; **82**, 9
Pharmacokinetics **1**, 3, 41, 121, 183, 213, 219, 233, 281, 323; **2**, 41, 127, 173, 337; **3**, 13, 247, 299, 341; **4**, 195; **5**, 25, 117, 335; **6**, 43, 225; **8**, 131, 277; **10**, 77, 125, 291; **11**, 119, 155, 323; **12**, 135, 153, 299; **13**, 99, 103, 227, 239, 263, 321, 345, 353; **14**, 59, 113, 173, 251; **15**, 235, 261; **16**, 23, 47, 105, 171, 191, 203, 285, 339; **17**, 23, 237, 357; **18**, 1, 117, 345, 353; **19**, 1, 75, 229, 263, 297, 345; **20**, 43, 53, 73119, 129, 139, 347; **21**, 3, 135, 187, 239; **22**, 31, 257, 291, 311; **23**, 25, 175, 345; **24**, 115, 259, 267; **25**, 155, 347; **26**, 15, 99, 145, 157; **27**, 45, 287; **28**, 33, 85, 249; **29**, 113, 191, 195, 259; **30**, 73, 143, 199; **31**, 83, 179; **32**, 103, 133, 159; **34**, 131; **35**, 187, 211; **36**, 105, 127, 185; **37**, 49, 97, 145, 163, 219; **38**, 123; **40**, 151, 257; **45**, 101; **57**, 9; **59**, 225; **63**, 77, 83, 101; **71**, 175, 237; **72**, 79; **76**, 71, 133; **78**, 35; **79**, 71; **80**, 63, 75, 119, R15; **81**, 49, 67; **82**, 1, 9, 47, 67, 157; **83**, 155; **84**, 273; **90**, 51, 81; **95**, 127; **98**, 9; **101**, 249; **102**, 35, 249; **103**, 1, 147; **104**, 215, 253; **105**, 19; **106**, 201; **108**, 21; **109**, 181; **111**, 295; **113**, 149, 175; **114**, 117; **119**, 213, 223; **120**, 51; **121**, 1, 55, 181; **123**, 113; **124**, 119; **125**, 17, 31, 73, 91; **127**, 147, 155; **128**, 45, 55, 145, 179, 215, 239, 287; **131**, 57, 73, 91, 229; **132**, 81, 175, 189, 271; **133**, 65, 107, 149, 223, 265; **134**, 37, 119, 129, 157; **136**, 37, 53, 147; **138**, 129, 207, 223; **139**, 1, 25, 25, 45, 79, 95, 187; **140**, 33, 51, 77; **141**, 227; **142**, 235; **143**, 265; **146**, 225; **147**, 207; **148**, 55, 63
Pharmacology **34**, 35, 111; **36**, 253; **37**, 19, 27, 33, 127
Pharmacotherapy **1**, 15, 121; **13**, 183, 353; **19**, 169; **22**, 321; **23**, 25; **24**, 259; **26**, 267, 275; **27**, 245; **28**, 85, 151; **30**, 95; **32**, 159; **34**, 75, 105; **36**, 233
Pharmacy **12**, 113, 199, 209; **13**, 75; **14**, 43, 299, 349; **17**, 39, 115; **19**, 27, 63, 89, 129, 139, 229
Pharmasol **110**, 155
Pharmatose **66**, 47; **81**, 153; **88**, 159; **96**, 59; **99**, 181
Pharmatose 325 M **135**, 53
Phase transition **58**, 185; **91**, 1; **126**, 209; **128**, 203; **132**, 231; **141**, 197; **142**, 189
Phenacetin **1**, 249; **7**, 343; **13**, 35; **16**, 345; **17**, 1; **25**, 57; **28**, 67; **30**, 161; **32**, 265, 269; **35**, 111; **36**, 243; **67**, 231; **69**, 255; **116**, 191; **125**, 157
Phenanthrene **87**, 47
Phenazine **37**, 163
Phenazone **5**, 109; **14**, 123; **28**, 67; **32**, 79; **37**, 163; **64**, 235; **71**, 223; **79**, 21; **129**, 211
Phenethanol **2**, 295
Phenethyl alcohol **29**, 29; **122**, 91; **129**, 33
Phenethylamine **30**, 63
Pheneticillin **9**, 273; **17**, 47
Phenformin **89**, 1
Phenindione **8**, 303; **14**, 263; **19**, 307
Pheniramine **35**, 129
Phenmetrazine **21**, 75
Phenobarbital **3**, 341; **4**, 99, 135; **6**, 205, 277; **7**, 343; **8**, 211; **17**, 39; **18**, 89; **20**, 73; **23**, 195; **31**, 107; **34**, 75; **36**, 1; **41**, 139; **42**, 97, 181; **51**, 161, 263; **60**, 235; **63**, 95; **65**, 141; **72**, 157; **83**, 87; **92**, 15; **98**, 93; **99**, 247; **125**, 327; **145**, 9
Phenol **7**, 189; **10**, 89; **16**, 127; **18**, 299; **20**, 203; **25**, 105; **34**, 115; **35**, 227; **56**, 169; **60**, 147; **81**, 253; **100**, 189; **126**, 49, 219; **132**, 23; **140**, 155
Phenol derivative **1**, 127; **9**, 49; **14**, 123; **19**, 239; **20**, 247; **28**, 1; **91**, 39; **103**, 163; **112**, 143; **123**, 209; **132**, 23; **140**, 155
Phenolsulfonphthalein **4**, 281; **7**, 317; **40**, 257; **44**, 15; **60**, 177; **80**, 119; **93**, 91
Phenomenology **17**, 237
Phenothiazine **5**, 273; **12**, 219; **13**, 227; **16**, 153; **36**, 185
Phenothiazine derivative **16**, 271; **94**, 135
Phenoxybenzamine **25**, 293, 313
Phenprocoumon **12**, 231, 355; **18**, 9; **19**, 307; **48**, 223; **59**, 137
Phensuximide **17**, 1
N-(3-Phenyl-2-propenylidene)-aniline derivative **85**, 199
Phenylacetic acid **64**, 127; **79**, 135
Phenylacetic acid derivative **79**, 135
Phenylalanine **28**, 119; **72**, 149; **85**, 7; **115**, 139; **121**, 157; **132**, 63
Phenylalkanoic acid **64**, 127
Phenylbutazone **2**, 283; **5**, 167; **6**, 205; **8**, 303; **17**, 1; **23**, 315; **28**, 11; **54**, 1; **67**, 207; **89**, 183; **91**, 1; **92**, 151; **93**, 111; **125**, 157
Phenylbutyric acid **64**, 127
4-Phenylbutyric acid **79**, 135
Phenylcarbamic acid derivative **112**, 143
Phenylenediamine derivative **93**, 13
Phenylephrine **1**, 165; **2**, 317; **12**, 57; **29**, 243; **32**, 47; **44**, 31; **101**, R1; **130**, 179
7-Phenylheptanoic acid **79**, 135
6-Phenylhexanoic acid **79**, 135
Phenylmercuric acetate **147**, 127
Phenylmercuric borate **9**, 49
Phenylmercuric nitrate **1**, 165; **9**, 1; **16**, 127; **25**, 105; **34**, 115
Phenylmercuric salt **1**, 165
8-Phenyloctanoic acid **79**, 135
5-Phenylpentanoic acid **79**, 135
Phenylpropanolamine **12**, 57; **15**, 25; **39**, 141; **41**, 55; **47**, 89; **48**, 1, 217; **53**, 91; **85**, 29, 65; **89**, 223; **95**, 67; **103**, 97; **140**, 131; **147**, 135
Phenylpropionic acid **64**, 127
Phenylpropionic acid derivative **139**, 205
4-Phenylquinazoline **1**, 173
4-Phenylquinazolinone **1**, 173
Phenytoin **1**, 151; **5**, 67, 167; **6**, 205; **7**, 83, 129; **9**, 17; **14**, 113; **17**, 1; **23**, 35; **24**, 297; **26**, 165; **28**, 193; **29**, 73; **32**, 171; **34**, 75; **35**, 7; **49**, 163; **54**, 1; **65**, 141; **75**, 201, 219; **79**, 171; **81**, 263; **83**, 241; **86**, 175; **92**, 15; **93**, 7; **96**, 139; **98**, 189; **102**,

109, 267; **105**, 189; **109**, 27; **115**, 139; **126**, 11, 21; **131**, 153; **140**, 25; **144**, 127
Phenytoin derivative **126**, 21
Phonophoresis **20**, 235; **40**, 105
Phosphate **81**, 1; **132**, 53
Phosphatidic acid **8**, 101
Phosphatidylcholine **36**, 147; **37**, 75, 127; **38**, 83; **42**, 19; **44**, 169, 257; **48**, 133; **50**, 1, 213; **56**, 181; **59**, 69; **63**, 65; **69**, 13; **70**, 43; **76**, 161; **80**, R1; **82**, 99; **84**, 107, 117, 161, R5; **85**, 97; **87**, 183; **88**, 1, 53, 171, 319; **89**, 147, R9; **90**, 59, 255; **91**, 157; **93**, 161; **94**, 89; **98**, 173; **102**, 151; **103**, 81; **105**, 259; **106**, 51, 89; **107**, 99, 159, 239; **108**, 57; **111**, 15, 63, 191; **113**, 175; **116**, 253; **117**, 243; **118**, 65, 207, 237; **120**, 41; **121**, 195; **124**, 195; **125**, 73, 107; **131**, 57, 159; **132**, 9; **134**, 119; **140**, 119; **142**, 199, 215; **143**, 67; **145**, 17, 87, 95; **146**, 81, 255; **148**, 47
Phosphatidylethanolamine **105**, 259; **106**, 89; **108**, 57; **125**, 73; **126**, 41; **131**, 57; **134**, 119
Phosphatidylglycerol **87**, 183; **111**, 103; **132**, 9; **143**, 1; **147**, 219
Phosphatidylinositol **90**, 59; **106**, 89
Phosphatidylserine **23**, 1, 299; **44**, 269; **90**, 59; **127**, 273
Phosphocitric acid **98**, 165
Phosphodiesterase **58**, 129
Phospholipan 100 **50**, 1
Phospholipid **1**, 141; **17**, 135; **42**, 257; **49**, 179; **51**, 39; **52**, 115; **54**, 191; **58**, 13; **62**, 75; **70**, 111; **75**, 171; **79**, 263; **84**, R5; **88**, 171, 201; **91**, 1; **96**, 51; **98**, 19; **102**, 25; **104**, 41; **105**, 259; **106**, 51, 89; **108**, 109; **111**, 63; **114**, 65, 141; **116**, 253; **117**, 159; **124**, 1, 271; **125**, 107, 231; **128**, 163; **131**, 57; **132**, 9, 231; **133**, 89, 171; **137**, 117; **141**, 151; **143**, 1; **145**, 95
Phospholipid bilayer **89**, 183; **138**, 139
Phospholipid vesicle **20**, 335
Phospholipon **87**, 183
Phosphonic acid derivative **107**, 169
Phosphoramide mustard **37**, 257
Phosphoramidon **95**, 171; **128**, 229
Phosphoric acid **141**, 71; **142**, 175
Phosphorylcholine **120**, 73
Photochemistry **29**, 67; **50**, 45; **56**, 213; **99**, 351, R5
Photodegradation **31**, 65; **41**, 227; **42**, 41, 217; **52**, 123, 179; **54**, 211; **56**, 175; **60**, 157; **63**, R5; **67**, 169; **72**, 11; **76**, 187, 261; **80**, 109; **86**, 25; **87**, 79, 133; **88**, 275; **92**, 151; **93**, 85; **100**, 15; **103**, 69; **104**, 247; **107**, 199; **110**, 55; **117**, 85; **119**, 115; **120**, 137; **121**, 95, 107; **124**, 279; **129**, 275; **132**, 53; **141**, 171; **142**, 75, 83, 251
Photolysis **15**, 149; **44**, 31; **57**, 217; **60**, 157; **76**, 187; **83**, 263; **137**, 143
Photooxidation **14**, 133; **38**, 247; **76**, 187; **145**, 87
Photoproduct **20**, 265
Photosensitivity **70**, 95, 129, 195; **76**, 187; **117**, 85
Photosensitization **5**, 273; **38**, 265
Photosensitizing agent **90**, 221
Phototoxicity **3**, 205
Phthalaldehyde derivative **45**, 47
Phthalic acid **140**, 169
Phthalic acid-bis(2-ethylhexyl)-ester **113**, 241; **127**, 65
Phthalic acid dibutyl ester **29**, 37; **34**, 259; **33**, 71; **55**, 247; **103**, 47; **120**, 205; **126**, 83**129**, 279; **133**, 161; **145**, 247
Phthalic acid ethylhexyl ester **29**, 233
Phthalimide **7**, 129
4-Phthalimidobenzoic acid ethyl ester **79**, 61
Phthalocyanine **80**, 1
Physical characterization of drug-polymer system **40**, 63
Physical model **107**, 9
Physical phenomena **127**, 95; **129**, 1, 79; **130**, 149; **132**, 95; **133**, 161; **134**, 105; **141**, 205; **149**, 131
Physicochemistry **18**, 47
Physostigmine **1**, 165; **13**, 183; **109**, 283; **118**, 129; **133**, 179
Physostigmine salicylate **30**, 47; **65**, 169
Phytomenadione **26**, 289; **48**, 223; **81**, 1
Picenadol **92**, 1
Picotamide **52**, 225
Pigment **45**, 245
Pilocarpic acid **10**, 281; **20**, 285; **39**, 113; **74**, 221; **76**, 171; **88**, 397
Pilocarpine **1**, 165; **7**, 169; **10**, 219, 281; **11**, 155; **12**, 147; **16**, 203; **20**, 285; **26**, 45; **31**, 261; **33**, 45; **37**, 19; **39**, 229; **44**, 97; **47**, 95; **51**, 203; **54**, 71; **72**, 131; **74**, 221; **75**, 249, 259; **76**, 171; **79**, 233, 243; **81**, 127; **86**, 159; **88**, 397, 409; **95**, 229; **103**, 19; **116**, 77; **117**, 243; **123**, 159; **127**, 85; **129**, 41; **133**, 171; **137**, 233; **142**, 153
Pimozide **32**, 187
Pindolol **36**, 165; **46**, 95; **53**, 7; **56**, 135; **63**, 73; **99**, 303, 327; **100**, 257; **111**, 283; **120**, 215; **123**, 149
β-Pinene **124**, 279
Pinene epoxide **74**, 157
Pipemidic acid **111**, 83
Piperacillin **11**, 71; **46**, 45; **137**, 1
Piperazine estrone sulfate **71**, 25
N-(Piperidinomethyl)acetamide **9**, 7
N-(Piperidinomethyl)dichloroacetamide **9**, 7
N-(Piperidinomethyl)salicylamide **9**, 7
N-(Piperidinomethyl)trichloroacetamide **9**, 7
N-Piperidinomethylbenzamide **7**, 119; **9**, 7
Piperonal curcuminoid **131**, 1
Pirampicillin **27**, 175
Pirarubicin **68**, 151; **146**, 81
Pirenzepine **28**, 85, 151; **77**, 47
Piretanide **136**, 117
Piribedil **80**, 39
Piromidic acid **111**, 83
Piroxicam **24**, 61, 101; **53**, 257; **56**, 249; **68**, 35; **71**, 193; **74**, 85; **83**, 73; **84**, 197; **93**, 233; **98**, 29; **100**, 257; **106**, 1; **111**, 1; **117**, 219; **121**, 113; **126**, 111; **129**, 289; **131**, 137; **132**, 33; **134**, 183; **135**, 63; **137**, 133; **143**, 59
Piroxicam-β-cyclodextrin **129**, 289
Piroximone **25**, 135
7-(Pivaloyloxymethyl)theophylline **12**, 299
6-Pivaloyloxymethylthiopurine derivative **57**, 253
α-Pivaloyloxystyrene **9**, 29
O-Pivaloyltimolol **33**, 15
Pivampicillin **6**, 225; **13**, 89; **27**, 175
Pivmecillinam **13**, 89

pK_a **28**, 157; **46**, 173; **61**, 173; **72**, 193; **88**, 15
Placebo **7**, 293; **84**, 59; **91**, 75; **101**, 89; **107**, 57, 211; **110**, 93; **114**, 1, 111; **138**, 67; **140**, 229; **145**, 203
Placenta **88**, 313
Plasma **1**, 3; **4**, 75; **26**, 15; **29**, 137; **33**, 1; **64**, R1
Plasma clearance **12**, 135
Plasma protein **7**, 179; **61**, 219
Plasma substitute **99**, 285
Plasminogen **129**, 191
Plasminogen[glutamic acid] **129**, 191
Plasmodium berghei **79**, 29
Plasmodium falciparum **34**, 137
Plastibase **48**, 247
Plastic **10**, 339; **16**, 181; **17**, 145; **36**, 199; **39**, 183; **42**, 243; **88**, 351; **99**, 59; **107**, 111; **113**, 133, 241; **127**, 255; **128**, 283; **138**, 125; **140**, 169
Plasticity **146**, 159
Plasticizer **29**, 233; **86**, 49; **103**, 47, 293; **104**, 203; **111**, 127; **112**, 191; **113**, 17, 241; **114**, 205; **120**, 205; **126**, 83; **127**, 203; **130**, 187; **133**, 97, 161, 253; **138**, 57; **144**, 37; **145**, 247
Plastoid 150 **122**, 91
Platinum 195 **1**, 41
Plurol **99**, 29
Pluronic **40**, 93
Pluronic F 88 **129**, 113
Pluronic F 108 **80**, 119; **113**, 175; **129**, 113
Pluronic F 127 **13**, 47; **32**, 223
Pluronic L 61 **21**, 277
Pluronic P 85 **129**, 113
Pluronic PE 6800 **139**, 87
Pneumococcus **34**, 241
Pneumocystis carinii **67**, 275
Pneumocystis carinii pneumonia **98**, 45
Pneumonia **67**, 275
Podophyllotoxin **109**, 73
Podophyllotoxin derivative **109**, 73
Polacrilin **44**, 177; **48**, 79
Polarography **33**, 1; **42**, 235; **75**, 161; **148**, 191
Policapram **35**, 91
Polidocanol **56**, 29; **65**, 243; **70**, 19; **76**, 61; **77**, 75; **90**, 187; **99**, 29; **110**, 29; **116**, 95; **127**, 239; **128**, 29; **130**, 93, 203; **142**, 103; **143**, 75, 83
Politef **60**, 197; **83**, 163; **133**, 161
Poloxalene **22**, 207
Poloxamer **1**, 275; **12**, 147; **18**, 269; **21**, 277; **26**, 25; **29**, 53; **30**, 17; **33**, 45, 65; **34**, 145; **39**, 121; **40**, 93; **43**, 67; **45**, 91; **48**, 125; **59**, 69, 131; **61**, 79; **63**, 65, 177; **65**, 201; **66**, 213; **67**, 29; **68**, 121; **69**, 155; **71**, 65; **75**, 171; **77**, 309; **80**, 1, 119; **83**, 273; **86**, 25; **89**, 25, 33; **96**, 41; **99**, 1; **100**, 279; **102**, 19; **107**, 85; **108**, 57; **110**, 29; **112**, 65, 79; **113**, 73, 175; **114**, 121, 177; **120**, 73, 179; **125**, 283, 309; **129**, 13, 113; **130**, 75; **131**, 91; **132**, 33, 123; **133**, 107; **135**, 115; **138**, 1; **139**, 87, 113; **142**, 9, 61; **143**, 133; **144**, 99; **145**, 129, 241; **146**, 255; **147**, 131; **148**, 47
Poloxamer 237 **63**, 177
Poloxamer 238 **80**, 1
Poloxamine **69**, 155; **70**, 147; **89**, 25, 33
Poloxamine 908 **34**, 145; **52**, 221; **67**, 29; **110**, 29
Poly(acrylate-methacrylate) **19**, 229; **27**, 233; **31**, 15; **47**, 37; **55**, 47, 157; **62**, 97, 105; **76**, 55
Poly(acrylic-acid-2,5-dimethyl-1,5-hexadiene)copolymer **26**, 203; **121**, 129, 249
Poly(acrylic acid divinyl glycol) copolymer **26**, 203
Poly(alkyl-2-cyanoacrylate) **39**, 213; **128**, 189
Poly(aspartic-acid-hydrazide) **127**, 165
Poly(butyl-2-cyanoacrylate) **8**, 35; **13**, 349; **16**, 105; **30**, 17; **34**, 145; **101**, 165; **103**, 303; **128**, 189
Poly(caprolactone) **7**, 1
Poly(cyanoacrylate) **13**, 349; **17**, 65
Poly(epsilon-caprolactone) **35**, 145; **45**, 1; **84**, 85; **103**, 283; **132**, 153; **135**, 103; **139**, 113; **143**, 25; **145**, 77
Poly(ethyl-2-cyanoacrylate) **13**, 349; **111**, 31; **125**, 283
Poly(ethyl methacrylate) **17**, 39
Poly(2-ethyl-2-oxazoline) **61**, 127
Poly(hexamethylenebiguanide) **117**, 123
Poly(hexyl-2-cyanoacrylate) **13**, 349; **17**, 65; **23**, 175; **54**, 149; **55**, 39
Poly(hexyl methacrylate) **33**, 45
Poly(3-hydroxybutyric-acid) **37**, 41; **51**, 103
Poly(2-hydroxyethyl-aspartamide) **7**, 1; **64**, 239; **70**, 43
Poly-α,β[*n*(2-hydroxyethyl)aspartamide] **136**, 31
Poly-α,-β[*n*(2-hydroxyethyl)aspartamide]-ketoprofen conjugate **135**, 177
Poly[*n*(2-hydroxyethyl)aspartamide] **57**, 55; **59**, 19; **90**, 195; **99**, 135
Poly(hydroxyvaleric acid) **37**, 41
Poly(isobutyl-2-cyanoacrylate) **28**, 125; **36**, 121; **35**, 121; **72**, 211; **138**, 1
Poly(methacrylamide) **39**, 173
Poly(methacrylate-methyl-methacrylate) **47**, 263
Poly(methyl-2-cyanoacrylate) **7**, 1; **13**, 349; **29**, 267
Poly(methyl-methacrylate) **13**, 109; **14**, 95; **33**, 45; **35**, 177; **45**, 91; **60**, 197; **61**, 85; **89**, 229; **110**, 29; **136**, 107
Poly(ortho ester) **25**, 1
Poly[thio-1-(diethylaminomethyl)ethylene] **26**, 113
Poly(vinyl acetate) **30**, 209
Poly(*n*-vinylpyrrolidone-co-maleic acid) **104**, 227
Polyacryl **30**, 95
Polyacrylamide **13**, 23; **31**, 261; **32**, 165; **80**, 9
Polyacrylic acid **12**, 107; **14**, 43; **15**, 13; **17**, 273; **19**, 107; **31**, 261; **38**, 65; **48**, 247; **51**, 203; **56**, 97; **61**, 35; **73**, 131; **81**, 59; **82**, 39; **83**, 1, 221; **86**, 113; **100**, 25, 65; **103**, 243, 303; **120**, 33; **124**, 173; **126**, 79, 83; **127**, 135; **130**, 83; **131**, 47; **133**, 1; **134**, 167, 203; **136**, 155; **138**, 13; **140**, 145; **142**, 169; **143**, 13; **146**, 247; **148**, 63
Polyacrylstarch **32**, 165; **36**, 253; **40**, 215; **46**, 149
Polyalkylcyanoacrylate **40**, 23
Polyamide 6 **12**, 87
Polyamine **12**, 87
Polyaminoacid **94**, 143
Polyasparthydrazide **99**, 285
Polyaspartic acid **19**, 107; **126**, 139
Polybutylmethacrylate **33**, 45
Polycarbonate **135**, 13; **145**, 231
Polycarbonic acid **17**, 215

Polycarbophil calcium **79**, 97
Polycarbophil **19**, 107; **78**, 43; **98**, 235; **107**, 91; **112**, 87; **113**, 209; **117**, 147; **133**, 1, 149; **136**, 37; **141**, 39, 251; **142**, 163; **145**, 231
Polycyclic aromatic hydrocarbon derivative **87**, 47
Polyether **65**, 57; **84**, 85; **121**, 211; **135**, 103; **139**, 187; **144**, 107; **147**, 173
Polyetherurethan **42**, 117; **65**, 57
Polyethoxylated castor oil **31**, 231, 239
Polyethylene **2**, 295; **6**, 349; **16**, 163; **32**, 71; **34**, 137; **35**, 91, 201; **36**, 99; **39**, 183; **45**, 1; **50**, 61; **82**, 71; **101**, 35; **103**, 87; **116**, 191; **144**, 127
Polyethylene glycol **40**, 111
Polyethylene glycol 1000 **31**, 55, 175
Polyethylene glycol 1500 **3**, 55; **28**, 11; **40**, 187
Polyethylene glycol 4000 **20**, 163
Polyethylene glycol 6000 **24**, 165
Polyethylene polypropylene glycol **1**, 275
Polyethylene terephthalate **135**, 13
Polyethyleneglycol 40 **108**, 207
Polyethyleneglycol 4000 **3**, 55; **28**, 11; **31**, 175
Polyethyleneglycol-25-glyceryl trioleate **106**, 15
Polyglactin **49**, 69; **70**, 77; **99**, 239, 247, 263; **107**, 41; **115**, 61, 259; **127**, 31, 43; **129**, 263; **131**, 73, 171; **134**, 37; **136**, 89; **137**, 57, 187; **138**, 1, 85; **141**, 81, 205; **145**, 77; **146**, 147; **147**, 173; **148**, 55
Polyglutamic acid **7**, 1; **19**, 107
Polyglutaraldehyde **50**, 15
Polyglycolic acid **7**, 1; **72**, 199; **89**, 235; **115**, 87; **126**, 169
Polyglycolide **98**, 157; **103**, 37; **106**, 201; **117**, 209; **119**, 239, 251; **121**, 45; **124**, 149; **126**, 129
Polyhydric alcohol **16**, 319
Polyhydroxybutyric acid **29**, 223
Polyhydroxyvaleric acid **29**, 223
Polyisobutylene **29**, 1; **82**, 129; **92**, 55
Polylactic acid **7**, 1; **27**, 145; **29**, 223; **30**, 215; **51**, 103; **67**, 67; **72**, 199; **78**, 165; **86**, 107; **87**, 247; **89**, 235; **94**, 1; **100**, 123; **107**, 41; **115**, 87; **122**, 9; **125**, 223; **126**, 169; **131**, 171; **136**, 23; **138**, 1, 237; **140**, 219; **143**, 133; **147**, 173
Polylactide **29**, 95; **43**, 179; **51**, 1; **55**, R1; **84**, 151; **93**, 153; **98**, 157; **103**, 37, 217; **106**, 77; **106**, 201; **107**, 41; **113**, 57; **115**, 259; **117**, 209; **119**, 239, 251; **121**, 45; **124**, 149; **126**, 129, 253; **129**,51, 95, 263; **132**, 195; **133**, 107; **134**, 37; **138**, 85, 237; **142**, 135; **144**, 107; **145**, 77
Polyloc 42 **94**, 223
Polylysine **19**, 107; **61**, 163; **79**, 77; **138**, 167
Polymacon **7**, 189; **45**, 1; **46**, 179; **48**, 1; **79**, 21; **92**, 225; **99**, 293; **138**, 121
Polymer **6**, 285; **19**, 107; **23**, 35; **24**, 79; **26**, 25; **27**, 233, 299; **28**, 157; **30**, 95, 133; **38**, 65; **48**, 1; **52**, 265; **56**, 197; **57**, 1, 55, 183; **58**, 31, 63, 89, 165; **59**, 173, 197, 245; **60**, 171; **61**, 15, 35, 109, 127, 157; **62**, 97; **63**, 183; **64**, 117, 207, 239; **65**, 19, 29; **66**, 87, 157; **67**, 51, 163, 265; **70**, 53; **71**, 229, 237, 245; **72**, 1, 35, 163, 211, 243; **73**, 111, 117, 163; **75**, 241; **76**, 77, 91, 193, R5; **77**, 81, 169, 211, 221, 255, 261, 309; **78**, 43, 115, 237; **79**, 21, 29, 61, 97; **80**, 1, 9, 161, 179, 213; **81**, 31, 59, 75, 103, 243; **82**, 129; **83**, 115, 139; **84**, 1, 13, 85, 217; **85**, 1, 141; **86**, 49, 159; **88**, 75, 405; **89**, 25; **90**, 89, 195, 255; **91**, 95, 235; **92**, 35, 55, 71, 211; **94**, 31, 235; **95**, 57, 153, 193, 219; **96**, 129; **97**, 111, 133; **98**, 63, 75, 157; **99**, 13, 181, 239, 247, 263, 285; **100**, 55; **101**, 23, 165; **102**, 47, 55, 207; **103**, 47, 205, 243, 259, 283; **104**, 115, 203, 227; **105**, 65; **106**, 77, 201, 255; **107**, 85, 91; **108**, 1, 133; **109**, 115; **110**, 21, 127, 169, 267, 285, 291; **111**, 31, 127, 217, 251, 295; **112**, 37, 47, 125, 249; **113**, 9, 113, 231; **114**, 13; **115**, 29, 61, 103; **116**, 1, 39; **117**, 75, 209; **118**, 169, 207, 243; **119**, 81, 149; **120**, 33, 63, 115, 205; **121**, 45, 113, 129, 141, 211, 239, 249; **122**, 9, 27, 91, 107, 121; **123**, 65; **124**, 27, 87, 97, 149; **125**, 151, 205, 223, 243, 283, 289, 297; **126**, 1, 65, 73, 83, 129, 169; **127**, 31, 43, 115, 165, 293; **128**, 9, 23, 189, 269; **129**, 21, 95, 147; **130**, 41, 83, 187; **131**, 9, 25, 101, 229, 247; **132**, 9, 153; **133**, 1, 97, 149, 253, 265; **134**, 37, 129, 167, 173, 203, 239; **135**, 63; **136**, 23, 31, 107; **137**, 199; **138**, 57, 67, 121; **139**, 113, 187; **140**, 97, 145, 169, 219; **141**, 109, 137, 205; **143**, 25, 133, 187, 195, 255; **144**, 37; **145**, 77, 197; **146**, 159, 233, 247; **148**, 1, 63, 85
Polymer matrix **40**, 33
Polymer surface **39**, 59
Polymeric gel **40**, 93
Polymerization **25**, 121; **59**, 9; **61**, 79; **68**, 69, 89; **70**, 77; **88**, 405; **92**, 71; **95**, 153; **106**, 249; **111**, 103; **125**, 283; **142**, 75; **146**, 55
Polymethacrylic acid **42**, 145; **53**, 67; **92**, 225; **103**, 293; **137**, 159; **149**, 93
Polymethacrylic acid derivative **120**, 33
Polymethane **65**, 57
Polymorphism **8**, 11; **15**, 199, 213; **26**, 123; **40**, 1
Polymyxin **1**, 165; **12**, 67
Polymyxin B **46**, 45; **82**, 107; **102**, 201
Polyolefin **127**, 65
Polyoxyethylene **10**, 153; **13**, 47; **22**, 207; **87**, 261, 265; **93**, 21; **94**, 89; **112**, 79
Polyoxyethylene alkyl ether **92**, 219; **110**, 179; **141**, 197; **142**, 103
Polyoxyethylene cetyl ether **14**, 279
Polyoxyethylene derivative **9**, 173; **15**, 13
Polyoxyethylene ester **15**, 13
Polyoxyethylene hydrogenated castor oil **2**, 245
Polyoxyethylene oleyl ether **122**, 27; **142**, 103; **146**, 81
Polyoxyethylene sorbitan trioleate **95**, 77
Polyoxyethylene stearate **17**, 99
Polyoxyethylene stearyl ether **142**, 103
Polyoxyethyleneglycerol monolaurate **75**, 155
Polyoxyethyleneglycerol monooleate **75**, 155
Polyoxypropylene **13**, 47; **22**, 207
Polyoxypropylene polyoxyethylene **8**, 89
Polyoxypropylene polyoxyethylene copolymer **26**, 25
Polypeptide **34**, 35; **59**, 173; **50**, 261; **79**, 77
Polypropylene ethylene copolymer **50**, 61
Polypropylene **34**, 137; **39**, 59; **45**, 65; **48**, 247; **103**, 87; **107**, 111; **110**, 197; **120**, 179; **127**, 65; **130**, 251; **142**, 115; **143**, 259; **146**, 213
Polysaccharide **73**, 117; **100**, 199; **112**, 125; **121**, 113; **128**, 113; **133**, 1
Polysorbate **12**, 77; **15**, 13; **45**, 91; **69**, 143; **79**, 251; **88**, 417

Polysorbate 20 **79**, 251; **91**, 23; **93**, 215; **109**, 45; **110**, 29, 223; **120**, 179; **123**, 33; **124**, 61; **125**, 7; **128**, 29, 73; **129**, 123; **140**, 97; **144**, 217; **145**, 175
Polysorbate 40 **79**, 251; **143**, 119
Polysorbate 60 **52**, 165; **79**, 251; **121**, 149; **124**, 183; **137**, 243; **143**, 119
Polysorbate 65 **78**, 95
Polysorbate 80 **37**, 49; **43**, 93; **44**, 187; **47**, 13, 67; **55**, 165; **69**, 221; **76**, 49; **77**, 1; **79**, 123, 135, 251; **82**, 67; **96**, 147; **106**, 15; **109**, 45; **110**, 29, 211; **113**, 89; **114**, 103; **121**, 211; **122**, 1; **124**, 61; **125**, 7, 73, 283; **127**, 191; **128**, 29, 73; **129**, 123; **134**, 119, 193; **139**, 87, 223; **140**, 25, 69; **142**, 163; **143**, 133; **144**, 99
Polysorbate 81 **79**, 251
Polysorbate 85 **27**, 335; **79**, 251
Polystyrene **12**, 135; **14**, 159; **15**, 251; **52**, 221; **68**, 121; **84**, 245; **86**, 239; **88**, 303; **89**, 25, 33; **90**, R9; **99**, 1; **102**, 261; **121**, 233; **132**, 123
Polystyrene beads **40**, 63
Polystyrene **29**, 53
Polystyrenesulfonate sodium **128**, 269
Polystyrenesulfonic acid **19**, 107; **99**, R5
Polysulfone **132**, 143
Polythiazide **70**, 187
Polyurethan **103**, 199
Polyvinyl acetate phthalate **26**, 215; **28**, 157; **79**, 61
Polyvinyl alcohol **15**, 25; **16**, 163; **20**, 187; **25**, 73; **26**, 243; **33**, 219; **44**, 177; **47**, 21; **48**, 1; **49**, 231; **54**, 95; **55**, 123; **58**, 63, 135; **81**, 59; **83**, 177; **86**, 159; **87**, 239, 247; **93**, 21; **100**, 107; **102**, 207; **106**, 201; **130**, 169; **134**, 173; **135**, 1; **138**, 1; **146**, 147
Polyvinyl derivative **19**, 107
Polyvinylchloride **17**, 145, 183; **28**, 211; **29**, 233; **30**, 29, 209; **34**, 137; **39**, 183; **50**, 61; **60**, 197; **73**, 191; **88**, 351; **100**, 189; **103**, 87; **105**, 83, 89; **107**, 111; **110**, 197; **113**, 241; **114**, 177; **127**, 65, 255; **139**, 243; **146**, 213
Polyvinylsulfate **19**, 107
Porfiromycin **52**, 239
Porosity **48**, 173
Postherpetic neuralgia **123**, 159
Postoperative infection **146**, 167
Potassium **105**, 209; **126**, 199
Potassium carbonate **105**, 103
Potassium chloride **1**, 141; **5**, 167; **13**, 9, 35; **14**, 1; **15**, 25; **29**, 103; **30**, 237; **35**, 201; **37**, 15; **67**, 21; **80**, 171, R21; **104**, 95; **105**, 103; **111**, 117; **121**, 141; **139**, 177; **144**, 187; **145**, 61
Potassium hydrogen phosphate **13**, 9; **19**, 177
Potassium iodide **11**, 131
Potassium sucrose octasulfate **129**, 103
Potassium sulfate **105**, 103
Potato starch **13**, 287; **56**, 207; **55**, 175; **134**, 27, 223
Potentiometric titration **46**, 173
Potentiometry **42**, 235; **75**, 161; **77**, 41; **97**, 141
Povidone **2**, 113; **5**, 229; **6**, 179, 285; **7**, 285; **9**, 59; **12**, 341; **15**, 25, 97; **19**, 107, 177, 197; **20**, 187; **24**, 165, 275; **25**, 73; **30**, 209; **31**, 99, 157; **32**, 13; **33**, 219; **35**, 1, 7, 201; **37**, 155, 167, 175; **38**, 39, 99, 255; **45**, 139; **48**, 207, 231; **49**, 157, 231, 241; **50**, 222; **52**, 149; **53**, 47, 75; **59**, 155; **60**, 203; **69**, 239; **72**, 243; **75**, 141; **77**, 261; **79**, 257; **80**, 63, 179; **91**, 23; **99**, 173, 181; **102**, 117; **103**, 259; **104**, 169; **109**, 173; **110**, 127, 169, 223; **112**, 17; **113**, 103; **114**, 55; **119**, 71, 149; **121**, 141; **123**, 247, 257; **125**, 1; **126**, 65, 261; **127**, 293; **130**, 169; **131**, 229; **134**, 79; **135**, 1; **136**, 155; **138**, 57; **139**, 177; **140**, 207, 247; **141**, 161, 197; **142**, 61, 115, 251; **143**, 59, 133; **144**, 195; **146**, 1; **148**, 139, 201
Povidone iodine **3**, 13; **49**, 205; **65**, 35
Powder **4**, 299; **5**, 139; **10**, 43; **12**, 41, 199; **14**, 1, 209, 231; **15**, 293; **23**, 55; **44**, 1; **47**, 121; **57**, 95, R1; **58**, 209; **59**, 145; **59**, 155; **61**, 145, 251; **63**, 189, 275; **64**, 55, 227; **68**, 157; **71**, R5; **72**, 43, 157; **73**, 209; **76**, 9; **77**, 47; **78**, 95; **79**, 257; **80**, 277; **82**, 145; **83**, 11, 147; **85**, 121; **89**, R17; **90**, 203, 213; **91**, 183; **92**, 81, 123, 137, 157, 211; **93**, 249; **94**, 49; **95**, 29; **97**, 101; **99**, 37, 239; **101**, 1, 63, 81; **102**, 1, 11, 127; **103**, 55, 103, 155; **104**, 135; **105**, 125; **108**, 233; **109**, 83, 155; **113**, 123; **114**, 85, 125; **116**, 179, 191; **117**, 253; **119**, 65, 91, 103; **120**, 21; **121**, 65, 187; **127**, 109, 123, 283; **130**, 147; **131**, 201; **131**, 209; **132**, 189; **133**, 117, 133; **134**, 137, 243; **136**, 175; **137**, 41; **138**, 45; **139**, 9, 139, 149; **141**, 101; **142**, 97; **143**, 107; **144**, 141, 187; **145**, 221; **146**, 101, 159; **148**, 33; **149**, 131
Pralidoxime **109**, 35
Pralidoxime chloride **40**, 157
Prasterone **80**, 253; **106**, 33; **112**, 249
Pravastatin **136**, 117; **143**, 265
Prazepam **61**, 189; **127**, 293
Praziquantel **103**, 237
Prazosin **33**, 201; **70**, 187; **81**, 169; **105**, 169; **125**, 99; **141**, 179
Precipitation **42**, 61; **58**, 197; **69**, 13; **75**, 201; **80**, 259; **102**, 185; **109**, 27; **139**, 87; **146**, 233
Predenema **25**, 191
Prednimustine **65**, 219; **99**, 275
Prednisolone **8**, 217; **10**, 1; **12**, 77; **17**, 1; **20**, 163; **23**, 247; **25**, 191, 225; **30**, 1, 9, 215; **35**, 1; **39**, 129; **46**, 69; **48**, 1; **49**, 163; **54**, 1; **59**, 79; **63**, 95; **68**, 265; **89**, 81; **91**, 235; **111**, 295; **120**, 197; **123**, 281; **130**, 83; **138**, 95
Prednisolone acetate **10**, 1; **63**, 49
Prednisolone 21 phosphate **120**, 197
Prednisolone sodium succinate **68**, 265
Prednisone **14**, 1; **48**, 1; **50**, 213; **53**, 35; **91**, 143; **110**, 169
Pregnancy **69**, 93
Pregnenolone **132**, 23; **138**, 149
Prematurity **24**, 259
Preservative **39**, 229; **56**, 169; **60**, 147; **66**, 201; **71**, 223; **72**, 29, 51; **119**, 183; **122**, 91; **124**, 271; **139**, 63, 113; **141**, 151; **147**, 127; **148**, 131
Preservative agent **25**, 105; **29**, 29
Presolar **108**, 11
Prevention **35**, 181
Prilocaine **44**, 65; **92**, 1; **136**, 165
Primaquine **40**, 215; **68**, 111; **79**, 29; **92**, 71; **96**, 23; **100**, 15; **110**, 161; **126**, 253; **132**, 71
Primidone **17**, 1; **36**, 1; **145**, 165
Proamipide **92**, 197, 203
Probenecid **6**, 205, 225; **95**, 1
Procainamide **2**, 155; **9**, 295; **16**, 47; **28**, 145; **47**, 209; **59**, R1; **93**, 13; **146**, 55; **148**, 73

Procainamide ethyl bromide **4**, 281
Procaine **24**, 231; **36**, 7; **51**, 189; **64**, 235; **67**, 97; **81**, 103; **133**, 17
Procaine penicillin **47**, 1
Procarbazine **108**, 69; **124**, 205; **145**, 9
Prochlorperazine **3**, 231; **12**, 219; **13**, 227; **16**, 271; **52**, 159
Prochlorperazine dimesylate **16**, 153
Prochlorperazine maleate **52**, 159
Procinonide **11**, 257
Prodrug **1**, 219, 233; **5**, 67; **6**, 19; **7**, 119, 129, 169; **8**, 183; **9**, 29; **10**, 29, 181, 239; **12**, 153, 299; **13**, 89; **14**, 113; **15**, 61, 235; **17**, 91; **18**, 67, 79, 325; **20**, 273; **21**, 251, 349; **22**, 45; **23**, 117, 163, 209; **24**, 43, 307; **25**, 27, 95; **26**, 227, 317; **27**, 71, 205, 219; **28**, 1; **29**, 9, 19; **30**, 111, 151; **31**, 165; **33**, 15; **35**, 39; **35**, 243; **36**, 41; **37**, 87, 135, 145; **39**, 75, 113; **43**, 101, 261; **45**, 47; **46**, 57, 77, 159; **47**, 103, 195; **48**, 91, 159, 195; **51**, 27, 233; **52**, 255; **55**, 1, 91; **56**, 159; **57**, 41, 55, 253, 263; **58**, 143, R1; **59**, 19; **60**, 163, 253; **61**, 43; **62**, 193; **63**, 129; **64**, 75; **65**, 109; **66**, 39, 183; **68**, 77; **70**, 43, 137; **71**, 31, 45, 105, 117; **73**, 221; **74**, 67, 195; **75**, 97, 117, 201, 219, 249, 259; **76**, 99, 113, 177, R1; **77**, 21, 265; **78**, 199; **79**, 171, 205, 233, 243; **81**, 253, 263; **82**, 85, 205; **84**, 253; **86**, 175; **87**, 59; **88**, 221, 229, 237, 243; **89**, 65; **91**, 39; **92**, 105; **93**, 27, 37, 49, 177; **95**, 1; **99**, 21, 135, 189; **100**, 99, 233; **101**, 145; **102**, 109, 231; **103**, 1, 147; **104**, 157, 175; **105**, 57, 147, 169; **106**, 115; **108**, 241; **109**, 181; **110**, 189; **111**, 223; **112**, 143, 181; **115**, 45; **116**, 39, 51; **118**, 31; **120**, 189, 197; **121**, 157; **122**, 107; **123**, 65, 95, 127, 209; **124**, 45, 149; **126**, 11, 139; **127**, 85, 155; **128**, 179; **129**, 203, 223; **131**, 263; **132**, 243; **133**, 171; **134**, 147; **135**, 91, 177; **136**, 31, 61; **141**, 125, 217; **144**, 61; **145**, 103; **146**, 55, 123; **148**, 139
Progabide **43**, 83, 93; **46**, 231; **52**, 11, 19
Progesterone **4**, 249; **6**, 195; **7**, 1; **10**, 1; **14**, 191; **15**, 115; **17**, 1; **20**, 139; **26**, 203; **29**, 73, 95; **42**, 161; **46**, 133; **49**, 163; **50**, 253; **55**, 199; **64**, 61; **65**, 109; **72**, 1; **76**, 77; **89**, 41, 81; **93**, 153; **93**, 233; **94**, 1, 103; **103**, 19; **110**, 169; **115**, 133; **124**, 213; **129**, 113; **132**, 23; **138**, 95; **145**, 241
Progesterone derivative **28**, 1; **47**, 195; **89**, 41; **138**, 95
Proguanil **7**, 177; **50**, 219; **60**, 185; **100**, 249
Prolactin **75**, 97
Proline **72**, 149; **95**, 153; **104**, 187
Proline derivative **82**, 91
Prolyl endopeptidase **82**, 91; **107**, 133
Prolylglycine **104**, 187
Promazine **12**, 219; **13**, 227; **14**, 123; **16**, 271
Promethazine **3**, 231; **9**, 353; **12**, 219; **13**, 227; **16**, 153; **24**, 327; **28**, 239; **35**, 129; **40**, 223; **66**, 223, 233; **71**, 95
Promulgen D **35**, 91
Propamidine isethionate **105**, 71
Propan-1-ol **1**, 15
Propan-2-ol **3**, 247
Propanediol **15**, 185
Propanol **68**, 285; **78**, 17; **80**, 17; **92**, 81; **106**, 51; **113**, 97; **125**, 107; **126**, 219; **132**, 23
2-Propanol **17**, 39; **21**, 119; **28**, 67; **33**, 225; **49**, 129; **80**, 17; **90**, 171; **95**, 23; **106**, 51; **110**, 137; **118**, 151; **121**, 19; **126**, 261; **129**, 51; **132**, 267; **135**, 151; **136**, 71; **141**, 71; **145**, 247
Propantheline bromide **5**, 167; **33**, 125; **37**, 227
Propellant **95**, 111; **106**, 69; **118**, 229; **123**, 291; **124**, 195; **147**, 207
Propicillin **17**, 47
Propidium iodide **103**, 283
Propiomazine **144**, 217
Propiomazine maleate **141**, 53; **144**, 217
Propionaldehyde **23**, 163
Propionic acid **11**, 131; **42**, 181; **86**, 43
Propionic acid derivative **43**, 101
Propionic acid-3-hydroxymethylphenytoin-ester **79**, 171
Propionic acid methyl ester **33**, 235; **40**, 257; **55**, 17
7-(Propionyloxymethyl)theophylline **12**, 299
α-Propionyloxystyrene **9**, 29
O-Propionyltimolol **33**, 15
Propisomide **45**, 13
Propofol **127**, 255; **130**, 251; **134**, 235; **139**, 215
Propolis **26**, 243; **28**, 113; **145**, 61
Propranolol **17**, 161; **24**, 339; **28**, 145; **30**, 225; **31**, 275; **32**, 79; **34**, 75; **36**, 165; **37**, 251; **39**, 39; **40**, 223; **42**, 51; **43**, 261; **46**, 95; **47**, 1, 133; **48**, 1; **49**, 141; **53**, 189; **54**, 1; **55**, 25; **56**, 135; **59**, 197; **62**, 113; **63**, 73; **64**, 171; **66**, 233; **68**, R1; **69**, 5, 169; **71**, 55, 213; **74**, 9; **81**, 193, 235; **83**, 197; **84**, 117; **86**, 221, 247; **88**, 423; **90**, R5; **92**, 1, 35, 177, 225, 243; **95**, 29; **98**, 235; **99**, 253, 303; **100**, 143, 155, 165, 175, 257; **101**, 145; **103**, 27, 55; **104**, 19; **105**, 189; **109**, 83, 283; **110**, 291; **111**, 251, 283; **116**, 159; **118**, 199; **119**, 203; **120**, 115, 215; **121**, 37; **123**, 1, 149; **125**, 165; **127**, 109, 115; **128**, 145; **130**, 137, 179, 187; **131**, 25, 97, 229; **133**, 179; **134**, 223, 239; **137**, 255; **139**, 15; **140**, 85, 131; **141**, 9, 125, 161, 217; **142**, 169; **144**, 159, 233; **147**, 135; **148**, 1, 73
Propranolol derivative **46**, 57; **48**, 159; **101**, 145; **141**, 125
Propranolol oxime **53**, 189
Propranololctd cetylpyridinium salt **118**, 121
Propyl paraben **19**, 1; **27**, 1, 163; **32**, 223; **33**, 105; **34**, 115; **39**, 47, 229; **80**, 203; **83**, 211; **90**, 203; **109**, 155; **119**, 183; **125**, 141; **127**, 27; **133**, 17; **147**, 51
4-Propylaniline **57**, 189; **100**, 1
Propylcaine **127**, 27
Propylene glycol **1**, 71; **3**, 247; **10**, 109; **15**, 185; **17**, 77; **19**, 149; **24**, 61; **25**, 85, 359; **27**, 29, 89; **28**, 113, 201; **29**, 177; **31**, 9; **32**, 187; **33**, 37, 225; **34**, 259; **36**, 211; **37**, 203; **38**, 71; **39**, 183; **43**, 1, 17, 31; **49**, 199; **53**, 13; **54**, 51; **56**, 43; **57**, 29, 87; **61**, 205; **68**, 127; **79**, 61; **82**, 145, 171; **85**, 7; **90**, 129; **91**, 85, 157; **95**, 57; **96**, 147; **98**, 101; **99**, 109; **101**, 57; **105**, 189; **106**, 1; **109**, 283; **110**, 137; **111**, 191; **112**, 165; **114**, 75; **115**, 255; **116**, 19; **119**, 183; **124**, 183; **125**, 63; **126**, 267; **127**, 9; **130**, 1, 225; **131**, 183; **132**, 71; **133**, 71; **137**, 125; **140**, 237; **144**, 99, 127, 217; **145**, 9, 175, 187; **146**, 63; **147**, 51; **148**, 41
Propylene glycol dinonanoate **98**, 37
Propyleneglycol stearate **25**, 199
Propylthiouracil **57**, 41
Propyphenazone **37**, 179; **80**, 263; **109**, 209, 221
Prostacyclin **35**, 193
Prostacyclin derivative **135**, 127
Prostaglandin **28**, 1
Prostaglandin A_2 **106**, 167

Prostaglandin B_2 **106**, 167
Prostaglandin derivative **4**, 1; **28**, 1; **83**, 171; **132**, 165
Prostaglandin E_1 **74**, 229; **115**, 113
Prostaglandin E_2 **22**, 131; **33**, 65; **48**, 1; **74**, 229; **106**, 135, 167; **132**, 165
Prostaglandin F_2-α **29**, 259; **37**, 27; **74**, 229; **132**, 165
Prostaglandin F_2-α benzyl ester **29**, 259
Prostaglandin F_{20}-α ester **29**, 259; **37**, 27
Prostate **46**, 89; **81**, 31
Prosthesis **52**, 139; **99**, 293
Protamine **46**, 45; **53**, 253; **72**, 51
Protamine sulfate **136**, 53
Protease inhibitor AG1284 **17**, 197
Protective agent **99**, 173
Protein **28**, 151; **59**, 173; **48**, 15; **75**, 97, 117; **81**, 133; **86**, 89; **101**, 63; **105**, 19; **112**, 215; **115**, 259; **120**, 179; **142**, 227; **144**, 47
Protein A **121**, 233; **124**, 165
Protein aggregation **134**, 193; **140**, 155
Protein binding **124**, 165
Protein conformation **132**, 143
Protein degradation **95**, 171; **97**, 171; **109**, 17; **136**, 61
Protein DNA binding **72**, 251
Protein modification **148**, 131
Protein synthesis **52**, 183
Proteinase **48**, 189; **104**, 151; **107**, 133; **112**, 133; **113**, 25; **128**, 229; **141**, 39
Proteinase inhibitor **78**, 1, 9; **104**, 151; **117**, 197; **126**, 245; **129**, 263; **144**, 99
Proteus mirabilis **82**, 107
Proteus vulgaris **12**, 295
Prothrombin **18**, 9
Prothrombin complex **7**, 293
Prothrombin time **48**, 223; **131**, 131
Protionamide **35**, 129
Protirelin **49**, 231; **66**, 39, 183; **69**, 69; **72**, 199; **74**, 67; **75**, 97, 117; **76**, R1; **105**, 241; **120**, 221; **131**, 19; **137**, 33; **138**, 167
Protirelin derivative **76**, R1; **138**, 167
Proton **81**, 179
Proton magnetic resonance spectrometry **35**, 259
Proton transport **89**, 261
Protriptyline **38**, 265; **50**, 45
Proxicromil **23**, 43
Proxyphylline **38**, 139; **58**, 63
Pruv **126**, 57
Pseudoephedrine **10**, 165; **37**, 211; **53**, 91; **91**, 75; **111**, 171; **114**, 171; **123**, 119; **127**, 53; **147**, 135
Pseudoephedrine salicylate **127**, 53
Pseudoerythromycin **113**, 215
Pseudomonas **23**, 357
Pseudomonas aeruginosa **2**, 325; **9**, 49; **12**, 295; **25**, 329; **32**, 55, 91; **46**, 45, 141; **53**, 253; **54**, 51; **72**, 51; **75**, 81; **82**, 107; **87**, 195; **101**, R7; **147**, 51
Psoralen derivative **133**, 245
Psoriasis **35**, 61; **55**, 91; **105**, 227
Psychotropic agent **28**, 1; **94**, 135
Pullulan **147**, 153
Pulse polarography **3**, 239
Pupil **10**, 219; **12**, 147; **25**, 73; **32**, 47; **37**, 19; **83**, 177
Purine nucleoside **59**, 45
Puromycin **30**, 73; **76**, 247; **121**, 217
Pyran derivative **123**, 95
Pyrazinamide **126**, 179; **134**, 15
Pyrene **19**, 107
Pyrene derivative **141**, 27; **145**, 187
1,3,6,8-Pyrenetetrasulfonic acid **141**, 27
Pyridine derivative **51**, 95; **94**, 153; **125**, 297; **137**, 149
Pyridine **14**, 263; **129**, 283; **137**, 149
Pyridine-3-curcuminoid **131**, 1
Pyridostigmine **97**, 55
Pyridoxal **77**, 315; **97**, 161
Pyridoxine **81**, 1; **113**, 47
2(3-Pyridyl)benzoxazole derivative **110**, 109
Pyrimethamine **111**, 283; **145**, 9
Pyrimidine **137**, 149
Pyrimidine derivative **137**, 149
Pyrogallol **24**, 231
Pyrogastrone **28**, 249
Pyroglutamic acid **61**, 205; **113**, 247
Pyroglutamic acid dodecyl ester **80**, 81
Pyrophosphate **63**, 183
Pyrophosphate sodium **1**, 141
Pyrophosphate technetium tc 99m **69**, 189
Pyroxylin **83**, 197
2-Pyrrolidinone **21**, 17; **22**, 291; **56**, 181
7-Pyrrolidinomethyltheophylline **21**, 251
2-Pyrrolidone derivative **44**, 15; **95**, 43
Pyruvic acid ethyl ester **33**, 235
Pyruvic acid **23**, 163; **108**, 225

Q

Quality control **15**, 115
Quantitative assay **97**, 55; **100**, 227
Quaternary ammonium derivative **36**, 191; **113**, 263; **126**, 281
Quatrimycin **1**, 185
Quercetin **86**, 17; **124**, 1; **145**, 87
Quinestrol **6**, 185; **125**, 99
Quingestrone **6**, 195
Quinidine **6**, 205; **13**, 1, 353; **28**, 145; **29**, 195; **34**, 207; **43**, 179; **51**, 1
Quinidine gluconate **13**, 1
Quinine **66**, 233; **105**, 189; **146**, 193
Quinobenzoic acid derivative **2**, 239
Quinoline **137**, 149
Quinoline derivative **137**, 149
Quinoline derived antiinfective agent **79**, 89; **83**, 267; **87**, 203; **88**, 379; **93**, 121, 131; **95**, 51; **106**, 229; **118**, 65; **121**, 123; **123**, 187; **131**, 131; **138**, 113
8-Quinolinol **35**, 187; **49**, 205; **123**, 145
Quinolone derivative **63**, 237; **111**, 83
Quinuclidinyl benzilate **123**, 137

R

Racemic mixture **86**, 221; **87**, 95; **92**, 1

γ-Radiation **4**, 299; **7**, 189; **9**, 359; **17**, 29; **32**, 151; **105**, 255; **144**, 123
Radiation sterilization **4**, 299; **7**, 279; **121**, 245; **142**, 247; **149**, 85
Radical **90**, 221
Radioactive material **131**, 243
Radioisotope **32**, 251; **69**, 77
Radioisotope distribution **79**, 77; **106**, 201
Radiolysis **63**, 155
Radioreceptor assay **63**, 135; **77**, 309
Radiosensitizing agent **102**, 63
Raffinose **134**, 193
Raman spectrometry **81**, R11
Ramoplanin **144**, 71
Ranitidine **81**, 1; **82**, 233; **100**, 253; **136**, 117; **148**, 73
Razoxane **107**, 67
Reaction analysis **103**, 253
Reagent **114**, 205
Receptor binding **96**, 1; **85**, 75; **123**, 137
Recombinant erythropoietin **111**, 197; **145**, 1
Recombinant granulocyte colony stimulating factor **101**, 89; **109**, 17; **110**, 47, 93; **114**, 177; **145**, 1
Recombinant growth hormone **90**, 81
Recombinant-α-interferon **84**, 273; **101**, 63
Recombinant interleukin 2 **113**, 73
Recombinant protein **145**, 1
Recombinant somatomedin C **85**, 51
Recombinant tumor necrosis factor-α **131**, 57
Recombinant vaccine **106**, 89
Rectum **3**, 299; **9**, 107; **14**, 279; **20**, 163; **21**, 239; **27**, 71; **31**, 185; **33**, 181; **79**, 159
Rectum absorption **15**, 13; **21**, 239; **22**, 147; **30**, 247
Rectum injury **133**, 107
Rectum mucosa **22**, 147; **40**, 181
Recycling **83**, 257
Red 3 lake **50**, 163
Reduction **59**, 211
Rehydration **140**, 13
Release mechanism of theophylline **40**, 63
Remoxipride **40**, 165
Renin inhibitor **65**, 195; **73**, 231; **102**, 177
Reproterol **41**, 55
Reserpine **17**, 161
Reservoir **2**, 265
Residue analysis **142**, 67
Resin **87**, 31, 215; **101**, 263; **121**, 239
Resomer R104 **107**, 41; **134**, 37; **138**, 85
Resomer R202 **107**, 41; **129**, 51
Resomer R206 **133**, 107
Resomer R208 **138**, 85
Resomer R504 **134**, 37
Resomer RG 502 **141**, 205
Resomer RG 503 **107**, 41
Resomer RG 504 **107**, 41; **138**, 85
Resomer RG 755 **107**, 41; **131**, 171
Resomer RG 858 **107**, 41
Resorcinol **24**, 231; **78**, 89; **132**, 23; **140**, 155
Respiratory system **97**, 205; **101**, 1; **102**, 159; **112**, 181; **123**, 291
Respiratory tract **37**, 239
Reticuloendothelial system **34**, 145; **82**, 99; **90**, 73; **131**, 91
Retinoic acid **54**, 175; **65**, R9; **98**, 131; **101**, 249; **106**, 125; **110**, 241; **111**, 111; **112**, 249; **114**, 43; **115**, 235; **127**, 73; **131**, 191; **133**, 89; **134**, 99; **138**, 191
Retinoic acid derivative **98**, 131
Retinoid **29**, 73; **101**, 249; **130**, 57
Retinol **11**, 21; **19**, 207; **31**, 1; **68**, 277, 281; **81**, 1; **86**, 147; **91**, 157; **101**, 249; **112**, 249; **147**, 31
Retinol palmitate **19**, 207; **31**, 1
Reverse osmosis **58**, 155
Reverse phase chromatography **10**, 119; **13**, 333; **24**, 27
Reversed phase chromatography **31**, 125
Reversed phase high performance liquid chromatography **123**, 149; **138**, 95
RG 506 **108**, 133
RG 755 **108**, 133
RG 14620 **104**, 29
Rheogel **120**, 63; **129**, 233
Rheology **2**, 27; **12**, 19; **18**, 269
Rheumatic disease **124**, 279
Rheumatoid arthritis **24**, 101; **42**, 251
Rhinitis **123**, 47
Rhinomer **123**, 47
Rhizoxin **43**, 191; **110**, 277
Rhodamine 6G **103**, 283
Rhodamine B **48**, 133
Rhodigel **139**, 79
Riboflavin **24**, 231; **35**, 157; **74**, 25; **81**, 1; **117**, 85; **125**, 151; **144**, 225
Riboflavin derivative **147**, 95
Ribonucleotide reductase **106**, 85
Ribozyme **136**, 23
Ricinomacrogol **101**, 89; **110**, 47, 155
Rifampicin **20**, 99; **78**, 59; **108**, 125; **126**, 179; **134**, 15; **144**, 27
Rifamycin **38**, 39; **144**, 27
Rifamycin B **144**, 27
Rifamycin derivative **144**, 27
Ringer lactate solution **27**, 17; **119**, 149
Ritodrine **140**, 97; **149**, 85
RNA **52**, 183
RO 04 3780 **101**, 249
RO 11 5036 **101**, 249
Roentgen analysis **1**, 49; **29**, 95
Roentgen diffraction **24**, 1; **28**, 95; **33**, 115
Roferose Ff **135**, 53
Roferose Sf **135**, 53
Rose bengal **30**, 17; **39**, 189; **84**, 23
Rosette forming cell **103**, 303
Rosmarinus officinalis extract **63**, 57
Roxatidine acetate **119**, 109
RS 1541 **110**, 277
RU 27987 **72**, 1
RU 58841 **126**, 235
Rubber **74**, 175; **77**, 47; **79**, 301; **103**, 125

Ruga **57**, 1
Ruthenium derivative **144**, 81

S

S-1033 **132**, 165
Saccharin sodium **142**, 251
Saccharomyces cerevisiae **30**, 241; **66**, 107
Safflower oil **24**, 173; **132**, 33
Salacetamide **30**, 111
Salazosulfapyridine **71**, 55; **76**, 55; **97**, 1; **98**, 63; **139**, 95
Salbutamol **35**, 81; **41**, 55; **44**, 57; **47**, 171; **51**, 39, 67; **60**, 83; **63**, 17, 259, 267; **71**, 187; **77**, 303; **80**, R1; **81**, 199; **91**, 111; **93**, 221; **100**, 227; **101**, 1; **102**, 11; **105**, 147; **107**, 211; **108**, 233; **113**, 123; **115**, 1; **116**, 113, 169; **117**, 189; **118**, 229; **124**, 53, 195; **125**, 133; **130**, 13; **134**, 15; **137**, 261; **139**, 45; **141**, 117; **142**, 129; **148**, 1; **149**, 63, 85
Salbutamol adipate diethanolate **130**, 13
Salbutamol derivative **105**, 147
Salbutamol stearate **130**, 13
Salbutamol sulfate **86**, 153; **111**, 241; **113**, 123; **130**, 13; **136**, 89; **137**, 57; **139**, 45; **149**, 63
Salcatonin **63**, 135; **113**, 1; **114**, 215; **127**, 31, 43; **131**, 101; **135**, 137; **144**, 19; **147**, 233; **149**, 93
Salicyl arginine **99**, 189
Salicyl curcuminoid **131**, 1
Salicylamide **3**, 39; **7**, 119, 343; **13**, 273; **14**, 1; **28**, 67; **29**, 19, 195; **30**, 111; **31**, 165; **33**, 125; **71**, 31; **78**, 199; **89**, 1; **92**, 15; **130**, 83; **147**, 1
Salicylamide derivative **64**, 45; **29**, 19; **30**, 111
Salicylamide hemiglutarate methyl ester **64**, 45
Salicylamide hemisuccinate ethyl ester **64**, 45
Salicylate **40**, 111
Salicylate sodium **5**, 161; **7**, 29, 111, 189; **8**, 143; **10**, 99; **11**, 131; **12**, 251; **13**, 67; **20**, 163; **21**, 239; **22**, 147; **23**, 87, 247; **27**, 233; **30**, 247; **31**, 185; **32**, 71, 143; **33**, 89, 253; **34**, 17, 35; **38**, 83, 91; **41**, 83; **44**, 125; **45**, 59, 217; **51**, 263; **52**, 203; **55**, 77; **57**, 205; **58**, 215; **62**, 27; **68**, 157; **72**, 35; **74**, 25; **77**, 81, 107; **80**, 145; **83**, 53; **84**, 117; **93**, 91; **98**, 181; **100**, 107, 115; **104**, 95; **106**, 103; **110**, 285; **113**, 231; **119**, 203; **123**, 1; **127**, 53, 177; **133**, 155; **134**, 47
Salicylic acid **7**, 99, 189, 317; **11**, 131, 271; **14**, 95, 343; **16**, 255; **17**, 99; **21**, 35; **23**, 247; **28**, 133; **32**, 257; **33**, 253; **38**, 71; **39**, 255; **41**, 115, 205; **42**, 97; **43**, 93; **47**, 13, 179, 261; **48**, 103; **49**, 47; **51**, 1, 47; **71**, 55; **75**, 97; **76**, 151; **78**, 77; **79**, 263; **80**, 227, 263; **85**, 129; **87**, 59; **88**, 23, 111, 293; **90**, 141; **92**, 151; **94**, 1; **95**, 193; **97**, 101; **99**, 189; **101**, 23; **104**, 95, 145; **106**, 255; **114**, 137; **117**, 57; **118**, 129; **121**, 107; **123**, 143; **126**, 35; **127**, 185
Salicylic acid derivative **35**, 73; **43**, 101; **54**, 1; **68**, 77; **123**, 1
Salicylic acid ethyl ester **21**, 35
Salicylic acid methyl ester **10**, 25; **15**, 185; **16**, 255; **21**, 35; **35**, 91; **83**, 73; **137**, 149
Salicylic acid propyl ester **21**, 35
Saliva **15**, 167; **51**, 103; **81**, 127; **88**, 221, 229
Salmeterol xinafoate **145**, 65, 221; **149**, 51
Salmeterol **101**, 1
Salmonella heidelberg **34**, 241
Salmonella typhimurium **34**, 115
Salmonella typhosa **9**, 49
Salsalate **16**, 115
Saluamine **22**, 105; **31**, 65
Samarium **131**, 243
Samarium oxide **76**, 55; **98**, 63, 75
Sanaq **91**, 183; **97**, 15
Sandoz 86823 **90**, 213
Sandoz 88828 **90**, 213
Sanguinaria canadensis **46**, 255
Sanguinarine **7**, 343; **46**, 255
Saponin **9**, 173; **142**, 103
Saran wrap **44**, 25
Sarcina lutea **31**, 137
Sarcoma 180 **146**, 31
Satiagel **121**, 11
Saturated fatty acid **110**, 155; **145**, 215
Scanning electron microscopy **40**, 129; **61**, 163; **100**, 123; **142**, 143
Scavenger **58**, 237; **63**, 155; **90**, 221; **92**, 151; **121**, 107; **131**, 1; **137**, 143
SCE 785 **23**, 117
Schistosoma mansoni **56**, 23
Schistosomiasis **53**, 249; **56**, 23; **103**, 237
Scintigraphy **11**, 323; **14**, 343; **16**, 31; **19**, 169; **20**, 347; **21**, 167; **25**, 191; **32**, 251; **34**, 145; **35**, 187
γ-Scintigraphy **22**, 1; **25**, 191; **34**, 145; **40**, 81, 119; **128**, 55
Scintillation calorimeter **16**, 305
Scintillation camera **17**, 59
Scintiscanning **1**, 41; **10**, 17
Scleroglucan **78**, 43; **112**, 125; **126**, 57; **136**, 43
Scopolamine butyl bromide **106**, 77
Scopolamine **32**, 159; **123**, 285; **138**, 149
Sea cure **78**, 43
Sea cure 200 **111**, 217
Sea water **123**, 47
Sebacic acid dibutyl ester **103**, 47; **126**, 83; **133**, 161
Sebum **16**, 255
Secbutabarbital **42**, 181; **65**, 141
Secobarbital **10**, 153; **29**, 103
Secretin **75**, 117
Secretory immunoglobulin a **29**, 53
Sedative agent **9**, 353
Sedimentation rate **76**, 207
Sefsol 318 **91**, 9
Segregation analysis **80**, 171
Selenium **55**, 99
Seminal vesicle **46**, 89
Senega **7**, 343
Senodin **7**, 343
Sensitization **45**, 101
Sephadex **128**, 9, 23; **129**, 175; **139**, 25
Serine **72**, 149
Serum **1**, 323; **45**, 165; **64**, R1; **72**, 51
Serum albumin **11**, 215, 227; **132**, 175
Serum protein **1**, 61
Sesame seed oil **9**, 17; **11**, 277
Sesquiterpene **70**, 87

Sex hormone **59**, 173; **80**, 253; **103**, 19
Shear rate **18**, 269
Shear stress **18**, 269; **45**, 39
Shikonin **120**, 137; **141**, 171
Sialic acid derivative **113**, 141
Sialidase **88**, 293
Sickle cell anemia **42**, 261
Silastic 382 **57**, 149, 155
Silastic Q7 4840 **50**, 181
Silastic **27**, 29; **30**, 1, 9; **39**, 59; **48**, 247; **130**, 83
Silicic acid **42**, 77
Silicon **127**, 147
Silicon dioxide **3**, 127; **44**, 43; **45**, 259; **50**, 147; **73**, 221; **99**, 21, 173; **119**, 71; **144**, 195
Silicone **7**, 83; **12**, 87; **18**, 235; **42**, 161; **57**, 149, 155; **74**, 175; **82**, 39; **87**, 117; **92**, 177; **101**, 81; **107**, 51; **113**, 165; **121**, 27; **130**, 83
Silicone oil **49**, 69; **132**, 243; **149**, 51
Siloxane **113**, 113
Siloxane derivative **28**, 1
Silver nitrate **75**, 81
Silylation **28**, 1
Simeco **47**, 209
Simethicone **28**, 151, 249; **34**, 207
Simulation **71**, R5
Simulator **71**, R5
Simvastatin **66**, 97; **89**, 133; **125**, 141
Sitosterol **94**, 1; **107**, 1; **114**, 33
SKF 106760 **120**, 127
SKF UK 52 **57**, 9
Skin **1**, 41; **3**, 247; **7**, 99; **8**, 193, 263; **11**, 119; **12**, 153, 299; **15**, 125, 185, 235; **17**, 23; **20**, 43, 139; **21**, 17; **22**, 31, 291; **23**, 43; **24**, 61, 115, 259, 267; **25**, 85; **26**, 89; **27**, 361; **28**, 219; **30**, 35; **31**, 247; **33**, 225; **34**, 179; **35**, 91; **43**, 9; **44**, 125; **46**, 19, 193; **49**, 15, 39, 57; **50**, 197; **51**, 183; **53**, 13, 43; **56**, 1, 13, 43; **62**, 9; **68**, 127, 239; **69**, 109; **86**, 201
Skin absorption **3**, 143; **10**, 125; **15**, 185; **16**, 255; **42**, 97; **56**, 181; **57**, 29, 73; **61**, 225; **64**, 7; **65**, 211, 265; **67**, 97; **73**, 125; **82**, 171, R1; **83**, 171; **92**, 243; **102**, 141; **106**, 1; **110**, 137, 231; **111**, 235; **118**, 129; **121**, 89, 169; **123**, 41, 127; **124**, 137; **125**, 63; **126**, 111, 119, 235; **128**, 169; **129**, 211; **130**, 121, 141, 225; **131**, 117; **131**, 137, 143, 243; **132**, 71; **133**, 245; **134**, 183; **137**, 75, 117, 125, 137, 217; **138**, 149; **139**, 161, 205; **143**, 119; **145**, 29, 103, 215; **147**, 165, 243; **148**, 41
Skin allergy **88**, 251
Skin atrophy **10**, 307
Skin conductance **26**, 275; **124**, 137
Skin cream **13**, 205
Skin defect **74**, 115; **128**, 251; **145**, 87
Skin discoloration **94**, 211
Skin disease **55**, 1
Skin enzyme **27**, 325; **39**, 23
Skin fibroblast **131**, 263
Skin flap **127**, 213
Skin homogenate **8**, 167
Skin hydration **26**, 275
Skin infection **48**, 141
Skin inflammation **83**, 73; **88**, 251
Skin irritation **38**, 265; **92**, 203; **93**, 1; **95**, 181; **99**, 67; **110**, 147; **111**, 111; **113**, 159; **115**, 119; **125**, 63; **128**, 251; **130**, 121; **131**, 143; **133**, 215
Skin lipid **79**, 141; **126**, 49; **127**, 9; **128**, 203; **131**, 103, 117; **137**, 217; **144**, 81; **146**, 41
Skin penetration **30**, 63, 151; **45**, 59; **48**, 103; **55**, 91; **58**, 93; **60**, 139; **61**, 179, 205; **67**, 1, 97; **68**, 1; **70**, 87, 159, 167; **71**, 167, 193, R9; **72**, 149, 219, R1; **73**, 125; **74**, 115, 157, 229; **75**, 89; **76**, 217; **78**, 209; **81**, 39, 89; **82**, 171, R1; **83**, 171; **84**, 93, 117, 253; **85**, 7; **86**, 69, 167; **87**, 67, 265; **88**, 89, 165; **89**, 81, R1, R13; **90**, R5; **90**, 129; **91**, 9, 85; **92**, 89, 219, 243; **93**, 1, 27; **94**, 189, 203; **95**, 43, 161; **96**, 79, 105, 111, 189, 249; **98**, 37, 93, 101, 113, 131; **99**, 43, 109, 219, 253; **100**, 1, 41, 99, 219, 233; **101**, 57, 105, 219; **102**, 19, 151; **103**, 77, 131, 179, R1; **104**, 19; **105**, 31, 47; **106**, 115; **107**, 169; **108**, 141; **109**, 251, 283; **111**, 73, 191, 223, 235; **113**, 165, 189; **115**, 255; **116**, 19, 101, 201, 211; **117**, 101, 165, 173, 219; **118**, 41, 113, 207; **123**, 41, 127, 181, 281, 285; **124**, 45; **125**, 265; **126**, 35, 111, 267; **127**, 1, 9; **129**, 41, 203, 211, 223; **130**, 169, 225; **131**, 229; **133**, 39, 71, 89, 179; **134**, 239; **135**, 91, 127; **137**, 117; **139**, 161, 197; **141**, 17; **143**, 37, 47; **145**, 37, 103, 145, 175, 187; **146**, 115, 255; **147**, 219
Skin penetration cell **40**, 101
Skin permeability **4**, 309; **18**, 299; **25**, 95; **27**, 89; **30**, 151; **36**, 61, 67; **41**, 91; **43**, 233; **44**, 71; **46**, 169; **48**, 247; **49**, 189; **50**, 21, 27, 53; **55**, 17; **57**, 253; **59**, 87; **60**, 177; **61**, 179; **65**, 29, 77, 115; **66**, 79, 157, 193, 243; **67**, 97; **68**, 285; **69**, 143, 155, 193; **70**, 87, 159, 167, 173, 219; **71**, 105, 193; **72**, 149; **73**, 97; **74**, 1, 59, 137, 157; **76**, 37, 77; **77**, 21, 231; **78**, 123, 137, 209; **79**, 21; **80**, 17, 81; **81**, 89; **84**, 93, 117, 129, 253; **85**, 7; **86**, 69; **87**, 67, 261, 265; **88**, 89, 165, 391; **89**, 81, 191, 199, R1, R13; **90**, 129, 171, R5; **92**, 89, 203, 219; **93**, 27; **94**, 189, 203; **95**, 161, 181; **96**, 79, 105, 189, 249; **97**, 39; **98**, 93, 101, 113, 131, 141; **99**, 43, 51; **100**, 1, 41, 219, 233; **101**, 57, 219; **102**, 19, 141; **103**, 77, 93, 131, 163, 179, R1; **104**, 19; **105**, 31, 103; **107**, 23, 169; **108**, 141; **109**, 283; **110**, 137, 231; **111**, 7, 191; **113**, 189; **114**, 75, 237; **116**, 101; **117**, 101; **120**, 221; **121**, 89; **124**, 45, 213, 285; **125**, 195, 231; **126**, 73, 119, 219, 267; **127**, 9; **129**, 33; **130**, 169; **131**, 117; **132**, 23; **138**, 149; **139**, 197, 205; **141**, 71; **143**, 119; **146**, 41
Skin protective agent **145**, 87
Skin sensation **130**, 121
Skin toxicity **15**, 235
Skin tumor **137**, 199
Skin ulcer **104**, 49; **127**, 73
Skin water loss **113**, 247
Sleep **78**, 69
SM 10902 **135**, 127
SM 10906 **135**, 127
Small intestine **29**, 169; **44**, 187; **48**, 231; **68**, 199; **107**, 223; **112**, 133
Smooth muscle relaxation **58**, 129
Sodium **33**, 181; **64**, 217; **64**, 67; **85**, 45; **105**, 209; **126**, 199
Sodium 22 **87**, 31
Sodium azide **110**, 127
Sodium bromide **105**, 103; **135**, 31

Sodium carbonate **109**, 237
Sodium chloride **1**, 141, 249; **2**, 145; **3**, 127; **7**, 189, 269; **10**, 209; **13**, 35; **13**, 9; **14**, 1, 223; **16**, 349; **20**, 203; **21**, 155; **22**, 105, 147; **24**, 91; **25**, 359; **27**, 17; **28**, 23; **29**, 195; **30**, 101; **31**, 65; **33**, 45; **36**, 29, 99; **37**, 15; **40**, 15; **42**, 89; **43**, 41; **50**, 141, 181; **54**, 131; **57**, 155; **58**, 229; **59**, 145; **64**, 27, 35; **68**, 157; **70**, 103, 241; **73**, 249; **79**, 223; **80**, 119; **81**, 1; **85**, 19; **87**, 31, 239; **88**, 39, 369; **90**, 1; **92**, 47; **98**, 101; **105**, 103; **106**, 187; **108**, 213; **109**, 59; **113**, 39, 199; **123**, 47; **125**, 45; **126**, 95; **127**, 43; **130**, 115; **132**, 95; **136**, 71; **138**, 215; **143**, 233; **145**, 9; **146**, 175; **147**, 233
Sodium diclofenac **40**, 125
Sodium dihydrogen phosphate **50**, 141; **125**, 17
Sodium hydroxide **80**, 119
Sodium iodide **49**, 115
Sodium iodide I 131 **49**, 115
Sodium ion **20**, 335; **22**, 147
Sodium metabisulfite **24**, 91; **54**, 51; **71**, 223; **90**, 19
Sodium nitrate **33**, 45
Sodium nitrite **16**, 127; **73**, 97; **105**, 103
Sodium nonivamide acetate **130**, 121; **145**, 175
Sodium salicylate **40**, 33
Sodium stearyl fumarate **126**, 57
Sodium sulfate **1**, 141; **33**, 45; **124**, 87, 97; **135**, 31
Sodium thiosulfate **99**, 343
Sodium transport **28**, 145
Solid **42**, 77; **44**, 1; **65**, 141; **75**, 211; **83**, 87; **114**, 85, 121; **125**, 157
Solid state **10**, 1
Solid state chemistry **40**, 129
Solid tumor **142**, 25; **144**, 11
Solivito **13**, 9
Solka-Floc BW20 **112**, 153
Soluamine **99**, 333
Solubility **10**, 109; **14**, 333; **18**, 47; **31**, 175; **40**, 1; **58**, 143; **63**, 259; **86**, 263; **91**, 195; **93**, 37; **94**, 1, 11, 15, 23, 31, 153; **101**, 199, 209; **130**, 159; **140**, 237
Solubilization **55**, 165; **81**, R1; **82**, 223, 229; **83**, 53; **92**, 191; **98**, 239; **105**, 77; **110**, 169; **126**, 95; **132**, 195
Soluflazine **46**, 173
Solulan **81**, R5
Solution **11**, 71; **25**, 191
Solvation **17**, 237
Solvent **6**, 261; **14**, 263; **17**, 1, 77; **21**, 61; **28**, 193, 219; **29**, 177; **31**, 209, 225; **33**, 37; **42**, 181; **44**, 141, 225; **59**, 211, 271; **60**, 99, 133; **65**, 141; **68**, 127, 285; **72**, 43; **76**, 91; **77**, 169, 211, 221, 231; **78**, 17, 115; **84**, 151, R1; **87**, 47; **88**, 437; **89**, 25; **93**, 183; **94**, 23; **95**, 23; **97**, 101, 111; **98**, 189; **101**, 199; **104**, 125; **108**, 149; **109**, 291; **111**, 117, 137; **114**, 185, 237, 257; **119**, 183; **120**, 189, 197; **121**, 19, 27; **126**, 129, 219; **127**, 27; **129**, 51, 283; **130**, 1; **131**, 25, 171; **133**, 97; **137**, 187; **138**, 149; **140**, 45, 237; **141**, 71, 257; **142**, 251; **144**, 127; **145**, 247; **146**, 233
Soman **109**, 35
Somatomedin C **85**, 51
Somatostatin **75**, 117; **126**, 129
Sorbate potassium **55**, 175; **80**, 267
Sorbic acid **28**, 113; **72**, 51; **148**, 1
Sorbimacrogol 40 **4**, 89; **9**, 315; **28**, 47; **33**, 45
Sorbimacrogol 60 **4**, 89; **28**, 47
Sorbimacrogol laurate **2**, 343; **4**, 89; **28**, 47; **33**, 45
Sorbimacrogol oleate **1**, 183; **4**, 89, 205; **10**, 109; **11**, 277; **14**, 1; **15**, 185; **16**, 345; **19**, 35, 177; **22**, 131; **24**, 165; **28**, 47; **29**, 169; **33**, 45; **34**, 51
Sorbitan **105**, 1, 77
Sorbitan 80 **11**, 277
Sorbitan ester **42**, 1; **105**, 227
Sorbitan laurate **10**, 43, 57; **105**, 219; **108**, 117; **121**, 239; **124**, 61; **140**, 97; **143**, 119
Sorbitan oleate **10**, 43, 57; **22**, 131; **28**, 113; **50**, 27; **81**, 67; **91**, 59; **103**, 267; **109**, 45; **118**, 229; **121**, 239; **124**, 61; **125**, 315; **135**, 1; **143**, 119; **148**, 41
Sorbitan palmitate **108**, 117
Sorbitan sesquioleate **2**, 245; **28**, 47
Sorbitan stearate **10**, 43, 57; **103**, 267; **108**, 117; **124**, 183; **143**, 119; **148**, 139
Sorbitan trioleate **10**, 43, 57; **42**, 267; **80**, R1; **93**, 221; **95**, 77; **103**, 267; **109**, 45; **110**, 9; **121**, 239; **125**, 133; **129**, 113, 123; **145**, 241
Sorbitan tristearate **103**, 267
Sorbitol **11**, 131; **16**, 319; **19**, 297; **22**, 127; **35**, 1; **52**, 165; **58**, 177; **61**, 205; **66**, 149; **67**, 231; **72**, 175; **85**, 19; **88**, 417; **101**, 81; **107**, 199; **119**, 71; **128**, 1; **140**, 185
Sorbopercuten **131**, 229
Sorivudine **103**, 69; **110**, 249
Sotalol **62**, 113; **63**, 73; **105**, 65; **136**, 107; **139**, 9
Soybean oil **1**, 141; **50**, 141; **78**, 217; **109**, 45; **113**, 175; **117**, 129; **119**, 173; **125**, 107; **132**, 33; **146**, 31, 81
Soybean protein **124**, 19
Soybean trypsin inhibitor **81**, 119; **101**, 89; **113**, 1; **129**, 263
Span **99**, 29
Spectinomycin **90**, 59
Spectrofluorometry **16**, 215
Spectrometry **28**, 113; **31**, 75; **34**, 23, 247; **35**, 1, 21, 145
Spectrophotometry **1**, 49, 95, 257; **13**, 23, 67, 89, 273; **14**, 113; **17**, 357; **22**, 197; **24**, 165, 275, 297, 327, 339; **34**, 23; **37**, 103, 135; **40**, 43; **72**, 193, 207; **88**, 15; **93**, 13; **127**, 279
Spectroscopy **16**, 93, 327; **48**, 125; **65**, 201; **83**, 139; **125**, 123; **128**, 261
Sperm **125**, 81
Spermicidal agent **125**, 81
Spermine **9**, 321; **108**, 39
Spheric bead **40**, 33
Spheroid cell **96**, 59
Spheronization **146**, 1; **148**, 201
Sphingomyelin **90**, 73
Spiramycin **110**, 223
Spirogermanium **60**, 41
Spiromustine **145**, 9
Spironolactone **5**, 167; **29**, 73; **63**, 95; **73**, 9
Spleen **8**, 131
SPM 751 **135**, 1
Spore germination **25**, 105; **34**, 51
Sporicidal agent **4**, 57; **16**, 127

SQ 33600 **108**, 195
Squalene **93**, 1; **113**, 159
STA RX 1500 **55**, 143; **64**, 35; **90**, 213
Stability and antitumor activity in vitro **40**, 51
Stabilography **63**, 183
Stachyose **22**, 137
Staining **72**, 259
Staphylococcus aureus **7**, 279; **9**, 49; **12**, 295; **29**, 29; **34**, 241; **36**, 147; **45**, 153; **46**, 141; **54**, 51; **57**, 1; **60**, 147; **65**, 35; **82**, 107; **147**, 51
Staphylococcus epidermidis **2**, 325; **34**, 241; **131**, 83
Starch **1**, 249; **2**, 145; **7**, 269; **12**, 341; **17**, 313; **22**, 127; **25**, 225; **31**, 43, 99, 277; **32**, 165; **37**, 15; **42**, 77; **44**, 151; **45**, 145; **47**, 89, 233; **48**, 51; **49**, 1; **51**, 9, 63, 161, 233; **52**, 149, 183; **55**, 135, 143, 175; **56**, 51, 65; **59**, 73, 145; **62**, 125, 187; **63**, 201, 251; **64**, 55; **65**, 43, 249; **66**, 207, 273, 283; **67**, 39, 79; **69**, 43; **71**, 201; **72**, 43, 111; **74**, 25; **75**, 181; **88**, 285; **89**, 1; **89**, R17; **90**, 203; **91**, 23; **92**, 123, 137; **94**, 181, 223; **97**, 119, 161; **98**, 219; **99**, 173; **102**, 81; **105**, 255; **110**, 257; **112**, 207; **114**, 23, 55; **115**, 35, 201; **116**, 179, 191; **119**, 71; **121**, 73; **128**, 1, 73; **130**, 41; **133**, 127, 155; **134**, 27, 243; **136**, 81, 107; **139**, 79; **142**, 61; **144**, 187; **146**, 271; **147**, 23; **148**, 103
Starch derivative **45**, 145
Starch glycolate sodium **13**, 287; **30**, 237; **38**, 255; **39**, 201; **44**, 177; **45**, 79; **50**, 147; **62**, 125; **83**, 65; **97**, 119; **99**, 173; **107**, 141; **110**, 257; **114**, 55; **130**, 41; **132**, 131; **136**, 155; **144**, 71; **146**, 271
Statistics **14**, 291; **79**, 1; **82**, 145
Steady state **81**, 143
Stearate aluminum **106**, 213
Stearate calcium **14**, 1; **21**, 99; **72**, 43; **90**, 203
Steareth 21 **125**, 315
Stearic acid **4**, 271, 281; **7**, 269; **14**, 1; **17**, 313; **21**, 99; **26**, 191; **37**, 127; **50**, 111; **51**, 161; **53**, 47; **70**, 241; **74**, 49; **79**, 257; **89**, R9; **98**, 203; **106**, 1; **111**, 213; **112**, 165; **117**, 243; **119**, 71; **126**, 119; **130**, 121; **137**, 117; **148**, 47
Stearin **37**, 127; **13**, 243
Stearyl fumarate sodium **2**, 145; **126**, 57
N-Stearyl lactobionamide **128**, 65
Stearylamine **20**, 139
Stereochemistry **2**, 127; **68**, 97
Stereoisomerism **92**, 1; **101**, 127
Sterilization **10**, 109; **17**, 351
Sterlization **40**, 173
Steroid **14**, 191; **15**, 235; **28**, 1; **29**, 73; **45**, 1; **66**, 63; **94**, 1; **133**, 203; **138**, 149
Steroid-5-α-reductase inhibitor **136**, 147
Steroidogenesis **54**, 231
Sterol **107**, 1; **114**, 33; **117**, 129; **124**, 19; **146**, 31
Sterol derivative **124**, 19
Sterotex **128**, 1
Stibogluconate sodium **53**, 129; **57**, 23; **83**, 251
Stigmasterol **107**, 1; **114**, 33
Stiripentol **74**, 77
Stoichiometry **12**, 185
Stomach **4**, 27; **19**, 169; **28**, 33; **105**, 97
Stomach acid secretion **28**, 249; **43**, 83
Stomach carcinoma **129**, 103
Stomach emptying **8**, 277; **12**, 315; **17**, 59; **21**, 341; **22**, 321; **32**, 85; **34**, 213; **35**, 187; **37**, 245; **43**, 45; **48**, 79; **52**, 1; **53**, 79; **57**, 17; **63**, 101; **69**, 189; **72**, 175; **73**, 171; **75**, 241; **76**, 123, 133; **84**, 59; **88**, 433; **92**, 167; **101**, 263; **109**, 91; **114**, 1; **119**, 47; **123**, 145; **125**, 185
Stomach epithelium **129**, 103; **131**, 159
Stomach juice **16**, 345; **17**, 47; **27**, 233; **32**, 143; **125**, 327
Stomach lesion **119**, 133
Stomach motility **12**, 315; **22**, 321
Stomach mucosa **57**, 17; **62**, 53; **64**, 127; **107**, 91; **108**, 109; **116**, 223
Stomach mucosa injury **125**, 309
Stomach mucus **64**, 161; **131**, 159; **134**, 15
Stomach pH **4**, 27; **10**, 143; **20**, 99; **23**, 277; **23**, 289; **29**, 113; **47**, 263; **81**, 49; **109**, 91
Stomach scintiscanning **19**, 169; **69**, 189
Stomach ulcer **60**, 27; **119**, 133; **138**, 75
Storage **143**, 93
Stratum corneum **49**, 39, 47; **50**, 197; **52**, 47; **53**, 13, 119; **55**, 129; **56**, 75, 87, 243; **61**, 205; **68**, 1, 127; **70**, 173; **71**, 245, R1; **72**, 227; **74**, 137; **78**, 123, 137, 209; **79**, 141, 263; **80**, 17, 81; **81**, 39, 89, R11; **83**, 171; **84**, 93, 205; **87**, 175, 265; **89**, 81; **91**, 51; **95**, 43; **98**, 203; **99**, 51, 219; **103**, 277; **104**, 61; **105**, 47, 103; **108**, 141; **111**, 7; **125**, 265; **126**, 49, 219; **128**, 203; **131**, 103; **137**, 137, 217; **140**, 51; **144**, 81; **146**, 41
Strength **45**, 39; **48**, 63; **61**, 251; **76**, 9; **109**, 115; **121**, 11; **141**, 161
Streptococcus faecalis **92**, 237
Streptococcus mutans **44**, 243
Streptococcus pyogenes **12**, 295
Streptomycin **2**, 101; **4**, 271; **46**, 169; **90**, 59; **123**, 1
Streptozocin diabetes **54**, 223; **78**, 9; **128**, 215
Stress ulcer **35**, 181
Strophanthin **18**, 259
Structure activity relation **1**, 15, 189; **3**, 275; **4**, 1; **10**, 219; **15**, 61, 223, 273; **17**, 47; **18**, 247, 325ql| **19**, 149; **20**, 129; **21**, 155, 349; **22**, 345; **23**, 163; **24**, 173, 307; **25**, 27, 95, 265; **26**, 227, 317; **28**, 11, 157; **29**, 237; **31**, 165; **32**, 199; **34**, 241; **37**, 65, 87, 145; **38**, 251; **56**, 273; **62**, 125; **72**, 29; **79**, 199; **85**, 199; **94**, 81, 153; **125**, 81; **129**, 295
Structure analysis **59**, 45; **96**, 13; **117**, 231; **118**, 11
Strychnine **17**, 1
Styrene **39**, 59; **103**, 163; **138**, 149
Styrene maleic anhydride copolymer **136**, 31
Substance P|**75**, 117; **128**, 229
Substance P[1–7] **128**, 229
Substance P[1–9] **128**, 229
Succinic acid **84**, 167; **98**, 121; **101**, 89; **106**, 109; **142**, 85
Succinimide derivative **8**, 183
Succinylamidophenylglucopyranoside insulin **54**, 241, 251
Succinylsulfathiazole **35**, 111; **36**, 243; **112**, 55
Sucralfate **22**, 257; **59**, R1; **96**, 183; **99**, 173; **129**, 103; **131**, 159
Sucrose **1**, 95, 249; **7**, 189; **14**, 1, 223; **16**, 319, 349; **23**, 299; **31**, 271; **32**, 79; **37**, 15; **44**, 151; **49**, 1; **54**, 131; **58**, 177, 185; **64**, 35; **66**, 149; **73**, 37, 249; **78**, 103; **90**, 187; **100**, 65; **101**,

237; **106**, 187; **107**, 99, 199; **108**, 213; **119**, 25; **125**, 157; **130**, 245; **132**, 95; **133**, 139; **134**, 193; **143**, 13, 233; **145**, 9
Sucrose C **8**, 101; **22**, 137; **32**, 79
Sucrose ester derivative **93**, 209; **121**, 205; **137**, 177
Sucrose laurate **92**, 197, 203; **93**, 209; **137**, 177
Sucrose palmitate **93**, 209; **137**, 177
Sucrose polyester **128**, 73
Sucrose stearate **93**, 209
Sufentanil **96**, 189
Sugar **31**, 43; **64**, 55; **90**, 187; **134**, 193
Sulbenicillin **7**, 317; **17**, 47
Sulconazole **13**, 205; **14**, 209; **72**, 89
Sulfabenzamide **15**, 199
Sulfacetamide **7**, 261; **12**, 67
Sulfadiazine **25**, 121; **52**, 129; **54**, 1; **68**, 19, 215; **75**, 81; **82**, 107; **88**, 437; **92**, 237; **100**, 257; **125**, 157; **140**, 237
Sulfadiazine silver **75**, 81
Sulfadimethoxine **46**, 45; **65**, 141
Sulfadimidine **3**, 73; **7**, 285; **9**, 59; **10**, 99; **65**, 141; **68**, 215; **88**, 437; **91**, 95; **113**, 47; **140**, 237
Sulfafurazole **3**, 13; **7**, 179; **23**, 345; **30**, 123; **54**, 1; **82**, 165; **88**, 437
Sulfaguanidine **56**, 207; **106**, 33
Sulfamerazine **35**, 111; **36**, 243; **39**, 51; **65**, 141; **68**, 215; **88**, 437; **131**, 41
Sulfamethizole **4**, 291; **15**, 87; **23**, 97; **29**, 223; **30**, 123; **37**, 41, 103; **88**, 437
Sulfamethoxazole **30**, 123; **59**, 255; **63**, 173; **68**, 215; **72**, 141; **80**, R15; **85**, 163, 209; **90**, 51; **95**, 135; **110**, 55, 169; **113**, 9; **127**, 293; **135**, 63
Sulfamethoxypyridazine **88**, 437; **140**, 237
Sulfamethylthiazole **88**, 437
Sulfanilamide derivative **6**, 185
Sulfanilamide **5**, 273; **7**, 261; **12**, 67; **29**, 177, 195; **39**, 255; **41**, 159; **45**, 123; **50**, 133, 147; **51**, 137; **55**, 115; **57**, 155; **81**, 49; **88**, 437; **129**, 79; **140**, 237; **148**, 1
Sulfanilic acid **4**, 281; **30**, 35; **81**, 39; **90**, 161
Sulfaphenazole **17**, 1; **88**, 437
Sulfaproxyline **88**, 437
Sulfapyridine **5**, 273; **35**, 111; **36**, 243; **39**, 51; **88**, 437; **98**, 75; **102**, 101
Sulfaquinoxaline **65**, 141
Sulfate **105**, 39
Sulfathiazole **5**, 229, 273; **8**, 11; **14**, 1; **65**, 141; **88**, 437; **112**, 55; **125**, 157; **129**, 79
Sulfide **111**, 153
Sulfinpyrazole **97**, 1
Sulfonamide derivative **5**, 273; **18**, 311; **29**, 177
Sulfonamide **37**, 87; **45**, 47; **51**, 27; **88**, 437; **112**, 55; **148**, 1
Sulfur 35 **105**, 39
Sulfuric acid **11**, 131; **123**, 71
Sulindac **6**, 225; **126**, 95
Sulisobenzone **143**, 75, 83
Sulphanilamide **40**, 73
Sulpiride **2**, 155; **22**, 257; **29**, 73; **37**, 227
Sultopride **2**, 155
Sumatriptan **136**, 117
Sumikagel **72**, 35
Sunlight **56**, 213
Sunscreen **108**, 57
Superoxide **58**, 237; **65**, 261; **113**, 159; **131**, 1
Superoxide dismutase **75**, 97; **105**, 19; **113**, 159; **132**, 263; **142**, 75
Suppocire **40**, 181, 187; **47**, 31; **55**, 53; **85**, 103; **89**, 111
Suppository **1**, 323; **2**, 317; **3**, 39, 179, 187; **7**, 29; **9**, 107; **13**, 243; **14**, 251, 343; **17**, 273; **19**, 177; **23**, 87; **30**, 247; **38**, 103; **40**, 187; **44**, 261; **61**, 1; **73**, 17; **77**, 71; **81**, 67; **85**, 103, 113; **86**, 231; **102**, 71, 213; **104**, 1, 87; **112**, 233; **113**, 65, 209; **117**, 147; **123**, 1; **124**, 27; **125**, 31; **133**, 107; **136**, 37; **142**, 163; **148**, 155
Suppository base **3**, 55; **89**, 111; **104**, 215; **113**, 209; **114**, 117; **123**, 1; **142**, 163
Surelease **103**, 55; **130**, 187
Surface **16**, 181; **21**, 75
Surface active agent **1**, 281; **9**, 165, 315; **10**, 109, 153; **21**, 277; **27**, 117; **29**, 169; **32**, 39
Surface charge **1**, 141; **58**, 229; **72**, 43, 157; **75**, 155, R9; **80**, R21; **131**, 83; **149**, 15
Surface contact **40**, 129
Surface energy **17**, 39; **47**, 121
Surface property **103**, 19; **111**, 241; **121**, 187; **124**, 253; **125**, 141; **132**, 231; **133**, 29; **134**, 27, 79; **136**, 101; **138**, 1; **141**, 93; **143**, 107; **145**, 65; **146**, 21
Surface tension **15**, 97; **16**, 345; **17**, 39; **53**, 1; **57**, 1, 211; **58**, 203; **59**, 155; **71**, 147; **72**, 43; **75**, 171; **83**, 163; **97**, 101; **98**, 173; **109**, 155; **117**, 75; **119**, 149; **125**, 157, 327; **129**, 123; **132**, 8, 123; **133**, 171; **134**, 59; **138**, 199; **145**, 197
Surfactant **37**, 49; **39**, 263; **40**, 73; **41**, 83; **42**, 1, 267; **57**, 189; **61**, 85, 213, R1; **64**, 239; **67**, 29; **69**, 143, 155, 221; **70**, 9, 19, 59, 153; **73**, 17; **74**, 137, 169; **75**, 155, 171; **79**, 123, 135, 251; **80**, R1; **81**, 67, 111, R1, R5; **82**, 215, 223; **83**, 53, 115; **84**, 107, R5; **85**, 19, R1; **86**, 25, 43; **87**, 261, 265; **88**, 63, 211, 417; **89**, 33, 119, 139; **90**, 105; **91**, 59, 95, 105; **92**, 191, 197, 203, 219; **93**, 61, 161, 221; **94**, 31; **95**, 77, 219; **96**, 231; **99**, 29; **101**, 89, 209; **102**, 257; **103**, 119, 267; **104**, 41; **105**, 1, 137, 189, 219; **106**, 51; **107**, 51, 159, 239; **108**, 117, 141; **109**, 197; **110**, 9, 29, 47, 147, 155, 179, 267; **111**, 63; **112**, 65, 79; **113**, 57; **114**, 177; **115**, 87; **116**, 95; **118**, 221, 229; **120**, 41, 73, 179; **121**, 239; **122**, 1, 27; **124**, 61, 97, 107, 149; **125**, 99, 107, 133; **126**, 103; **128**, 29; **129**, 13, 113, 123; **130**, 93; **131**, 33; **132**, 89, 123; **135**, 73; **137**, 85, 177; **138**, 185; **139**, 113; **140**, 25, 45, 69; **141**, 197; **142**, 103, 189; **143**, 67, 75, 83, 119; **144**, 115; **145**, 175, 241; **148**, 1, 47
Survival **30**, 241
Suspension **2**, 195; **3**, 73; **7**, 285; **9**, 17, 59; **13**, 145, 159, 171; **16**, 115; **19**, 177; **42**, 267; **49**, 29; **58**, 229; **59**, 255; **71**, 223; **75**, 25; **78**, 217; **80**, 161, 271, R11; **83**, 163; **84**, 107, 129, 283; **85**, 149; **89**, 9; **90**, 1; **96**, 183; **103**, 119, 259; **108**, 117; **109**, 271; **110**, 9; **113**, 89; **115**, 95; **116**, 45; **118**, 229; **124**, 195; **128**, 73; **131**, 257; **133**, 107, 203; **134**, 71; **136**, 147; **138**, 129
Sustained release **40**, 93, 125
Sustained release preparation **1**, 197; **6**, 11; **8**, 153; **11**, 323; **13**, 1, 23; **15**, 235; **16**, 47; **18**, 1; **19**, 107, 229; **20**, 25, 53, 87; **21**, 3; **22**, 137; **24**, 327, 339, 355; **25**, 113, 121, 199; **26**, 99,

203; **27**, 157, 219, 245; **29**, 1, 37, 83, 195, 223; **30**, 1, 47, 215; **31**, 15, 43, 169; **32**, 21, 71, 85; **33**, 45, 65, 71, 147, 181; **34**, 93, 259; **35**, 21, 145, 157, 201; **36**, 73, 81; **37**, 97; **38**, 91, 231; **40**, 151; **42**, 257; **44**, 243; **45**, 65, 145; **46**, 69, 133; **47**, 37, 95; **48**, 1; **49**, 121; **50**, 133, 155; **51**, 175, 233; **52**, 249; **53**, 79; **54**, 123; **55**, 25, 47, 157, 199; **56**, 51, 65, 227; **57**, 9; **58**, 49, 73, 197; **59**, 173, 197; **60**, 125, 151, 191; **61**, 235, 241; **62**, 15; **63**, 77, 143, 201; **65**, 85, 133; **66**, 111; **67**, 113, 259, 265; **68**, 167, 255; **69**, 69, 123; **70**, 59, 77, 153; **73**, 51, 89, 157, 239; **75**, 25, 55; **76**, 133, 161; **77**, 71, 303; **78**, 35; **79**, 11; **80**, 119, 151; **81**, 49; **83**, 59, 95, 155; **84**, 59; **85**, 65, 141; **86**, 79, 123, 159; **87**, 21, 31, 117, 183, 229, 247; **88**, 99, 201; **89**, 9, 19, 177, 223, 229, 235; **90**, 89, 151, 255; **91**, 75, 95, 151, 235; **92**, 211; **94**, 59, 103, 235; **95**, 1, 127; **96**, 139, 175; **97**, 61, 93; **98**, 57, 181, 235; **99**, 13, 229, 247, 311; **100**, 33, 81, 133, 263; **102**, 47, 101, 185; **103**, 115, 243; **104**, 11, 95; **105**, 65, 181, 189; **106**, 103, 141, 213; **107**, 41, 179; **108**, 77, 133; **109**, 229, 271; **110**, 291; **111**, 89, 99, 117, 127, 171, 181, 295; **112**, 125, 215, 241; **113**, 9, 39, 65, 73, 209, 231; **114**, 117, 171, 257; **115**, 1, 17, 87, 201, 217; **117**, 31, 113, 119, 151; **119**, 1, 11, 47, 165, 223; **120**, 229; **121**, 239; **122**, 9; **123**, 1, 85, 113, 119, 237; **124**, 149; **125**, 223, 243, 297; **126**, 79, 83, 161; **128**, 113; **129**, 233, 243, 279; **130**, 187; **131**, 101, 183; **132**, 243; **133**, 1, 17, 65, 223; **134**, 37, 129, 239, 247; **135**, 73; **136**, 23, 31, 43, 107; **137**, 11, 95, 133, 225; **138**, 215, 225; **139**, 53, 79, 95, 237; **140**, 33, 131, 145; **141**, 117, 179; **142**, 169; **143**, 101; **146**, 147

Suxamethonium **39**, 263

Suxamethonium chloride **14**, 123

Sweetening agent **107**, 199

Swine **29**, 259

Syloid **99**, 173

Symperonic **69**, 155

Synaptosome **123**, 137

Synperonic **48**, 125; **65**, 201; **79**, 251; **112**, 65

Synperonic A 50 **81**, R5

Synperonic T 304 **81**, R1

Synthesis **100**, 99

Syringe pump **39**, 183

Syrup **2**, 155; **77**, 41

T

Tablet **1**, 183, 249; **2**, 145, 155; **7**, 177, 261, 269; **8**, 153; **10**, 57, 209; **14**, 313; **15**, 97, 115, 167; **21**, 167; **35**, 187; **37**, 119, 155; **40**, 125; **43**, 155; **44**, 43, 249; **45**, 19, 111; **48**, 63, 173; **49**, 1; **57**, 17; **58**, 209; **62**, 125, 153, 229; **63**, 95, R9; **64**, 27, 35, 55; **66**, 169; **67**, 21, 217, 231; **69**, 123, 239; **71**, 127, 213; **73**, 37; **75**, 241; **77**, 287; **78**, 49, 69; **79**, 83; **80**, 263; **82**, 71, 233; **86**, 99, 131; **87**, 215, 229; **88**, 23, 99, 229; **89**, 1, 19; **90**, R1; **92**, 123, 137; **93**, 111; **94**, 235; **95**, 93; **96**, 183; **97**, 15, 29, 55, 149; **98**, 219; **99**, 173, 229, 327; **101**, 35; **101**, 45, 89; **102**, 117; **103**, 55, 259; **105**, 65; **106**, 103; **108**, 11; **111**, 83; **113**, 39; **118**, 151, 191; **125**, 45; **127**, 95, 279; **129**, 1, 279; **132**, 95; **133**, 65; **134**, 79, 231; **135**, 147; **137**, 11, 225; **138**, 185, 225; **139**, 79, 177; **140**, 229; **142**, 61; **143**, 93; **144**, 153

Tablet compression **144**, 1, 147; **147**, 199

Tablet disintegration **13**, 287; **35**, 145; **37**, 245; **41**, 159; **44**, 177; **46**, 211; **51**, 63, 77; **52**, 1; **55**, 115; **56**, 207; **64**, 223; **66**, 175, 233; **67**, 39, 139; **69**, 175; **73**, 37, 171, 249; **73**, 249; **78**, 49; **80**, 129; **86**, 131, 259; **87**, 149; **88**, 9; **89**, 1; **91**, 173; **91**, 217, 227, 241; **94**, 125; **96**, 119; **97**, 15, 119, 149; **102**, 117; **120**, 33, 63; **136**, 71, 107; **143**, 13; **147**, 11

Tablet manufacture **12**, 41, 163; **13**, 35; **16**, 349; **17**, 313; **23**, 79; **25**, 207; **26**, 191, 329; **30**, 209, 237; **31**, 271; **35**, 145; **39**, 93; **41**, 1, 55; **46**, 35; **49**, 209; **52**, 213; **54**, 131; **58**, 209; **60**, 197; **63**, 23; **64**, 27, 35, 55, 223; **66**, 47, 143, 175, 233; **67**, 51, 113, 139, 217, 231; **68**, 51, 157, 199, 255; **69**, 129, 175, 239; **70**, 77, 103, 241, 261; **71**, 81, 127; **72**, 111, 117; **73**, 37, 239, 249; **76**, 9; **77**, 287; **81**, 187, 243; **83**, 187; **84**, 191; **85**, 57, 65; **85**, 29; **86**, 49; **87**, 117, 133, 149; **88**, 9, 99, 261, 285, 345; **89**, 1, 19, 177, 223, R17; **90**, 151, 213; **91**, 143, 151, 167, 173, 183, 217, 227, 241; **92**, 29, 35, 55, 157; **93**, 111, 249; **94**, 15, 39, 49, 125, 181; **95**, 201; **96**, 119, 167, 175, 197, 205, 217, 225; **97**, 15, 29, 79, 119, 149, 183, 195; **98**, 57; **99**, 7; **100**, 49, 107, 115, 155, 165, 175, 241, 257, 263; **101**, 71; **102**, 1, 87, 117, 207; **103**, 115, 205; **104**, 115; **105**, 11, 65, 97, 255; **106**, 103, 141, 149, 157, 187; **107**, 179, 247; **108**, 155, 207; **109**, 1, 59, 229; **110**, 37, 75, 203; **111**, 89; **112**, 37, 125, 173, 225; **114**, 23, 85; **115**, 29, 35, 53, 201; **116**, 65, 131, 179, 191; **117**, 13, 57; **119**, 65; **120**, 13; **121**, 65, 205; **123**, 85, 119; **124**, 69, 231; **125**, 257; **128**, 1, 113; **129**, 21, 233, 253; **132**, 95, 183, 207; **133**, 253; **134**, 79; **136**, 13, 107; **139**, 177; **140**, 185, 195, 207; **141**, 101; **143**, 151, 233; **146**, 71

Tablet weight **40**, 15

Tablettose **48**, 149; **56**, 51

Tacrine **95**, 161; **109**, 283; **114**, 75; **133**, 179

Tagat 02 **75**, 155

Tagat l2 **75**, 155

Tail flick test **38**, 171

Tak 029 **146**, 147

Takazyma **28**, 239

Talampicillin **75**, 117

Talc **22**, 127, 265, 273; **24**, 355; **25**, 225; **31**, 15, 43; **37**, 119, 203, 211; **48**, 51, 63, 113; **50**, 163; **88**, 261; **99**, 173; **128**, 1; **133**, 139

Tamarind gum **133**, 1; **134**, 173

Tamoxifen **99**, 59

Tamoxifen citrate **99**, 59; **108**, 69; **124**, 205

Tannin **24**, 355; **133**, 1

Tapioca **55**, 175

Target **23**, 333

Target cell **46**, 31

Tartaric acid **84**, 167; **101**, 89

Taurinamide **49**, 135

Taurine **24**, 61

Taurocholate sodium **12**, 1; **27**, 117

Taurocholic acid **4**, 219, 271; **6**, 137; **8**, 303; **9**, 173; **11**, 27; **31**, 193; **47**, 13; **64**, 161; **69**, 221; **74**, 95; **83**, 197; **84**, 223; **89**, 81; **91**, 105; **93**, 91; **96**, 1; **98**, 141; **101**, 15, 115; **103**, 27; **104**, 145, 187; **105**, 137; **107**, 159; **109**, 197; **111**, 147; **116**, 147; **118**, 55; **122**, 91; **137**, 85

Taurodeoxycholic acid **8**, 143, 303; **12**, 219; **28**, 33; **31**, 193; **38**, 211; **56**, 269; **74**, 95; **91**, 157; **111**, 147; **114**, 65; **127**, 239; **142**, 103; **148**, 47

Taurodihydrofusidate **59**, 263; **65**, 243; **76**, 61; **103**, 27

Taurolidine **48**, 167; **49**, 135; **64**, R1; **126**, 261

Tauromustine **56**, 37; **71**, 73
Taurultam **48**, 167; **49**, 135
Taxol **108**, 69; **124**, 205; **127**, 65; **132**, 195; **133**, 191; **137**, 199
Tazifylline **95**, 161
TC 5 **81**, 49
Technetium 99m **4**, 175; **35**, 187; **40**, 207; **42**, 211; **64**, 1; **66**, 169; **68**, 199; **74**, 203, 209; **80**, R1; **87**, 215; **91**, 111; **96**, 183; **100**, 81; **102**, 11, 101, 127; **108**, 125; **114**, 1; **115**, 209
Technetium sulfur colloid Tc 99m **8**, 217; **12**, 315; **83**, 177; **128**, 55
Tefose **99**, 29
Tegafur **25**, 347; **38**, 239; **41**, 21; **48**, 1; **108**, 69, 241; **123**, 1; **124**, 205; **145**, 9
Tego betain l7 **75**, 155
Teicoplanin **142**, 235; **146**, 167
Temafloxacin **63**, 237; **83**, 267; **88**, 379; **93**, 121
Temarotene **101**, 249; **106**, 15; **130**, 57
Temazepam **22**, 311; **31**, 9; **34**, 17; **78**, 189; **110**, 169; **136**, 81
Temperature dependence **135**, 13
Temperature measurement **92**, 157
Teniposide **105**, 89; **108**, 69; **109**, 73; **124**, 205
Tenoxicam **37**, 219; **147**, 219
Tenside **1**, 183
Tensile strength **10**, 43, 57; **37**, 119; **61**, 101; **63**, 23; **67**, 59; **95**, 29; **118**, 169; **125**, 157; **126**, 189; **132**, 207; **135**, 103; **141**, 101
Terbutalin sulfate **1**, 213
Terbutaline sulfate **106**, 213; **119**, 91, 103; **149**, 85
Terbutaline **1**, 213; **3**, 335; **8**, 1; **41**, 55; **74**, 209; **90**, 171; **97**, 47; **101**, 1; **113**, 47, 123; **116**, 169; **119**, 91, 103; **124**, 149; **130**, 1; **134**, 137; **142**, 247
Terephthaloyl chloride **103**, 253; **128**, 197
Terfenadine **141**, 257
Terodiline **98**, 113; **124**, 9
Terpene **116**, 237; **127**, 9; **130**, 141; **143**, 37; **145**, 37, 49
1-Tert-butyl-7(2,5-diazabicyclo[2,2,1]heptan-2-yl)-6-fluoro-1,4-dihydro-4-oxo-1,8-naphthyridine-3-carboxylic acid **140**, 1
2-Tert-butylcyclohexanone **68**, 239
3[3-Tert-butylthio-1(4-chlorobenzyl)-5-isopropyl-2-indolyl]-2,2 **103**, 259
3[3-Tert-butylthio-1(4-chlorobenzyl)-5(2-quinolinylmethoxy)-2-indolyl]-2,2-dimethylpropionic acid **121**, 73
Testosterone **4**, 249; **6**, 185; **7**, 99; **10**, 1; **11**, 119; **14**, 325, 333; **16**, 255; **17**, 1; **20**, 43; **24**, 173; **25**, 265; **29**, 73; **33**, 65; **42**, 235; **49**, 163; **56**, 197; **59**, 173; **61**, 135; **63**, 281; **71**, 55; **76**, 77; **77**, 231; **79**, 257; **80**, 243, 253; **83**, 197; **94**, 1; **99**, 209; **100**, 41; **103**, 19; **105**, 189; **116**, 147; **125**, 157, 165; **131**, 263; **132**, 23, 267; **133**, 47; **138**, 149, 191; **140**, 237; **148**, 1
Testosterone acetate **24**, 173
Testosterone derivative **28**, 1; **131**, 263
Testosterone enantate **24**, 173
Testosterone ester **131**, 263
Testosterone metabolism **131**, 263
Testosterone oleate **24**, 173
Testosterone palmitate **24**, 173
Testosterone propionate **53**, 47
Testosterone undecanoate **24**, 173
Tetanus toxoid **8**, 217; **65**, 235; **88**, 335
Tetrabid **56**, 151
Tetrabutylammonium salt **8**, 143
Tetracaine **11**, 131; **51**, 183; **62**, 9; **78**, 209; **94**, 75; **98**, 101
Tetrachlorsalan **1**, 127
Tetracosactide **131**, 171
Tetracycline **1**, 165, 185; **5**, 273; **13**, 23; **14**, 133; **26**, 259; **28**, 33; **34**, 75; **35**, 211; **38**, 211; **40**, 223; **51**, 103; **56**, 151; **66**, 233; **89**, 75; **92**, 15; **98**, 231; **100**, 155, 165; **142**, 9
Tetracycline derivative **123**, 187
Tetradecane **42**, 97
Tetradecylmaltoside **132**, 107
Tetradecyltrimethylammonium bromide **57**, 189; **124**, 107
1,3,4,14b-Tetrahydro-2,7-dimethyl-2h-dibenzo[b,f]pyrazino [1,2d]-1,4 **3**, 39
1,2,3,4-Tetrahydro-2,4-quinazolinedione derivative **50**, 117
Tetrahydrocannabinol **6**, 195; **74**, 59
δ-8-Tetrahydrocannabinol **43**, 9, 17
Tetrahydrofuran **34**, 217; **129**, 51
Tetrahydrouridine **31**, 185
Tetrahymena **7**, 89
Tetrakis(4-amidinophenoxy)neopentane **72**, 251
Tetramethylammonium chloride **48**, 159
Tetramethylammonium hydroxide **17**, 325
Tetramethylurea **14**, 263
Tetramisole **39**, 255
Tetrandrine **109**, 135
Tetrapeptide **101**, 57
Tetrazole derivative **30**, 101
Tetronic **69**, 155
Tetronic 1508 **75**, 231
Tetrylammonium **98**, 141; **131**, 257
Tetrylammonium bromide **49**, 57
Tetryzoline **35**, 129
Texofor A30 **82**, 223
Thalidomide **92**, 1
Thebaine **9**, 245; **32**, 7
Theo SR **52**, 63
Theobroma oil **3**, 39; **9**, 107; **22**, 131; **47**, 31
Theobromine **17**, 1; **18**, 127, 259; **32**, 187; **140**, 237
Theophylline **3**, 299; **4**, 135; **6**, 11; **8**, 153; **10**, 177; **12**, 299; **13**, 23; **14**, 1, 79, 299; **18**, 127, 259; **21**, 3; **24**, 259; **25**, 113; **26**, 99; **28**, 67; **29**, 73, 195; **30**, 143; **31**, 15; **33**, 55, 71; **34**, 75, 153; **35**, 243; **36**, 73; **37**, 97, 119, 211; **38**, 139; **40**, 63, 223; **42**, 15, 35; **43**, 139; **44**, 87; **45**, 145; **47**, 31; **48**, 1, 195; **49**, 179; **50**, 35, 79, 205; **51**, 195; **52**, 63, 109; **53**, 47; **54**, 1, 123; **55**, 143; **56**, 65; **58**, 31, 129; **59**, 105; **60**, 125, 171; **62**, 15; **63**, 201; **65**, R5; **66**, 111, 223; **69**, 123; **70**, 159; **72**, 175; **73**, 89, 97; **77**, 183, 255; **79**, 67, 257; **81**, 39; **82**, 117; **83**, 59; **85**, 149; **86**, 231; **87**, 117; **88**, 75, 141; **92**, 115; **95**, 135; **97**, 61, 101, 183; **98**, 51, 181; **103**, 115; **104**, 95; **106**, 33, 149; **108**, 149, 155; **111**, 271; **112**, 241; **114**, 247; **116**, 65; **117**, 23; **118**, 251; **119**, 165, 203; **121**, 205; **125**, 157; **126**, 57, 83; **129**, 243; **132**, 1; **133**, 17; **135**, 151; **136**, 117; **137**, 75; **138**, 199; **139**, 95; **140**, 131, 237; **141**, 53; **143**, 93; **144**, 209; **147**, 23, 41, 135; **148**, 1, 73, 85

Theophylline derivative **13**, 89; **21**, 251; **43**, 119; **44**, 87; **55**, 1
Theoretical study **97**, 149
Theory **46**, 35
Thermal analysis **5**, 109; **16**, 327; **26**, 123; **29**, 95; **31**, 25, 157; **33**, 115; **40**, 129; **45**, 13; **68**, 179; **74**, 43; **77**, 193, 287; **89**, 119; **126**, 287; **135**, 13
Thermal conductivity **113**, 133
Thermodynamics **3**, 319; **10**, 25; **17**, 313; **25**, 135; **58**, 107, 177; **62**, 157; **63**, 73; **87**, 47, 95; **94**, 135; **101**, 199; **104**, 125, 239; **111**, 51, 159; **129**, 51
Thermography **91**, 1
Thermogravimetric analysis **40**, 129
Thermostability **96**, 41, 243; **97**, 161, 219; **134**, 193
Thiamine **48**, 1; **57**, 41; **81**, 1
Thiamphenicol glycinate **40**, 85
Thiazinamine **3**, 39
Thiazinamium methyl sulfate **3**, 55
Thiazole derivative **67**, 265
1,3,4-Thiadiazole derivative **50**, 117
Thickening agent **133**, 265
Thickness **133**, 253
Thiethylperazine maleate **16**, 271
Thin layer chromatography **1**, 257; **11**, 329; **14**, 113; **15**, 149; **20**, 273; **31**, 157; **34**, 247; **35**, 7
Thiobarbituric acid **142**, 199
Thiocyanate **69**, 197
Thiocyanate ammonium **11**, 131
Thiocyanate potassium **63**, 155; **124**, 173
Thioglycolic acid **28**, 119
Thioglycolic acid ethyl ester **28**, 119
Thioglycolic acid methyl ester **28**, 119
Thioguanine **19**, 17
Thiol group **134**, 147
Thiomalic acid **28**, 119
Thiomersal **23**, 357; **89**, 213; **121**, 217; **139**, 113
3-Thiomorpholinone **76**, 217
Thiopental **10**, 153, 339; **14**, 123; **17**, 183
Thiophene **137**, 149
Thiophene derivative **137**, 149
Thiopropazate **5**, 273
Thioproperazine methanesulfonate **16**, 271
Thioridazine **3**, 231; **16**, 271; **36**, 185; **94**, 135
Thiorphan **43**, 23; **112**, 133; **121**, 217
Thiosulfate **69**, 197
Thiourea **29**, 177; **32**, 79; **36**, 61, 67; **63**, 155; **145**, 9
Thiourea derivative **50**, 117
Thoracic duct **24**, 185
Thorax **115**, 87
Threonine **72**, 149
Throat **101**, 81
Thrombin **9**, 321; **90**, 95; **91**, 235
Thrombin time **9**, 321
Thrombocyte aggregation **52**, 101
Thromboplastin **9**, 321
Thrombospirin **108**, 11
Thrush **130**, 257
Thymidine **116**, 217
Thymidylate synthase inhibitor **105**, 227
Thymocyte **16**, 153
Thymol **126**, 219; **128**, 81; **130**, 141
Thymopentin **70**, 29; **98**, 19; **132**, 259
Thymopoietin **98**, 19
Thymostimulin **80**, 93
Thymus **15**, 235
Thymus hormone **80**, 93
Thyrotropin **107**, 231
Thyroxine **51**, 175; **92**, 1
Tiapride **2**, 155; **9**, 295
Tiaprofenic acid **83**, 155
Ticarcillin **26**, 133; **32**, 151; **37**, 113; **46**, 45
Ticaxan **128**, 113
Tiemonium iodide **16**, 241
Tight junction **123**, 13
Tilisolol **93**, 49
Tiludronic acid **123**, 13, 71
Timefurone **57**, 223
Timegadine **46**, 149
Timobesone acetate **26**, 145
Timolol **32**, 159; **33**, 15; **38**, 9; **48**, 247; **57**, 163; **61**, 235, 241; **63**, 73; **65**, 19; **75**, R1; **78**, 237; **81**, 59; **84**, 217; **86**, 137; **92**, 1; **95**, 219; **99**, 253; **100**, 257; **123**, 149; **124**, 107; **128**, 179; **142**, 103
Timolol derivative **46**, 77
Timolol maleate **81**, 59; **89**, 235; **126**, 79; **133**, 171
Tin chloride **26**, 303
Tin derivative **86**, 107
Tincture **48**, 119
Tinidazole **99**, 189
Tioconazole **19**, 149
Tioguanine **95**, 51; **124**, 261; **143**, 171
Tiopinac **7**, 245
Tirilazad **66**, 63
Tissue distribution **90**, 73; **126**, 235
Tissue fixation **34**, 75
Tissue level **45**, 101; **47**, 265; **53**, 67
Tissue plasminogen activator **129**, 191
Tissue regeneration **130**, 93
Tissue slice **72**, 227
Titanium dioxide **22**, 17, 265, 273; **48**, 63, 113; **50**, 163; **53**, 47; **79**, 257; **87**, 133; **105**, 157
Titrimetry **87**, 11; **97**, 141; **134**, 1
Tixocortol-17-butyrate-21-propionate **46**, 49
Tobramycin **81**, 1; **101**, R7; **108**, 39; **123**, 173
Tobramycin sulfate **26**, 133
Tocainide **12**, 163
Tocopherol **148**, 41
α-Tocopherol **8**, 101; **22**, 299; **26**, 289; **32**, 39; **78**, 95; **84**, 93; **86**, 137; **88**, 211; **91**, 209; **93**, 1; **104**, 49; **111**, 261; **127**, 73; **138**, 191; **140**, 251
Tocopherol acetate **26**, 289
Tofisopam **61**, 189
Tolazamide **5**, 223
Tolazoline **101**, R5; **123**, 181

Tolbutamide **5**, 167, 223; **14**, 1; **42**, 27; **58**, 221; **94**, 69; **117**, 247; **125**, 157; **129**, 63; **133**, 117; **140**, 237; **146**, 93
Tolfenamic acid **54**, 1; **108**, 109
Toliprolol **36**, 165; **46**, 95
Tolmetin **2**, 283; **86**, 131; **103**, 217
Tolnaftate **35**, 129
Toluene **16**, 127; **17**, 77; **76**, 91; **126**, 219; **129**, 51, 283; **138**, 149
Toluenesulfonamide **7**, 129; **37**, 87
Toluenesulfonamide derivative **47**, 103
Toluenesulfonic acid **8**, 143
4-Tolylacetic acid **27**, 39
N-4-Tolylsulfonyl-dimethylsulfimine **37**, 87
Tolylsulfonylurea derivative **37**, 87
Tomato **107**, 223
Tonic **22**, 311
Tooth enamel **15**, 177; **16**, 171
Topical agent **18**, 53; **19**, 245; **35**, 243; **39**, 23; **60**, R1; **82**, 21
Topical drug administration **56**, 243
Topotecan **127**, 229
Toprolol XL **140**, 229
Total parenteral feeding **2**, 325
Total parenteral nutrition **54**, 143, 191; **55**, 99; **58**, 13; **64**, 67; **83**, 227; **87**, 167
Toxicity **40**, 23
Toxicity testing **58**, 41; **59**, 231; **119**, 37
Toxicokinetics **82**, 9
Trace element **16**, 57; **81**, 1
Trachea **12**, 57, 67, 77
Trachea mucosa **12**, 57, 67, 77
Tramazoline **12**, 57
Transcutol **49**, 199
Transdermal drug administration **43**, 201; **56**, 13, 181; **58**, 93; **61**, 179; **67**, 1; **71**, 237; **82**, 39, R1; **86**, 167; **123**, 127; **135**, 1; **137**, 177; **148**, 177
Transdermal drug delivery **24**, 259, 267; **30**, 63; **32**, 159, 243
Transdermal drug delivery system **55**, 147
Transplatin **118**, 85
Transport kinetics **60**, 133
Trasyl **95**, 101
Trehalose **115**, 193; **134**, 193
Tri tab **110**, 37
Triacetin **37**, 203; **73**, 197; **86**, 113; **121**, 19; **133**, 161; **138**, 57; **145**, 247
Triacylglycerol **38**, 103; **43**, 241; **85**, 103; **104**, 1, 87; **110**, 155; **119**, 173; **124**, 271; **125**, 107; **129**, 159; **131**, 159; **136**, 147
Triacylglycerol lipase **115**, 45; **118**, 55; **142**, 1; **144**, 115
Triamcinolone **10**, 1; **46**, 69; **77**, 199; **138**, 95
Triamcinolone acetonide **3**, 247; **10**, 1; **16**, 339; **17**, 1; **19**, 245; **27**, 145; **44**, 25; **60**, 139; **66**, 53; **70**, 87; **77**, 199; **95**, 51, 105; **109**, 283; **110**, 169; **121**, 249; **124**, 195; **127**, 1; **133**, 179
Triamcinolone acetonide H 3 **3**, 247; **16**, 339; **27**, 145
Triamcinolone derivative **121**, 129
Triamcinolone diacetate **10**, 1; **16**, 93
Triamterene **6**, 205; **57**, 9; **76**, 187; **123**, 25
Triazinetrione triglycidide **10**, 109
Triazole **137**, 149
Triazole derivative **137**, 149
1,2,4-Triazole derivative **81**, 103
Tribenoside **6**, 43
Tributyrin **78**, 217
Tricaprin **13**, 243
1,1,1-Trichloro-2-(2-chlorophenyl)-2-(4-chlorophenyl) ethane **25**, 57; **30**, 161
N-(2,2,2-Trichloro-1-hydroxyethyl)benzamide **22**, 45
N-(2,2,2-Trichloro-1-hydroxyethyl)thiobenzamide **22**, 45
1,1,2-Trichloro-1,2,2-trifluoroethane **124**, 195
Trichloroacetic acid **110**, 127
α-Trichloroacetoxystyrene **9**, 29
1,1,1-Trichloroethane **28**, 75
1,1,2-Trichloroethane **28**, 75
Trichloroethanol **44**, 31
Trichloroethylene **28**, 75
1,1,1-Trifluoro-2,4-pentanedione **108**, 95
Trichlorofluoromethane **110**, 9; **124**, 195
5-Trifluoromethyl-2-benzimidazolecarbamic acid methyl ester **77**, 265
Trichomonas vaginalis **60**, 253
Tricyclic antidepressant agent **94**, 135
Triethanolamine **15**, 185; **137**, 143; **145**, 175
Triethylene glycol **15**, 185
Trifluoperazine **3**, 231; **16**, 271; **33**, 89
N-Trifluoroacetyldoxorubicin 14 valerate **56**, 103; **68**, 271
Trifluorothymidine **28**, 201
Triflupromazine **3**, 231; **5**, 273; **16**, 153, 271; **35**, 129; **36**, 185
Trifluridine **28**, 201
Triglycine **71**, 117
11β,17α,21-Trihydroxypregn-4-en-20-one-3-spiro[2′–4] **10**, 307
11β,17α,21-Trihydroxypregn-4-en-20-one-3-spiro[2′-thiazolidine]-21 **10**, 307
Trilaurin **2**, 317; **13**, 243; **99**, 337; **115**, 129; **146**, 123
Trimethoprim **46**, 89; **59**, 255; **63**, 173; **72**, 141; **75**, 81, 161; **85**, 209; **90**, 51; **92**, 237
2,2,4-Trimethylpentane **15**, 347; **17**, 77; **22**, 177; **31**, 209
Trimethylpsoralen **133**, 245
Trimethylsilyl derivative **28**, 1
17β-Trimethylsilyloxytestosterone **28**, 1
Trimipramine **35**, 129
Trimyristin **13**, 243
Trinitrobenzenesulfonic acid **96**, 85; **103**, 253
Trioctanoin **4**, 219; **78**, 217
Triolein **37**, 127; **78**, 217; **4**, 271, 281
Tripalmitin **13**, 243; **129**, 159
Tripelennamine **116**, 77
Tripelennamine hydrochloride **12**, 77
Tripeptide **71**, 45; **115**, 79
Triphenyl phosphate **14**, 263
Tripolyphosphate **131**, 101
Triptorelin **49**, 69; **59**, 173; **62**, 119; **108**, 49; **115**, 235; **127**, 31, 43
N,n,n′-Tris(2-hydroxyethyl)-*n*-octadecyl-1,3-diaminopropane dihydrofluoride **131**, 33
3,4,β Tris(trimethylsilyl)noradrenalin H 3 **28**, 1

Tri-tab **110**, 37
Tritium oxide **28**, 219
Triton X-100 **45**, 91; **103**, 19; **113**, 263; **117**, 159; **123**, 33; **125**, 283; **126**, 281; **128**, 29
Triton X-114 **123**, 33
Trometamol **19**, 177; **82**, 135; **89**, 213; **104**, 41
Tropicamide **20**, 187; **25**, 73; **43**, 67; **51**, 203
Trypan blue **49**, 205
Trypsin **12**, 295; **78**, 1; **81**, 119; **88**, 293; **95**, 171; **97**, 171; **113**, 1; **117**, 95; **138**, 13
Trypsin inhibitor **78**, 1, 9; **81**, 133
Tryptophan **72**, 149
Tumor **22**, 137; **43**, 267; **61**, R1
Tumor cell **17**, 65; **47**, 129; **78**, 103
Tumor marker **17**, 65
Tumor necrosis factor **111**, 15
Turbidity **117**, 159
Turbulent flow **9**, 337
Tween **69**, 143; **99**, 29; **104**, 145; **105**, 77
Tylosin **85**, 87
Tyloxapol **11**, 277; **39**, 263; **102**, 91; **103**, 225
Tyrosine **72**, 149; **76**, 99; **87**, 59; **104**, 157
Tyrosine peptide **12**, 271
Tyrphostin **104**, 29
Tyrphostin derivative **104**, 29

U

U-83-836E **103**, 103
U-88-943E **103**, 103
Ubidecarenone **86**, 25; **93**, 189
Ubiquinone **122**, 57
Ubiquinone 10 **26**, 289
Ulcerative colitis **48**, 79; **68**, 199
Ulcerogenesis **120**, 145
Ulex europaeus agglutinin **142**, 227
Ultrafiltration **32**, 171; **94**, 115
Ultrasonics **13**, 47; **20**, 235
Ultrasound **40**, 105; **44**, 65; **65**, 127; **96**, 249; **118**, 129; **133**, 39; **137**, 217
Ultraviolet irradiation **145**, 87
Ultraviolet radiation **1**, 173; **3**, 205; **12**, 265; **19**, 207; **31**, 1; **50**, 219; **56**, 213; **70**, 9; **113**, 113
Ultraviolet spectrophotometry **13**, 1, 333; **15**, 61; **18**, 235, 325; **20**, 273; **21**, 251, 349; **22**, 45; **23**, 209, 223; **24**, 43, 219, 223; **25**, 27; **26**, 123, 203; **27**, 185; **28**, 11, 23, 47, 95; **29**, 1; **31**, 25, 43; **32**, 123; **33**, 225, 235; **35**, 7, 47, 61, 181, 243; **36**, 73, 147; **37**, 185; **49**, 79; **53**, 35; **65**, 177; **86**, 99, 153; **93**, 245; **102**, 271
Ultraviolet spectroscopy **35**, 13; **63**, 173; **127**, 185; **134**, 213
Umbelliferone **12**, 355
Undecanoic acid **20**, 3; **148**, 41
Unimac **66**, 143; **91**, 123, 143
Unimac MG 100 **80**, 179; **83**, 147; **131**, 201
Unimac MG 200 **131**, 201
Unindexed drug **92**, 1; **94**, 81; **105**, 189; **123**, 1; **124**, 149; **132**, 23; **136**, 117; **148**, 1
Unsaturated fatty acid **98**, 203; **126**, 267
Uracil **103**, 93; **116**, 217
Urea **7**, 189; **8**, 211; **11**, 131; **14**, 1; **16**, 127; **22**, 357; **24**, 61; **32**, 79; **35**, 1, 7; **36**, 61, 67; **44**, 71; **48**, 207; **56**, 43; **61**, 205; **65**, 29; **66**, 157; **67**, 97; **72**, 97; **82**, 229; **89**, 111; **99**, 109; **112**, 117; **113**, 247; **124**, 173; **126**, 267; **137**, 125; **145**, 9; **146**, 255; **148**, 41
Urea derivative **45**, 47
Urease **22**, 137
Urethan **38**, 261
Uric acid **86**, 17
Urinary excretion **8**, 175; **40**, 125
Urine **1**, 3, 323; **2**, 173; **47**, 89; **49**, 173
Urine level **45**, 101; **50**, 175; **57**, 177
Urine pH **36**, 233
Urokinase **134**, 193
Urothelium **59**, 205
Ursodeoxycholic acid **74**, 127; **109**, 173; **115**, 249; **118**, 77; **130**, 41; **142**, 103; **149**, 1
Uterine cervix dysplasia **115**, 235
Uterine cervix mucosa **84**, 69
Uterus **111**, 197
Uvinul D 50 **82**, 223
Uvinul N 35 **82**, 223
Uxepam **61**, 189

V

Vaccine **75**, 1, 97, 117; **102**, 261; **103**, 37; **116**, 1; **128**, 99
Vaccinia virus **49**, 205
Vagina **2**, 215; **12**, 107; **24**, 209
Vagina cytology **77**, 75
Vagina epithelium **77**, 75
Vagina flora **43**, 77
Vagina mucosa **106**, 237; **125**, 81
Vagina pessary **22**, 131
Valeric acid **86**, 43; **126**, 219
Valine **72**, 149; **85**, 7
Valproate calcium **115**, 245
Valproic acid **16**, 285; **20**, 53; **23**, 25; **24**, 297; **27**, 45; **36**, 1; **54**, 1; **81**, 67; **115**, 245
Valpromide **23**, 25
Vamin N **27**, 17
Vanadium derivative **146**, 115
Vancomycin **89**, 261; **109**, 261; **117**, 119; **139**, 243; **144**, 71
Vantocil 1B **117**, 123
Vapor **42**, 77; **130**, 195; **81**, 153
Vapreotide **127**, 31; **127**, 43
Vascular endothelium **45**, 249
Vaselin **27**, 29; **28**, 47
Vasoconstriction **18**, 299; **26**, 145; **27**, 29; **86**, 201
Vasodilator agent **110**, 1; **123**, 181
Vasopressin **48**, 15; **49**, 189, 199; **81**, 119; **86**, 35; **106**, 255; **116**, 211; **127**, 115
Vasopressin derivative **114**, 151
Vehicle **18**, 269
Velocity **13**, 47
Verapamil **47**, 209; **53**, 145; **54**, 1; **59**, 87; **68**, 43; **79**, 199; **86**, 167; **87**, 223; **89**, 111; **91**, 167; **92**, 1, 23; **102**, 267; **130**, 169; **131**, 153; **136**, 43

Veratryl curcuminoid **131**, 1
Vibration **32**, 177
Vicilin **142**, 227
Vidarabine **24**, 209; **59**, 45; **95**, 153; **123**, 159
Vidarabine phosphate **123**, 159
Vigabatrin **61**, 265; **93**, 7
Vinblastine **47**, 239; **77**, 279; **103**, 87; **108**, 69; **124**, 205
Vinblastine sulfate **43**, 131
Vincristine **50**, 189; **68**, 151; **77**, 279; **103**, 87; **108**, 69; **111**, 25; **124**, 205; **145**, 9
Vindesine **50**, 189; **77**, 279
Vinyl acetate **17**, 39
Vinyl acetate vinyl chloride copolymer **30**, 209; **60**, 197
Vinyl chloride copolymer **17**, 39
1-Vinyl-2-pyrrolidinone **138**, 57
Vinylbenzaldehyde **81**, 103
Virus infection **65**, 183; **115**, 235
Virus transmission **58**, 25
Visceral leishmaniasis **57**, 23; **83**, 251
Viscoelasticity **55**, 247; **57**, 95; **58**, 135; **67**, 217, 231; **92**, 123, 137; **95**, 57; **107**, 51; **130**, 147; **131**, 201
Viscometer **83**, 147; **133**, 133
Viscometry **76**, 239
Viscosity **10**, 133; **11**, 309; **12**, 173; **13**, 67, 75, 243; **24**, 231; **27**, 219; **29**, 243; **40**, 93; **44**, 141; **47**, 179; **53**, 219; **55**, 77; **58**, 73, 203; **61**, 15, 35, 213; **66**, 23; **70**, 147; **72**, 65; **73**, 117, 163; **76**, 91, 239; **77**, 261; **80**, 179; **83**, 147, 221; **84**, R5; **85**, 19; **90**, 195; **95**, 23; **107**, 85; **109**, 115; **111**, 205, 261; **126**, 189; **130**, 245; **146**, 233
Viskaldix **99**, 327
Vitacel **91**, 183; **97**, 15
Vitamin **5**, 167; **81**, 1
Vitamin mixture **13**, 9
Vitlipid **13**, 9
Vivacel pH 101 **101**, 169
Vivacel pH 102 **101**, 169
Volatile agent **77**, 47

W

Walker carcinoma **41**, 213; **113**, 141
Warfarin **2**, 283; **6**, 205; **10**, 339; **12**, 231, 355; **17**, 183; **19**, 307; **20**, 335; **54**, 1; **56**, 249; **59**, 137; **65**, 149; **71**, 55; **81**, 1; **92**, 1; **110**, 197; **118**, 85; **131**, 131
Water **1**, 71; **10**, 109; **11**, 277; **21**, 155; **22**, 131, 177; **25**, 57, 85; **26**, 25, 243; **27**, 1, 29, 335; **28**, 113, 193; **29**, 103, 177; **30**, 161; **31**, 9, 33, 157; **32**, 187; **33**, 235, 249; **43**, 31, 45; **44**, 71, 141; **48**, 173; **50**, 147; **55**, 17; **58**, 155; **62**, 153; **63**, 251; **76**, 91; **79**, 47, 113; **81**, 21; **82**, 223; **83**, 11; **90**, R1; **92**, 47, 81; **95**, 57; **98**, 173; **99**, 109; **103**, 225; **112**, 117; **114**, 55, 75; **120**, 189, 197; **121**, 27; **122**, 49; **135**, 31, 73, 151; **136**, 71; **139**, 33; **144**, 127; **145**, 9
Water absorption **55**, 115; **77**, 269; **120**, 33; **135**, 31
Water analysis **82**, 185, 195
Water content **69**, 233; **82**, 117; **134**, 79
Water flow **136**, 141
Water loss **72**, 227
Water oil cream **91**, 59; **92**, 191; **107**, 51; **121**, 169; **123**, 199; **124**, 183; **125**, 73, 315; **126**, 129; **132**, 259; **140**, 97; **141**, 27; **148**, 47
Water permeability **73**, 197, 209; **79**, 301; **99**, 181
Water quality **58**, 155
Water transport **50**, 61; **51**, 63; **73**, 111, 239; **79**, 301; **112**, 47, 153; **122**, 49
Water vapor **17**, 237; **31**, 247; **101**, 35, 45; **126**, 83; **135**, 31; **137**, 1
Wax **38**, 91; **123**, 85; **133**, 155; **138**, 215
Wettability **125**, 327; **126**, 21; **135**, 31; **138**, 199
Wetting agent **119**, 149
Wheat germ agglutinin **22**, 137
Win 63843 **117**, 231
Witepsol **23**, 87; **148**, 155
Witepsol E 75 **89**, 111
Witepsol H 12 **40**, 181, 187
Witepsol H 15 **33**, 249; **38**, 103, 191; **89**, 111; **148**, 155
Witepsol H 19 **40**, 181
Witepsol H 35 **3**, 179; **22**, 131
Witepsol H 37 **22**, 131
Witepsol H 5 **3**, 187; **47**, 157
Witepsol N 15 **3**, 39, 55; **9**, 107
Witepsol S 55 **22**, 131; **47**, 157; **148**, 155
Witepsol W 35 **47**, 157; **148**, 155
Wound dressing **91**, 51; **112**, 29
Wound infection **75**, 81

X

Xanthan **86**, 159; **120**, 63; **129**, 233; **133**, 1; **134**, 173; **139**, 53, 79
Xanthan CX 90 **112**, 125
Xanthan gum **9**, 315; **11**, 309; **78**, 43; **86**, 159; **95**, 229; **112**, 125; **118**, 207; **120**, 63; **128**, 73, 113; **133**, 1; **134**, 173
Xanthan SM **112**, 125
Xanthine **18**, 259, 277
Xanthine derivative **58**, 129
Xanthine oxidase **86**, 17
XD 405 **94**, 11
Xenobiotic agent **57**, 189; **64**, 127; **69**, 221; **77**, 1; **79**, 135; **80**, 51; **101**, 209; **109**, 197
Xenobiotic metabolism **109**, 197
Xerostomia **12**, 277
Xilobam **33**, 201
X-ray **46**, 179
X-ray analysis **103**, 125
X-ray diffraction **45**, 139; **74**, 77; **77**, 287; **81**, 169; **84**, 205; **139**, 9; **144**, 81, 147
X-ray fluorescence **131**, 243
X-ray spectrometry **57**, 183; **72**, 157
Xylazine **14**, 123
Xylene **16**, 127; **76**, 91; **99**, 93
Xylenol **10**, 89
2,4-Xylenol **10**, 89
2,6-Xylenol **10**, 89
Xylitol **64**, 223; **66**, 71; **67**, 139; **90**, 105
Xylometazoline **12**, 57; **86**, 59; **123**, 47; **135**, 137

O,o(1,4-Xylylene)-bispilocarpic acid ester derivative **75**, 249; **75**, 259

Y

Yellow 6 lake **50**, 163
Ylang ylang oil **111**, 235
Yoshida sarcoma **41**, 21; **121**, 195

Z

Zankiren **120**, 127
Zanoterone **122**, 17
Zearalenone **128**, 129
Zein **141**, 137
Zeta potential **58**, 229; **63**, 183
Zidovudine **50**, 175; **57**, 73; **77**, 71; **95**, 1; **96**, 249; **123**, 1; **125**, 17, 31; **129**, 243; **137**, 187; **144**, 61; **146**, 123
Zidovudine derivative **144**, 61
Zidovudine 5′(1,4-dihydro-1-methylnicotinate) **125**, 17, 31
Zidovudine palmitate **146**, 123
Zileuton **140**, 119; **143**, 179
Zinc **22**, 283; **60**, 27; **105**, 209; **108**, 173; **120**, 145
Zinc acetate **33**, 105
Zinc chloride **136**, 53
Zinc oxide **132**, 267
Zinc sulfate **104**, 247
Zirconium aluminum trichlorohydroxide **1**, 49
Zomepirac **19**, 177; **54**, 1
Zwitterion **8**, 25; **25**, 135